10 – ORIGINAL ARTICLE CLINICAL INVESTIGATION

DNA damage and antioxidant status in medical residents occupationally exposed to waste anesthetic gases¹

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ABSTRACT

PURPOSE: To investigate the effects of occupational exposure to waste anesthetic gases on genetic material and antioxidant status in professionals during their medical residency.

METHODS: The study group consisted of 15 medical residents from Anesthesiology and Surgery areas, of both genders, mainly exposed to isoflurane and to a lesser degree to sevoflurane and nitrous oxide; the control group consisted of 15 young adults not exposed to anesthetics. Blood samples were drawn from professionals during medical residency (eight, 16 and 22 months of exposure to waste anesthetic gases). DNA damage was evaluated by comet assay, and antioxidant defense was assessed by total thiols and the enzymes glutathione peroxidase (GPX), superoxide dismutase (SOD) and catalase (CAT).

RESULTS: When comparing the two groups, DNA damage was significantly increased at all time points evaluated in the exposed group; plasma thiols increased at 22 months of exposure and GPX was higher at 16 and 22 months of exposure.

CONCLUSION: Young professionals exposed to waste anesthetic gases in operating rooms without adequate scavenging system have increased DNA damage and changes in redox status during medical residency. There is a need to minimize exposure to inhalation anesthetics and to provide better work conditions.

Key words: Anesthetics. Occupational Exposure. Toxicity.

Introduction

Worldwide, healthcare professionals who work in environments contaminated by waste anesthetic gases (surgeons, anesthesiologists, nurses, technicians, dentists, and dental assistants) besides students are occupationally exposed to halogenated anesthetics and nitrous oxide (N₂O). Occupational exposure may result in adverse health effects such as fatigue, irritability and headache^{1,2}; abnormalities of the liver, kidneys and hematopoietic system^{3,4}; and neurobehavioral changes⁵. Furthermore, exposure to waste anesthetic gases are also related to an increased abortion/miscarriage incidence^{6,7}, reduced fertility⁸, and birth defects, which are especially related to N₂O⁹.

Although healthcare workers are exposed to much lower anesthetic concentrations than patients, such exposure can extend over many years. Studies have shown increased DNA damage, assessed by sister chromatid exchange (SCE), chromosomal aberration (CA) and micronuclei (MN) tests in operating room personnel exposed for several years to waste anesthetic gases ¹⁰⁻¹⁷. However, there is still no report about the effect of waste anesthetic gases in professionals occupationally exposed for a shorter time.

Free radicals and reactive oxygen species (ROS) are important molecules known to defend the organism against microorganisms, but their exacerbate release can lead to nucleic acid, lipid and protein damages¹⁸. Oxidative stress is defined as the imbalance between ROS formation and antioxidant defense. Additionally, ROS are formed during the metabolism of drugs, including anesthetics. Only few studies have linked exposure to waste anesthetic gases with oxidative stress. Nonetheless, increased plasma lipid peroxidation, but no changes in antioxidant capacity, has been observed in health professionals exposed to anesthetic gases¹⁹. Antioxidant enzymes and trace elements in operating room personnel exposed to a mixture of volatile anesthetics have been reported to be lower than in a non-exposed group²⁰. Moreover, anesthesiologists, nurses and surgeons exposed to halothane, isoflurane, sevoflurane, desflurane and N₂O for a mean of seven years have shown increased DNA damage and oxidative status²¹.

Long-term exposure to anesthetics causes toxic effects. However, the effects on DNA breaks and oxidative stress in medical residents exposed to inhaled anesthetics have not yet been investigated. Therefore, this study evaluated the DNA damage and antioxidant status in professionals exposed to waste anesthetic gases during their medical residency.

Methods

This study was approved by the Human Research Ethics Review Board, Federal University of Amazonas - UFAM (0044.0.115.000-10), in accord with the Helsinki Declaration of 1975 as revised in 2008. Written informed consent was obtained from all subjects, who were asked to respond to a standardized questionnaire on lifestyle, health status and previous exposure to environmental pollutants. Thirty adults, of both genders, aged 25-32 years, were enrolled in the study. The exposed group consisted of 15 medical residents in the fields of Anesthesiology, General surgery, Neurosurgery and Orthopedics mainly exposed to isoflurane and to a lesser degree to sevoflurane and N2O, from eight months up to 22 months of exposure. The control group comprised 15 volunteers not exposed to waste anesthetic gases or other pollutants, who did not work in a hospital area. The UFAM Hospital in Manaus (Northern, Brazil) has seven operating rooms with no active scavenging system.

Subjects with any disease, smokers, and alcoholics, those recently exposed to radiation, under medication or vitamin supplements/antioxidants, and those with any kind of occupational exposure other than waste anesthetic gases (exposed group) were excluded from the study.

Blood sampling

Venous blood samples were drawn eight, 16 and 22 months of exposure to waste anesthetic gases from the exposed residents, and once from the controls. All samples were coded, and subjected to blind analysis. Each step was carried out under indirect light to prevent additional damage. Whole blood was used for glutathione peroxidase (GPX) detection, erythrocytes for the assessment of superoxide dismutase (SOD) and catalase (CAT) activities, lymphocytes for comet assay, and plasma for total thiols measurement. Butylated hydroxytoluene (BHT) was used as an antioxidant preservative for biochemical analysis.

DNA damage

Lymphocytes were separated from whole blood and freshly used for the comet assay as previously described 22 , with some modifications. Lymphocytes (20 μL) were mixtured with 90 μL of low-melting point agarose, and embedded in agarose gel on microscope slides, and kept in lysing solution. The slides were left in a high-pH buffer for 20 min to allow DNA unwinding. Electrophoresis was conducted at 25 V and 300 mA for 20 min.

Following neutralization, the slides were fixed with ethanol and stained with 60 μ L SYBR® green and immediately analyzed under a fluorescent microscope (Leica DMI 6000, Switzerland) by visual scoring as previously described²³. One hundred nucleoids were randomly analyzed and assigned to one of five classes, from 0 (no tail) to 4 (almost all DNA in tail). The total score followed the calculation formula 1 x $n1 + 2 \times n2 + 3 \times n3 + 4 \times n4$, where n represents nucleoid number attributable to each damage class (score). Thus, the total score was between 0 and 400 arbitrary units.

Biochemical analyses

Total thiols interacted with 5, 5'-dithiobis-(2-nitrobenzoic acid) (DTNB), forming a yellow complex with absorbance measured at 412 nm in spectrophotometer²⁴.

Antioxidant defense was also evaluated through the most important enzymes. CAT activity was evaluated as previously described²⁵. SOD activity was determined by using a commercially available assay - Ransod kit (Randox Laboratories, UK), according to instructions, and GPX was evaluated using the Ransel kit according to the manufacturer's protocol (Randox Laboratories, UK). For biochemical analysis, the DTX 800 Multimode Detector (Beckman Couter, Germany) and UV/Visible Spectrophotometers model T70 (PG Instruments Limited, UK) were used.

Statistics

The characteristics of the study subjects and biochemical data are expressed as means and standard deviations (X \pm SD). Student's t-test was used to compare groups and ANOVA was applied in the analysis of exposure times (exposed group), followed by Tukey's test. The distribution of gender in each group was compared using the Chi-square test. For comet assay data (expressed as median, 25% and 75%), the non-parametric Mann-Whitney test was used to compare the groups, while the test of Friedman was applied to compare exposure times within the same group. Pearson's correlation was used to correlate DNA damage with thiols and GPX only at 22 months of exposure, since this time point presented the most significant alterations. p<0.05 was considered statistically significant.

Results

Demographic characteristics of subjects are shown in Table 1. There were no significant differences between the groups in age, gender, weight, height or body mass index (p>0.05). The

average length of exposure to waste anesthetic gases among medical residents was 30h/week.

TABLE 1 - Demographic data of the groups (X \pm SD or absolute number).

Characteristics	Groups		p value
	Control	Exposed (residents)	
Age (years)	26.8 ± 1.9	27.9 ± 2.3	0.57
Gender (female/male)	4 / 11	1 / 14	0.14
Weight (kg)	75.1 ± 18.8	77.8 ± 14.0	0.66
Height (m)	1.71 ± 0.1	1.74 ± 0.1	0.24
Body mass index (kg/m²)	25.6 ± 5.5	25.5 ± 3.8	0.94

Lymphocyte DNA damage assessed by comet assay in medical residents exposed to waste anesthetic gases was significantly increased at all time points evaluated, in comparison with controls. In the exposed group, DNA damage scores were significantly higher at 16 months than at eight months of exposure (Figure 1).

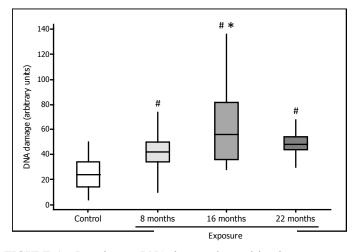


FIGURE 1 - Lymphocyte DNA damage detected by the comet assay (boxplot) in residents exposed to waste anesthetic gases and in unexposed volunteers (control). *# p=0.001 compared to the control group; * p<0.01 in relation to eight months of exposure.

Thiols content was significantly higher at 22 months of exposure in medical residents than in controls. In the exposed group, plasma thiol values were increased at 22 months of exposure compared with the other previous samplings (p<0.05) (Figure 2).

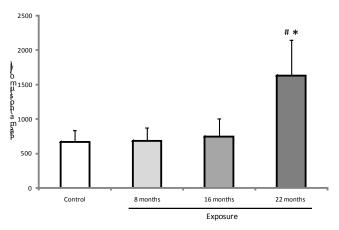


FIGURE 2 - Plasma thiols-SH ($X \pm SD$) in both groups. #p<0.001 versus control group. *p<0.001 compared to eight and 16 months.

GPX was higher at 16 and 22 months of exposure among residents than in the non-exposed group (p<0.01) (Figure 3). At 22 months of exposure, GPX activity was significantly enhanced in the exposed group compared with the other two samplings (p<0.001). CAT enzyme activity showed no difference between groups (p>0.05), but it was lower at 22 months of exposure when compared with other exposure times in the resident group (p=0.012) (Figure 4). SOD activity did not significantly differ between groups or among exposure times (p>0.05) (Figure 5).

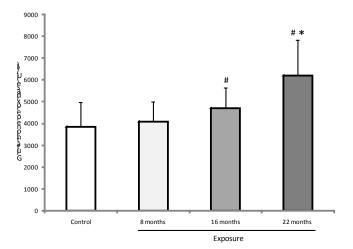


FIGURE 3 - Antioxidant GPX ($X \pm SD$) in both groups. # p<0.05 versus control group; * p<0.001 compared to eight and 16 months.

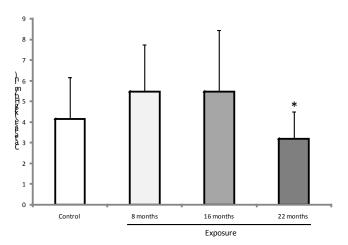


FIGURE 4 - CAT enzyme ($X \pm SD$) in both groups. * p=0.012 in relation to eight and 16 months.

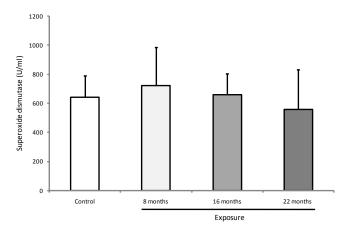


FIGURE 5 - SOD acticity ($X \pm SD$) in both groups. p>0.05.

The correlations of genotoxicity with antioxidant protection were determined at 22 months of exposure in the resident group. There was a significant negative correlation between DNA damage and thiols (p=0.03) and also between DNA damage and GPX (p=0.001) (Figures 6 and 7, respectively). There were no significant correlations between DNA damage and CAT (p=0.7) or SOD (p=0.2).

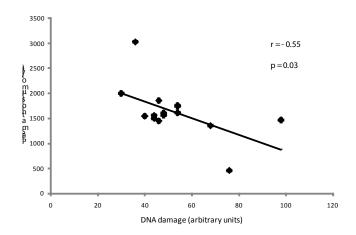


FIGURE 6 - Negative correlation between genetic damage and total thiols in the exposed group (residents) at 22 months of exposure (p<0.05).

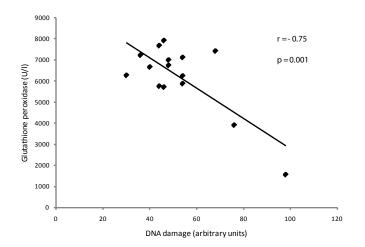


FIGURE 7 - Negative correlation between DNA damage and GPX activity in medical residents exposed to waste anesthetic gases at 22 months (p<0.05).

Discussion

Our study is the first to show that young professionals exposed to waste anesthetic gases already during medical residency have increased DNA damage and altered antioxidant protection.

A large number of studies conducted in professionals who worked for several years in operating rooms with no active scavenging system have related exposure to waste anesthetic gases with genetic modifications. Thus, increased DNA damage has been observed in professionals occupationally exposed for an average of 19 years to volatile anesthetics and N₂O¹⁷. Operating room personnel exposed for 12 years to halothane and N₂O have shown increased frequency of MN and CA26. Occupational exposure to halothane, enflurane, isoflurane, sevoflurane, desflurane and N₂O for around 11 years has been reported in professionals with increased DNA damage levels, MN formation and CA¹³. Anesthetits/nurses/ technicians exposed for 15 years to waste anesthetic gases have had more genetic damage including SCE, structural CA and MN than female workers not occupationally exposed or even female radiologists¹². Exposure to halothane has shown clastogenic effect in lymphocytes from anesthesiologists¹¹. Our results indicate that occupational exposure to anesthetic gases in operating rooms without an adequate scavenging system is genotoxic, even during a lower exposure time. We have observed that DNA lesions were increased from 8 months up to 22 months of exposure, with the highest levels at 16 months when subjects were exposed most of the time to halogenated and N₂O anesthetics. It is noteworthy that among the residents evaluated in our study the length of exposure time was considerably long.

To minimize potential health risks, public authorities have recommended threshold values to waste anesthetic gases. Thus, the exposure limits recommended by the National Institute of Occupational Safety and Health (NIOSH) are 2 ppm for volatile anesthetics and 25 ppm for $\rm N_2O$. The value of halogenated anesthetic is reduced to 0.5 ppm when used concomitantly with $\rm N_2O^{27}$. Low-level exposure to halogenated anesthetics (< 0.5 ppm) and $\rm N_2O$ (12 ppm) does not cause MN in operating room personnel, but high-level exposure to halogenated (4 ppm) and $\rm N_2O$ (170 ppm), for 20 h/week, causes genotoxic effects²⁸. However, among individuals occupationally exposed for 8 h/day to anesthetic gases, even if values were within the normal range of $\rm N_2O$ and halogenated, the frequency of SCE was increased²⁹.

The possible mechanisms underlying the genotoxic effects of halogenated anesthetics have been speculated. These drugs, including isoflurane, can directly damage DNA or be metabolized, giving rise to reactive metabolites³⁰. Another suggested mechanism is that anesthetic gases could act similarly to radiomimetic drugs, such as S-independent compound, inducing damage in all cell cycle phases¹¹. Besides that, N₂O promotes reduction of cyanocobalamin (vitamin B12) molecule of methionine synthase followed by formation of superoxide and hydroxyl radicals and inactivation of the enzyme that catalyzes the remethylation of homocysteine to methionine³¹.

Operating room personnel exposed to halothane and N₂O showed increased lipid peroxidation, decreased thiols content, with no antioxidant capacity changes, compared to non-exposed individuals¹⁹. In our study, medical residents were mainly exposed to isoflurane, but not to halothane, and only sometimes to sevoflurane and N₂O. The antioxidant enzymes, such as SOD and CAT activities remained unchanged in the medical residents compared with control group. Within the exposed group, CAT was consumed, especially at 22 months of exposure, showing increased oxidative stress. On the other hand, thiols started to increase at 16 months of exposure, with significant elevation at 22 months. Similarly, GPX activities enhanced at 16 and 22 months of exposure to anesthetics. Thus, there is a negative correlation between DNA damage and antioxidant status, especially after 22 months of exposure. This antioxidant mechanism is likely to be important as DNA damage occurred throughout exposure. Differently, decreased plasma and erythrocyte SOD and GPX, as well as trace elements, such as selenium, have been observed in anesthesia and surgery personnel working at least 6 hours daily for at least 3 years in operating theaters with no active scavenging systems²⁰. Cytochrome P450 2E1 is the most important enzyme for phase 1 metabolism of halogenated anesthetics. ROS, such as superoxide and hydrogen peroxide, are generated during this

stage^{32,33}. The binding of reactive intermediates to macromolecules may adversely affect cellular metabolism, protein synthesis, nucleic acids and lipids, producing a variety of injuries such as mutagenesis and carcinogenesis³⁴. Thus, our data indicate that both GPX and CAT activities, but not SOD, are directly related to hydrogen peroxide metabolism, and that exposure to waste anesthetic gases affects this antioxidant defense system. Our findings suggest that damage on DNA stimulated antioxidant defense, such as thiols and GPX to defend the organism against the aggression. Depletion of glutathione, which can promote injury leading to cell death, has been reported in professionals exposed to waste anesthetic gases for several years^{20,35}. In this study, however, the subjects evaluated were young healthy adults newly exposed to waste anesthetic gases, who were still capable of providing adequate antioxidant defense responses.

Technical anesthesiology staff presented a higher level of DNA breaks than controls. Even after a12-week supplementation with ascorbic acid (500 mg/day) and tocopherol (300 mg/day), the exposed group, although with a reduction of DNA damage, still showed more damage than the controls³⁶. Nurses have been reported to have increased oxidative DNA damage, lipoperoxidation and decreased GPX activity without alteration of alpha-tocopherol levels, when exposed from five to 27 years to high levels of N₂O³⁵. Based on that, the same authors suggest that increased oxidative stress may represent a mechanism link between chronic exposure to N₂O and genotoxicity.

Considering the outcome of this study, i.e. an increase of genetic material damage/oxidative stress caused by waste anesthetic gases, it is necessary an awareness of professionals to minimize the occupational exposure. In light of this, an active scavenging system is the most important asset for operating theaters. Additionally, better anesthesia equipment under constant maintenance, as well as the use of low fresh gas flow and total intravenous anesthesia can reduce the risks from occupational exposure.

Conclusion

The professionals occupationally exposed to waste anesthetic gases, such as isoflurane, sevoflurane and N_2O , in operating rooms without adequate scavenging system may have DNA damage and changes in antioxidant defense already during medical residency.

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