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Research article

Electrochemically assisted photocatalysis: Highly efficient treatment using thermal titanium oxides doped and non-doped electrodes for water disinfection





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ABSTRACT

Electrochemically assisted photocatalysis (by electronic drainage) is a highly promising method for disinfection of water. In this research, the efficiency of photolytic oxidation using UV-A radiation and electrochemically assisted photocatalysis (with electric potential of 1.5 V) was studied by using electrodes prepared by thermal treatment and doped with silver, for inactivation of *Escherichia coli* and *Staphylococcus aureus*. The Chick-Watson microorganism inactivation model was applied and the electrical energy consumption of the process was calculated. It was observed no significant inactivation of microorganisms when UV-A light or electric potential were applied separately. However, the electrochemically assisted photocatalytic process, with Ag-doped electrode completely inactivated the microbial population after 10 (*E. coli*) and 60 min (*S. aureus*). The best performing non-doped electrodes achieved 52.74% (*E. coli*) and 44.09% (*S. aureus*) inactivation rates after 60 min. Thus, electrochemically assisted photocatalytic activity was not only effective for the inactivation of microorganisms, but also notably low on electrical energy consumption during the treatment due to small current and low electric potential applied.

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1. Introduction

Alternative microorganism inactivation processes are highly encouraged in current applications to substitute chlorine-based methods (Li et al., 2015). The introduction of nanostructured materials applied to water treatment processes has yielded highly efficient water decontamination (Molina et al., 2014). The Advanced Oxidation Processes (AOP), considered by Wang et al. (2015a) as a "green" advanced oxidation technology is a promising, efficient and versatile form of disinfection, based on the modification, degradation or mineralization of organic compounds through processes where electrons are the only reagent involved. The AOP is characterized by the formation of hydroxyl radicals (*OH) with high oxidation potential (Chaplin, 2014; Claro et al., 2016). Among various AOP, heterogeneous photocatalysis is one of the most widely used method to effectively destruct toxic compounds and

* Corresponding author. E-mail address: ederio@rc.unesp.br (E.D. Bidoia). inactivate microorganisms (Brugnera et al., 2013; Augugliaro et al., 2015; Byrne et al., 2015).

Heterogeneous photocatalysis is based on the activation of a semiconductor by sunlight or artificial light. Semiconductors such as titanium dioxide (TiO₂), zinc oxide (ZnO), tungsten trioxide (WO₃), strontium peroxide (SrO₂), zirconium dioxide (ZrO₂), iron(III) oxide (Fe₂O₃), cerium(IV) oxide (CeO₂), etc. have been extensively studied due to their catalytic properties and photocatalytic potential of contaminants degradation. The TiO₂ is recognized as a promising semiconductor material for water purification (Scott-Emuakpor et al., 2012; Fan et al., 2014; Augugliaro et al., 2015).

From the photocatalytic standpoint, there are two main polymorphous phases of TiO₂: anatase and rutile (Yuangpho et al., 2015). The anatase phase is most commonly used due to its high photoactivity. Other studies demonstrated that pure rutile does not have photocatalytic activity (Ding et al., 2000). However, according to Molina et al. (2014), rutile photoactivity is dependent on three factors: preparation procedure (i), precursor (ii) or the organic



compound used as the reducing agent (iii). Very little is known about the rutile phase as opposed to the extensive studies on the anatase phase.

These commonly used photocatalysts are often found immobilized or powdered. When a TiO₂ photocatalyst film is immobilized on a conductive surface, a fixed positive electric potential is allowed under UV illumination. This process is known as electrochemically assisted heterogeneous photocatalysis (Dunlop et al., 2008).

The efficiency of photocatalytic processes is usually compromised by charge recombination. Hence, a reasonable strategy to improve catalyst efficiency is to modify the semiconductor structure by doping it with transition metal nanoparticles (Ag, Cr, Fe for example). When the electrons from TiO₂ after photonic excitation are imprisoned, there is a decrease followed by inhibition in the electron-hole pair (e^-)/(h^+) generated in photocatalyst surface (Hosseini et al., 2015; Bahadur et al. (2016). In addition, transition metals such as Ag have been extensively applied to the inactivation of microorganisms due to their antibacterial properties and biocompatibility (Brugnera et al., 2013; Pham and Lee, 2014; Parr and Kim, 2016).

Photocatalysis and electrochemically assisted photocatalysis have been widely described in the literature as an efficient alternative for inactivation of microorganisms such as *Escherichia coli* (Baram et al., 2007, 2011; Yu et al., 2008; Sun et al., 2014; Parr and Kim, 2016), *Bacillus subitilis, Staphylococcus aureus, Klebsiella pneumonia, Cryptosporidium parvum* oocysts (Cho et al., 2011; Vijay et al., 2013), *Enterococcus faecalis* and *Clostridium perfringens* spores (Dunlop et al., 2008).

The purpose of this research is therefore to evaluate the efficiency of photolytic oxidation (UV-A radiation only) and electrochemically assisted photocatalysis using non-doped Ti/TiO₂ either Ag-doped electrodes under UV-A irradiation and fixed electric potential and water samples containing *Escherichia coli* and *Staphylococcus aureus*.

2. Material and methods

2.1. Preparation of the Ti/TiO₂ and silver-doped Ti/TiO₂ semiconductor

The semiconductor (working electrode) was prepared according to Harper et al. (2001); Christensen et al. (2003, 2005) and Lopes et al. (2012). A titanium plate (5 cm \times 5 cm) was washed, immersed in acetone, dried and then calcined at 750 °C. The entire procedure was repeated four times. The TiO₂ film underwent heat treatment as proposed by Lopes et al. (2012). The Ti/TiO₂ electrode was removed from the muffle and cooled down to room temperature according to Christensen et al. (2003). Three electrodes were then setup with different calcination intervals to verify its impact during electrochemically assisted photocatalytic process, in which: Electrode 1: 10 min; Electrode 2: 5 min; Electrode 3: 15 min.

A fourth electrode was doped with silver nitrate (AgNO₃). The electrode doping was performed with a 1.7% AgNO₃ solution (Merck), where the electrode remained immersed for approximately 5 min and then calcined at 750 °C for 10 min. The procedure was repeated twice.

The thermal electrodes described were characterized using scanning electron microscopy (SEM) and X-ray diffraction spectrometry (XRD) by our research group (Lopes et al. (2012). Both techniques identified and characterized the formation of pure TiO_2 films and TiO_2 doped with silver (Ag-TiO₂).

2.2. Determining the photocurrent

The electrical potential applied to photoelectrochemical

processes was determined according to the photoactivity of the Ti/ TiO₂ electrodes calcined for 10 min. The photocurrent output was plotted against a 0–2.5 V potential range, at 10 mV. s⁻¹, in the presence of 0.1 mol.L⁻¹ Na₂SO₄. Measurements were taken using Autolab Potentiostat/Galvanostat. The Ag/AgCl electrode (KCl 1 mol.L⁻¹) was used as reference electrode. The control current was established under the same potential, in the absence of light.

2.3. Electrochemically assisted photocatalyst reactor

The electrochemically assisted photocatalysis setup is shown in Fig. 1. It was composed of a working electrode and a counter electrode (circular 170.1 cm² titanium net with 8.5 cm diameter) immersed in 250 mL of 0.08 N sodium sulfate solution (Na₂SO₄ Merck PA) with 0.4 mL of bacterial suspension (final colony counts within the range of 3×10^2 colony-forming units – CFU mL⁻¹). The Ti/TiO₂ anode (5 cm × 5 cm) was irradiated by two lamps in the UV-A spectrum range (8 W black light bulb – Foxlux). Maximum wavelength peaked at 365 nm, as shown in Fig. S1- supplementary materials. The electric potential was kept constant at 1.5 V.

2.4. UV-A radiation dose

Bacterial inactivation using UV-A lamp had its radiation dose defined as the product of the energy intensity and the exposure time (Equation (1)).

$$D = I \cdot t \tag{1}$$

where D is the dose of ultraviolet radiation (mW.s cm^{-2}), I the radiation intensity (mW cm^{-2}) and t is the exposure time (s).

The radiation intensity was measured by a spectrometer (Ocean Optic, USB-2000), at 0.2390 mW cm⁻² in each lamp, as shown in Fig. 2. In total, 0.48781 mW cm⁻² of radiation intensity was applied.

2.5. Photocatalytic activity test

The best microbial inactivation efficiency was verified through the different experimental conditions in Table 1.

2.6. Bacterial inactivation experiments

Inactivation assays were conducted with *Escherichia coli* CCT1457 and *Staphylococcus aureus* ATCCE538. It was inoculated with an inoculation loop, each bacterium in 50 mL of culture media



Fig. 1. Experimental setup: (A) Black light lamps UV-A; (B) Titanium counter electrode; (C) Ti/TiO₂ electrode; (D) Magnetic stirring.



Fig. 2. Photocurrent density as a function of potential, point by point, obtained for the electrode produced thermally ($____$). Currents measured under illumination conditions ($____$) currents measured in the absence of light conditions.

Table 1Tests performed for inactivation of microorganisms.

light and fixed amount of catalysts. Consequently, equation (2) can be rewritten as equation (3).

$$\ln\left(\frac{N}{N_0}\right) = -k' t \tag{3}$$

The constant (k) was obtained by linear regression analysis, and linearized by the OriginPro 9.0 software (OriginLab). According to the Chick-Watson Model, the slope is negative (-k). The N/No is the ratio of surviving bacteria after the contact time t, which can be expressed by Equation (4).

$$\frac{N}{N_0} = e^{-k t} \tag{4}$$

Thus, the inactivation percentage was calculated using equation (5):

Inactivation percentage (%) =
$$\left(1 - \frac{N}{N_0}\right) \times 100$$
 (5)

| Tests | Experimental conditions | | |
|---|--|--|--|
| Test 1 – photolysis Test 2 – electric potential Test 2 – electrochemically assisted photocatalyst | UV-A radiation TiO ₂ /electric potential, 1.5 V Electrode/UV-A/electric potential, 1.5 V | | |
| | Electrode 1: Ti/TiO ₂ $-$ 10 min of calcination Electrode 2: Ti/TiO ₂ $-$ 5 min of calcination Electrode 3: Ti/TiO ₂ $-$ 15 min of calcination Electrode 4: Ag-Ti/TiO ₂ $-$ 10 min of calcination | | |

(nutrient broth - NB, Himedia, for *E. coli* and brain heart infusion - BHI, Criterion, for *S. aureus*). The cultures were incubated for 24 h in orbital shaking at 180 rpm at 35 °C. Subsequently, 5 mL of each inoculum was centrifuged for 10 min at 80 rpm (0.500 g) and the precipitate was washed with 5 mL of solution containing 0.08 N Na₂SO₄.

Aliquots were collected at 0, 10, 20, 30, 40, 50 and 60 min, which were then diluted in 0.85 (m/v) saline solution and subsequently plated by the Pour Plate method. The dilution factor for microbial count was established at 10^{-4} in *E. coli* and 10^{-5} in *S. aureus* cultures. The plates were kept at 35 °C for 24 h and growth was quantified in CFU mL⁻¹. All experiments were performed in triplicate.

2.7. Efficiency of inactivation level – kinetic study

The classic microorganism inactivation model is described by Chick-Watson. According to this model the removal rate is directly proportional to the concentration of bacteria as a first-order reaction (Equation (2)).

$$\ln\left(\frac{N}{N_0}\right) = -k[c]^n t \tag{2}$$

where N is the colony count of the UV irradiated sample and N_0 is the colony count of the sample before UV irradiation, k is the microorganism inactivation kinetic constant, c is the concentration of the disinfecting agent at time t and n the reaction order.

The concentration of the disinfecting agent can be considered constant though time in photocatalytic processes using artificial

2.8. Experimental design

The experimental approach used factorial design aiming at enhanced inactivation efficiency. The microbial inactivation factors were planned using the software Minitab 17 1.0 (Minitab Inc.). The 3 independent variables were: microorganisms (A), treatments type (B) and contact time (C) according to Table 2. The inactivation efficiency of microorganisms (%) was selected as the response output (dependent variable).

The regression curve was rated by their coefficient of determination (R^2 , R^2 adj). Statistical significance was verified by ANOVA calculation through F-value and p-value. Model parameters were selected based on their probability values insertion within 95% confidence levels. Hence, it was evaluated the effects of each of the factors and their interaction on the overall efficiency of the photolysis, electric potential and electrochemically assisted photocatalytic with different electrodes.

3. Results and discussion

3.1. Electrode photoactivity

The electrode photoactivity was determined according to photocurrent, and measured in light and dark conditions. The Ti/TiO₂ electrode without irradiation presented no current up to the 1.5 V potential, thus guaranteeing the catalyst effect (Fig. 2). Electrochemical reactions (photoactivity) are added to the photocurrent above 1.5 V. An increase in current occurred when the system was irradiated, hence indicating presence of photoactivity within the electrodes. Any potential higher than 1.5 V can change the

| Tab | le 2 |
|-----|------|
|-----|------|

| | | C .1 C . | | C E 1 | 1.0 | | | • • |
|-------------|-------------|-------------|--------------|---------------|--------|------------------|---------------|--------------|
| Independent | variables o | of the fact | orial design | i for E. coli | and S. | <i>aureus</i> in | iactivation e | experiments. |

| Independent variable | Factor | Range | | | | | | |
|----------------------|--------|------------|--------------------|-------------|-------------|-------------|-------------|----|
| Microorganisms | А | E. coli | S. aureus | | | | | |
| Treatments | В | Photolysis | Electric potential | Electrode 1 | Electrode 2 | Electrode 3 | Electrode 4 | |
| Time (min) | С | 0 | 10 | 20 | 30 | 40 | 50 | 60 |

electrode properties due to the deposition of undesirable materials, leading to losses in its photochemical capacity.

Thus, the electronic drainage at 1.5 V does not produce any changes in the working electrode, which in turn allows it to be used for several treatments. The Ag-doped electrode similarly presents no release of Ag ions in the solution due to a practically null electrolytic current. It was observed point by point photocurrent approach determined that the electrode was photoactive. Therefore, the estabilished electric potential (1.5 V) was set on further electrochemically assisted photocatalysis experiments.

3.2. Bacterial inactivation

The regression parameters for each treatment time according to using the Chick-Watson model is shown in Table 3. The k parameter is defined by min^{-1} , due to its first order relation to microbial concentration.

According to Table 3, the results follow the Chick-Watson model. The N/N0 and inactivation efficiency values for *E. coli* and *S. aureus* were calculated and plotted in Figs. 3 and 4, respectively.

Treatments using only photolysis and electric potential show that UV-A light applied separately to an electric potential result in negligible inactivation of the *E. coli* and *S. aureus* (Fig. 4a and b and Fig. 6). The UV-A lamps used in the experiment had low power (8 W) and low energy irradiation, thus unable to damage to microbial cells. UV lamps that emit radiation at 254 nm (UV-C) are the ones with the highest germicidal effect (Adhikari et al., 2015b). The sole application of electric potential on the electrodes also did not inactivate bacterial cells due to the low voltage (1.5 V) and current.

On the other hand, Fig. 3a and b, Figs. 4 and 5 show that significant bacterial inactivation occurred when electrical potential was applied to Ti/TiO_2 electrodes combined with UV-A light. The treatment with Ag-doped electrode 4 yielded higher inactivation for both microorganisms. It was achieved complete inactivation after 10 and 60 min for *E. coli* and *S. aureus*, respectively. Electrode 1 had a 46.23% inactivation rate of *E. coli*, compared to the 16.23% inactivation of *S. aureus* after 60 min. Electrode 2 showed 39.12% inactivation of *E. coli* and 26.15% of *S. aureus*, while electrode 3 showed 52.74% and 44.09% inactivation of *E. coli* and *S. aureus* after 60 min, respectively.

Certainly, TiO₂ can be found in three polymorphic phases: anatase, rutile and brookite (Yuangpho et al., 2015; Wang et al., 2015b). From the catalytic point of view, anatase and rutile are the most studied forms. However, Lopes et al. (2012) produced Ti/ TiO₂ thermal electrodes following the same protocols applied by Bidoia's research group and found out by XRD that their thermally produced TiO_2 film was in rutile form. The anatase form was not identified in the electrodes used. Thus, the electrodes produced thermally according to the same methodology used by Lopes et al. (2012), at a temperature of 750 °C, were also in rutile form. Li et al. (2009) and Hanaor and Sorrell (2011) reported that the anatase phase is formed from lower temperatures (about 400 °C) of calcination, whereas at 600 °C, the anatase-rutile phase transformation begins to emerge (Yuangpho et al., 2015).

The TiO₂ semiconductor is characterized by valence bands and conduction bands. The region between bands is called band-gap. When a semiconductor is irradiated with ultraviolet radiation at energy values greater than its band-gap energy, electrons are generated in the conduction band and gaps in the valence band, thus creating the electron-hole pair (e-/h⁺) (Chong et al., 2010; Vijay et al., 2013; Chaplin, 2014).

The anatase phase is well known for its photocatalytic activity and its band-gap of 3.2 eV is greater than in rutile forms (3.0 eV), thus lowering the probability of the recombination between $e^{/h^+}$ (Scanlon et al., 2013). However, according to Molina et al. (2014), rutile displays photoactivity depending on the preparation procedures, the precursors or the organic compounds used as reducing agent.

As seen in Figs. 3 and 4, the thermally produced electrodes presented photocatalytic activity for inactivation of both *E. coli* and *S. aureus*, which contradicts previous studies performed by Ding et al. (2000). In their study, pure rutile did not present photocatalytic activity, hence relying on the anatase phase for microorganism inactivation.

Molina et al. (2014) carried out a study producing glassimmobilized TiO_2 thermal films for inactivation of *E. coli* ATCC 25922, with UV light at 364 nm. When XRD was performed, the authors confirmed the presence of rutile phase in their film. They also confirmed the photocatalytic effect of the rutile phase for *E. coli* inactivation after 60–90 min.

Liu et al. (2003) produced TiO₂ film in the rutile phase at a calcination temperature of 600 °C using photo-assisted sol-gel method. This method, compared to the traditional sol-gel method, accelerated the transformation of the anatase to the rutile phase. Rutile particles showed remarkable photocatalytic activity towards the decomposition of organic pollutants under ultraviolet irradiation. Furthermore, Butterfield et al. (1997) observed that the electrical potential coupled to a photocatalytic system subjected to UV irradiation caused an increase in the conductivity of titanium electrodes covered with TiO₂. Krýsa et al. (2007) have pointed out

Table 3

Disinfection constant and R-squared values obtained by the Chick-Watson Model.

| Treatment | Escherichia coli | | | Staphylococcus aureus | | |
|--|--------------------------------------|---------------------|----------------|---------------------------------------|----------------|----------------|
| | Inactivation efficiency in 60 min (% | ($k (min^{-1})$) | R ² | Inactivation efficiency in 60 min (%) | $k (min^{-1})$ | R ² |
| Photolysis | 18.65 | -0.0034 | 0.751 | 13.62 | -0.0024 | 0.754 |
| Electric potential | 12.79 | -0.0022 | 0.961 | 16.68 | -0.0030 | 0.9678 |
| Electrochemically assisted photocatalysis Electrod | e 1 47.48 | -0.0109 | 0.986 | 16.23 | -0.0029 | 0.953 |
| Electrod | 2 39.12 | -0.0082 | 0.983 | 26.15 | -0.0050 | 0.957 |
| Electrod | 2 3 52.74 | -0.0124 | 0.985 | 44.09 | -0.0096 | 0.994 |
| Electrod | e 4 100 | -0.7106 | 1 | 100 | -0.1272 | 0.608 |



Fig. 3. Inactivation of *E. coli* (a) and *S. aureus* (b) versus to treatment time. Experimental conditions (____): Photolysis (___); Electric potential (___, __); Electrochemically assisted photocatalysis with electrode 2 (____); Electrochemically assisted photocatalysis with electrode 3 (____); Electrochemically assisted photocatalysis with electrode 4.



Fig. 4. Efficiency of bacterial inactivation after 60 min of photolysis, electrical potential and electrochemically assisted photocatalysis with different electrodes, where (): *E. coli* (); *S. aureus*.

that TiO_2 layers thermally grown on a titanium substrate have high electrical conductivity, thus making them with better photocatalytic efficiency together with an electrical potential.

The applied potential in electrochemically assisted photocatalysis establishes a potential gradient into TiO_2 films, favoring the separation of generated charges (e-/h⁺) and hence inhibiting the recombination process, which is the main limiting factor of quantum efficiency. This is due to the ratio of the number of electrons formed and the number of photons incident on the photocatalyst (Yang et al., 2005). As a result, electron drainage occurs, as well as an increase in the current density, both due to the migration of photogenerated electrons from the working electrode to the counter electrode (Quan et al., 2004). The concentration gradient favors the diffusion of species in solution to the surface of the working electrode (TiO₂) (Byrne et al., 2002). The decreasing rate of recombination of e/h⁺ pair causes an increase in the availability of more gaps and consequent formation of •OH (Balasubramanian et al., 2004). This process ultimately leads to the increasing inactivation rate of microorganisms.

Harper et al. (2001) and Christensen et al. (2003) compared the efficiency of thermal and sol-gel electrodes for inactivation of *E. coli*. Both studies verified that the most efficient microorganism inactivation scenarios were promoted by thermal electrodes, due to thermal electrodes optimization effects caused by the electric potential, i.e. the decrease in e^{-/h^+} recombination rates. In the sol-gel electrode, the particles are smaller and the electric potential may not be sufficient to decrease the e^{-/h^+} recombination rates.

In addition to the electrical potential associated with Ti/TiO_2 electrodes, the Ag dopant is also important. The modification of the TiO_2 surface with Ag nanoparticles provides an increase in the photocatalytic activity. The proposed effect of Ag⁺ towards adsorption at catalytic surfaces is described by Equation (6). Under the presence of oxygen, the photocatalytic efficiency is even greater. According to Litter (1999) and Kumar et al. (2015) the Ag⁰ deposited on the TiO_2 captures the electrons (used as surface traps) decreasing the recombination rates between the e/h^+ pair. This process increases the number of "active sites" for adsorption of •OH, which consequently contributes to increase the oxidation efficiency of microorganisms (Parr and Kim, 2016; Ding et al., 2010). This is the reasoning for Ti/TiO_2 electrodes doped with Ag to be more efficient than non-doped thermal electrodes, as shown in Fig. 6 and 6 plots.

$$Ag^{+}_{ads} + e^{-}bc Ag_{ads}$$
(6)

Principal effects and interaction plots between treatment variables during inactivation of *E. coli* and *S. aureus* are shown in Figs. 5 and 6. The data output from electrochemically assisted photocatalytic process showed greater *E. coli* (gram-negative) inactivation than *S. aureus* (gram-positive), as evidenced by the bacterial inactivation rate constants (k) calculated by Chick-Watson's law. The k parameter was higher for *E. coli* than *S. aureus* in all treatments (Table 3).

Such inactivation results are explained by variations in cell wall thickness between the two microorganisms. In gram-negative bacteria *E. coli*, the plasma membrane is surrounded by a cell wall composed of peptideoglycan, followed by an outer membrane of proteins, lipids and lipopolysaccharides. Its structure is often thinner than the cell wall of gram-positive bacteria *S. aureus* (Feng



Fig. 5. Interaction plot between microorganisms, different treatments (photolysis, electric potential and electrochemically assisted photocatalysis with different electrodes) and the contact time for inactivation of *E. coli* and *S. aureus*. Where A: Microorganisms; B: Different treatments; C: contact time (min).

et al., 2000; Jung et al., 2008; Nakano et al., 2013). The microbial inactivation pattern found in this study was consistent with the ones found by other authors, such as Feng et al. (2000); Nakano et al. (2013); Pal et al. (2007) and Amininezhad et al. (2015).

According to Rincón and Pulgarin (2004), certain bacteria, especially gram-positive, have a very viscous capsule, which functions as a protection to the cellular wall. This external structure can be formed in response to environmental stimuli, exerting great influence on survival during the photocatalytic treatment with electrical potential. This additional bacterial resource may have been another factor responsible for an increased resistance of gram-positive *S. aureus* over gram-negative *E. coli* cells.

The electroassisted photocatalytic treatment combined processes related to bacterial inactivation, such as the direct absorption of UV radiation by the cells and the cellular destruction by •OH, with a powerful oxidizing agent generated by the degradation on the photocatalyst surface. Changes in the cell wall were therefore expected (Cho et al., 2011; Gusmão et al., 2009; Foster et al., 2011; Adhikari et al., 2015a; Grosser and Richardson, 2015). According to Matsunaga et al. (1985), the microbial death is due to the oxidation of coenzyme A (CoA) by the positively charged gaps, leading to the formation of dimeric CoA. In this study, the oxidation of CoA may have caused an inhibition of the cellular respiration, in the metabolic activity and consequent inactivation of the cells.

In this work, bacterial exposure to Ti/TiO_2 electrodes doped with Ag lead to many of those processes involving bacterial inactivation, mostly caused by electrochemically assisted photocatalysis. The inactivation by Ag nanoparticles also yielded higher inactivation rates in comparison to other electrodes (Figs. 5 and 6). Studies by Sondi and Salopek-Sondi (2004), Pal et al. (2007) and Adhikari et al.

(2015a) showed that, similarly to •OH, the Ag damages cell membrane constituents. The Ag promotes structural changes and ruptures in the cell wall, thus interfering with the intracellular metabolic activity, which may be responsible to the cellular death results.

Liu et al. (2008) synthesized thin Ag/TiO₂ films and evidenced their bactericidal effects in *E. coli*, which was mostly related to ruptures in cell membranes. Five years later, Brugnera et al. (2013) determined that Ag nanoparticles exhibited enhanced antibacterial properties and biocompatibility among all antibacterial materials.

The disinfection of drinking water is well-known for reducing waterborne disease, as it is the greatest achievement of the 20th century on sanitation (Cutter and Miller, 2005; Plewa et al., 2017). The on-going search for alternative disinfection methods is therefore a relevant challenge (Meireles et al., 2016). The development of novel methods for water and wastewater recycling are worldwide priorities since its implementation for the disinfection of drinking water are crucial for public health (Plewa and Richardson, 2017).

3.3. Electric energy consumption

Electroassisted photocatalysis can be commercially applicable towards water treatment, as long as the studies focus photoefficiency improvements (Parent et al., 1996). As seen in Table 4, the inactivation of 100% of *E. coli* with the Ag-doped electrode required 10 min of treatment and 286.86 mWs cm⁻² of ultraviolet radiation. The energy consumption of this process from the UV-A lamp was 0.018 kWh m⁻³. The inactivation of 100% *S. aureus* with the Ag-doped electrode took 60 min and 1721.16 mWs cm⁻² radiation dose with 0.108 kWh m⁻³ energy consumption from UV- A.



Fig. 6. Principal effects plot between microorganisms, different treatments (photolysis, electric potential and electrochemically assisted photocatalysis with different electrodes) and the contact time for inactivation of *E. coli* and *S. aureus*. Where A: Microorganisms; B: Different treatments; C: contact time (min).

| Energy consumption of the lamps used in the electrochemically assisted photocatalyst reactor in 60 min of treatment with a dose of 1721.16 mWs $\rm cm^{-2}$. | Table 4 | |
|--|--|--------------------------------|
| | Energy consumption of the lamps used in the electrochemically assisted photocatalyst reactor in 60 min of treatment with a dose of | 1721.16 mWs cm ⁻² . |

| Treatments E. coli | | S. aureus | | | |
|---|---|--|---|---|--|
| Power consumption of the lamps (kWh m ⁻³) | Energy consumption of applied electric potential (kWh m ⁻³) | Total consumption (kWh m ⁻³) | Power consumption of the lamps (kWh m ⁻³) | Energy consumption of applied electric potential (kWh m ⁻³) | Total consumption (kWh m ⁻³) |
| Electrode 1 0.108 Electrode 2 0.108 Electrode 3 0.108 Electrode 4 0.018 ^a | $\begin{array}{l} 10.8 \times 10^{-3} \\ 10.8 \times 10^{-3} \\ 10.8 \times 10^{-3} \\ 1.8 \times 10^{-3a} \end{array}$ | 0.119 0.119 0.119 0.019 | 0.108 0.108 0.108 0.108 | $\begin{array}{l} 10.8 \times 10^{-3} \\ 10.8 \times 10^{-3} \\ 10.8 \times 10^{-3} \\ 10.8 \times 10^{-3} \end{array}$ | 0.119 0.119 0.119 0.119 0.119 |

^a Energy consumption in 10 min of treatment considering a dose of 286.86 mWs cm⁻².

The same radiation dose (1721.16 mWs cm⁻²) was used to all other electrodes, but neither reached 100% inactivation values for both microorganisms after 60 min of treatment.

The energy consumption related to the electric potential during 10 min of treatment was 1.8×10^{-3} kWhh m⁻³ and for the whole 60 min treatment was 10.8×10^{-3} kWhh m⁻³. Therefore, the total electric energy consumption of lamps and electrical potential during treatment was 0.019 kWh m⁻³ for 10 min and 0.119 kWh m⁻³ for 60 min.

The total energy consumption achieved in this work to apply electrochemically assisted photocatalysis in 60 min (0.119 kWh m⁻³) is priced at 1.49 US cents (EIA, 2016), which is considered reasonably low for a water treatment procedure.

Most of this energy consumption is due to low-power and lowirradiation from UV-A black light bulbs. UV-A lamps do not harm humans, unlike UV-C or other high-output bulbs. The use of these UV-A lamps is interesting from operational standpoint, as they do not cause burns or skin cancer. Moreover, the titanium used in the electrodes are extremely resistant to corrosion, improving the viability of electrochemically assisted photocatalytic procedures.

4. Conclusions

The electrochemically assisted photocatalytic activity was shown to be highly effective for treating water contaminated with gram-negative and gram-positive bacteria in comparison to photolysis. The thermally produced electrodes in a specific temperature, in rutile form, showed remarkable photocatalytic activity and were successfully used as photocatalysts for microbial inactivation using an electron drainage potential.

Among the electrodes tested in all electrochemically assisted photocatalysis, the most efficient inactivation occurred with the Ti/

TiO₂ thermal electrode doped with Ag combining drainage electron from the applied potential plus the electron acceptor caused by the Ag itself. The inactivation was greater with gram-negative *E. coli* instead of gram-positive *S. aureus*.

The treatment also required low electrical power after 60 min due to the use of UV-A lamps, which provides much safer handling protocols compared to UV-C lamps, besides lower operation costs in terms of electric power consumption. In addition, the working electrode has a long lifespan. Thus, electrochemically assisted heterogeneous photocatalysis using silver-doped Ti/TiO₂ electrodes can be considered a highly promising and commercially viable alternative for the treatment of water contaminated with microorganisms.

This disinfection of water and development of new methods for the disinfection of drinking water are crucial for public health.

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Appendix A. Supplementary data

Supplementary data related to this article can be found at https://doi.org/10.1016/j.jenvman.2017.09.006.

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