

See discussions, stats, and author profiles for this publication at: <https://www.researchgate.net/publication/286638091>

Effects of vaccination against foot-and-mouth disease virus on reproductive performance of beef cows

Article in *Journal of Animal Science* · January 2015

DOI: 10.2527/jas.2015-9537

CITATIONS

2

READS

756

8 authors, including:



Reinaldo Fernandes Cooke

Texas A&M University

203 PUBLICATIONS 1,523 CITATIONS

[SEE PROFILE](#)



Rodrigo S. Marques

Oregon State University

82 PUBLICATIONS 244 CITATIONS

[SEE PROFILE](#)



Carlos Eurico Dos Santos Fernandes

Universidade Federal de Mato Grosso do Sul

67 PUBLICATIONS 386 CITATIONS

[SEE PROFILE](#)



Gumercindo Lorian Franco

Universidade Federal de Mato Grosso do Sul

68 PUBLICATIONS 218 CITATIONS

[SEE PROFILE](#)

Some of the authors of this publication are also working on these related projects:



Ganho compensatório, hormônios, composição corporal e avaliação econômica em bovinos conduzidos a pé ou por transporte rodoviário [View project](#)



Pathological aspects of lactococcosis in hybrid surubins: Hematological, biochemical and histopathological parameters [View project](#)



**Effects of vaccination against foot-and-mouth disease virus
on reproductive performance of *Bos indicus* beef cows**

Journal:	<i>Journal of Animal Science</i>
Manuscript ID	E-2015-9537.R1
Manuscript Type:	Animal Production
Date Submitted by the Author:	30-Oct-2015
Complete List of Authors:	Cooke, Reinaldo; Oregon State University, Eastern Oregon Agricultural Research Center
Key Words:	Acute-phase response, beef cattle, foot-and-mouth disease, inflammation, pregnancy loss

SCHOLARONE™
Manuscripts

Review

Running Head: Vaccination and pregnancy loss in beef cattle

Effects of vaccination against foot-and-mouth disease virus on reproductive performance of *Bos indicus* beef cows¹

L. C. L. Ferreira,* R. F. Cooke,† R. S. Marques,† H. J. Fernandes,‡ C. E. Fernandes,§ R. Stelato,# G. L. Franco,* and R. A. A. Lemos*

* Universidade Federal de Mato Grosso do Sul - Faculdade de Medicina Veterinária e Zootecnia, Campo Grande 79074-460, Brazil

† Oregon State University - Eastern Oregon Agricultural Research Center, Burns, OR 97720

‡ Universidade Estadual de Mato Grosso do Sul – Unidade Universitária de Aquidauana, Aquidauana 79200-000, Brazil

§ Universidade Federal de Mato Grosso do Sul - Centro de Ciências Biológicas e da Saúde, Campo Grande 79074-460, Brazil

Laboratório Zoetis Ltda., Campo Grande 79074-460, Brazil

¹ Corresponding author: reinaldo.cooke@oregonstate.edu. Dr. Reinaldo Cooke is also affiliated as graduate professor to the Programa de Pós-Graduação em Zootecnia / Faculdade de Medicina Veterinária e Zootecnia, UNESP - Univ. Estadual Paulista, Botucatu, SP, Brazil, 18618-970..

ABSTRACT: This study compared reproductive performance of *Bos indicus* cows vaccinated against the foot-and-mouth-disease (FMD) virus prior to timed-AI or during early pregnancy (Exp. 1), as well as rectal temperature (RT) and plasma concentrations of the acute-phase protein haptoglobin in cattle vaccinated or not against the FMD virus (Exp. 2). Cattle utilized in Exp. 1 and 2 were originated from herds with no historical occurrences of FMD, and that received vaccination against the FMD virus biannually. In Exp. 1, 604 lactating, multiparous, non-pregnant Nelore cows were randomly assigned on d -31 of the experiment to receive: 1) **VACPRE** = vaccination against FMD virus on d -31 ($n = 291$), and 2) **VACGEST** = vaccination against FMD virus on d 30 ($n = 313$). From d -11 to 0, all cows were assigned to an estrus synchronization + timed-AI (d 0) protocol. Pregnancy status to AI was verified on d 30 and 90 via transrectal ultrasonography. A treatment \times day interaction was detected ($P < 0.01$) for pregnancy rates to AI, which were similar ($P = 0.17$) between VACPRE and VACGEST on d 30 (61.8 vs. 56.2%, respectively; SEM = 2.8), but greater ($P < 0.01$) for VACPRE on d 90 (59.4 vs. 46.9%, respectively; SEM = 2.8). Pregnancy loss from d 30 to 90 was greater ($P < 0.01$) in VACGEST compared with VACPRE (16.5 vs. 3.9%, respectively; SEM = 2.2). In Exp. 2, 40 pregnant Nelore females (20 nulliparous and 20 multiparous cows; BCS = 4.73 ± 0.12) were ranked by parity and assigned to receive (VAC; $n = 20$) or not (NOVAC; $n = 20$) vaccination against FMD virus. Blood samples were collected and RT recorded prior to (h 0) and 24, 72, 120, and 168 h after treatment administration. Treatment \times day interactions were detected ($P < 0.01$) for RT and plasma haptoglobin. The RT was greater ($P < 0.01$) in VAC compared with NOVAC at 24 h after treatment administration, and similar ($P \geq 0.31$) between treatments at all other sampling hours. Plasma haptoglobin concentration was similar ($P = 0.98$) between VAC and NOVAC prior to treatment administration ($P = 0.48$), and greater ($P < 0.01$) in VAC at 24,

72, 120, and 168 h after treatment administration. In summary, vaccinating *B. indicus* beef cows against FMD virus resulted in a 4-fold increase in pregnancy loss when the vaccine was administered 30 d after timed-AI compared with 31 d prior to timed-AI. These outcomes can be associated with inflammatory and acute-phase reactions elicited by the FMD vaccine, which are known to impair pregnancy maintenance in cattle.

Key Words: Acute-phase response, beef cattle, foot-and-mouth disease, inflammation, pregnancy loss

INTRODUCTION

Foot-and-mouth disease (FMD) is a severe, highly contagious viral disease that affects cloven-hoofed livestock species including cattle (Grubman and Baxt, 2004), and has been recognized as a major constraint to international trade in animals and animal products (Leforban, 1999). While FMD has been eradicated in North America and Western Europe, this disease is still endemic in Africa, South America, and Asia (USDA-APHIS, 2013). Vaccination against the FMD virus successfully reduced FMD outbreaks in many parts of the world (Brown, 1992; Kahn et al., 2002); therefore, vaccination is a common and often mandatory strategy used to mitigate FMD in endemic regions (Rodriguez and Gay, 2011).

Early pregnancy loss, particularly during the first trimester of gestation, is a major reproductive challenge in cow-calf systems (Humbolt, 2001). Hence, strategies to alleviate early pregnancy losses are warranted for optimal reproductive and overall efficiency of cow-calf operations. The majority of FMD vaccines used worldwide contain inactivated FMD virus serotypes and an oil-based adjuvant to elicit a greater immune protection to target antigens (Rodriguez and Grubman, 2009). In general, adjuvants elicit innate immune responses associated with antigen presentation to T cell lymphocytes, including inflammatory and acute-phase reactions (Tizard, 2004; Rodrigues et al., 2015) known to result in pregnancy losses in cattle (Hansen et al., 2004). Based on this rationale, we hypothesized that administration of a FMD vaccine during early pregnancy stimulates an acute-phase protein reaction and results in increased pregnancy loss in vaccinated cattle. To test this hypothesis, Exp. 1 compared reproductive performance of *Bos indicus* cows vaccinated against FMD virus prior to timed-AI or during early pregnancy, whereas Exp. 2 compared rectal temperature and plasma

concentrations of the acute-phase protein haptoglobin in cattle vaccinated or not against the FMD virus.

MATERIALS AND METHODS

Experiment 1 was conducted on a commercial cow-calf operation located in Miranda, MS, Brazil, and cattle were cared for in accordance with the practices outlined in the *Guide for the Care and Use of Agricultural Animals in Agricultural Research and Teaching* (FASS, 2010). Experiment 2 was conducted at the Universidade Federal do Mato Grosso do Sul, located in Terenos, MS, Brazil, and cattle were cared for in accordance with acceptable practices and experimental protocols reviewed and approved by the Universidade Federal do Mato Grosso do Sul - Animal Ethics Committee. Cattle utilized in Exp. 1 and 2 originated from herds with no historical occurrences of FMD, and that received vaccination against the FMD virus twice a year. In addition, all cattle utilized herein were vaccinated against FMD approximately 6 mo prior to the beginning of each experiment.

Experiment 1

Animals and treatments. A total of 604 lactating, multiparous, non-pregnant Nelore cows (approximately 65 to 95 d days post-partum; BCS = 3.85 ± 0.05 according to Wagner et al., 1988), maintained in 2 groups of 266 and 338 cows each, were assigned to the experiment (d -31 to d 90 **relative to timed-AI**). Groups were maintained in individual *Brachiaria brizanta* pastures with ad libitum access to water and a commercial mineral-vitamin mix (DSM Produtos Nutricionais Brasil, São Paulo, SP, Brazil), and independently assigned to experimental procedures 1 d apart. Within each group, cows were randomly assigned on d -31 of the experiment to receive: 1) **VACPRE** = vaccination against the FMD virus (5 mL s.c. of Ourovac Aftosa; Ourofino Saúde Animal, Cravinhos, SP, Brazil) on d -31 of the experiment (**31 d prior to**

101 **timed-AI**), and 2) **VACGEST** = vaccination against FMD virus (5 mL s.c. of Ourovac Aftosa;
102 Ourofino Saúde Animal) on d 30 of the experiment (**30 d after timed-AI**).

103 On d -11, both groups were assigned to the same estrus synchronization + timed-AI
104 protocol (Meneghetti et al, 2009; d -11 to 0). **More specifically, cows received a 2 mg injection**
105 **(i.m.) of estradiol benzoate (Gonadiol; Zoetis, São Paulo, SP, Brazil) and an intravaginal**
106 **progesterone releasing device (CIDR, containing 1.9 g of progesterone; Zoetis) on d -11. On d -**
107 **2, CIDR was removed, cows received a 12.5 mg injection (i.m.) of PGF_{2α} (Lutalyse; Zoetis), in**
108 **addition to 0.6 mg injection (i.m.) of estradiol cypionate (ECP; Zoetis) and 300 IU injection**
109 **(i.m.) of eCG (Novormon; Zoetis). On d 0, cows were assigned to timed-AI. All cows were**
110 inseminated with semen from a single sire. Within each group, cows were inseminated by 1 of 2
111 technicians, and the distribution of cows inseminated by each technician was equal within each
112 treatment.

113 ***Sampling.*** Cow BCS (Wagner et al., 1988) was assessed at timed-AI on d 0. Pregnancy
114 status to AI was verified on d 30 and 90 of the experiment by detecting a viable **conceptus** with
115 transrectal ultrasonography (5.0-MHz transducer; Chison 600, Chison Medical Imaging Co.,
116 Ltd., Wuxi City, Jiang Su Province, China). Cows were not exposed to bulls or to additional AI
117 services between timed-AI and pregnancy evaluation on d 30. **On d 31, cows were exposed to**
118 **mature bulls for 50 d (1:25 bull to cow ratio).** Cows diagnosed as pregnant on d 30, and then
119 non-pregnant or with **an estimated conceptus age** of ≤ 60 d on d 90 (Curran et al., 1986) were
120 considered to have lost the AI pregnancy.

121 ***Statistical analysis.*** Quantitative and binary data were analyzed, respectively, with the
122 MIXED and GLIMMIX procedures of SAS (SAS Inst., Inc.; version 9.3) and Satterthwaite
123 approximation to determine the denominator df for the tests of fixed effects, using cow as the

experimental unit and cow(treatment \times group) as random variable. The model statement used for analysis of cow BCS on d 0 and pregnancy loss contained the effects of treatment, group, and the resultant interaction. The model statement used for analysis of pregnancy rates to timed-AI contained the effects of treatment, group, day of pregnancy diagnosis (d 30 or 90), and all resultant interactions. Results are reported as least square means and separated using LSD. Significance was set at $P \leq 0.05$, and tendencies were determined if $P > 0.05$ and ≤ 0.10 . Results are reported according to treatment effects if no interactions were significant, or according to the highest-order interaction detected.

Experiment 2

Animals and treatments. A total of 40 pregnant Nelore females, being 20 non-lactating nulliparous and 20 lactating multiparous cows, were assigned to the experiment (BCS = 4.73 ± 0.12 according to Wagner et al., 1988). All cows were maintained in a single *B. brizanta* pasture with ad libitum access to water and a commercial mineral-vitamin mix (DSM Produtos Nutricionais Brasil). At the beginning of the experiment (d 0), cows were ranked by parity and assigned to receive: 1) **VAC** = vaccination against the FMD virus (5 mL s.c. of Ourovac Aftosa; Ourofino Saúde Animal) on d 0, and 2) **NOVAC** = no vaccination against the FMD virus on d 0.

Sampling. Rectal temperature was recorded (G-Tech digital thermometer; G-Tech; São Paulo, SP, Brazil) and blood samples were collected immediately prior to (h 0) and 24, 72, 120, and 168 h after treatment administration. Blood was collected via jugular venipuncture into commercial blood collection tubes (Vacutainer, 10 mL; Becton Dickinson, Franklin Lakes, NJ) with 158 USP units of freeze-dried sodium heparin, placed immediately on ice, centrifuged ($2,500 \times g$ for 30 min; 4°C) for plasma harvest, and stored at -20°C on the same day of collection. All plasma samples were analyzed for haptoglobin concentration according to

colorimetric procedures described by Cooke and Arthington (2013). The intra- and inter-assay CV were, respectively, 2.4 and 7.6%.

Statistical analysis. Data were analyzed with the MIXED procedure of SAS (SAS Inst., Inc.; version 9.3) and Satterthwaite approximation to determine the denominator df for the tests of fixed effects, using cow as the experimental unit and cow(treatment \times parity) as random variable. The model statement used for analysis of plasma haptoglobin and rectal temperature contained the effects of treatment, parity, hour, and all resultant interactions. The specified term for the repeated statements was hour, cow(treatment \times parity) as subject, and the covariance structure utilized was autoregressive based on the Akaike information criterion. Results are reported as least square means and separated using LSD. Significance was set at $P \leq 0.05$, and tendencies were determined if $P > 0.05$ and ≤ 0.10 . Results are reported according to treatment effects if no interactions were significant, or according to the highest-order interaction detected.

RESULTS

Experiment 1

No treatment differences were detected ($P = 0.87$) for cow BCS at timed-AI (3.87 vs. 3.84 of BCS for VACPRE vs. VACGEST cows; SEM = 0.10),

A treatment \times day interaction was detected ($P < 0.01$) for pregnancy rates to AI (Table 1), which were similar ($P = 0.17$) between treatments on d 30, but greater ($P < 0.01$) for VACPRE compared with VACGEST cows on d 90. Accordingly, pregnancy loss from d 30 to 90 was greater ($P < 0.01$) in VACGEST compared with VACPRE cows (Table 1).

Experiment 2

A treatment \times hour interaction was detected ($P < 0.01$) for rectal temperature, which was similar between VAC and NOVAC cows prior to treatment administration ($P = 0.48$), greater (P

< 0.01) in VAC compared with NOVAC cows at 24 h after treatment administration, and similar ($P \geq 0.31$) between treatments at 72, 120, and 168 h after treatment administration (Figure 1).

A treatment \times hour interaction was also detected ($P < 0.01$) for plasma haptoglobin concentration, which was similar ($P = 0.98$) between VAC and NOVAC cows prior to treatment administration ($P = 0.48$), and greater ($P < 0.01$) in VAC compared with NOVAC cows at 24, 72, 120, and 168 h after treatment administration (Figure 2).

DISCUSSION

In FMD endemic regions, cattle are vaccinated against the FMD virus every 6 mo due to the vaccine immunoprotection length (Parida, 2009). Based on the productive cycle of cow-calf operations (Hixon and Sanson, 2012), vaccination against the FMD virus often occurs during or shortly after the annual breeding season. Although previous research reported that FMD vaccines impair cattle production traits, including decreased milk production (Yeruham et al., 2001) and increased carcass lesions (Leal et al., 2014), the impacts of vaccination against the FMD virus on reproductive performance of beef cows still warranted investigation. Therefore, results from Exp. 1 are novel and support our hypothesis that administering a FMD vaccine during early pregnancy increased incidence of pregnancy loss in beef cows by 4-fold when compared with vaccine administration prior to timed-AI. It is important to mention that these outcomes were independent of cow BCS, and should not be associated with cow nutritional status during the estrus synchronization + timed-AI protocol (Cooke et al., 2009).

The majority of FMD vaccines utilized worldwide contain inactivated FMD virus serotypes, and an oil-based adjuvant that elicits innate immune responses associated with antigen presentation to T cell lymphocytes, including inflammatory and acute-phase reactions (Tizard, 2004; Rodriguez and Grubman, 2009; Rodrigues et al., 2015). These immune responses,

however, have been negatively associated with pregnancy maintenance (Hansen et al., 2004) and overall reproductive performance in cattle (Cooke et al., 2009). More specifically, adjuvants stimulate synthesis of pro-inflammatory cytokines (Rodrigues et al., 2015), which in turn elicit 2 major acute-phase responses: 1) synthesis of prostaglandins that lead to hyperthermia, and 2) altered liver metabolism and gene regulation, favoring hepatic synthesis of acute-phase proteins such as haptoglobin (Carroll and Forsberg, 2007). Pro-inflammatory cytokines are known to impact pregnancy maintenance via direct embryotoxic effects, reduced endometrial cell proliferation, in addition to increased body temperature and endometrial $\text{PGF}_{2\alpha}$ synthesis to levels that interrupt early pregnancy (Hansen et al., 2004). Conversely, haptoglobin does not have detrimental effects on cattle productive and reproductive functions, although this acute-phase protein is widely used to monitor inflammatory and acute-phase responses in cattle (Horadagoda et al., 1999; Cooke and Arthington, 2013).

Supporting this rationale, results from Exp. 2 demonstrated that administering a FMD vaccine elicited inflammatory and acute-phase responses, represented by treatment effects on rectal temperature and plasma haptoglobin concentrations, which can be directly associated with treatment effects detected for pregnancy loss in Exp. 1. Supporting findings from Exp. 2, Arthington et al. (2013) and Rodrigues et al. (2015) also reported that administering a vaccine containing inactivated pathogens + adjuvant to beef cattle receiving increased plasma haptoglobin concentrations for up to 120 h, and associated these outcomes with reduced performance traits. It is important to note that inflammatory and acute-phase responses may also impair cattle fertility parameters such as follicle development and ovulation (Peter et al., 1989; Battaglia et al., 2000; Williams et al., 2001), which were not directly assessed in the present experiment although pregnancy rates to AI on d 30 were similar between treatments. Given that

the FMD vaccine utilized herein increased plasma haptoglobin concentrations for at least 7 d, it seems plausible that beef cows should be vaccinated against the FMD virus at least 1 wk prior to the beginning of the breeding season to prevent fertility and pregnancy losses, which supports that practical application of the VACPRES treatment evaluated in Exp. 1. Nevertheless, research is still warranted to determine the most appropriate timing for FMD vaccination to beef females.

In summary, administering a FMD vaccine to Nelore beef cows resulted in a 4-fold increase in pregnancy loss when vaccination occurred 30 d after timed-AI compared with 31 d prior to timed-AI. These outcomes can be associated with the inflammatory and acute-phase reactions elicited by the FMD vaccine, which are known to impair pregnancy maintenance in cattle (Hansen et al., 2004). Therefore, beef cows should not receive FMD vaccines based on inactivated virus and an oil-based adjuvant during early gestation; this should be administered prior to the beginning of the breeding season to prevent early pregnancy losses and optimize reproductive and overall efficiency of cow-calf operations.

REFERENCES

- Arthington, J. D., R. F. Cooke, T. D. Maddock, D. B. Araujo, P. Moriel, N. DiLorenzo, and G. C. Lamb. 2013. Effects of vaccination on the acute-phase protein response and measures of performance in growing beef calves. *J. Anim. Sci.* 91:1831-1837.
- Battaglia, D. F., H. B. Krasa, V. Padmanabhan, C. Viguie, and F. J. Karsch. 2000. Endocrine alterations that underlie endotoxin-induced disruption of the follicular phase in ewes. *Biol. Reprod.* 62:45-53.
- Brown, F. 1992. New approaches to vaccination against foot-and-mouth disease. *Vaccine* 10:1022-1026.

- 239 Carroll, J. A., and N. E. Forsberg. 2007. Influence of stress and nutrition on cattle immunity. *Vet.*
240 *Clin. Food. Anim.* 23:105-149.
- 241 Cooke, R. F., and J. D. Arthington. 2013. Concentrations of haptoglobin in bovine plasma
242 determined by ELISA or a colorimetric method based on peroxidase activity. *J. Anim.*
243 *Physiol. Anim. Nutr.* 97:531-536.
- 244 Cooke, R. F., J. D. Arthington, D. B. Araujo, and G. C. Lamb. 2009. Effects of acclimation to
245 human interaction on performance, temperament, physiological responses, and pregnancy
246 rates of Brahman-crossbred cows. *J. Anim. Sci.* 87:4125-4132.
- 247 Curran, S., R. A. Pierson, and O. J. Ginther. 1986. Ultrasonographic appearance of the bovine
248 conceptus from days 20 through 60. *J. Am. Vet. Med. Assoc.* 189:1295–1302.
- 249 FASS. 2010. Guide for the Care and Use of Agricultural Animals in Agricultural Research and
250 Teaching. (3rd ed.). Federation of Animal Science Societies, Savoy, IL.
- 251 Grubman, M. J., and B. Baxt. 2004. Foot-and-Mouth disease. *Clin. Microbiol. Rev.* 17:465-493.
- 252 Hansen, P. J., P. Soto, and R. P. Natzke. 2004. Mastitis and fertility in cattle – possible
253 involvement of inflammation or immune activation in embryonic mortality. *Am. J.*
254 *Reprod. Immunol.* 51:294–301.
- 255 Hixon, D. L., and D. W. Sanson. 2012. In *Cattle Producer's Handbook* 3rd ed., 400. JRAAdams
256 Publishing, Boise, ID.
- 257 Horadagoda, N. U., K. M. Knox, H. A. Gibbs, S. W. Reid, A. Horadagoda, S. E. Edwards, and P.
258 D. Eckersall. 1999. Acute phase proteins in cattle: Discrimination between acute and
259 chronic inflammation. *Vet. Rec.* 144:437–441.

- 260 Humbolt, P. 2001. Use of pregnancy specific proteins and progesterone assays to monitor
261 pregnancy and determine the timing, frequencies and sources of embryonic mortality in
262 ruminants. *Theriogenology* 56:1417–1433.
- 263 Kahn, S., D. W. Geale, P. R. Kitching, A. Bouffard, D. G. Allard, and J. R. Duncan. 2002.
264 Vaccination against foot-and-mouth disease: The implications for Canada. *Can. Vet. J.*
265 43:349–354.
- 266 Leal, P.V., R. C. Pupin, A. C. Santos, T. C. Faccin, E. Surdi, C. R. B. Leal, R. C. Brumatti, and
267 R. A. A. Lemos. 2014. Estimates of economic losses caused by local granulomatous
268 reaction after use of an oily vaccine against FMD in cattle of Mato Grosso do Sul. *Pesq.*
269 *Vet. Bras.* 34:738-742.
- 270 Leforban, Y. 1999. Prevention measures against foot-and-mouth disease in Europe in recent
271 years. *Vaccine* 17:1755–1759.
- 272 Meneghetti, M., O. G. Sa Filho, R. F. G. Peres, G. C. Lamb, and J. L. M. Vasconcelos. 2009.
273 Fixed-time artificial insemination with estradiol and progesterone for *Bos indicus* cattle:
274 I. Basis for development of protocols. *Theriogenology* 72:179-189.
- 275 Parida, S. 2009. Vaccination against foot-and-mouth disease virus: strategies and effectiveness.
276 *Expert Rev. Vaccin.* 8:347–365.
- 277 Peter, A. T., W. T. K. Bosu, R. J. DeDecher. 1989. Suppression of preovulatory luteinizing
278 hormone surges in heifers after intrauterine infusions of *Escherichia coli* endotoxin. *Am.*
279 *J. Vet. Res.* 50:368-373.
- 280 Rodrigues, M. C., R. F. Cooke, R. S Marques, B. I. Cappellozza, S. A. Arispe, D. H. Keisler, and
281 D. W. Bohnert. 2015. Effects of vaccination against respiratory pathogens on feed intake,

- 282 metabolic, and inflammatory responses in beef heifers. *J. Anim. Sci.*
283 doi:10.2527/jas2015-9277.
- 284 Rodriguez, L. L., and C. G. Gay. 2011. Development of vaccines toward the global control and
285 eradication of foot-and-mouth disease. *Expert Rev. Vaccines* 10:377–387.
- 286 Rodriguez, L. L., and M. J. Grubman. 2009. Foot and mouth disease virus vaccines. *Vaccine*
287 27:D90-D94.
- 288 Tizard, I. R. 2004. Vaccines and their production. In: T. Merchant, editor, *Veterinary*
289 *immunology*. 7th ed. Elsevier, Philadelphia, PA. p. 247.
- 290 USDA-APHIS. 2013. Foot-and-mouth disease. USDA-APHIS Publications. [http://www.aphis.](http://www.aphis.usda.gov/publications/animal_health/2013/fs_fmd_general.pdf)
291 [usda.gov/publications/animal_health/2013/fs_fmd_general.pdf](http://www.aphis.usda.gov/publications/animal_health/2013/fs_fmd_general.pdf). Accessed July 7, 2015.
- 292 Wagner, J. J., K. S. Lusby, J. W. Oltjen, J. Rakestraw, R. P. Wettemann, and L. E. Walters.
293 1988. Carcass composition in mature Hereford cows: Estimation and effect on daily
294 metabolizable energy requirement during winter. *J. Anim. Sci.* 66:603-612.
- 295 Williams, C. Y., T. G. Harris, D. F. Battaglia, C. Viguie, and F. J. Karsch. 2001. Endotoxin
296 Inhibits Pituitary Responsiveness to Gonadotropin-Releasing Hormone. *Endocrinology*
297 142:1915-1922.
- 298 Yeruham, I., H. Yadin, M. Haymovich, and S. Perl. 2001. Adverse reactions to FMD vaccine.
299 *Vet. Dermatol.* 12:197-201.

300

Table 1. Reproductive performance of *Bos indicus* beef cows vaccinated against foot-and-mouth disease virus (5 mL s.c. of Ourovac Aftosa; Ourofino Saúde Animal, Cravinhos, SP, Brazil) on d -31 (**VACPRE**; n = 291) or d 30 (**VACGEST**; n = 313) relative to timed-AI (d 0). ¹

Item	VACPRE	VACGEST	SEM	P-value
Pregnancy rates to timed-AI, ² %				
d 30	61.8 (180/291)	56.2 (176/313)	2.8	0.17
d 90	59.4 (173/291)	46.9 (147/313)	2.8	< 0.01
Pregnancy loss from d 30 to 90, ³ %	3.9 (7/180)	16.5 (29/176)	2.2	< 0.01

¹ On d -11, groups were assigned to the following estrus synchronization + timed-AI protocol. Cows received a 2 mg injection (i.m.) of estradiol benzoate (Gonadiol; Zoetis, São Paulo, SP, Brazil) and an intravaginal progesterone releasing device (**CIDR**, containing 1.9 g of progesterone; Zoetis) on d -11. On d -2, CIDR was removed, cows received a 12.5 mg injection (i.m.) of PGF_{2α} (Lutalyse; Zoetis), in addition to 0.6 mg injection (i.m.) of estradiol cypionate (ECP; Zoetis) and 300 IU injection (i.m.) of eCG (Novormon, Zoetis). On d 0, cows were assigned to timed-AI. Pregnancy status to AI was verified on 30 and 90 d after timed-AI by detecting a viable **conceptus** with transrectal ultrasonography (5.0-MHz transducer; Chison 600, Chison Medical Imaging Co., Ltd., Wuxi City, Jiang Su Province, China).

² Values within parenthesis represent number of pregnant cows divided by number of total cows assigned to timed-AI.

³ Values within parenthesis represent number of cows that lost AI pregnancy divided by number of diagnosed as pregnant to timed-AI on d 30.

Figure 1. Rectal temperature of *Bos indicus* beef cows vaccinated (VAC; n = 20) or not (NOVAC; n = 20) against foot-and-mouth disease virus (5 mL s.c. of Ourovac Aftosa; Ourofino Saúde Animal, Cravinhos, SP, Brazil). A treatment \times hour interaction was detected ($P < 0.01$). Treatment comparison within hour; ** $P \leq 0.01$.

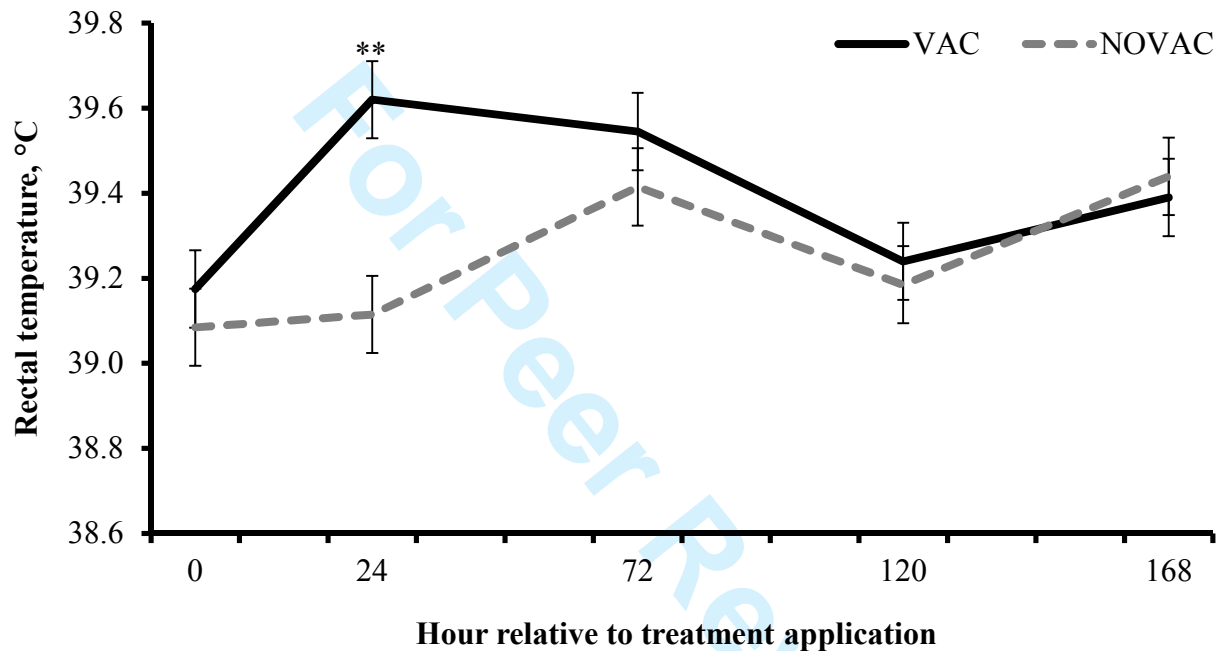


Figure 2. Plasma haptoglobin concentrations in *Bos indicus* beef cows vaccinated (VAC; n = 20) or not (NOVAC; n = 20) against foot-and-mouth disease virus (5 mL s.c. of Ourovac Aftosa; Ourofino Saúde Animal, Cravinhos, SP, Brazil). A treatment × hour interaction was detected ($P < 0.01$). Treatment comparison within hour; ** $P \leq 0.01$.

