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**PROGRAMA DE PÓS-GRADUAÇÃO EM CIÊNCIAS BIOLÓGICAS
(MICROBIOLOGIA APLICADA)**

**SYNERGY BETWEEN CHEMICAL ADDITIVES AND PRETREATMENTS OF
SUGARCANE BIOMASS FAVORING ENZYMATIC HYDROLYSIS AND
PRODUCTION OF FUEL ETHANOL**

ALISON ANDREI SCHMATZ

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SUGARCANE BIOMASS FAVORING ENZYMATIC HYDROLYSIS AND PRODUCTION
OF FUEL ETHANOL**

ALISON ANDREI SCHMATZ

Tese apresentada ao Instituto de Biociências do Câmpus de Rio Claro, Universidade Estadual Paulista, como parte dos requisitos para obtenção do título de Doutor em Ciências Biológicas (Microbiologia Aplicada).

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TÍTULO DA TESE: ANTIOXIDANTS FAVOR LIGNIN REMOVAL FROM SUGARCANE BIOMASS AND INCREASE ENZYME HYDROLYSIS YIELDS AND ETHANOL VIA SIMULTANEOUS FERMENTATION AND SACCHARIFICATION

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Dedico esse trabalho à minha família.

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Você não sabe o quanto eu caminhei
Pra chegar até aqui
Percorri milhas e milhas antes de dormir
Eu nem cochilei
Os mais belos montes escalei
Nas noites escuras de frio chorei, ei, ei, ei
Ei, ei, ei, ei, ei, ei, ei

A vida ensina e o tempo traz o tom
Pra nascer uma canção
Com a fé do dia a dia encontro a solução
Encontro a solução

Quando bate a saudade eu vou pro mar
Fecho os meus olhos e sinto você chegar
Você chegar
Psicon! Psicon! Psicon! Psicon!

Quero acordar de manhã do teu lado
E aturar qualquer babado
Vou ficar apaixonado
No teu seio aconchegado
Ver você dormindo e sorrindo
É tudo que eu quero pra mim
Tudo que eu quero pra mim
Quero!

Você não sabe o quanto eu caminhei
Pra chegar até aqui
Percorri milhas e milhas antes de dormir
Eu nem cochilei
Os mais belos montes escalei
Nas noites escuras de frio chorei, ei, ei, ei
Ei, ei, ei, ei, ei, ei, ei

Together, together

Meu caminho só meu pai pode mudar
Meu caminho só meu pai
Meu caminho só meu pai

Together, together

A estrada – Cidade Negra

ABSTRACT

Sugarcane bagasse is a feedstock material in the production of second-generation ethanol (2G) and value-added products with biotechnological potential. However, biomass needs pretreatments to break down the cellulose-hemicellulose-lignin complex. During acid and hydrothermic pretreatments, can occur formation of sugar degradation products (toxic to fermenting microorganisms), which can condense with lignin fragments forming the pseudo-lignin, harmful to enzymatic hydrolysis. Therefore, this study aimed evaluate the hemicellulose and lignin removal from sugarcane biomass during diluted acid and organosolv pretreatment with addition of low-cost antioxidants, and their influences on the suppression the inhibitors formation and fermentable sugars production in simultaneous saccharification and fermentation (SSF). Different sugarcane fractions showed different recalcitrance. External fraction showed lower yields in the enzymatic hydrolysis, and 5 g L⁻¹ of acetic acid in the hydrolysate from acid pretreatment. The content of extractives and lignin were identified as key factor contributing to the formation of pseudo-lignin. Partial delignified and acid pretreated biomass showed yields of approximately 90% of cellulose conversion by enzymatic hydrolysis (up to 12 g L⁻¹). Butylated hydroxytoluene (BHT) demonstrated a potential effect on the removal of lignin from biomass in acid pretreatment (121 °C). With 50% (v/v) ethanol, BHT induced 10 g 100 g⁻¹ more lignin removal. Addition of BHT in acid pretreatment resulted in lower formation of inhibitors in liquid fraction (0.01 g L⁻¹ of furfural, 0.01 g L⁻¹ of HMF and 0.68 g L⁻¹ of acetic acid) and low residual hemicellulose and lignin content in the pretreated biomass. All the 6 chemical compounds (tert-butylhydroquinone; 3-tert-butyl-4-hydroxyanisole; methyl 3,4,5-trihydroxybenzoate; Tween 20; Tween 80 and dimethyl sulfoxide) individually improved lignin removal and enzyme hydrolysis yield. The 3-tert-butyl-4-hydroxyanisole showed 71% lignin removal, while Tween 80 used in the pretreatment resulted in a material with high digestibility, 98.9% glucose yield by enzymatic hydrolysis. After 48 h of SSF, 17.06 g L⁻¹ of glucose and 15.17 g L⁻¹ of ethanol were observed in delignified-acid pretreated biomass (DL-PT). Removal of hemicellulose and lignin resulted in higher yields conversion of sugars. Antioxidants and surfactants were identified as potential lignin removal effect, indicating that they can be applied in biotechnological process.

Keywords: Saccharification. Fermentation. Bioethanol. Pseudo-lignin. Biorefinery. Lignocellulosic biomass.

RESUMO

O bagaço de cana-de-açúcar é matéria-prima na produção de etanol de segunda geração (2G) e produtos de valor agregado com potencial biotecnológico. No entanto, a biomassa precisa de pré-tratamentos para quebrar o complexo celulose-hemicelulose-lignina. Durante os pré-tratamentos ácido e hidrotérmico, pode ocorrer a formação de produtos de degradação de açúcares (tóxico para microrganismos fermentadores), que podem se condensar com fragmentos de lignina formando a pseudo-lignina, prejudicial à hidrólise enzimática. Portanto, este estudo teve como objetivo avaliar a remoção de hemicelulose e lignina da biomassa da cana-de-açúcar durante o pré-tratamento ácido diluído e organosolv com adição de antioxidantes de baixo custo, e suas influências na supressão da formação de inibidores e produção de açúcares fermentáveis na fermentação e sacarificação simultânea (SSF). Diferentes frações da cana-de-açúcar apresentaram diferentes recalcitrâncias. A fração externa apresentou menores rendimentos na hidrólise enzimática e 5 g L⁻¹ de ácido acético no hidrolisado do pré-tratamento ácido. O conteúdo de extrativos e lignina foram identificados como fatores chave contribuindo para a formação de pseudo-lignina. A biomassa parcialmente deslignificada e pré-tratada com ácido apresentou rendimentos de aproximadamente 90% de conversão de celulose na hidrólise enzimática (até 12 g L⁻¹). O hidroxibutil tolueno (BHT) demonstrou potencial efeito na remoção de lignina da biomassa no pré-tratamento ácido (121 ° C). Com 50% etanol (v/v), o BHT induziu 10 g 100 g⁻¹ a mais de remoção de lignina. A adição de BHT no pré-tratamento ácido resultou em menor formação de inibidores na fração líquida (0,01 g L⁻¹ de furfural, 0,01 g L⁻¹ de HMF e 0,68 g L⁻¹ de ácido acético) e baixo teor residual de hemicelulose e lignina na biomassa pré-tratada. Todos os 6 compostos químicos (terc-butil hidroquinona; 3-terc-butil-4-hidroxianisol; 3,4,5-trihidroxibenzoato de metila; Tween 20; Tween 80 e dimetilsulfóxido) melhoraram individualmente a remoção de lignina e o rendimento da hidrólise enzimática. O 3-terc-butil-4-hidroxianisol apresentou 71% de remoção de lignina, enquanto o Tween 80 utilizado no pré-tratamento resultou em um material com alta digestibilidade, 98,9% de rendimento de glicose na hidrólise enzimática. Após 48 h de SSF, 17,06 g L⁻¹ de glicose e 15,17 g L⁻¹ de etanol foram observados na biomassa deslignificada e pré-tratada com ácido (DL-PT). A remoção de hemicelulose e lignina resultou em maiores rendimentos de conversão de açúcares. Antioxidantes e surfactantes foram identificados como potenciais removedores de lignina, indicando que podem ser aplicados em processos biotecnológicos.

Palavras-Chave: Sacarificação. Fermentação. Bioetanol. Pseudo-lignina. Biorrefinaria. Biomassa lignocelulósica.

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ABBREVIATIONS

2G	Second generation
ATR	Attenuated total reflectance
BHT	Butylated hydroxytoluene
BHT-PT	Butylated hydroxytoluene pretreated
CCoAOMT	CaffeoylCoA O-methyltransferase
COMT	Caffeic acid O-methyltransferase
DL	Partial delignified
DL-PT	Partial delignified acid pretreated
DMSO / DS	Dimethyl sulfoxide
EF	Extractive-free
EF-PT	Extractive-free acid pretreated
F5H	Ferulate 5-hydroxylase
FPU	Filter paper units
FTIR	Fourier Transform Infrared
HPLC	High Performance Liquid Chromatography
HMF	5- hydroxymethylfurfural
IN	 <i>In natura</i>
IN-PT	 <i>In natura</i> acid pretreated
IU	International Units
MR	Mass recovery
MT	Methyl 3,4,5-trihydroxybenzoate
PT	Diluted acid pretreatment
SEM	Scanning electron microscopy
SSF	Simultaneous Saccharification and Fermentation
T2	Tween 20
T8	Tween 80
TB	Tert-butylhydroquinone
TH	3-tert-butyl-4-hydroxyanisole
WT	Wild-type

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CHAPTER 1 INTRODUCTION AND MOTIVATION

The burning of fossil fuels (oil, natural gas and coal) increases the atmospheric concentrations of gases that cause global warming (SO_2 , CO_2 and NO_x). Oil, gas and coal may continue to exist for decades to come, but the shift in energy matrices to low-carbon fuels is critical to slow climate change. Renewable energy sources are fundamental alternatives in the energy transition, migrating from fossil fuels to sustainable alternatives. Biotechnological processes have made it possible to convert biomass (plant and animal waste) into biofuels, which have become popular around the world (ABAS; KALAIR; KHAN, 2015).

Lignocellulosic biomass is abundant in the world and consists of carbohydrates (cellulose and hemicellulose) and lignin that can be converted into second generation-ethanol and value-added products (2G ethanol) (VIEIRA et al., 2020). Brazil is the largest producer and consumer of sugarcane in the world. In 2018, the country was responsible for the production of 746 million tons, almost double the second place (India, with 376 million tons) (FAOSTAT, 2018). For each ton of cane processed, 140 kg of dry bagasse and 140 kg of dry straw are generated (CARVALHO et al., 2014). In the state of São Paulo, the largest national producer, it was stipulated the gradual elimination of sugarcane straw burning during the harvest period, until 2021 in mechanizable areas and until 2031 in non-mechanizable areas (State Decree n ° 47.700 /2003; State Law n° 11241/2002), reducing the suspension of soot in the air and reducing the emission of carbon dioxide into the atmosphere (GONÇALVES et al., 2017). For this reason, Brazil is faced with a new abundant lignocellulosic material, the straw, which has potential in the production of 2G ethanol.

Due complex and recalcitrant structure of lignocellulosic biomass, pretreatments are necessary to fractionate the cellulose-hemicellulose-lignin complex and solubilize the sugar fraction of the biomass, converting it into fermentable sugars (glucose), an energy currency for various metabolic routes of microorganisms with biotechnological application (VIEIRA et al., 2020). However, in acidic and hydrothermal pretreatments, particularly in conditions of high severity (temperature, reaction time and acid loading), the formation of sugar degradation products can occur that have a negative impact on hydrolysis and fermentation yields. The degradation of glucose can generate 5-hydroxymethylfurfural (HMF), which can undergo reactions resulting in formic and levulinic acid, while the degradation of xylose can generate furfural. Aldehydes, aliphatic

and aromatic acids can also form and these compounds have an inhibitory impact on enzymes and yeasts (RASMUSSEN; SØRENSEN; MEYER, 2014).

Repolymerization of polysaccharide degradation products (furfural and HMF) and/or polymerization with lignin monomers can form pseudo-lignin. However, the formation of pseudo-lignin can be generated from carbohydrates without the contribution of lignin in diluted acid pretreatments. This molecule formed of carbonyl, aromatics, methoxy and aliphatic structures is harmful in the conversion of sugars. Pseudo-lignin tends to irreversibly bind to enzymes through hydrophobic interactions, resulting in loss in their activities. Therefore, to avoid pseudo-lignin in reaction medium could contribute to the process success (HU; JUNG; RAGAUSKAS, 2012).

Hemicellulose and lignin also have a direct influence on yields of enzymatic hydrolysis, acting as a barrier protecting cellulose. The diluted acid pretreatment partially removes hemicelluloses and modifying the structure of lignin (SHIMIZU et al., 2020). Lignin is responsible for the plant's structural integrity. Polysaccharides are naturally "protected" in the plant cell walls by lignin, which acts as a chemical barrier against external threats. Organosolv pretreatments can remove lignin, and this complete removal from biomass results in extremely digestible material. Delignification yields can be improved with addition of chemicals in pretreatment. The removal or modification of hemicellulose and lignin is essential to increase cellulose accessibility of enzymes (SHIMIZU et al., 2020; ZENG et al., 2014).

The removal of lignin from biomass is a fundamental step for the development of biofuels. Separating the components of sugarcane bagasse under mild conditions in addition to increasing the accessibility of cellulose, also enables the recovery and extraction of lignin, which can be used in the synthesis of new resins to replace those based on chemical compounds derived from the petrochemical industry (lignin valorization) (VALDIVIA et al., 2016).

Therefore, this study aimed to evaluate the contribution of biomass heterogeneity (fractions of internode, node, external fraction and leaf) in pseudo-lignin formation and sugar degradation products. Furthermore, the effect of low-cost antioxidants and surfactants (butylated hydroxytoluene; tert-butylhydroquinone; 3-tert-butyl-4-hydroxyanisole; methyl 3,4,5-trihydroxybenzoate; Tween 20; Tween 80 and dimethyl sulfoxide) were evaluated in lignin removal during organosolv and diluted acid pretreatment and subsequent conversion of sugars by enzymatic hydrolysis and SSF.

Removal of hemicellulose and lignin can generate value-added products, improving pretreatments efficiency and collaborating with the biorefinery use of biomass.

8. CONCLUSION

This study evaluated the addition of low-cost antioxidant compounds, applied during the pretreatment, in the removal of hemicellulose and lignin from the sugarcane biomass, pseudo-lignin remotion, enzymatic hydrolysis (glucose yield), and ethanol from SSF. The heterogeneity of sugarcane fractions (bagasse, leaf, external fraction, node and internode) resulted in different responses to the diluted acid pretreatment. The formation of pseudo-lignin was influenced by the presence of extractives and lignin from biomass, and its removal increased glucose yields in the enzymatic hydrolysis. The diversity in glucose yields may reflect the removal of lignin and hemicellulose by effect of BHT. In the organosolv pretreatments (50% ethanol v/v at 121 °C for 30 min) BHT action improved the lignin removal and resulted in lower yields of degradation products/inhibitors. In the pretreatments at 160 °C (50% ethanol), the individual addition of antioxidants/surfactants resulted in a positive effect on the removal of lignin, corroborating with an increase in the enzymatic hydrolysis glucose yield. The addition of Tween 80 in the pretreatment generated benefits in the enzymatic hydrolysis conversion yields, besides help to remove compounds of interest. The partial delignification of biomass, followed by diluted acid pretreatment resulted in high yields in the enzymatic hydrolysis and also in the best ethanol yields in the SSF. Better practical (EF-PT) and global ethanol yields (DL-PT) were observed in the biomass without extractives, indicating that the contribution to the formation of pseudo-lignin can negatively affect alcoholic yield. However, a slower ethanol growth curve may due to the assimilation of sugars during yeast growth (number of cells). In general, the addition of antioxidant/surfactant chemical compounds in the pretreatment resulted in an improvement in the removal of lignin from biomass, reflecting positively on the sugar conversion yields of enzymatic hydrolysis. Its use can bring biotechnological benefits in research and industries.

REFERENCES

ABAS, N.; KALAIR, A.; KHAN, N. Review of fossil fuels and future energy technologies. **Futures**, v. 69, p. 31–49, 2015.

ABNT. **ABNT NBR 16550:2018 Sugarcane bagasse - chemical characterization. Brazilian National Standards Organization**, 2018.

AGNIHOTRI, S. et al. Ethanol organosolv pretreatment of softwood (*Picea abies*) and sugarcane bagasse for biofuel and biorefinery applications. **Wood Science and Technology**, v. 49, n. 5, p. 881–896, 2015.

ALVIRA, P. et al. Pretreatment technologies for an efficient bioethanol production process based on enzymatic hydrolysis: A review. **Bioresource Technology**, v. 101, n. 13, p. 4851–4861, 2010.

BATISTA, G. et al. Effect of severity factor on the hydrothermal pretreatment of sugarcane straw. **Bioresource Technology**, v. 275, n. December 2018, p. 321–327, 2019.

BENJAMIN, Y.; CHENG, H.; GÖRGENS, J. F. Evaluation of bagasse from different varieties of sugarcane by dilute acid pretreatment and enzymatic hydrolysis. **Industrial Crops & Products**, v. 51, p. 7–18, 2013.

BENJAMIN, Y.; GARCÍA-APARICIO, M. P.; GÖRGENS, J. F. Impact of cultivar selection and process optimization on ethanol yield from different varieties of sugarcane. **Biotechnology for Biofuels**, v. 7, n. 60, p. 1–17, 2014.

BENJAMIN, Y.; GÖRGENS, J. F. Improving Sugar Conversion and Bioethanol Yield through Transgenic Sugarcane Clone in South Africa. **Int. J. Environ. Bioener.**, v. 10(1), n. October, p. 9–25, 2015.

BENJAMIN, Y.; GÖRGENS, J. F.; JOSHI, S. V. Comparison of chemical composition and calculated ethanol yields of sugarcane varieties harvested for two growing seasons. **Industrial Crops and Products**, v. 58, p. 133–141, 2014.

BEWG, W. P. et al. RNAi downregulation of three key lignin genes in sugarcane improves glucose release without reduction in sugar production. **Biotechnology for Biofuels**, v. 9, n. 270, p. 1–13, 2016.

BEZERRA, T. L.; RAGAUSKAS, A. J. Review A review of sugarcane bagasse for second-generation bioethanol and biopower production. **Biofuels, Bioprod. Bioref.**, v. 10, p. 634–647, 2016.

BOTTCHER, A. et al. Lignification in sugarcane: Biochemical characterization, gene discovery, and expression analysis in two genotypes contrasting for lignin content. **Plant Physiology**, v. 163, n. 4, p. 1539–1557, 2013.

BRIENZO, M. et al. Comparison Study on the Biomass Recalcitrance of Different Tissue Fractions of Sugarcane Culm. **Bioenerg. Res.**, v. 7, p. 1454–1465, 2014.

BRIENZO, M. et al. Relationship between physicochemical properties and enzymatic hydrolysis of sugarcane bagasse varieties for bioethanol production. **New Biotechnology**, v. 00, n. 00, 2015.

BRIENZO, M. et al. Characterization of anatomy, lignin distribution, and response to pretreatments of sugarcane culm node and internode. **Industrial Crops and Products**, v. 84, p. 305–313, 2016.

BRIENZO, M. et al. Influence of pretreatment severity on structural changes, lignin content and enzymatic hydrolysis of sugarcane bagasse samples. **Renewable Energy**, v. 104, p. 271–280, 2017.

BURTON, R. A.; FINCHER, G. B. Plant cell wall engineering: applications in biofuel production and improved human health. **Current Opinion in Biotechnology**, v. 26, p. 79–84, 2014.

CANDIDO, J. P. et al. Detoxification of sugarcane bagasse hydrolysate with different adsorbents to improve the fermentative process. **World Journal of Microbiology and Biotechnology**, v. 36, n. 3, p. 1–12, 2020.

CARVAJAL, J. C.; GÓMEZ, Á.; CARDONA, C. A. Comparison of lignin extraction processes: Economic and environmental assessment. **Bioresource Technology**, v. 214, p. 468–476, 2016.

CARVALHO, D. M. DE et al. Ethanol-soda pulping of sugarcane bagasse and straw. **Cellulose Chemistry and Technology**, v. 48, n. 3–4, p. 355–364, 2014.

CARVALHO, D. M. DE et al. Assessment of chemical transformations in eucalyptus, sugarcane bagasse and straw during hydrothermal, dilute acid, and alkaline pretreatments. **Industrial Crops and Products**, v. 73, p. 118–126, 2015.

CARVALHO, D. M. DE; COLODETTE, J. L. Comparative study of acid hydrolysis of lignin and polysaccharides in biomasses. **BioResources**, v. 12, n. 4, p. 6907–6923, 2017.

CESARINO, I. et al. An overview of lignin metabolism and its effect on biomass recalcitrance. **Brazilian Journal of Botany**, v. 35, n. 4, p. 303–311, 2012.

CHANDEL, A. K. et al. Sugarcane bagasse and leaves: Foreseeable biomass of biofuel and bio-products. **J Chem Technol Biotechnol**, v. 87, n. 1, p. 11–20, 2012.

CHANG, V. S.; HOLTZAPPLE, M. T. Fundamental factors affecting biomass enzymatic reactivity. **Applied Biochemistry and Biotechnology**, v. 84, n. 86, p. 5–37, 2000.

COSTA, T. H. F. et al. The enzymatic recalcitrance of internodes of sugar cane hybrids with contrasting lignin contents. **Industrial Crops and Products**, v. 51, p. 202–211, 2013.

DEL RÍO, J. C. et al. Differences in the chemical structure of the lignins from sugarcane bagasse and straw. **Biomass and Bioenergy**, v. 81, p. 322–338, 2015.

ESPIRITO SANTO, M. et al. Structural and compositional changes in sugarcane bagasse

subjected to hydrothermal and organosolv pretreatments and their impacts on enzymatic hydrolysis. **Industrial Crops and Products**, v. 113, n. October 2017, p. 64–74, 2018.

FAOSTAT. **Food and Agriculture Organization of the United Nations - FAO**. Disponível em: <http://www.fao.org/faostat/en/#rankings/countries_by_commodity>. Acesso em: 21 out. 2020.

FAUSTINO, H. et al. Antioxidant activity of lignin phenolic compounds extracted from kraft and sulphite black liquors. **Molecules**, v. 15, n. 12, p. 9308–9322, 2010.

FENGEL, D.; WEGNER, G. **Wood: Chemistry, Ultrastructure, Reactions**. Berlin; New York: [s.n.].

GONÇALVES, C. et al. Size-segregated aerosol chemical composition from an agro-industrial region of São Paulo state, Brazil. **Air Qual Atmos Health**, v. 10, n. 4, p. 483–496, 2017.

GOVENDER, M. et al. An accurate and non-labor intensive method for the determination of syringyl to guaiacyl ratio in lignin. **Bioresource Technology**, v. 100, n. 23, p. 5834–5839, 2009.

GUILHERME, A. A. et al. Evaluation of composition, characterization and enzymatic hydrolysis of pretreated sugar cane bagasse. **Brazilian Journal of Chemical Engineering**, v. 32, n. 1, p. 23–33, 2015.

GUILHERME, A. DE A. et al. Ethanol production from sugarcane bagasse: Use of different fermentation strategies to enhance an environmental-friendly process. **Journal of Environmental Management**, v. 234, p. 44–51, 2019.

HIMMEL, M. E. et al. Biomass recalcitrance: Engineering plants and enzymes for biofuels production. **Science**, v. 315, n. 5813, p. 804–807, 2007.

HOANG, N. V. et al. Potential for Genetic Improvement of Sugarcane as a Source of Biomass for Biofuels. **Frontiers in Bioengineering and Biotechnology**, v. 3, n. November, p. 1–15, 2015.

HONGDAN, Z.; SHAOHUA, X.; SHUBIN, W. Enhancement of enzymatic saccharification of sugarcane bagasse by liquid hot water pretreatment. **Bioresource Technology**, v. 143, p. 391–396, 2013.

HU, F.; JUNG, S.; RAGAUSKAS, A. Pseudo-lignin formation and its impact on enzymatic hydrolysis. **Bioresource Technology**, v. 117, p. 7–12, 2012.

HU, F.; JUNG, S.; RAGAUSKAS, A. Impact of pseudolignin versus dilute acid-pretreated lignin on enzymatic hydrolysis of cellulose. **ACS Sustainable Chemistry and Engineering**, v. 1, n. 1, p. 62–65, 2013.

HU, F.; RAGAUSKAS, A. Suppression of pseudo-lignin formation under dilute acid pretreatment conditions. **RSC Adv.**, v. 4, n. 9, p. 4317–4323, 2014.

JAIN, S.; SHARMA, M. P. Stability of biodiesel and its blends : A review. **Renewable and Sustainable Energy Reviews**, v. 14, p. 667–678, 2010.

JIAMBOONSRI, P. et al. A validated liquid chromatography – tandem mass spectrometry method for the determination of methyl gallate and pentagalloyl glucopyranose : Application to pharmacokinetic studies. **Journal of Chromatography B**, v. 986–987, p. 12–17, 2015.

JÖNSSON, L. J.; MARTÍN, C. Pretreatment of lignocellulose: Formation of inhibitory by-products and strategies for minimizing their effects. **Bioresource Technology**, v. 199, p. 103–112, 2016.

JUNG, J. H. et al. RNAi suppression of lignin biosynthesis in sugarcane reduces recalcitrance for biofuel production from lignocellulosic biomass. **Plant Biotechnology Journal**, v. 10, n. 9, p. 1067–1076, 2012.

JUNG, J. H. et al. RNA interference suppression of lignin biosynthesis increases fermentable sugar yields for biofuel production from field-grown sugarcane. **Plant Biotechnology Journal**, v. 11, p. 709–716, 2013.

JUNG, J. H. et al. Precision breeding for RNAi suppression of a major 4-coumarate:coenzyme A ligase gene improves cell wall saccharification from field grown sugarcane. **Plant Molecular Biology**, v. 92, p. 505–517, 2016.

JUNIOR, C. S. et al. The effects of lignin removal and drying on the porosity and enzymatic hydrolysis of sugarcane bagasse. **Cellulose**, v. 20, p. 3165–3177, 2013.

KAUR, R.; UPPAL, S. K. Structural characterization and antioxidant activity of lignin from sugarcane bagasse. **Colloid Polym Sci**, v. 293, p. 2585–2592, 2015.

KAUR, R.; UPPAL, S. K.; SHARMA, P. Antioxidant and Antibacterial Activities of Sugarcane Bagasse Lignin and Chemically Modified Lignins. **Sugar Tech**, v. 19, n. 6, p. 675–680, 2017.

KUMAR, R. et al. Carbohydrate derived-pseudo-lignin can retard cellulose biological conversion. **Biotechnology and Bioengineering**, v. 110, n. 3, p. 737–753, 2012.

LAURITO-FRIEND, D. F. et al. Sugarcane hybrids with original low lignin contents and high field productivity are useful to reach high glucose yields from bagasse. **Biomass and Bioenergy**, v. 75, p. 65–74, 2015.

LI, M.; PU, Y.; RAGAUSKAS, A. J. Current Understanding of the Correlation of Lignin Structure with Biomass Recalcitrance. **Frontiers in Chemistry**, v. 4, n. November, p. 1–8, 2016.

LIU, Q.; LUO, L.; ZHENG, L. Lignins: Biosynthesis and biological functions in plants. **International Journal of Molecular Sciences**, v. 19, n. 335, p. 1–16, 2018.

LORENCINI, P. et al. Biohydrogen Production from Liquid and Solid Fractions of Sugarcane Bagasse After Optimized Pretreatment with Hydrochloric Acid. **Waste and**

Biomass Valorization, v. 7, n. 5, p. 1017–1029, 2016.

MA, X. et al. Toward a further understanding of hydrothermally pretreated holocellulose and isolated pseudo lignin. **Cellulose**, v. 22, n. 3, p. 1687–1696, 2015.

MARTINS, L. H. S.; RABELO, S. C.; DA COSTA, A. C. Effects of the pretreatment method on high solids enzymatic hydrolysis and ethanol fermentation of the cellulosic fraction of sugarcane bagasse. **Bioresource Technology**, v. 191, p. 312–321, 2015.

MASARIN, F. et al. Chemical composition and enzymatic digestibility of sugarcane clones selected for varied lignin content. **Biotechnology for Biofuels**, v. 4, n. 55, p. 1–10, 2011.

MASARIN, F. et al. Evaluation of a simple alkaline pretreatment for screening of sugarcane hybrids according to their in vitro digestibility. **Industrial Crops and Products**, v. 51, p. 390–395, 2013.

MELATI, R. B. et al. Sugarcane Bagasse: Production, Composition, Properties, and Feedstock Potential. In: RACHEL MURPHY (Ed.). **Sugarcane: Production Systems, Uses and Economic Importance**. New York: Nova Science Publishers, 2017. p. 1–38.

MELATI, R. B. et al. Key Factors Affecting the Recalcitrance and Conversion Process of Biomass. **Bioenergy Research**, v. 12, n. 1, p. 1–20, 2019.

MENG, X.; RAGAUSKAS, A. J. Pseudo-Lignin Formation during Dilute acid Pretreatment for Cellulosic Ethanol. **Recent Advances in Petrochemical Science**, v. 1, n. 1, p. 1–6, 2017.

MESA, L. et al. An approach to optimization of enzymatic hydrolysis from sugarcane bagasse based on organosolv pretreatment. **J Chem Technol Biotechnol**, v. 85, n. 8, p. 1092–1098, 2010.

MESA, L. et al. Restructuring the processes for furfural and xylose production from sugarcane bagasse in a biorefinery concept for ethanol production. **Chemical Engineering & Processing**, v. 85, p. 196–202, 2014.

MESA, L. et al. Techno-economic evaluation of strategies based on two steps organosolv pretreatment and enzymatic hydrolysis of sugarcane bagasse for ethanol production. **Renewable Energy**, v. 86, p. 270–279, 2016.

MILÉO, P. C. et al. Thermal and chemical characterization of sugarcane bagasse cellulose/lignin-reinforced composites. **Polym. Bull.**, v. 73, p. 3163–3174, 2016.

MOSIER, N. et al. Features of promising technologies for pretreatment of lignocellulosic biomass. **Bioresource Technology**, v. 96, n. 6, p. 673–686, 2005.

MUSHRIF, S. H.; CARATZOULAS, S.; VLACHOS, D. G. Understanding solvent effects in the selective conversion of fructose to 5-hydroxymethyl-furfural: A molecular dynamics investigation. **Physical Chemistry Chemical Physics**, v. 14, n. 8, p. 2637–2644, 2012.

NEVES, P. V.; PITARELO, A. P.; RAMOS, L. P. Production of cellulosic ethanol from sugarcane bagasse by steam explosion: effect of extractives content, acid catalysis and different fermentation technologies. **Bioresource Technology**, v. 208, p. 184–194, 2016.

PEDERSEN, J. F.; VOGEL, K. P.; FUNNELL, D. L. Impact of reduced lignin on plant fitness. **Crop Science**, v. 45, p. 812–819, 2005.

PHITSUWAN, P.; SAKKA, K.; RATANAKHANOKCHAI, K. Improvement of lignocellulosic biomass in planta: A review of feedstocks, biomass recalcitrance, and strategic manipulation of ideal plants designed for ethanol production and processability. **Biomass and Bioenergy**, v. 58, p. 390–405, 2013.

PINGALI, S. V. et al. Breakdown of cell wall nanostructure in dilute acid pretreated biomass. **Biomacromolecules**, v. 11, n. 9, p. 2329–2335, 2010.

PRASAD, S. S. et al. Tween 20-/H₂O Promoted Green Synthesis, Computational and Antibacterial Activity of Amino Acid Substituted Methylene Bisphosphonates. **Phosphorus, Sulfur, and Silicon**, v. 190, p. 2040–2050, 2015.

PU, Y. et al. Assessing the molecular structure basis for biomass recalcitrance during dilute acid and hydrothermal pretreatments. **Biotechnology for Biofuels**, v. 6, n. 15, p. 2–13, 2013.

RAI, P. K. et al. Biohydrogen production from sugarcane bagasse by integrating dark- and photo-fermentation. **Bioresource Technology**, v. 152, p. 140–146, 2014.

RASMUSSEN, H.; SØRENSEN, H. R.; MEYER, A. S. Formation of degradation compounds from lignocellulosic biomass in the biorefinery: Sugar reaction mechanisms. **Carbohydrate Research**, v. 385, p. 45–57, 2014.

ROCHA, G. J. DE M. et al. Dilute mixed-acid pretreatment of sugarcane bagasse for ethanol production. **Biomass and Bioenergy**, v. 35, p. 663–670, 2011.

ROCHA, G. J. M. et al. Pilot scale steam explosion and diluted sulfuric acid pretreatments: Comparative study aiming the sugarcane bagasse saccharification. **Industrial Crops and Products**, v. 74, p. 810–816, 2015.

SALIBA, E. D. O. S. et al. Ligninas – Métodos De Obtenção E Caracterização Química Lignins. **Ciência Rural**, v. 31, n. 5, p. 917–928, 2001.

SANNIGRAHI, P. et al. Pseudo-lignin and pretreatment chemistry. **Energy Environ. Sci.**, v. 4, n. 4, p. 1306–1310, 2011.

SANNIGRAHI, P.; RAGAUSKAS, A. J.; MILLER, S. J. Effects of Two-Stage Dilute Acid Pretreatment on the Structure and Composition of Lignin and Cellulose in Loblolly Pine. **BioEnergy Research**, v. 1, n. 3–4, p. 205–214, 2008.

SANT'ANNA, C.; SOUZA, W. DE; BRIENZO, M. The Influence of the Heterogeneity, Physicochemical and Structural Properties on the Recalcitrance and Conversion of

Sugarcane Bagasse. In: WEBB, E. (Ed.). . **SUGARCANE: PRODUCTION, CONSUMPTION AND AGRICULTURAL MANAGEMENT SYSTEMS**. New York: Nova Science Publishers, 2014. p. 1–361.

SANTOS, A. B. DOS et al. Water Stress Alters Lignin Content and Related Gene Expression in Two Sugarcane Genotypes. **Journal of Agricultural and Food Chemistry**, v. 63, p. 4708–4720, 2015.

SANTOS, F. A. et al. Potencial da palha de cana-de-açúcar para produção de etanol. **Química Nova**, v. 35, n. 5, p. 1004–1010, 2012.

SANTOS, N. C. et al. Multidisciplinary utilization of dimethyl sulfoxide: pharmacological, cellular, and molecular aspects. **Biochemical Pharmacology**, v. 65, p. 1035–1041, 2003.

SCHMATZ, A. A. et al. Pseudo-Lignin Content Decreased with Hemicellulose and Lignin Removal, Improving Cellulose Accessibility, and Enzymatic Digestibility. **BioEnergy Research**, 2020.

SCHMATZ, A. A.; TYHODA, L.; BRIENZO, M. Sugarcane biomass conversion influenced by lignin. **Biofuels, Bioprod. Bioref.**, v. 14, p. 469–480, 2020.

SERPEN, J. Y. Comparison of Sugar Content in Bottled 100% Fruit Juice versus Extracted Juice of Fresh Fruit. **Food and Nutrition Sciences**, v. 03, n. 11, p. 1509–1513, 2012.

SHIMIZU, F. L. et al. Acid , alkali and peroxide pretreatments increase the cellulose accessibility and glucose yield of banana pseudostem. **Industrial Crops & Products**, v. 115, n. February, p. 62–68, 2018.

SHIMIZU, F. L. et al. Minimum Lignin and Xylan Removal to Improve Cellulose Accessibility. **BioEnergy Research**, v. 13, p. 775–785, 2020.

SHINDE, S. D. et al. Recent advances in understanding the pseudo-lignin formation in a lignocellulosic biorefinery. **Green Chemistry**, v. 20, n. 10, p. 2192–2205, 2018.

SINGH, R. et al. Lignin–carbohydrate complexes from sugarcane bagasse: Preparation, purification, and characterization. **Carbohydrate Polymers**, v. 62, p. 57–66, 2005.

SIQUEIRA, G. et al. Topochemical distribution of lignin and hydroxycinnamic acids in sugar-cane cell walls and its correlation with the enzymatic hydrolysis of polysaccharides. **Biotechnology for Biofuels**, v. 4, n. 7, p. 1–9, 2011.

SOARES, L. C. S. R. et al. Screening of Yeasts for Selection of Potential Strains and Their Utilization for In Situ Microbial Detoxification (ISMD) of Sugarcane Bagasse Hemicellulosic Hydrolysate. **Indian Journal of Microbiology**, v. 56, n. 2, p. 172–181, 2016.

SOLARTE-TORO, J. C. et al. Acid pretreatment of lignocellulosic biomass for energy vectors production: A review focused on operational conditions and techno-economic

assessment for bioethanol production. **Renewable and Sustainable Energy Reviews**, v. 107, n. March, p. 587–601, 2019.

SRITRAKUL, N.; NITISINPRASERT, S.; KEAWSOMPONG, S. Evaluation of dilute acid pretreatment for bioethanol fermentation from sugarcane bagasse pith. **Agriculture and Natural Resources**, v. 51, n. 6, p. 512–519, 2017.

SUN, F. F. et al. Industrially relevant hydrolyzability and fermentability of sugarcane bagasse improved effectively by glycerol organosolv pretreatment. **Biotechnology for Biofuels**, v. 9, n. 59, p. 1–13, 2016.

TRČEK, J.; MIRA, N. P.; JARBOE, L. R. Adaptation and tolerance of bacteria against acetic acid. **Appl Microbiol Biotechnol**, v. 99, p. 6215–6229, 2015.

VALDIVIA, M. et al. Biofuels 2020: Biorefineries based on lignocellulosic materials. **Microbial Biotechnology**, v. 9, n. 5, p. 585–594, 2016.

VIEIRA, S. et al. Sustainability of sugarcane lignocellulosic biomass pretreatment for the production of bioethanol. **Bioresource Technology**, v. 299, n. 122635, p. 1–9, 2020.

VINZANT, T. B. et al. Simultaneous saccharification and fermentation of pretreated hardwoods effect of native lignin content. **Applied Biochemistry and Biotechnology**, v. 62, n. 1, p. 99–104, 1997.

WAN, G. et al. How Pseudo-lignin Is Generated during Dilute Sulfuric Acid Pretreatment. **J. Agric. Food Chem**, v. 67, n. 36, p. 10116–10125, 2019.

XIN, D. et al. Improving the Hydrolytic Action of Cellulases by Tween 80: Offsetting the Lost Activity of Cellobiohydrolase Cel7A. **ACS Sustainable Chem. Eng.**, v. 5, p. 11339–11345, 2017.

YANG, J. et al. TG-FTIR-MS study of pyrolysis products evolving from peat. **Journal of Analytical and Applied Pyrolysis**, v. 117, p. 296–309, 2016.

YU, N. et al. Bioethanol from sugarcane bagasse: Focused on optimum of lignin content and reduction of enzyme addition. **Waste Management**, v. 76, p. 404–413, 2018.

ZENG, Y. et al. Lignin plays a negative role in the biochemical process for producing lignocellulosic biofuels. **Current Opinion in Biotechnology**, v. 27, p. 38–45, 2014.

ZHANG, H. et al. Enhancing enzymatic hydrolysis of sugarcane bagasse by ferric chloride catalyzed organosolv pretreatment and Tween 80. **Bioresource Technology**, v. 258, p. 295–301, 2018.

ZHANG, H.; WU, S. Generation of lignin and enzymatically digestible cellulose from ethanol-based organosolv pretreatment of sugarcane bagasse. **Cellulose**, v. 22, n. 4, p. 2409–2418, 2015.

ZHAO, X. et al. The fate of lignin during atmospheric acetic acid pretreatment of sugarcane bagasse and the impacts on cellulose enzymatic hydrolyzability for bioethanol

production. **Renewable Energy**, v. 128, p. 200–209, 2018.

ZHUANG, J. et al. Formation and deposition of pseudo-lignin on liquid-hot-water-treated wood during cooling process. **Wood Science and Technology**, v. 51, p. 165–174, 2017.