



## Aphid vectors of *Papaya ringspot virus* and their weed hosts in orchards in the major papaya producing and exporting region of Brazil



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### ARTICLE INFO

#### Article history:

Received 6 February 2016

Received in revised form

25 August 2016

Accepted 28 August 2016

#### Keywords:

*Carica papaya*

Epidemiology

Management tactics

Potyvirus

Virus transmission

Weed hosts

### ABSTRACT

Papaya ringspot, one of the most important diseases of papaya, is caused by *Papaya ringspot virus* type P (PRSV-P), which is transmitted by various aphid species. Objectives of this research were to identify the aphid species and their weed hosts of papaya orchards in the major papaya producing and exporting region of Brazil in northern Espírito Santo State. Aphids were collected from weed hosts, and with Möericke-type traps in papaya orchards of twenty farms in Linhares, Sooretama and Aracruz municipalities of Espírito Santo, Brazil. A total of forty aphid species were collected, of which 22 are new records in the State. Thirty-eight aphid species were identified from traps, including *Aphis craccivora* Koch, *A. gossypii* Glover, *A. spiraecola* Patch, *Myzus persicae* (Sulzer), and *Toxoptera citricidus* (Kirkaldy), known vectors of PRSV-P. *Cryptomyzus* Oestlund (Aphididae) is recorded for the first time in Brazil. Thirteen aphid species associated with 22 weed species of 14 families were collected. *Aphis gossypii*, which has been reported to be one of the most efficient vectors of PRSV-P, infested the greatest number of weed hosts. *Chamaesyce hyssopifolia* (L.) Small, *Euphorbia hirta* L. (Euphorbiaceae), *Commelina benghalensis* L. (Commelinaceae), *Lepidium virginicum* L. (Brassicaceae) and *Sida* sp. (Malvaceae) were the most frequently infested weeds. *Commelina benghalensis* was infested by the greatest number of aphid species during this study. Integrated management of weed hosts of aphids to reduce aphid populations in papaya orchards may be useful for control of PRSV-P. Based on accurate knowledge of the aphid and weed species present in papaya orchards obtained in this study, integrated pest management efforts may be more efficiently directed toward eliminating the most frequently infested weed species that serve as hosts for aphid vectors of PRSV-P.

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### 1. Introduction

Papaya ringspot, caused by *Papaya ringspot virus* type P (PRSV-P), genus *Potyvirus* (Potyviridae) is one of the major diseases that limits papaya production in most countries (Fermin et al., 2010).

The loss of production may reach 100% depending on the level of incidence of the disease and age of infected plants (Singh et al., 2005; Martins and Ventura, 2007; Tennant et al., 2007; Tripathi et al., 2008). Diseased papaya trees exhibit symptoms of mosaic, foliar deformation, oily ringspots on the fruits, and oily spots or streaks on the leaf petioles. Plants may appear stunted, and infected seedlings present distinct leaf vein-clearing with the youngest leaves curving downwards in 1–2 weeks after infection; after several weeks, the leaves become mottled and distorted, with lobes

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greatly deformed (Purcifull et al., 1984; Lima et al., 2001). Infected young plants may not be killed by this virus, and may produce fruits, and they become important reservoirs of inoculum (Ventura et al., 2004; Costa, 2005). Dissemination of PRSV-P in papaya orchards is very rapid, particularly in regions with high population of aphids. From the first foci, all plants in an orchard may be infected within a period of 4–7 months. Thus, infected plants must be immediately removed (roguing) after the first appearance of disease symptoms to prevent further spread of the virus to adjacent plants or nearby orchards (Ventura et al., 2004).

Aphids (Hemiptera: Aphidoidea) are vectors of PRSV-P, and their rapid life cycle, high reproductive rate, and dispersal ability make these insects suitable for rapid dissemination of this virus (Costa, 2005; Fereres and Moreno, 2009). Twenty-one species of aphids have been confirmed as vectors of PRSV-P in a non-persistent virus-vector relationship (Purcifull et al., 1984; Gonsalves, 1998; Culik et al., 2003). Aphid species that have been reported to transmit PRSV-P include *Acyrtosiphon malvae* (Mosley), *Aphis craccivora* Koch, *A. fabae* Scopoli, *A. coreopsidis* Thomas, *A. gossypii* Glover, *A. medicaginis* Koch, *A. nerii* Boyer de Fonscolombe, *A. rumicis* Linnaeus, *A. spiraeicola* Patch, *Uroleucon sonchi* (Linnaeus), *Myzus persicae* (Sulzer), *Pentalonia nigronervosa* Coquerel, *Rhopalosiphum maidis* (Fitch), *Toxoptera aurantii* (Boyer de Fonscolombe), and *T. citricidus* (Kirkaldy) (Jensen, 1949; Capoor and Varma, 1958; Lana, 1980; Namba and Higa, 1981; Wang, 1981; Vegas et al., 1985; Prasad and Sarkar, 1989; Culik et al., 2003; Kalleshwaraswamy et al., 2007). *Aphis coreopsidis*, *A. fabae*, *A. gossypii*, *M. persicae*, and *T. citricidus* have been confirmed as vectors of PRSV-P in Brazil (Martins et al., 2009).

Aphids do not colonize papaya trees, but the virus is acquired and transmitted by aphids during brief periods of feeding probes to locate suitable hosts for colonization (Kalleshwaraswamy et al., 2007), and Fermin et al. (2010) proposed that it is necessary to evaluate the different aphid species that are able to transmit PRSV-P to develop effective management methods for the most abundant and prevalent species. The efficiency of PRSV-P transmission by aphids depends on the aphid species and viral isolate (Adsuar, 1947; Jensen, 1949; Capoor and Varma, 1958; Lana, 1980). Many aphid species and their host plants are exotic in the tropics, and these insects are commonly associated with herbaceous plant hosts (Dixon, 1987; Peronti et al., 2002). Most commercial papaya orchards in the Brazilian producing and exporting region are maintained with spontaneous vegetation (weeds) among the rows of papaya trees, based on requirements for plant groundcover of GlobalGap and Brazilian Integrated Papaya Production (Martins et al., 2009; Mansilla et al., 2013). Therefore, because little is known of the aphid species and their weed hosts that may be present in papaya orchards in Brazil, the objectives of this study were to identify the aphid species and their host plants in commercial papaya orchards to improve understanding of their importance in the epidemiology of papaya ringspot for development of integrated pest management (IPM) strategies.

## 2. Materials and methods

Aphids species were sampled during two years in twenty papaya producing farms with a total of 333 ha cropped with 'Golden', 'Sunrise Solo' and 'Tainung 01' cultivars. Fifteen farms were sampled in Linhares (19°22'55,8"S; 40°01'43,2"W), three in Sooretama (19°04'18,0"S; 40°08'52,0"W), and two in Aracruz (19°46'04,7"S; 40°10'34,8"W) municipalities, located in northern Espírito Santo State, the main papaya producing and exporting region of Brazil (Fig. 1).

Aphids were collected in papaya orchards with Möericke-type yellow traps (Oliveira et al., 1977) mounted on wood stands 0.50 m

above the ground, at a rate of one trap per hectare, with a maximum of 10 traps per orchard. Traps were placed in the orchards for 48 h, at weekly intervals for two years. All winged aphids specimens collected were fixed in 100% ethyl alcohol. Aphids were also collected from infested weeds of five 1 m<sup>2</sup> plots, randomly located between the rows of papaya trees of each orchard as follows: the number of weed plants in the plots was recorded and aphid infested weeds found in the plots were removed with their roots, placed in kraft paper bags and transported to the Laboratory of Entomology of the Instituto Capixaba de Pesquisa, Assistência Técnica e Extensão Rural (Incaper) where they were potted and the number of aphids specimens counted. Subsequently, the infested plants were transferred to screened cages in climate control chambers with temperature 25 ± 2 °C, relative humidity 70 ± 10%, and L:D 14:10 to obtain winged forms of the aphids. Winged aphid specimens that developed on the weed samples were collected with forceps and fixed in 100% ethyl alcohol for identification.

Aphids were identified by the authors (A.L.B.G. Peronti and R.C.Z. Carvalho) and Dr. Susan Halbert, Florida Department of Agriculture, USA. Weed hosts were preserved and identified at Incaper with nomenclature based on Tropicos.org - Missouri Botanical Garden (Tropicos, 2013). Voucher specimens are deposited in the arthropod collection of the Instituto Capixaba de Pesquisa, Assistência Técnica e Extensão Rural (Incaper), Vitória, Espírito Santo, Brazil, and in the entomological collection of the Department of Entomology, Universidade Federal de Viçosa (UFV).

## 3. Results

A total of 356 samples with aphids were obtained from traps, including 3874 aphid specimens belonging to 38 species of six families (Table 1). *Aphis spiraeicola*, *A. gossypii*, *T. citricidus*, and *A. craccivora* were the most frequent and constant species found in the papaya orchards (each in more than 20% of the samples and making up from 5.1 to 52.0% of the total number of aphids collected). Of the 38 aphid species collected from traps, 21 were recorded for the first time in Espírito Santo (Table 1).

A total of 42 weed species from 16 families were identified in the papaya orchards, and 22 of these weed species (55%) in 21 genera of 14 families were identified as aphid hosts (Table 2). These aphid host plants represented 9.4% of the total number of weed plants growing among the rows of papaya trees. Aphididae was the only family of the 13 aphid species found on the weed hosts. *Aphis gossypii* was the most frequent species, found in 55% of samples, and colonized 14 weed host species (Table 2). *A. solanella* Theobald found on *Solanum americanum* Mill. (Solanaceae), and *Cryptomyzus* sp. found on *Commelina benghalensis* L. (Commelinaceae), and on *Leonotis nepetaefolia* (L.) R. Br. (Labiatae) were not collected in traps. The genus *Cryptomyzus* Oestlund (Aphididae) was recorded for the first time in Brazil. The most frequent weed hosts of aphid species in papaya orchards were *Commelina benghalensis* (Commelinaceae), *Sida* sp. (Malvaceae), *Chamaesyce hyssopifolia* (L.) Small, *Euphorbia hirta* L. (Euphorbiaceae) and *Lepidium virginicum* L. (Brassicaceae). *Cucumis anguria* L. was the only species of Cucurbitaceae found as an aphid host in this study.

## 4. Discussion

Results of this research indicate that a high diversity of aphids is present in papaya orchards in northern Espírito Santo, including at least 40 species, of which 22 were recorded for the first time in this State. Of the 40 species found in papaya orchards in this study, *Aphis coreopsidis*, *A. gossypii*, *M. persicae* and *T. citricidus* have been reported to be efficient vectors of PRSV-P (Martins and Ventura, 2007). *Aphis craccivora*, *A. nerii*, *A. spiraeicola*, *L. erysimi*,



Fig. 1. Location of the research area in the main papaya producing-exporting region of Brazil in the State of Espírito Santo.

*P. nigronervosa*, *R. maidis*, *T. aurantii*, *U. ambrosiae* (Thomas), and *U. sonchi* were also collected in this study, and are vectors of PRSV-P (Jensen, 1949; Capoor and Varma, 1958; Higa and Namba, 1971; Lana, 1980; Culik et al., 2003). At least 16 species of aphids have previously been reported in this region, and *A. gossypii*, *A. spiraeicola*, *L. erysimi* (Kaltenbach), *M. persicae*, *T. citricidus* and *U. sonchi* (L.) have been considered to be the most common aphid species in this region (Oliveira et al., 1977; Souza-Silva and Ilharco, 1995).

The PRSV-P transmission efficiency by aphids has been studied in papaya-production regions worldwide, and *A. gossypii*, *M. persicae*, and *A. rumicis* Linnaeus, were the most efficient vectors; *Acyrtosiphon malvae*, *A. medicaginis* Koch, *Macrosiphum euphorbiae* (Thomas), *Micromyzus formosanus* (Takahashi), *R. maidis* (Fitch), *T. aurantii*, *T. citricidus*, and *U. sonchi* also transmitted PRSV-P (Jensen, 1949; Capoor and Varma, 1958; Vegas et al., 1985; Kalleshwaraswamy et al., 2007).

The most frequent and constant aphid species (number of individuals of species/total, and number of samples with species/total; Table 1) found in this study, *A. spiraeicola*, *A. gossypii*, *T. citricidus*,

and *A. craccivora*, are polyphagous species, abundant in anthropogenic environments, and are considered to be pests in many crops (Leal and Oliveira, 1983; Lazzarotto and Lazzari, 1998; Peronti et al., 2002). The three most frequent species found in papaya orchards in this study, *A. spiraeicola*, *A. gossypii*, and *T. citricidus*, are also considered to be efficient vectors of PRSV-P (Adsuar, 1947; Jensen, 1949; Videla, 1953; Capoor and Varma, 1958; Higa and Namba, 1971; Lana, 1980; Kalleshwaraswamy and Krishna Kumar, 2008), and their presence in papaya orchards represents a serious problem for the production of papaya in Brazil (*A. craccivora* has been shown to be an inefficient vector of PRSV-P) (Lana, 1980; Ventura et al., 2004).

Information on the diversity of aphids and their weed hosts in papaya orchards is useful for understanding the epidemiology of papaya ringspot, and for the development of integrated management strategies for this disease. The most frequent weed and host of the highest number of aphid species in papaya orchards, *C. benghalensis* is a very common and widely distributed pest. This weed species occurs throughout the year in this papaya region, and

**Table 1**  
Species of aphids collected in Möericke-type yellow traps in commercial papaya orchards, in the major Brazilian papaya producing and exporting region, Northern Espírito Santo State.

Species	Municipality <sup>b</sup>	No. of samples with the species <sup>c</sup>	Constancy (%) <sup>d</sup>	No. of specimens	Frequency (%) <sup>e</sup>
<b>Aphididae</b>					
<i>Aphis coreopsidis</i> (Thomas, 1878) <sup>a,g</sup>	A, L, S	25	7.02	52	1.34
<i>Aphis craccivora</i> Koch, 1854 <sup>f</sup>	A, L, S	81	22.75	198	5.11
<i>Aphis</i> cf. <i>fabae</i> Scopoli, 1763	A, L, S	27	7.58	167	4.31
<i>Aphis gossypii</i> Glover, 1877 <sup>g</sup>	A, L, S	159	44.66	453	11.69
<i>Aphis nerii</i> Boyer de Fonscolombe, 184 <sup>f</sup>	A, L, S	35	9.83	69	1.78
<i>Aphis spiraecola</i> Patch, 1914 <sup>f</sup>	A, L, S	260	73.03	2015	52.01
<i>Brachycaudus helichrysi</i> (Kaltenbach, 1843) <sup>a</sup>	A, L, S	8	2.25	9	0.23
<i>Capitophorus hippophaes</i> (Walker, 1852) <sup>a</sup>	A, L, S	10	2.81	12	0.31
<i>Carolinaia caricis</i> Wilson, 1911 <sup>a</sup>	S	1	0.28	1	0.03
<i>Hyperomyzus lactucae</i> (Linnaeus, 1758) <sup>f</sup>	S	1	0.28	1	0.03
<i>Hysteroneura setariae</i> (Thomas, 1878) <sup>a</sup>	L, S	10	2.81	12	0.31
<i>Lipaphis erysimi</i> (Kaltenbach, 1843) <sup>f</sup>	A, L, S	30	8.43	76	1.96
<i>Longiunguis sacchari</i> (Zehntner, 1897)	L, S	11	3.09	20	0.52
<i>Microparsus vignaphilus</i> (Blanchard, 1922) <sup>a</sup>	L, S	2	0.56	3	0.08
<i>Myzus persicae</i> (Sulzer, 1776) <sup>g</sup>	A, L, S	11	3.09	12	0.31
<i>Pentalonia nigronervosa</i> Coquerel, 1859 <sup>a,f</sup>	A, L, S	13	3.65	17	0.44
<i>Protaphis terricola</i> (Rondani, 1847)	L, S	37	10.39	55	1.42
<i>Rhopalosiphum maidis</i> (Fitch, 1856) <sup>f</sup>	A, L, S	25	7.02	27	0.70
<i>Rhopalosiphum nymphaeae</i> (Linnaeus, 1761) <sup>a</sup>	L	1	0.28	1	0.03
<i>Rhopalosiphum padi</i> (Linnaeus, 1758) <sup>a</sup>	L, S	2	0.56	2	0.05
<i>Rhopalosiphum rufiabdominale</i> (Sasaki, 1899)	L, S	5	1.40	5	0.13
<i>Schizaphis graminum</i> (Rondani, 1852) <sup>a</sup>	L, S	15	4.21	27	0.70
<i>Tetraneura nigriabdominalis</i> (Sasaki, 1899) <sup>a</sup>	A, L, S	26	7.30	43	1.11
<i>Toxoptera aurantii</i> (Boyer de Fonscolombe, 1841) <sup>f</sup>	A, L, S	51	14.33	82	2.12
<i>Toxoptera citricidus</i> (Kirkaldy, 1907) <sup>g</sup>	A, L, S	96	26.97	390	10.07
<i>Uroleucon ambrosiae</i> (Thomas, 1878) <sup>a,f</sup>	A, L, S	13	3.65	15	0.39
<i>Uroleucon compositae</i> (Theobald, 1915) <sup>a</sup>	A, L, S	35	9.83	64	1.65
<i>Uroleucon sonchi</i> (Linnaeus, 1767) <sup>f</sup>	A, S	3	0.84	3	0.08
<b>Drepanosiphidae</b>					
<i>Anoecia cornicola</i> (Walsh, 1863) <sup>a</sup>	S	1	0.28	1	0.03
<i>Lizerius cermelii</i> Quednau, 1974 <sup>a</sup>	L	1	0.28	1	0.03
<i>Lizerius tuberculatus</i> (Blanchard, 1939) <sup>a</sup>	S	1	0.28	1	0.03
<i>Sipha flava</i> (Forbes, 1885) <sup>a</sup>	L	2	0.56	2	0.05
<b>Greenideidae</b>					
<i>Brasilaphis bondari</i> Mordvilko, 1930 <sup>a</sup>	S	1	0.28	1	0.03
<b>Hormaphididae</b>					
<i>Cerataphis brasiliensis</i> (Hempel, 1901) <sup>a</sup>	A, L, S	9	2.53	9	0.23
<i>Cerataphis orchidearum</i> (Westwood, 1879) <sup>a</sup>	L, S	11	3.09	11	0.28
<b>Lachnidae</b>					
<i>Cinara atlantica</i> (Wilson, 1919) <sup>a</sup>	L	1	0.28	1	0.03
<i>Cinara fresai</i> (Blanchard, 1939)	S	1	0.28	2	0.05
<b>Pemphigidae</b>					
<i>Geopemphigus floccosus</i> (Moreira, 1925)	L, S	14	3.93	14	0.36

<sup>a</sup> First record of species in the State of Espírito Santo, Brazil.

<sup>b</sup> Occurrence: Aracruz (A), Linhares (L), Sooretama (S).

<sup>c</sup> Number of samples with the species present in relation to the total collected.

<sup>d</sup> Presence of the species in the total samples.

<sup>e</sup> Number of individuals of each species in relation to the total specimens collected.

<sup>f</sup> Species confirmed as vectors of PRSV-P (Jensen, 1949; Capoor and Varma, 1958; Namba and Higa, 1981; Lana, 1980; Wang, 1981; Vegas et al., 1985; Prasad and Sarkar, 1989; Culik et al., 2003).

<sup>g</sup> Species confirmed as vectors of PRSV-P in Brazil (Martins et al., 2009).

this study demonstrates that it is one of the most important natural reservoirs of aphid vectors of PRSV-P. Aphid species found on this weed species include *A. gossypii*, a vector of PRSV-P in Brazil, and considered one of the most efficient PRSV vectors worldwide (Rezende and Martins, 2005). *Tetraneura nigriabdominalis*, and *U. ambrosiae* were also found on this weed, however their transmission efficiency of PRSV-P is not known. *Blainvillea rhomboidea* Cass. is a host of *A. spiraecola*, thus it may be considered another important weed in PRSV-P disease management in papaya orchards because this aphid species is a relatively efficient for transmission of PRSV-P (Adsuar, 1947; Videla, 1953). Such weed hosts of aphids may be eradicated as a strategy for disease management (Martins et al., 2009; Ventura et al., 2004). *Cucumis anguria* is a common weed in Brazilian crops, particularly in papaya orchards, and it has not been related with natural infection of PRSV-P; however, other

cucurbits such as *Momordica charantia* L., and *Cucurbita maxima* Duchesne cv. Exposição (pumpkin) were not infected by PRSV-P though mechanical or aphid inoculations, and field exposure of plants (Spadotti et al., 2013; Mansilla et al., 2013). These data indicate that these cucurbits species do not seem to have a role in the epidemiology of the papaya ringspot in Brazil. However, these plant species may be reservoirs of aphid populations.

Several strategies have been adopted to reduce the incidence of PRSV-P such as the use of certified seedlings, planting in virus free areas whenever available, avoiding intercropping with Cucurbitaceae, systematic roguing of infected papaya trees, use of resistant varieties, cross protection (Rezende and Müller, 1995; Lima et al., 2001; Rezende and Martins, 2005), and use of resistant transgenic plants (Gonsalves, 1998; Lima et al., 2001). The commercial Brazilian papaya varieties are not resistant to PRSV-P (Singh et al.,

**Table 2**

Species of aphids colonizing weeds in commercial papaya orchards in the major Brazilian papaya producing and exporting region, Northern Espírito Santo State.

Aphid Species	Samples		Botanical family	Host plant
	No	Frequency (%)		
<i>Aphis coreopsidis</i> (Thomas, 1878)	5	6.58	Asteraceae	<i>Bidens pilosa</i> L.
<i>Aphis craccivora</i> Koch, 1854	6	7.89	Asteraceae	<i>Emilia coccinea</i> (Sims) G. Don
			Amaranthaceae	<i>Amaranthus deflexus</i> L.
<i>Aphis gossypii</i> Glover, 1877	42	55.26	Euphorbiaceae	<i>Chamaesyce hyssopifolia</i> (L.) Small
			Loranthaceae	<i>Psiittacanthus cordatus</i> (Hoff.) Blume
			Malvaceae	<i>Sida cordifolia</i> L.
			Malvaceae	<i>Sida</i> sp.
			Commelinaceae	<i>Commelina benghalensis</i> L.
			Lamiaceae	<i>Leonotis nepetaefolia</i> (L.) R. Br.
			Asteraceae	<i>Siegesbeckia orientalis</i> L.
			Asteraceae	<i>Ageratum conyzoides</i> L.
			Portulacaceae	<i>Portulaca oleracea</i> L.
			Convolvulaceae	<i>Ipomoea ramosissima</i> (Poir.) Choisy
			Cucurbitaceae	<i>Cucumis anguria</i> L.
			Asteraceae	<i>Bidens pilosa</i> L.
			Asteraceae	<i>Gnaphalium spicatum</i> Mill.
Euphorbiaceae	<i>Euphorbia hirta</i> L.			
Asteraceae	<i>Emilia coccinea</i> (Sims) G. Don			
<i>Aphis nerii</i> Boyer de Fonscolombe, 1841	2	2.63	Euphorbiaceae	<i>Chamaesyce hyrta</i> (L.) Millsp.
<i>Aphis solanella</i> Theobald, 1914	4	5.26	Solanaceae	<i>Solanum americanum</i> Mill.
<i>Aphis</i> sp.	2	2.63	Asteraceae	<i>Ageratum conyzoides</i> L.
<i>Aphis spiraeicola</i> Patch, 1914	2	2.63	Commelinaceae	<i>Commelina benghalensis</i> L.
			Asteraceae	<i>Blainvillea rhomboidea</i> Cass.
<i>Brachycaudus helichrysi</i> (Kaltenbach, 1843)	2	2.63	Asteraceae	<i>Ageratum conyzoides</i> L.
<i>Cryptomyzus</i> sp. <sup>a</sup>	4	5.26	Labiatae	<i>Leonotis nepetaefolia</i> (L.) R. Br.
<i>Lipaphis erysimi</i> (Kaltenbach, 1843)	1	1.32	Commelinaceae	<i>Commelina benghalensis</i> L.
			Brassicaceae	<i>Lepidium virginicum</i> L.
<i>Tetraneura nigriabdominalis</i> (Sasaki, 1899)	1	1.32	Commelinaceae	<i>Commelina benghalensis</i> L.
<i>Uroleucon ambrosiae</i> (Thomas, 1878)	3	3.95	Asteraceae	<i>Acanthospermum australe</i> (Loef.)
			Commelinaceae	<i>Commelina benghalensis</i> L.
			Boraginaceae	<i>Echium</i> sp.
<i>Uroleucon compositae</i> (Theobald, 1915)	2	2.63	Malvaceae	<i>Hibiscus</i> sp.
Total	76	100.00		

<sup>a</sup> First record of this genus in Brazil.

2005), and none of them are transgenic (Costa et al., 2013). Cucurbitaceae crops such as squash intercropped or in the vicinity of papaya orchards should be avoided because cucurbits are susceptible to PRSV-P (Mansilla et al., 2013). The use of live barriers, such as sugarcane and intercropping of corn or sorghum may also be used to inhibit the migration of aphids into papaya orchards (Paéz-Redondo, 2003). However, spontaneous vegetation that is maintained among the rows of papaya trees, based on groundcover requirements of international standards for production and export of Brazilian papaya, such as GlobalGap and Brazilian Integrated Fruit Production, may be one of the major problems for the integrated management of aphids to prevent the spread of PRSV-P. Weed hosts that serve as hosts of vectors such as *A. gossypii* could be eradicated, and replaced with aphid non-host plants that also protect the soil. Information generated in this research may be used to contribute to a management plan for papaya ringspot disease including management of groundcover of papaya orchards to eliminate weeds, such as *C. benghalensis*, that are major hosts of aphid vectors.

Thus, strategies for the management of aphid weed hosts in papaya orchards should be incorporated into the integrated pest management tactics in papaya orchards. This will prevent the multiplication and the dispersion of PRSV-P vectors, and may contribute to the reduction in the spread of the virus and reduce the percentage of eradicated papaya trees in mandatory operations of systematic roguing of virus infected plants.

## Acknowledgements

We thank the Fundação de Amparo à Pesquisa e Inovação do

Espírito Santo (FAPES, grant no: 53236092/11), Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq, grant no: 307752/2012-7), and Banco do Nordeste do Brasil (BNB) for financial support; and many thanks to Susan Halbert, Florida Department of Agriculture (USA), for identification of aphid species.

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