

1 **Association of zein nanoparticles with botanical compounds for effective pest**
2 **control systems**

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4 **Short running title: Botanical compound-loaded zein nanoparticles: a promising**
5 **system aiming pest control**

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Abstract

BACKGROUND: Botanical compounds from plant species are known to have pesticidal activity and have been used in integrated pest management programs. The varied spectrum of pesticidal action of these compounds can also avoid selection of resistance in pest populations. In this study, mixtures of the botanical compounds geraniol, eugenol and cinnamaldehyde were encapsulated in zein nanoparticles to improve their stability and efficiency. Biological effects of the nano-scale formulations of the botanical compounds were evaluated against two agricultural pests - the two-spotted spider mite (*Tetranychus urticae*) and the soybean looper (*Chrysodeixis includes*). **RESULTS:** The formulations were stable over time (120 days) with a high encapsulation efficiency (>90%). Nanoencapsulation also provided protection against degradation of the compounds during storage and led to a decrease in toxicity to non-target organisms. The release of the compounds (especially eugenol and cinnamaldehyde) from the nanoparticles was directly influenced by temperature, and the main mechanism of release through diffusion-based process. Nanoencapsulated compounds also showed superior efficiency than the emulsified compounds in terms of repellency and insecticidal activity. **CONCLUSION:** The findings of this study indicate that the convergence of botanical compounds with nano-scale formulation has the promise to improve efficacy for their sustainable use in integrated pest management in agriculture.

Keywords: botanical pesticides, nano-scale, environmentally friendly formulations

74 1. Introduction

75 There have been enormous scientific and technological changes (use of pesticides
76 and fertilizers, mechanization of production, **transgenesis**) in agriculture since the 40's.
77 As a result, food production has increased significantly ¹. Despite the great advancements,
78 such practices have also brought several health and environmental impacts (soil and water
79 contamination, toxicity to non-target organisms) ². In this context, there is an increasing
80 emphasis on development of practices, methods and technologies that can contribute to
81 increasing safety and sustainability of agriculture ³.

82 Botanical pesticides have been sought as an important tool for sustainable
83 agriculture. These compounds are produced in the secondary metabolism of various plant
84 species to form a defense against pests and diseases ⁴. Being degradable, these compounds
85 generally present minimal adverse effects on human and animal health and the
86 environment. With few exceptions, they can therefore be considered safer than most
87 synthetic pesticides ⁵.

88 Geraniol is a compound derived from different essential oils (citronella, palmrose,
89 among others), and is classified as an acyclic alcohol having a vapor pressure of 2.21×10^{-2} mm Hg at 25°C, water solubility of 100 mg.L⁻¹ and boiling point at 230° C. Several
90 applications of geraniol are reported in literature, including in the control of agricultural
91 pests ⁶⁻⁹. Eugenol, the main component of clove essential oil, belongs to the chemical
92 class of phenylpropanoids. It has a vapor pressure of 2.89×10^{-2} mm Hg at 25°C, solubility
93 in water of 1.460 mg.L⁻¹ and a boiling point of 225°C. Due to its anesthetic properties, it
94 is used as pain relief agent in dental applications¹⁰. In addition to bactericidal and
95 antifungal properties ¹¹, it is also known for pest control properties ^{12,13}. Cinnamaldehyde
96 (vapor pressure 3.2×10^{-2} mm Hg at 25°C, solubility in water of 1,420 mg.L⁻¹ and boiling
97 point of 246°C) is also a phenylpropanoid found in essential oil of cinnamon bark
98 (*Cinnamomum zeylanicum* J.Presl) and other *Cinnamomum* spp. It is known for
99 antifungal as well as pest control properties ¹⁴⁻¹⁶.

101 The use of a combination of different compounds from plants is an important
102 strategy to enhance biological activity and to develop novel formulations that have a
103 mixture of active principles that are not normally present together in one plant ¹⁷. For
104 example, a mixture of geraniol and cinnamaldehyde in equivalent proportions does not
105 occur naturally, since the compounds originate from different plants. Therefore, this
106 strategy may also contribute towards increased effectiveness (because of a combined
107 efficacy underpinned by different modes of action) and retarding the development of

108 resistance by pests ¹⁸. Despite such potentials of natural compounds, the existing
109 agricultural applications also face certain limitations. Natural substances are generally
110 sensitive to degradation by light, humidity and temperature in the field ¹⁹.

111 In this context, innovative formulations based on nanoencapsulation have been
112 shown to improve stability and efficacy of natural compounds ²⁰. Protection against
113 premature degradation coupled with sustained release and increased solubility of active
114 compounds have been reported in numerous studies for nanoencapsuled botanical
115 pesticides ^{9,21-23}. Such nanostructured systems can be produced from different matrices
116 (natural and synthetic). A particular example is zein, which belongs to a class of
117 prolamins, and is the main storage protein of maize that makes up around 50% of the total
118 protein content. Among its desirable properties are: high coating capacity,
119 biodegradability and biocompatibility. Zein is extensively investigated in the production
120 of biodegradable nanoparticles, including encapsulates of botanical pesticides ²⁴.

121 In view of this context, the objective of the present study was to develop
122 nanopesticide formulations that contain mixtures of botanical compounds that are known
123 to be active against insect pests (geraniol+eugenol and geraniol+cinnamaldehyde). The
124 study used zein as the biodegradable matrix for nano-scale encapsulation. The
125 nanostructured carrier systems were prepared and characterized for stability and rate of
126 release of the compounds. Biological efficacy was evaluated in terms of cytotoxicity
127 against two cell lines, and two species of agricultural pests: the two-spotted spider mite
128 (*Tetranychus urticae* Koch) and the soybean looper [*Chrysodeixis includes* (Walker,
129 1858)]. The approach adopted in this study is likely to contribute towards the
130 development of safe and sustainable pest control systems for use in agriculture.

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142 **2. Materials and Methods**

143 **2.1. Materials**

144 Geraniol (GRL), Eugenol (EGL), Trans-cinnamaldehyde (CND), Zein and
145 Pluronic F-68 were obtained from Sigma-Aldrich, EUA. Ethanol was purchased from
146 Labsynth (Brazil). Acetonitrile (Grade HPLC) was obtained from J.T. Baker (USA).
147 Other reagents (analytical or higher) were purchased from local vendors.

148 **2.2. Preparation of Zein nanoparticles**

149 Zein nanoparticles containing the actives were prepared according to the anti-
150 solvent precipitation method described by (Hu e McClements, 2014) ²⁵, with certain
151 modifications. Initially, a solution of zein (2% w/v) was prepared in hydroethanolic
152 solution (85% v/v) and stirred overnight. The zein solution was then centrifuged for 30
153 minutes at 4,500 rpm and subsequently subjected to a heat treatment (15 minutes at 75°C).
154 Finally, the solution was filtered through syringe filters (0.45 µm - Milipore). To prepare
155 the particles containing the active compounds 600 mg of each active were added to 10
156 mL of the zein solution. Formulations containing the geraniol mixture with eugenol
157 (NP_GRL + EGL) and the mixture of geraniol and cinnamaldehyde (NP_GRL + CND),
158 both formulations containing 2% (w/v) of each active compound, were prepared. An
159 aqueous solution of 1% Pluronic F68 (w/v) (Poly(ethylene glycol)-block-poly(propylene
160 glycol)-block-poly(ethylene glycol) surfactant was prepared and pH was adjusted to 4.
161 With the aid of a syringe the zein solution (10 mL) was rapidly injected into the solution
162 of Pluronic F68 under magnetic stirring. The colloidal dispersion was stirred for ethanol
163 evaporation (room temperature). Control nanoparticles were also prepared without the
164 addition of the active compounds in zein solution. The loss of active compound(s) during
165 the preparation process was investigated by high performance liquid chromatography
166 (HPLC).

167 **2.3. Physicochemical characterization of zein nanoparticles**

168 Analysis of size distribution and polydispersity was performed using the dynamic
169 light scattering (DLS) technique. The zeta potential was determined by the
170 microelectrophoresis method. For both techniques a ZetaSizer Nano ZS90 system
171 (Malvern Instruments, UK) was used at a fixed angle of 90° and 25°C, the samples were
172 diluted about 100 to 500 times. In addition, the nanoparticle tracking analysis technique
173 was used to measure size distribution and nanoparticle concentration. For this, NanoSight

174 LM 10 cell (green laser, 532 nm) and a sCMOS camera controlled by NanoSight v. 3.1
175 were used. The results were expressed as the average of three determinations. The
176 formulations were stored in amber bottles at room temperature and their stability was
177 investigated as a function of time (after 0, 15, 30, 60, 90 and 120 days).

178 The morphology of the nanoparticles was investigated by atomic force microscopy
179 (AFM). For this, the nanoparticles were diluted (2,000 times) and deposited on silicon
180 plates that were dried in a desiccator. The analysis were performed using an atomic force
181 microscope Easy Scan 2 Basic BT02217 (Nanosurf, Switzerland), operated in contactless
182 mode with the TapAl-G (BudgetSensors, Bulgaria) cantilevers and a scan rate of 90 Hz.
183 The images (256x256 pixels, TIFF format) were captured in time mode and analyzed
184 using Gwyddion software.

185 **2.4. Quantification of botanical compounds and determination of encapsulation** 186 **efficiency (EE)**

187 The quantification of the botanical compounds was carried out using high
188 performance liquid chromatography (HPLC). For geraniol, a Phenomenex Gemini C18
189 reverse phase column (150 mm×4.6 mm, 5.0 μm) maintained at 30°C was used, the
190 mobile phase was composed of acetonitrile: water (60:40, v/v) and flow rate of 1 mL.min⁻¹.
191 The injection volume was 100 μL and the wavelength of the detector was set at 210
192 nm. For eugenol and cinnamaldehyde compounds, Phenomenex Kinetex C18 reverse
193 phase column (150 mm×4.6 mm, 3.0 μm) was used. For eugenol, the mobile phase was
194 composed of acetonitrile: water (50:50 v/v), whereas for cinnamaldehyde it was
195 methanol: water (65:35 v: v), at a flow rate of 1mL.min⁻¹ for both. The wavelength for
196 the detection of the compounds was set at 210 nm and the injection volume was 100 μL.

197 It is noteworthy that all chromatographic analysis were performed in a UltiMate
198 3000 system (Thermo Scientific), operated by Chromeleon 7.2 software, which was used
199 for the acquisition and analysis of the chromatograms. All analytical curves showed
200 correlation coefficients (r^2) higher than 0.99.

201 The ultrafiltration/centrifugation method was used to quantify botanical compounds
202 encapsulated in zein nanoparticles⁹. The technique is based on the use of Microcon 10
203 kDa regenerated cellulose ultrafilters (Millipore), which allows the passage of only the
204 non-encapsulated substances. Thus, the difference between the quantity initially added
205 and the quantity not encapsulated gives the encapsulation efficiency (EE). It should be

206 noted that the total amounts of botanical compounds (100%) present in the formulations
207 were calculated considering the total amount added minus any losses during the
208 preparation process.

209 **2.5. Release assays and assessment of release mechanisms**

210 The *in vitro* release assay was performed according to Chang et al. (2017)²⁶ with
211 some modifications. The nanoparticle suspension (2 mL) containing the botanicals was
212 placed in dialysis membrane bags (1 kDa exclusion pore, SpectraPore) and immersed in
213 100 mL solution of 3% Pluronic F68 (w/v). Over time, aliquots were collected and
214 subjected to HPLC quantification. The containers were kept closed to avoid losses by
215 evaporation and were only opened during sampling (in triplicate). In order to investigate
216 the influence of temperature on the release of the actives from the nanoparticles, the tests
217 were performed at three different temperatures (20, 25 and 30°C). The release data were
218 submitted to mathematical modeling using the order zero, first order, Higuchi and
219 Korsmeyer-Peppas models to investigate the mechanism of release of the active
220 substances through the nanoparticles²⁷.

221 **2.6. Cytotoxicity assays**

222 Cytotoxicity assays were conducted according to the cell viability method,
223 measured in terms of reduction of tetrazolium dye (MTT test)²⁸. For this, two cell lines
224 were used: pulmonary fibroblast permanent cell line (v79) and a fibroblast cell line (3T3).
225 Cells were maintained in continuous culture using DMEM medium and 10% fetal bovine
226 serum. A supplementation with 100 IU mL⁻¹ of penicillin and 100 µg.mL of streptomycin
227 sulphate was added and cells were maintained at pH 7.4, 37 °C, under humidified
228 atmosphere with 5% CO₂. To perform the assays, the plates containing 1x10⁴ viable cells
229 were incubated (37 °C) for 48h until semiconduction, and the cells were then exposed (for
230 24h) to the following solutions: zein nanoparticles (NP), zein nanoparticles containing
231 geraniol and eugenol (NP_GRL+EGL), geraniol and eugenol emulsified with Pluronic
232 (EM_GRL+EGL), geraniol and cinnamaldehyde emulsified with Pluronic
233 (EM_GRL+CND) and zein nanoparticles containing geraniol and cinnamaldehyde
234 (NP_GRL+CND). The absorbance was measured using a plate reader at 570 nm, and cell
235 viability was determined in triplicate and results expressed in terms of percentage means
236 and standard deviation.

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238 **2.7. Biological activity assays**

239 **2.7.1. Repellency against the two-spotted spider mite (*Tetranychus urticae*)**

240 The bioassays with *T. urticae* were carried out in the Laboratory of Acarology
241 (UNESP/FCAV, Jaboticabal Campus). First stage of the experiment was performed in a
242 greenhouse (mean temperature 25.3°C, 79.3% relative humidity). Initially, seeds of
243 *Canavalia ensiformes* (L) DC. were planted in pots of 5 L capacity, containing soil, sand
244 and bovine manure (1:1:1 w/w/w) as substrate. After germination of the seeds, only one
245 plant per pot was kept. Thirty (30) days after germination, the formulations (treatments)
246 were applied to the plants. For each treatment, three (3) plants were distributed in a
247 completely randomized design in the greenhouse. The treatments comprising 5 mg.mL⁻¹
248 of active compound were applied with manual sprayer (500 mL capacity) until complete
249 coverage of the plants. The products were carefully applied so that all top and bottom
250 surfaces of the plants were covered with the product. On average 15 mL of each
251 formulation per plant was used. After 12, 24, 72, 120 and 168 h, leaflets were removed
252 from the plants, placed in plastic trays, and sent to the laboratory. Circular leaf arenas (2.5
253 cm diameter) were removed. The leaflets were placed in Petri dishes 9.0 cm in diameter
254 x 2.0 cm in height on a layer of moist foam and hydrophilic cotton. For each treatment,
255 eight (8) arenas were used, corresponding to eight (8) repetitions. In the sequence, 10
256 adults female of the two-spotted spider mite were transferred to each arena with the aid
257 of a brush and a stereoscope microscope (Zeiss® Stemi DV4). Evaluation of the live and
258 dead mites was carried out as well as of those trapped in the glue barrier 24 hours after
259 the transfer of mites to the arenas.

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261 **2.8. Soybean looper (*Chrysodeixis includes*) assays**

262 Bioassays with *C. includes* were carried out at the Laboratory of Microbial Control
263 of Arthropods-Pest (UNESP/FCAV, Jaboticabal campus). Aliquots of 800 µL (sufficient
264 to wet the whole diet surface) of the nanoparticle formulations and of the emulsified
265 compounds (GRL, EGL and CND) were applied to the artificial diet discs (4.8 cm³), and
266 packed in clear acrylic plates (10 cm x 1.2 cm). The control diet disc was treated with the
267 same volume of sterilized water. After complete drying, ten (10) second instar larvae were
268 transferred to the plates and ten replicates were performed. The plates were incubated in
269 a BOD (biological oxygen demand) incubator at 25 ± 1 ° C and 70 ± 10% relative

270 humidity, with photoperiod of 12 hours. Larval mortality was assessed on the seventh
 271 day. In addition, sublethal effects of the formulations were evaluated by weighing the
 272 larvae 15 days after the end of the mortality evaluation. The evaluation of oviposition was
 273 performed in PVC cages.

274

275 3. Results and Discussion

276 3.1. Characterization and physicochemical stability

277 The results of characterization of nano-formulations in terms of mean diameter
 278 (MD, nm), polydispersity index (PDI), zeta potential (ZP, mV), nanoparticle
 279 concentration (CT, particles/mL) and encapsulation efficiency (EE, %) are presented in
 280 Table 1. Data on physicochemical stability are presented in Figure S1 (supplementary
 281 material).

282 **Table 1:** Characterization of zein nanoparticles containing the botanical compounds (geraniol, eugenol and
 283 cinnamaldehyde). Values are expressed as mean of three determinations.

Formulations	MD (nm)		PDI	ZP (mV)	CT (x10 ¹² particles/mL)	EE (%)
	DLS	NTA				
NP	302 ± 8	232 ± 9	0.52 ± 0.09	-15 ± 1	0.8 ± 0.1	-
NP_GRL+CND	234 ± 5	156 ± 6	0.38 ± 0.02	43 ± 2	3.3 ± 0.8	GRL 99 ± 1 CND 97 ± 2
NP_GRL+EGL	282 ± 3	160 ± 8	0.34 ± 0.05	41 ± 2	3.2 ± 0.7	GRL 99 ± 1 EGL 98 ± 1

284 MD – Mean diameter; PDI - Polydispersity index; ZP - zeta potential; CT - Nanoparticle concentration;
 285 EE - Encapsulation efficiency

286 The control nanoparticles had a larger mean diameter compared to the other
 287 formulations and a high value polydispersity index and a relatively low zeta potential
 288 (Table 1). This indicates a low stability of these formulations, which prevented the
 289 continuation of the analysis after 15 days of storage due to precipitation and phase
 290 separation, and were therefore not included in the extended stability analysis (Figure S1).
 291 According to Da Rosa et al. (2015b), the presence of active compounds in the dispersion
 292 can play a stabilizing role that can prevent aggregation and consequent increase of particle
 293 size.

294 With the two techniques used for analysis of average diameter, a significant
 295 increase was observed in the values for both formulations of nanoparticles as a function
 296 of time, especially with 120 days, indicating gradual particle aggregation (Figure S1).
 297 However, no significant changes were observed in both the polydispersity index and the
 298 nanoparticle concentration. The zeta potential decreased within 15 days but remained

299 stable until the final analysis. Furthermore, the prepared nanoparticle formulations
300 showed few changes in size distribution as a function of time (Figure 1 A and B). The
301 AFM micrographs (Figure 1 - Ab and Bb), show spherical morphology and smooth
302 surface of the nanoparticles. They also show high polydispersity of the formulations, with
303 particle size distribution between 90 and 550 nm. This corroborates with the
304 polydispersity index (Figure S1-C) that shows values higher than 0.3. This indicates good
305 physicochemical stability of the prepared nanoparticle formulations, which is extremely
306 important for commercial applications that require storage over long periods. These
307 results also corroborate a previous study by our research group ⁹ that prepared zein
308 nanoparticles for encapsulation of geraniol and R-citronellal separately. The results of
309 that study had also shown that the prepared nanoparticle formulations were stable over
310 time, with mean size of 200 nm, polydispersion index of 0.3, and zeta potential of -20
311 mV.

312 The encapsulation efficiency was also investigated in the current study (Figure 1-
313 Ac and Bc). It was observed that, as shown in the previous work by Oliveira et al., (2018)⁹,
314 the encapsulation index of the botanical compounds was high (>98%) in zein
315 nanoparticles. This is likely to be due to the strong interaction between the studied
316 compounds and the hydrophobic part of the zein. For the encapsulated compounds, it was
317 observed that there was **a** significant decrease in encapsulation efficiency only at the
318 extended storage times (90 and 120 days). This is most likely because of the loss of
319 compounds due to volatilization, and/or degradation of the particles and release of the
320 compounds. This is still a major improvement in stability as there was a much greater
321 degradation when preparations were made only by emulsifying the substances with a
322 surfactant. For encapsulated GRL and CND, 92 ± 2 and 90 ± 2 % of the active substance
323 were available after 120 days, while emulsions has 65 ± 3 and 44 ± 2 % of the active
324 substances, respectively. Similar results were obtained for encapsulated formulations
325 containing GRL and EGL, that had 94 ± 1 and 92 ± 2 % of the substances, while
326 emulsified forms had 68 ± 3 and 61 ± 4 % of the compounds respectively. It appears that
327 when the compounds are not encapsulated, they are more prone to loss due to
328 volatilization and degradation than when they are encapsulated in nanoparticles. The
329 results of this study therefore provide further evidence that encapsulation can protect
330 active ingredients against rapid volatilization and degradation ^{9,30,31}. Scremin et al.,
331 (2018)³² also observed that encapsulation of eugenol in rice-bran protein based

332 microcapsules provided protection to the active substance against degradation (around
333 30% compared to non-encapsulated compound).

334

335 [Figure 1]

336

337 The formulations containing mixture of active compounds were more stable as a
338 function of time (Table 1, Figure 1 and Figure S1). In addition, the formulations were
339 able to protect the active substance against degradation in solution. Thus, the nanoparticle
340 formulations containing mixtures of geraniol with eugenol, and geraniol with
341 cinnamaldehyde, were stable from physicochemical point of view. These findings are
342 very important for the formulations to be useable in agricultural applications since a stable
343 shelf life of active substance is essential to maintain efficacy.

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346 **3.2. *In vitro* release and release mechanism**

347 Figure 2 shows release data for the zein nanoparticle formulations containing
348 mixtures of the active substances: geraniol with eugenol and geraniol with
349 cinnamaldehyde at different temperatures (25, 30 and 35°C). Geraniol, both when
350 encapsulated with eugenol (Fig. 2-A) and encapsulated with cinnamaldehyde (Fig. 2-C),
351 exhibited the same release profile, averaging at 48 ± 3 % within 1440 minutes. In addition,
352 no differences were observed with increasing temperature. On the other hand, eugenol
353 (Fig. 2-C) showed a greater release compared to geraniol, and differences as a function
354 of temperature increases. At 1440 minutes, the release of eugenol was 55 ± 1 %, 58 ± 2
355 % and 68 ± 3 % at temperatures of 25, 30 and 35°C, respectively. The highest release was
356 observed for cinnamaldehyde, which under the same experimental time released 52 ± 2
357 %, 76 ± 1 % and 88 ± 2 % at temperatures of 25, 30 and 35°C, respectively. The increase
358 in the release of active compounds with increasing temperature reflects the differences in
359 physicochemical characteristics, such as volatility and solubility. The tendency of a
360 substances to evaporate is depicted in terms of vapor pressure, and a higher vapor pressure
361 indicates the substance to be more volatile ³³. Among the substances studied,
362 cinnamaldehyde has the highest vapor pressure (3.2×10^{-2} mm Hg at 25°C), followed by
363 eugenol (2.89×10^{-2} mm Hg at 25°C) and geraniol (2.21×10^{-2} mm Hg at 25°C). Differences
364 in the release of active eugenol and cinnamaldehyde have also been observed by Gomes

365 et al., (2011)³⁴ for poly(lactic-co-glycolic acid) (PLGA) nanoparticles containing eugenol
 366 and cinnamaldehyde. The release assays showed differences in the release profile of the
 367 substances from nanoparticles, with around 80% of cinnamaldehyde released after 5
 368 hours, compared to 45% of eugenol. According to the authors, the steric conformation of
 369 eugenol and greater lipophilicity than trans-cinnamaldehyde probably makes it more
 370 difficult for eugenol to diffuse from inside the nanoparticles to the external medium.

371 [Figure 2]

372 Mathematical models are widely used to predict the time release patterns of the
 373 encapsulated molecules to understand the mechanisms of release and assist in the design
 374 of formulations³⁵. This study used different mathematical models to evaluate the
 375 mechanism of release of the active compounds through zein nanoparticles (Table 2).

376

377 **Table 2:** Constants (*k*) and correlation coefficients (*r*²) for different mathematical models applied
 378 to evaluate the release of active compounds from zein nanoparticles at different temperatures.

379

	Mathematical model								
	Zero order		First order		Higuchi		Korsmeyer-Peppas		
	<i>k</i> (h ⁻¹)	<i>r</i> ²	<i>k</i> (h ⁻¹)	<i>r</i> ²	<i>k</i> (h ^{-1/2})	<i>r</i> ²	<i>k</i> (h ⁻¹)	<i>n</i>	<i>r</i> ²
25 °C									
NP_GRL+EGL									
GRL	0.0081	0.4594	2.82x10 ⁻⁴	0.3414	1.166	0.7123	1.372	0.3897	0.8441
EGL	0.0011	0.2751	1.61x10 ⁻⁴	0.1872	1.031	0.5185	1.695	0.2612	0.6799
NP_GRL+CND									
GRL	0.0084	0.4439	3.71x10 ⁻⁴	0.2783	1.285	0.6888	0.4261	0.5493	0.7899
CND	0.0096	0.4482	3.87x10 ⁻⁴	0.2672	1.446	0.6904	2.1551	0.5805	0.7762
30 °C									
NP_GRL+EGL									
GRL	0.0105	0.5072	3.58x10 ⁻⁴	0.3228	1.287	0.7493	0.6727	0.4045	0.8288
EGL	0.0025	0.3081	1.46x10 ⁻⁴	0.2957	1.059	0.5613	1.709	0.2107	0.7937
NP_GRL+CND									
GRL	0.0082	0.4329	3.46x10 ⁻⁴	0.2688	1.321	0.6812	0.6825	0.5184	0.7781
CND	0.3631	0.8971	5.95x10 ⁻⁴	0.7196	6.206	0.9117	2.8282	0.7321	0.9633
35 °C									
NP_GRL+EGL									
GRL	0.0103	0.4952	3.45x10 ⁻⁴	0.2502	1.308	0.7375	0.7175	0.4138	0.8187
EGL	0.0842	0.6865	1.19x10 ⁻⁴	0.7252	3.311	0.9192	1.3512	0.3183	0.9791
NP_GRL+CND									
GRL	0.0069	0.3979	2.99x10 ⁻⁴	0.2967	1.322	0.6433	1.262	0.5273	0.7825
CND	0.3821	0.8971	6.33x10 ⁻⁴	0.7441	6.474	0.9152	3.132	0.6849	0.9654

380

381 According to the data presented in Table 2, it is possible to observe that the
 382 mathematical model that best fits for all active compounds was the Korsmeyer-Peppas
 383 model. Through use of this model, it is possible to determine if the release of the active
 384 substances followed Fick's law of diffusion, or a different mechanism such as

385 swelling/relaxation phenomena (Case-II transport). It can be seen that for zein
386 nanoparticles containing geraniol and eugenol, the value of n was <0.45 , which indicates
387 that the diffusion is the main mechanism that controls release of the active substance in
388 the system. For zein nanoparticles containing geraniol and cinnamaldehyde, the value of
389 n was between 0.45 and 0.89, indicating an anomalous transport kinetics, which indicates
390 a combination of two mechanisms (diffusion and transport of Case II)²⁷. However,
391 diffusion is the main form of release in both systems, which leads to the compound
392 passing through the zein protein chain matrix to the external environment. In such a type,
393 the rate of release usually decreases with time, since more internalized compound has a
394 greater distance to cross, which requires more time. This is supported by the results shown
395 in Figure 2, which show a faster release of the active substances within the first 60
396 minutes. This is due to diffusion of the most superficial layers of the encapsulates as well
397 as any adsorbed substances on the outer surface of the nanoparticles. After this period,
398 the internalized active compounds diffuse into the nanoparticle matrix. These results also
399 corroborate previous work described in the literature. For example, diffusion has been
400 suggested as the main mechanism of release of geraniol from chitosan/gum arabic
401 nanoparticles³⁶. Campos et al., (2018)³⁷ evaluated the mechanism of release of carvacrol
402 and linalool through chitosan nanoparticles functionalized with β -cyclodextrin. The
403 authors also found that diffusion, was the main mechanism for the release of the active
404 substances, along with relaxation of the polymer chains (Case Transport II). This shows
405 that mathematical models can be important tools in the study of the release of active
406 compounds from nanoparticle based formulations.

407

408 **3.3. Cytotoxicity**

409 Toxicity tests are important in order to assess the safety of these systems for non-
410 target organisms In this study, two cell lines (V79-4 and 3T3) were used (cytotoxicity
411 assays performed to determine cell viability). Both the surfactant used (Pluronic F-68)
412 and the control nanoparticles (without addition of the active compounds) did not cause
413 any significant decrease in cell viability (Figure 3). The emulsions, as well as the active
414 compounds encapsulated in the nanoparticles, showed a decrease in cellular viability with
415 increasing concentration. According to A. Al-Tamimi et al., (2016)³⁸ essential oils and
416 their active components can have cytotoxic effects amongst other biological activities.

417 Indeed, cytotoxicity has been reported in literature for geraniol ^{39,40}, eugenol ^{41,42} and
418 cinnamaldehyde, especially in tumor cell lines.

419 In this study for 3T3 cell line (Figure 3-A) and for V79 (Figure 3-B), the
420 encapsulation of the active compounds in zein nanoparticles decreased IC₅₀ values. For
421 the 3T3 line, the emulsion containing geraniol and eugenol showed IC₅₀ (obtained
422 through the probit analysis) of 0.0362 ± 0.0012 mg.mL⁻¹, whereas the emulsion
423 containing geraniol and cinnamaldehyde showed IC₅₀ of 0.0348 ± 0.0042 mg.mL⁻¹. When
424 the compounds were encapsulated in nanoparticles, the IC₅₀ values were 0.0780 ± 0.0114
425 and 0.0661 ± 0.0135 mg.mL⁻¹, respectively. For the V79 line, the emulsion containing
426 geraniol and eugenol, showed IC₅₀ of 0.0361±0.0110 mg.mL⁻¹, while the emulsion
427 containing geraniol and cinnamaldehyde showed a value of 0.0266±0.0094 mg.mL⁻¹. For
428 3T3 cell line, the IC₅₀ values were higher for the compounds when they were in
429 encapsulated form (0.0841±0.0185 and 0.0640±0.0121 mg.mL⁻¹ respectively). This
430 indicates that encapsulation of the substances in nanoparticles not only had a protective
431 effect, but also reduced their cytotoxicity. This is likely to be due to that the compounds
432 are encapsulated in the protein matrix, which reduces the amount available freely to cause
433 immediate toxic effects. Similar results have also been observed in previous studies of
434 our research group. The encapsulation of geraniol and R-citronelal in zein nanoparticles
435 caused a decrease in cytotoxic activity ⁹. Campos and co-workers (2018)³⁵ have also
436 shown that the encapsulation of carvacrol and linalool compounds in chitosan
437 nanoparticles functionalized with β-cyclodextrin significantly increased IC₅₀ values.
438 Chen et al. (2009) ²⁵ also observed a reduction in the cytotoxic activity of eugenol when
439 encapsulated in chitosan nanoparticles. According to the authors the fibroblasts exhibited
440 >80% viability when treated with the encapsulated compound, whereas for the free
441 compound the viability values were < 20%.

442

443 [Figure 3]

444

445 3.4. Biological activity assays

446 3.4.1. Two-spotted spider mite (*Tetranychus urticae*)

447 Figure 4 shows results of the repellency assays of the formulations containing
448 blends of the botanical compounds against the two-spotted spider mite (*T. urticae*). The
449 formulations were tested at a concentration of 5 mg/mL (0.5%) of each botanical

450 repellent, based on previous work of our research group (Oliveira et al., 2018)⁹, that
451 showed no toxic effects at this concentration. From the repellency curves, an adjustment
452 was applied to the area under the curve (Fig. 4), and data are presented in Table 3.

453 The emulsions showed a significantly higher repellency against the mite than the
454 encapsulated compounds two hours after application of the products (Figure 4). However,
455 whilst repellency of the formulation decreased as a function of time, the repellent effect
456 of the encapsulated compounds increased significantly. This is likely to be due to a
457 sustained release of the encapsulated compounds, and protection of the compounds from
458 premature degradation. This is evident in the area under curve (AUC) values (Table 3).
459 Geraniol and eugenol showed an AUC of 19.9 ± 1.4 repellency x time when emulsified,
460 and 24.2 ± 1.0 when they were encapsulated. Geraniol and cinnamaldehyde showed an
461 AUC of 16.1 ± 1.1 repellency x time when emulsified, and 25.5 ± 0.9 when they were
462 encapsulated. The higher AUC of the encapsulated botanicals than the emulsified
463 compounds indicates an increase in overall effectiveness. The control, as well as the
464 nanoparticles in the absence of the botanical compounds (Figure 4 - inset) did not present
465 repellent effect, and no significant differences were found between them.

466 In a previous study, Tak e Isman, (2017)⁴³ evaluated acaricidal and repellent
467 activity of different terpenes derived from plant essential oils, in addition to the effect of
468 binary mixtures against *T. urticae*. The authors tested twice the concentration used in the
469 present study (10 mg mL^{-1}) and obtained repellency value of $66.7 \pm 6.7\%$ for trans-
470 cinnamaldehyde, $62.4 \pm 10.5\%$ for eugenol and $74.3 \pm 6.1\%$ for geraniol. Also, in the
471 binary mix effect tests, the authors studied synergistic effects between the eugenol, trans-
472 cinnamaldehyde and geraniol compounds, and reported that only vanillin had any
473 significant synergistic effect. The authors however noted a significant increase in the
474 acaricidal effects of the compound mixtures. Other studies have also described the
475 repellent activity of botanical compounds^{7,44,45}. In a previous study, our research group
476 (Oliveira et al., 2018)⁹ also observed repellent effects of geraniol and R-citronellal
477 compounds against *T. urticae* at the same concentrations (5 mg.mL^{-1}) used in the present
478 study. A repellent effect of about 60% was observed for R-citronellal and 35% for
479 geraniol, with the repellency of the encapsulated compound superior to that of the
480 emulsified compound. It needs to be emphasized that, as in the previous study, the
481 repellent activity of these compounds was evaluated under controlled conditions. Other
482 studies have carried out evaluation under semi-field conditions, where formulations were
483 applied to plants in greenhouse under the action of light, humidity and uncontrolled

484 temperatures, to demonstrate that the processes are also dependent on environmental
 485 factors. Furthermore, it has been reported that other factors, such as vapor pressure and
 486 interaction with treated surfaces, also have a significant influence on the repellent effect
 487 ⁴⁶⁻⁴⁸. In the present study, for example, the speed and degree of metabolism in addition to
 488 the penetration of the compounds into the leaf structure may have played a major role in
 489 the repellent effect, since the leaf arenas were only removed from the plants for the tests.

490 Overall the results of this study showed a promising enhancement of efficacy of the
 491 nanoparticle formulations containing the botanicals, which still had a repellent effect of
 492 ~15% after 7 days, as compared to emulsified formulations.

493

494 **Table 3:** Area values on the curve for the repellent activity assays of the formulations containing the mixture
 495 of the active compounds, as well as the respective controls. Nanoparticles of zein (NP); geraniol and
 496 eugenol emulsified with surfactant (EM_GRL+EGL); geraniol and cinnamaldehyde emulsified with
 497 surfactant (EM_GRL+CND); zein nanoparticles loaded with geraniol and eugenol (NP_GRL+EGL); zein
 498 nanoparticles loaded with geraniol and cinnamaldehyde (NP_GRL+CND). Significance level of $p < 0.05$
 499 (OneWay ANOVA) for the differences between groups, where in a * there is a significant difference in
 500 relation to the control; in b * a significant difference in relation to geraniol and eugenol emulsified and c *
 501 a significant difference in relation to geraniol and cinnamaldehyde emulsified.

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Formulation	Area under the curve (repellency x time)
Control	1.7 ± 0.3
NP	2.8 ± 0.4
EM_GRL+EGL	19.9 ± 1.4 ^a
NP_GRL+EGL	24.2 ± 1.1 ^{a,b}
EM_GRL+CND	16.2 ± 1.1 ^a
NP_GRL+CND	25.5 ± 0.9 ^{a,c}

511

512

[Figure 4]

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3.4.2. Soybean looper (*Chrysodeixis includes*)

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The effects of the formulations on the larvae were evaluated considering mortality rates as well as sublethal effects determined in terms of larval and pupal weight (Table 3). It is noteworthy that for the bioassays mortality was assessed after 7 days and the sublethal effects 15 days after the end of the mortality assessment.

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520

It was observed (Table 4) that all the treatments showed mortality rate significantly higher than the control. Except for the formulation of nanoparticles containing the mixture

521 of geraniol and eugenol, the other treatments presented mortality above 80% (index
522 recommended as satisfactory). However, when evaluated for sublethal effects, the larval
523 and pupal weights treated with the nanoparticle formulations containing the active
524 compound mixtures was significantly lower than the emulsified compounds. Except for
525 the control and for the emulsion containing geraniol and eugenol, all the other treatments
526 prevented adult oviposition. Thus, the results indicate that the effects of the nanoparticle
527 formulations are longer term, most likely due to the sustained release of the active
528 compounds. For example, nanoparticle encapsulated formulations containing geraniol
529 and eugenol caused mortality rates lower than the emulsified formulations. However, it
530 manifested not only in higher sublethal effects but also prevention of oviposition, whereas
531 adult oviposition was observed for the emulsions.

532 These results show that the nanoencapsulation improved efficacy of the botanical
533 compounds. Such improvements have also been reported by other researchers. Campos
534 et al. (2018)³⁵ studied sublethal effects of the chitosan nanoparticle formulations
535 containing the carvacrol and linalool mixture against *H. armigera*. The encapsulated
536 compounds also had a greater sublethal effect than the emulsified compounds, as
537 demonstrated in this study. A. Al-Tamimi et al., (2016)⁴⁹ evaluated the effect of
538 nanoparticles of chitosan containing botanical pesticide Ponneem[®] (neem oil and karanj
539 oil) against *H. armigera*. The formulations produced growth and developmental
540 abnormalities in *H. armigera* larvae. However, the nanoformulations showed more
541 effectiveness, and a lower concentration of 0.3% caused 9.1% of defective pupae,
542 compared to 7.8% of the free compound. The mean weight of the pupae was also
543 significantly reduced in the treatment with the nanoformulations containing the botanical
544 pesticides compared to other treatments, and for the control group.

545 The greater effects of nanoformulations under larvae development may be a result
546 of higher uptake and accumulation in the larvae after feeding. Koo et al., (2015)⁵⁰
547 investigated biomagnification of quantum dot functionalized polymer nanoparticles
548 (QD). For this, they used *Arabidopsis thaliana* (L.) Heynh. ingestion by cabbage looper
549 [*Trichoplusia ni* (Hübner)]. After feeding the larvae for 7 days, the authors observed a
550 high level of fluorescence in the tissues of the larvae fed with the leaves treated with the
551 nanoparticles compared to those fed with the control plants. This showed accumulation
552 of nanoparticles in the larvae that led to a weight reduction of about 1.5 time in
553 comparison to control.

554

555 **Table 4:** Biological effects on mortality and mass of larvae and pupae of *Chrysodeixis includes* fed
556 with artificial diets treated with emulsified and nanoencapsulated botanicals. Laboratory evaluation at $25 \pm$
557 2 °C, $70 \pm 10\%$ relative humidity and 12-hour photoperiod. Nanoparticles of zein (NP); geraniol and
558 eugenol emulsified with surfactant (EM_GRL+EGL); geraniol and cinnamaldehyde emulsified with
559 surfactant (EM_GRL+CND); zein nanoparticles loaded with geraniol and eugenol (NP_GRL+EGL); zein
560 nanoparticles loaded with geraniol and cinnamaldehyde (NP_GRL+CND). Significance level of $p < 0.05$
561 (OneWay ANOVA) for the differences between groups, where in a * there is a significant difference in
562 relation to the control; in b * a significant difference in relation to geraniol and eugenol emulsified and c *
563 a significant difference in relation to geraniol and cinnamaldehyde emulsified.
564

Formulations	Mortality (%)	Larvae mass (mg)	Pupae mass (mg)	Oviposition
Control	0	198.4 ± 4.6	212.4 ± 3.2	YES
NP	47.6 ± 3.1 ^a	172.1 ± 3.1 ^a	191.3 ± 2.1 ^a	NO
EM_GRL+EGL	81.8 ± 3.5 ^a	163.4 ± 2.3 ^a	178.4 ± 1.9 ^a	YES
NP_GRL+EGL	76.4 ± 2.2 ^{a,b}	151.1 ± 2.1 ^{a,b}	167.1 ± 2.5 ^{a,b}	NO
EM_GRL+CND	88.4 ± 1.5 ^a	160.8 ± 2.2 ^a	174.1 ± 3.3 ^a	NO
NP_GRL+CND	82.2 ± 1.9 ^{a,b}	147.8 ± 4.3 ^{a,b}	165.1 ± 1.5 ^{a,c}	NO

565 **4. Conclusions**

566 Our studies have shown that both nanoparticle formulations containing blends of
567 the botanic compounds - geraniol, eugenol and cinnamaldehyde - had physicochemical
568 properties suitable for the colloidal stability over 120 days. The encapsulation of the
569 compounds not only offered protection against degradation but also enabled a sustained
570 release of the actives over time. The nanoencapsulation also led to a decrease in IC₅₀
571 values for the cell viability indicating that the nanoparticles lowered the acute toxic effect
572 of the botanical compounds. Testing of the systems demonstrated effectiveness in the
573 control of two species of agricultural pests: the two-spotted spider mite and the soybean
574 looper. For both organisms, significant efficacy improvements were observed for the
575 nanoencapsulated formulation compared to the emulsified compounds. Thus, zein based
576 nanoparticles enabled effective encapsulation of the blends of botanicals, provided
577 protection against their rapid degradation, decreased acute toxic effect, and increased
578 longer-term effectiveness to the target organisms. The use mixtures of active compounds
579 from different plants may also aid in the prevention of resistance selection in pest species.
580 Thus, a convergence between nanotechnology based formulations and botanical control
581 agents offers a promising new approach to the sustainable management of pests in
582 agriculture and reduce negative impacts on the human health and the environment.

583

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589 **6. Author contribution**

590 JLO, EVRM and LFF performed experimental delineation for carrier systems
591 preparation and characterization. TG and RL performed the cytotoxicity assays. JFDV,
592 STS and DJA performed evaluation of biological activity against *Tetranychus urticae*.
593 KGC, JN and RAP performed the evaluation of biological activity against *Chrysodeixis*
594 *includes*. JLO led the writing of the manuscript. All authors contributed critically to the
595 drafts and gave approval for the final version.

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598 **7. Compliance with ethical standards**

599 Conflict of interest: The authors declare that they have no conflict of interest.

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