

RESEARCH ARTICLE

# Cytogenetic and Molecular Data Demonstrate that the Bryconinae (Ostariophysi, Bryconidae) Species from Southeastern Brazil Form a Phylogenetic and Phylogeographic Unit

Natália Martins Travenzoli<sup>1</sup>, Priscilla Caroline Silva<sup>1</sup>, Udson Santos<sup>1</sup>, José Cola Zanuncio<sup>1</sup>, Claudio Oliveira<sup>2</sup>, Jorge Abdala Dergam<sup>1\*</sup>

**1** Laboratório de Sistemática Molecular-Beagle, Departamento de Biologia Animal, Universidade Federal de Viçosa, CEP 36570–000, Viçosa, Minas Gerais, Brazil, **2** Instituto de Biociências, Departamento de Morfologia, Universidade Estadual Paulista (UNESP), CEP 18618–970, Botucatu, São Paulo, Brazil

\* [jdergam@gmail.com](mailto:jdergam@gmail.com)



OPEN ACCESS

**Citation:** Travenzoli NM, Silva PC, Santos U, Zanuncio JC, Oliveira C, Dergam JA (2015) Cytogenetic and Molecular Data Demonstrate that the Bryconinae (Ostariophysi, Bryconidae) Species from Southeastern Brazil Form a Phylogenetic and Phylogeographic Unit. PLoS ONE 10(9): e0137843. doi:10.1371/journal.pone.0137843

**Editor:** Jose Luis Balcazar, Catalan Institute for Water Research (ICRA), SPAIN

**Received:** March 25, 2014

**Accepted:** August 24, 2015

**Published:** September 15, 2015

**Copyright:** © 2015 Travenzoli et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

**Data Availability Statement:** All relevant data are within the paper and its Supporting Information file.

**Funding:** This work was supported by the Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES), Conselho Nacional de Pesquisa (CNPq), and Fundação de Amparo à Pesquisa do Estado de Minas Gerais (FAPEMIG).

**Competing Interests:** The authors have declared that no competing interests exist.

## Abstract

*Brycon* spp. occur in Neotropical watersheds to the west and east of the Andes, and as they are sensitive to anthropogenic changes, many these species are endangered in southeastern Brazil. Coastal rivers in southeastern Brazil are characterized by the presence of relatively few freshwater fish species and high endemism of this fauna. The objective of this study was to examine whether *Brycon* spp. occurring in the coastal basins of southeastern Brazil are monophyletic, using cytogenetic data, mitochondrial, and nuclear molecular markers. All the species showed a diploid number of 50 chromosomes, a conserved number within the subfamily Bryconinae. However, the karyotypic formulas were unique to most species, including *Brycon devillei* (26m+22sm+2st), *Brycon ferox* (26m+12sm+12st), *Brycon insignis* (22m+20sm+8st), *Brycon opalinus*, and *Brycon vermelha* (24m+20sm+6st), indicating the prevalence of pericentric and paracentric inversions in the chromosomal evolution of these species. All of them had nucleolar organizer regions in the first pair of subtelocentric chromosomes and no equilocal distribution of heterochromatin in the first pair of chromosomes of the karyotype. These two features, not seen in any other *Brycon* spp. examined to date, indicate that Bryconinae species from the Brazilian southeastern coastal basins, including the monotypic genus *Hemichilus*, are monophyletic. Also, this is the first study that reports NOR location and C-banding patterns as synapomorphies for a Neotropical fish species group. The monophyly was also supported by a phylogenetic analysis of 16S rDNA (16S), cytochrome oxidase subunit I (COI), alpha-myosin (MYH6) genes and S72 intron molecular data. Our results partially corroborate the “*Brycon acuminatus*” group proposed by Howes in 1982: our proposed clade keeps *B. devillei*, *B. ferox*, and *B. insignis*; but it also includes *B. opalinus*, *B. vermelha*, and *H. weatlandii* whereas it excludes *B. nattereri*. The phylogeographic unit formed by Bryconinae species in southeastern Brazil reflects the

long and isolated paleohydrological history of these coastal basins relative to the continental watersheds.

## Introduction

The genus *Brycon* Müller & Troschel 1844 occurs in the watersheds that drain into the Caribbean Sea and in most of the rivers of South America [1]. The main morphological characteristics that define *Brycon* spp. are: presence of three (rarely four) series of teeth on the premaxilla, larger teeth in the inner than the outer premaxillary series, and presence of a pair of dental symphysean teeth which are uncommon in other Characidae [1]. Bryconidae are migratory fish and bioindicators of high quality habitat because they preferentially occur in rivers of clean water with high oxygen levels [1–3]. In Brazil, these species are distributed in the major river systems and are sensitive to anthropogenic changes [1, 4]. The coastal basins of southeastern Brazil are small to medium-sized watersheds characterized by the occurrence of relatively few freshwater fish species and high levels of endemism [5–7]. To the east, the coastal basins are isolated from the continental basins by the Serra do Espinhaço and Serra da Mantiqueira reliefs, the two major barriers that also represent the distribution ranges of many coastal fish populations [8, 9]. The paleohydrological history of these watersheds has been mainly influenced by local geomorphological processes and eustatic changes in sea level, which account for vicariant events and geodispersal processes affecting the freshwater fish faunas [8–10].

To date, no phylogenetic studies have encompassed all Bryconinae from the coastal basins of southeastern Brazil; previous studies have included *Brycon ferox*, *Brycon opalinus*, *Brycon insignis*, *Brycon vermelha* and *Henochilus wheatlandii* [11–14]. Molecular phylogenies based on mitochondrial and nuclear DNA sequences have already revealed the paraphyletic condition of *Brycon*, as some coastal species of this genus are phylogenetically closely related to *H. wheatlandii* [11, 14].

A morphological group comprising bryconins from southeastern Brazil was suggested by Howes [2], who proposed that *Brycon* might be divided into five groups, two are trans-Andean and three of them are cis-Andean. The trans-Andean groups are the group *Brycon alburnus* and *Brycon guatemalensis*. The former is composed of *Brycon alburnus* and *Brycon atrocaudatus* and the latter is composed of *B. guatemalensis*, *Brycon meeki*, *Brycon oligolepis*, *Brycon striatulus*, and *Brycon rubricauda*. The cis-Andean groups are *Brycon falcatus*, *Brycon orbignyanus* and *Brycon acuminatus*. The first group is represented by *Brycon amazonicus*, *Brycon bicolor*, *Brycon cephalus*, *B. falcatus*, *Brycon hilarii*, *Brycon moorei*, *Brycon orthotaenia*, and *Brycon bahiensis* (later synonymized to *B. opalinus*); the second group is composed of *B. hilarii* and *B. orbignyanus*; finally, the third group is composed of *B. insignis* (Howes' *B. acuminatus*), *B. ferox*, *Brycon reinhardti* (later synonymized to *B. nattereri*), and *B. devillei*. The latter group included many southeastern Brazilian bryconins. Besides *B. nattereri*, that occurs in the Upper Paraná Basin, the *B. acuminatus* group species share some morphological characters such as long maxilla with many teeth, small dental, a simple color pattern characterized by humeral and caudal spot, and long and pointed snout. With the exception of the humeral spot, *B. vermelha* can be included in this group because it has all the other characters [4]. Although Howes [2] did not indicate that "*Brycon acuminatus*" is monophyletic, Lima and Castro [4] suggested that *B. vermelha* is a member of that group, and that the *Brycon* that occur in the southeastern Brazil might be a monophyletic group based on morphological characters.

Few studies on Neotropical freshwater fish species have been conducted to test phylogeographic hypotheses with cytogenetic marks [10, 13, 15, 16]. Most cytogenetic studies on Neotropical fishes aimed detecting cryptic species and chromosomal evolution in large taxonomic groups. These studies have routinely applied two classic chromosome banding techniques (Ag-NOR and C-banding) because they are cost-effective for characterization of numerical and structural chromosomal alterations. The Ag-NORs technique allows to identify nucleolus organizer regions (NORs) active on the last cell interphase and involves silver nitrate precipitation on proteins involved in the transcription of rDNA cistrons with 18S rDNA [17]. The C-banding technique allows to identify constitutive heterochromatin regions and it involves differential degradation of euchromatic and heterochromatic chromosomal regions by alternating treatments with acids and bases [18]. Together, these techniques have successfully contributed to our knowledge of the chromosomal evolution of Neotropical freshwater fishes [16, 19, 20, 21, 22, 23, 24].

Cytogenetic studies on Brazilian coastal species of *Brycon* have been restricted to Paraíba do Sul River basin populations of *Brycon insignis* [22]. In continental watersheds, cytogenetic studies have been carried out in *Brycon* spp. from the Amazon River Basin (*B. cephalus*), Paraguay River Basin (*Brycon hilarii*), Tocantins River Basin (*B. falcatulus*), Magdalena-Cauca River Basin (*Brycon henni*), Paraná River Basin (*B. orbignyanus*), Orinoco River Basin (*Brycon amazonicus*) and São Francisco River Basin (*B. orthotaenia*) [22–26]. A study on “*B. nattereri*” from the Paraíba do Sul Basin [22] seems to be a misplacement or misidentification, because this species is restricted to the Upper Paraná Basin. All studies have indicated a conserved diploid number of 50 chromosomes and a karyotype composed of metacentric chromosomes (characterized by the median location of the centromere and chromosome arms of similar length); submetacentric chromosomes (characterized by the displaced position of the centromere and rather unequal chromosome arms), and subtelocentric chromosomes (characterized by a centromere that is much closer to one of the chromosome telomeric regions and very unequal chromosome arms) [27]. Patterns of chromosome evolution that do not alter the diploid number are usually interpreted as the outcome of chromosome rearrangements called pericentric inversions (chromosome break/fusions that occur around the chromosome’s centromere) and paracentric inversions (chromosome break/fusions that occur within the same chromosome arm) that do not involve the centromere. Because they alter the chromosome morphology, pericentric inversions alter the chromosome formulae, whereas paracentric inversions keep the chromosome morphology [19, 28, 29]. Also, all these *Brycon* spp. have their NORs in the terminal region of the long arm of the second pair of submetacentric chromosomes [22, 23, 25–28, 30]. Based on the distribution patterns of heterochromatin, Margarido and Galetti Jr. [23] proposed the existence of two groups of *Brycon*. The first group is characterized by heterochromatic pericentromeric markings, predominantly on the submetacentric chromosomes, whereas the second group shows heterochromatic blocks on the telomeres of metacentric chromosomes, and the presence of a distinct equilocal heterochromatic block on the first pair of metacentric chromosomes. A third, divergent pattern occurs in the coastal bryconin *H. wheatlandii*, which has a pair of NORs located in a pair of subtelocentric chromosomes, heterochromatic blocks in subtelocentric chromosomes, and non-equilocal heterochromatin in its first chromosome pair [13]. Thus, based on the cytogenetic differences between *H. wheatlandii* and other Bryconinae, we hypothesized that at least some of these characteristics could also be shared with other Bryconinae from southeastern Brazil, suggesting the existence of a phylogeographic unit within this subfamily. The aim of this study was to examine this hypothesis with a combined approach using cytogenetic data, fragments of mitochondrial DNA (cytochrome oxidase subunit I–*COI* and *16S* rDNA) and two nuclear DNA fragments (the cardiac muscle myosin heavy chain 6 alpha-*MYH6* gene and the *S72* intron) on

six southeastern bryconins: *B. devillei*, *B. ferox*, *B. insignis*, *B. opalinus*, *B. vermelha*, and *H. wheatlandii*.

## Materials and Methods

### Sampling, Preparation of Chromosomes, Banding, and Karyotypic Analysis

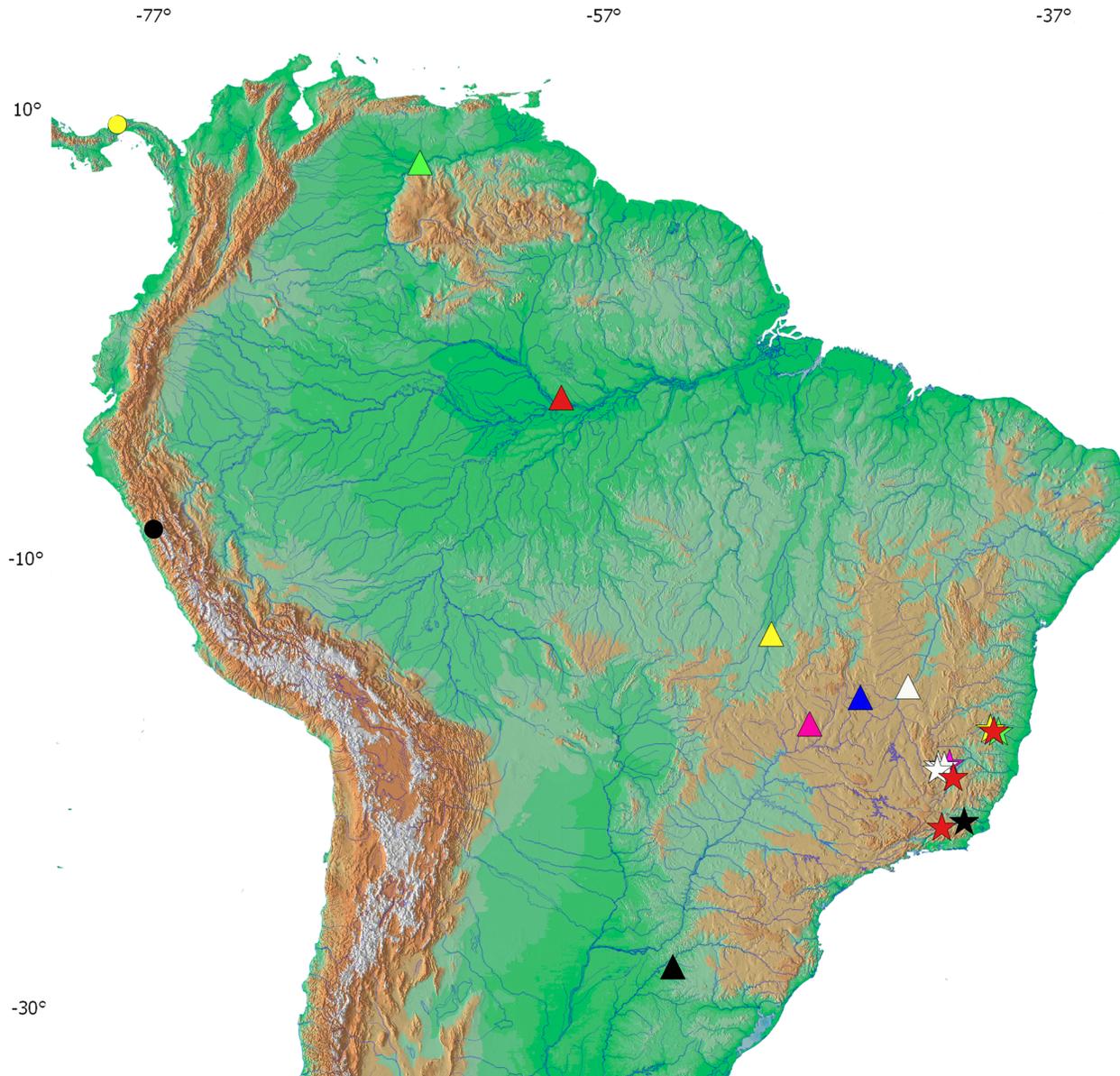
A total of 78 samples of *Brycon* spp. were collected on three coastal basins of eastern Brazil (Fig 1) and were compared with other species from seven South American river basins comprising four of Howe's bryconine groups (Table 1). The collections were carried out with collecting permit from the Instituto Chico Mendes de Biodiversidade (ICMBio) (SISBIO14975-1) issued to JAD. The specimens were deposited in the João Moojen de Oliveira Museum of Zoology at the Universidade Federal de Viçosa, Viçosa, Minas Gerais State, Brazil (MZUFV3564, MZUFV3969, MZUFV4008, MZUFV4012, MZUFV4027, MZUFV4049-4051, MZUFV4092 and MZUFV4144), Universidade Federal do Rio Grande do Sul, Porto Alegre, Rio Grande do Sul State, Brazil (UFRGS12687, UFRGS17127 and UFRGS11377) and Universidade Estadual Paulista, Botucatu, São Paulo State, Brazil (LBP13782; LBP4211; LBP2750; LBP12818; LBP3130; LBP3027; LBP1356).

The collected specimens were anesthetized with clove oil at a concentration of 0.3 gL<sup>-1</sup> [31], as approved by the Universidade Federal de Viçosa Ethics Committee (permit 032/2013). Mitotic chromosomes were obtained from the anterior kidney of fishes collected on coastal river basins, following Bertollo *et al.* [32], stained by conventional staining (Giemsa), and classified according to their arm ratio (longer chromosome arm/shorter chromosome arm) in metacentrics (1,00–1,69), submetacentrics (1,70–2,99) and subtelocentrics (3,00–6,99) following Levan *et al.* [33].

The NORs that were active in the last cell interphase were identified using silver nitrate precipitation [17]. The morphology of the NOR-bearing chromosome (S1 Fig) was estimated from using the arms ratio of 10 metaphase plates per specimen. The regions of constitutive heterochromatin were evidenced using C-banding [18]. The images of metaphases were obtained with a Olympus BX53 microscope with Olympus CellSens Imaging Software and measured using Image Pro Plus<sup>®</sup> software.

### DNA Extraction, Amplification, and Sequencing

The DNA was extracted from the gill filaments, muscle, or liver tissue of the samples, and fixed in 95% ethanol following Boyce *et al.* [34]. The *COI* gene was amplified with primers cocktail FishF1t1 and FishR1t1 [35, 36], the *16S* gene was amplified with primers Sar-5 and Sbr-3 [37], the *MYH6* gene was amplified with nested-PCR using the primers F459 and R1325 (1<sup>st</sup> PCR) and F507 and R1322 (2<sup>nd</sup> PCR) [38] and the *S72* gene was amplified with primers S72F and S73R [39]. The PCR reactions for *COI* and *16S* genes were carried out in a reaction volume of 12.5 µL [8.76 µL of H<sub>2</sub>O, 1.2 µL of 10× reaction buffer (200 mM Tris-HCl and 500 mM KCl; pH 8.4), 0.3 µL of MgCl<sub>2</sub> (100 mM), 0.05 µL of dNTPs (20 mM), 0.12 µL of each primer (10 µM), 0.0625 µL (2.5 U) of *Taq* polymerase (Phonectria<sup>®</sup>), and 200 ng of template DNA]. For *COI* and *16S* reaction (PCR) conditions were as follows: 94°C (2 min), and 35 cycles of 94°C (30 s), 52°C (40 s), and 72°C (1 min), and 72°C (10 min). PCR reactions for *MYH6* and *S72* intron were carried out in a reaction volume of 20 µL [10.3 µL of H<sub>2</sub>O, 2 µL of 10× reaction buffer (Platinum<sup>®</sup> *Taq*—Invitrogen), 0.6 µL of MgCl<sub>2</sub> (50 mM), 2 µL of dNTPs (2 mM), 2 µL of each primer (2 µM), 0.1 µL (5 U) of Platinum<sup>®</sup> *Taq*, and 100 ng of template DNA]. For *MYH6*, the first PCR was as follows: 94°C (3 min), 35 cycles of 94°C (30 s), 53°C (45 s), 72°C (1 min and



- |   |                                 |  |
|---|---------------------------------|--|
| ▲ <i>Brycon nattereri</i>               | ★ <i>Brycon devillei</i>        | ● <i>Brycon petrosus</i>                 |
| ▲ <i>Brycon</i> sp.                     | ☆ <i>Brycon opalimus</i>        | ● <i>Brycon</i> aff. <i>atrocaudatus</i> |
| △ <i>Brycon orthotaenia</i>             | ★ <i>Hemichilus wheatlandii</i> |  |
| ▲ <i>Brycon gouldingi</i>               | ★ <i>Brycon ferox</i>           |  |
| ▲ <i>Brycon orbignyianus</i>            | ★ <i>Brycon vermelha</i>        |  |
| ▲ <i>Brycon</i> cf. <i>melanopterus</i> | ★ <i>Brycon insignis</i>        |  |
| ▲ <i>Brycon falcatus</i>                |                                 |  |

**Fig 1. Coastal and continental basins in southeastern Brazil and collect local of the Bryconine species examined on this study.**

doi:10.1371/journal.pone.0137843.g001

**Table 1. Bryconin species, number of samples used on cytogenetic and molecular analyses with geographical coordinates of sampling sites.**

Species	Sample size			GPS coordinates	Locality (Hydrographical basin)
	Cytogenetics			Molecular	
	♂ ♀				
<i>Brycon amazonicus</i>	00	00	02	21° 58'07"S 43° 07'43"W	Doce River, Santana do Deserto, MG (Doce River basin—Brazil).
<i>Brycon aff. atrocaudatus</i>	00	00	01	08°40'40"S78° 09'163"W	Rio Santa (Pacific—Peru).
<i>Brycon devillei</i>	01	00	01	19°45'24"S 42°37'13"W	Carioca Lake, Dionísio, MG (Doce River basin—Brazil).
	06	05	03	21° 58'07"S 43° 07'43"W	Doce River, Santana do Deserto, MG (Doce River basin—Brazil).
	00	02	02	17°41'09"S 40°50'33"W	Mucuri River, Carlos Chagas, MG (Brazil).
<i>Brycon falcatus</i>	00	00	01	07°38'11.6"S 66° 19'04.2" W	Orinoco River, Caicara del Orinoco (Venezuela)
<i>Brycon ferox</i>	04	03	02	17°41'09"S 40°50'33"W	Mucuri River, Carlos Chagas, MG (Mucuri basin—Brazil).
<i>Brycon gouldingi</i>	00	00	01	13°20'051"S50° 42'162"W	Lagoa da Égua, Mato Grosso (Araguaia River basin—Brazil)
<i>Brycon insignis</i>	07	02	02	21°42'35"S 42°07'55"W	Paraíba do Sul River, Itaocara, RJ (Paraíba do Sul River basin—Brazil).
<i>Brycon cf. melanopterus</i>	00	00	01	28°37'44"S 60°58'44"W	Balneary Adão e Maria, Manaus, AM (Amazonas River basin—Brazil).
<i>Brycon moorei</i>	00	00	01	not available	Magdalena River, Antioquia, (Colombia).
<i>Brycon nattereri</i>	00	00	02	17°20'00"S 49°0'05"W	Córrego Coqueiro, Cachoeira das Piracanjubas, GO (Paraíba do Sul/Alto Paraná Rivers basin—Brazil).
<i>Brycon opalinus</i>	01	02	02	19°13'02"S 42°53'03"W	Santo Antônio River, Sete Cachoeiras, MG (Doce River basin—Brazil).
	07	03	00	19°13'24"S 42° 52'12"W	Esmeralda Stream, Sete Cachoeiras, MG (Doce River basin—Brazil).
	02	02	02	19°25'11"S43°19'22"O	Preto/Itambé do Mato Dentro River, MG (Doce River basin—Brazil).
Species	Sample size			GPS coordinates	Locality(Hydrographicalbasin)
	Cytogenetics			Molecular	
	♂♀				
<i>Brycon orthotaenia</i>	00	00	02	15°40'18"S 44° 37'43"O	River Pandeiros, Januária, MG (São Francisco River basin—Brazil).
<i>Brycon orbignyanus</i>	00	00	01	28°08'33"S 55°04'44"O	River Ijuí, Roque Gonzales, RS (Uruguai River—Brazil).
<i>Brycon petrosus</i>	00	00	01	09°19'262"N79° 46'082"O	Río Llano Sucio, Panama (Atlantic)
<i>Brycon sp.</i>	00	00	01	16°26'66"S 46°12'67"O	Palmeirinha, MG (São Francisco River basin—Brazil).
<i>Brycon vermelha</i>	04	02	02	17°41'09"S 40°50'33"O	Mucuri River, Carlos Chagas, MG (Mucuri River basin—Brazil).
<i>Henochilus wheatlandii</i>	00	00	01	19°13'02"S 42°53'03"O	Santo Antônio River, Sete Cachoeiras, MG (Doce River basin—Brazil).

doi:10.1371/journal.pone.0137843.t001

30 s), and 72°C (10 min). The second PCR was: 94°C (3 min), 35 cycles of 94°C (30 s), 62°C (45 s), 72°C (1 min and 30 s), and 72°C (5 min). For *S72*, PCR conditions were as follows: 95°C (2 min), 35 cycles of 95°C (30 s), 54°C (30 s), 72°C (1 min), and 72°C (10 min). The PCR products were purified by using enzymatic method Exosap (25% exonuclease, 25% Shrimp Alkaline Phosphatase and 50% of deionized water) or PEG 8000 (20% polyethyleneglycol, 2.5 M NaCl), and sequencing was performed on a Macrogen sequencing platform of Macrogen, Seoul, South Korea. Sequences of each locus were independently aligned using Clustal W [40] with sequencing chromatograms checked by eye and alignments realigned with Muscle [41] in MEGA 5.0 software [42]. Standard genetic summaries were calculated for each gene using MEGA 5.0 [42].

DNA alignments were concatenated and analysed using a partitioned Bayesian approach. The molecular evolution models were selected based on the Bayesian information criterion BIC in Partition Finder v1.1.0 [43]. The data set was divided into eight sections corresponding to 16S gene and S72 intron and first, second and third positions for the genes COI and MYH6. Bayesian inference was performed two times by using MrBayes software version 3.2.3 [44] with four independent Markov chain Monte Carlo (MCMC) that were run 30,000,000 replicates with a phylogenetic tree sampled every 1,000 generations. The first 25% generations were discarded as burn-in and the remaining trees were used to generate statistics and topology. The distribution of log likelihood scores was examined to determine stationarity for each search and to decide whether extra runs were required to achieve convergence using the program Tracer 1.4 [45].

Maximum likelihood analysis was performed with RAxML version 8.1.1 [46], using a mixed partition model indicated by Partition Finder v1.1.0 [43] with all parameters set to default values. Topological robustness was assessed using 1,000 nonparametric bootstrap replicates. MrBayes and RAxML analyses were run on computational resources provided by Cyberinfrastructure for Phylogenetic Research (CIPRES) [47].

Maximum parsimony analyses were conducted with the concatenated alignment within PAUP\* 4.0b10 [48]. Heuristic searches were performed with 100 random addition replicates and TBR branch swapping. All characters were unordered, all character transformations were equally weighted, and gaps were treated as missing data. Clade robustness was verified with 1,000 bootstrap pseudoreplicates [49]. The posterior probability values of 1–0.91 output by MrBayes and bootstrap values over 90% output by RAxML and maximum parsimony analyses were considered as well supported [50]. The sequences amplified in this study were deposited in GenBank (Access No. XXXX).

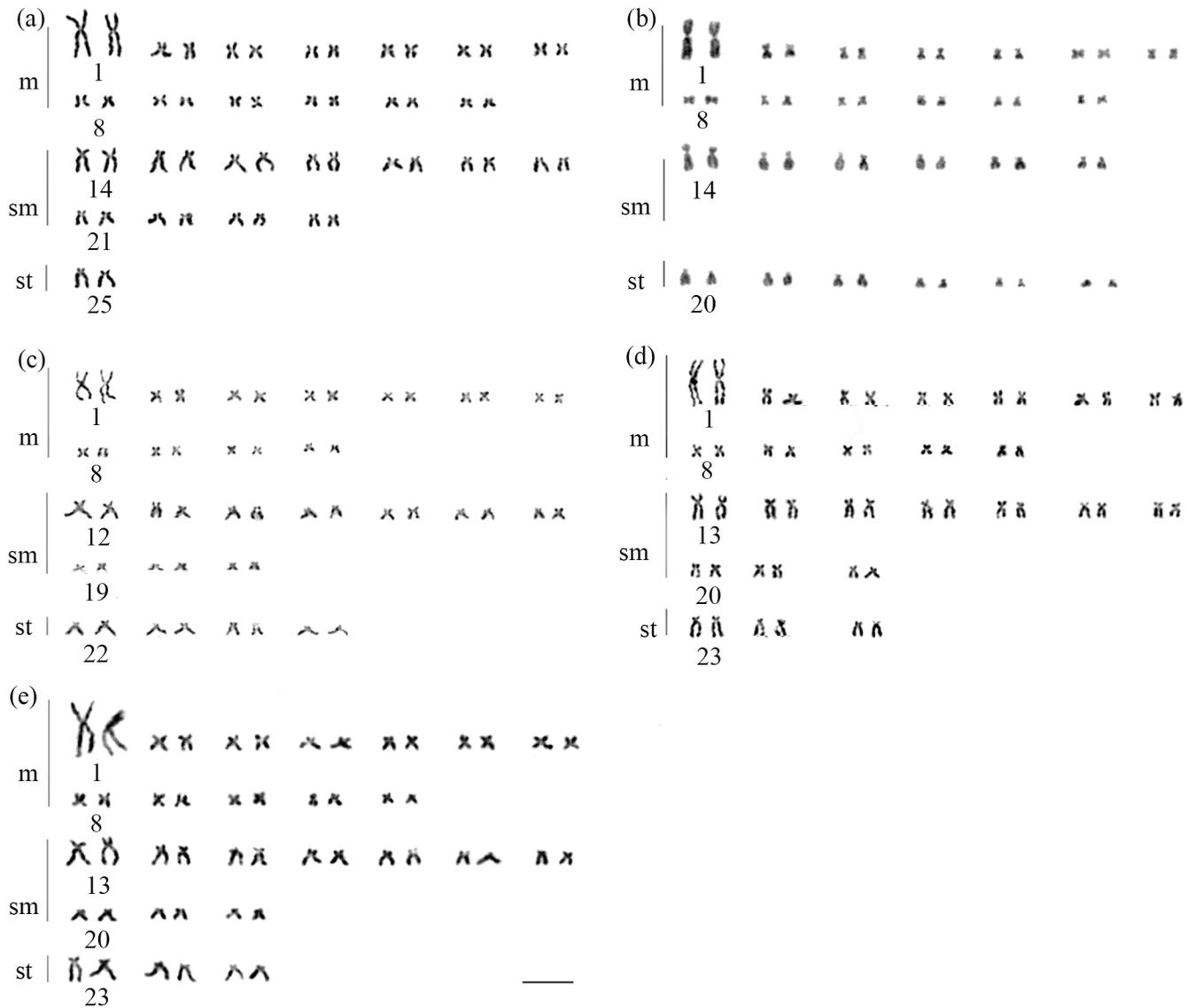
## Results

### Karyotypic Analyses

All species of *Brycon* from the coastal basins of southeastern Brazil had diploid number of 50 chromosomes and a fundamental number equal to 100, though the karyotypic formulae varied among most of the species; no polymorphisms were observed within each species: *B. devillei*, 26m+22sm+2st; *B. ferox*, 26m+12sm+12st; *B. insignis*, 22m+20sm+8st; and *B. opalinus* and *B. vermelha*, 24m+20sm+6st (Fig 2). In all samples, NORs occurred in the terminal region of the long arm on the first pair of subtelocentric chromosomes (Fig 3). All samples exhibited a pericentromeric heterochromatic block on the long arm of the first pair of the metacentric chromosomes. Additionally, in *B. vermelha* a centromeric heterochromatin block was also observed on the first chromosome pair (Fig 4). On the other hand, the presence of pericentromeric heterochromatic regions varied among samples and characterized different chromosome pairs: *B. devillei*: 14, 15, 17, 18, and 25 (Fig 4a); *B. ferox*: 14, 20, and 21 (Fig 4b); *B. insignis*: 12, 13, 22, 23, and 24 (Fig 4c); *B. opalinus*: 14, 15, 16, 18, and 20 (Fig 4d); and *B. vermelha*: 13, 14, 23, and 24 (Fig 4e) (Table 2).

### Molecular Analyses

Sequence alignment of the 418-bp 16S gene fragment yielded 100 variable sites, 62 of them were parsimony informative; the average nucleotide composition was  $\pi_T = 0.20$ ;  $\pi_C = 0.24$ ,  $\pi_A = 0.34$ , and  $\pi_G = 0.22$ . The estimated average transition/transversion rate was 4. Sequence alignment of the 599 bp- COI gene fragment was obtained for all species, except for *Brycon melanopterus*, which was included as missing data in the multiple alignments. This fragment yielded 193 variable sites, 147 of them were parsimony informative, the average nucleotide



**Fig 2. Karyotype of *Brycon* species from eastern Brazil.** *Brycon devillei* (a), *Brycon ferox* (b), *Brycon insignis* (c), *Brycon opalinus* (d), and *Brycon vermelha* (e). The bar represents 10  $\mu$ m.

doi:10.1371/journal.pone.0137843.g002

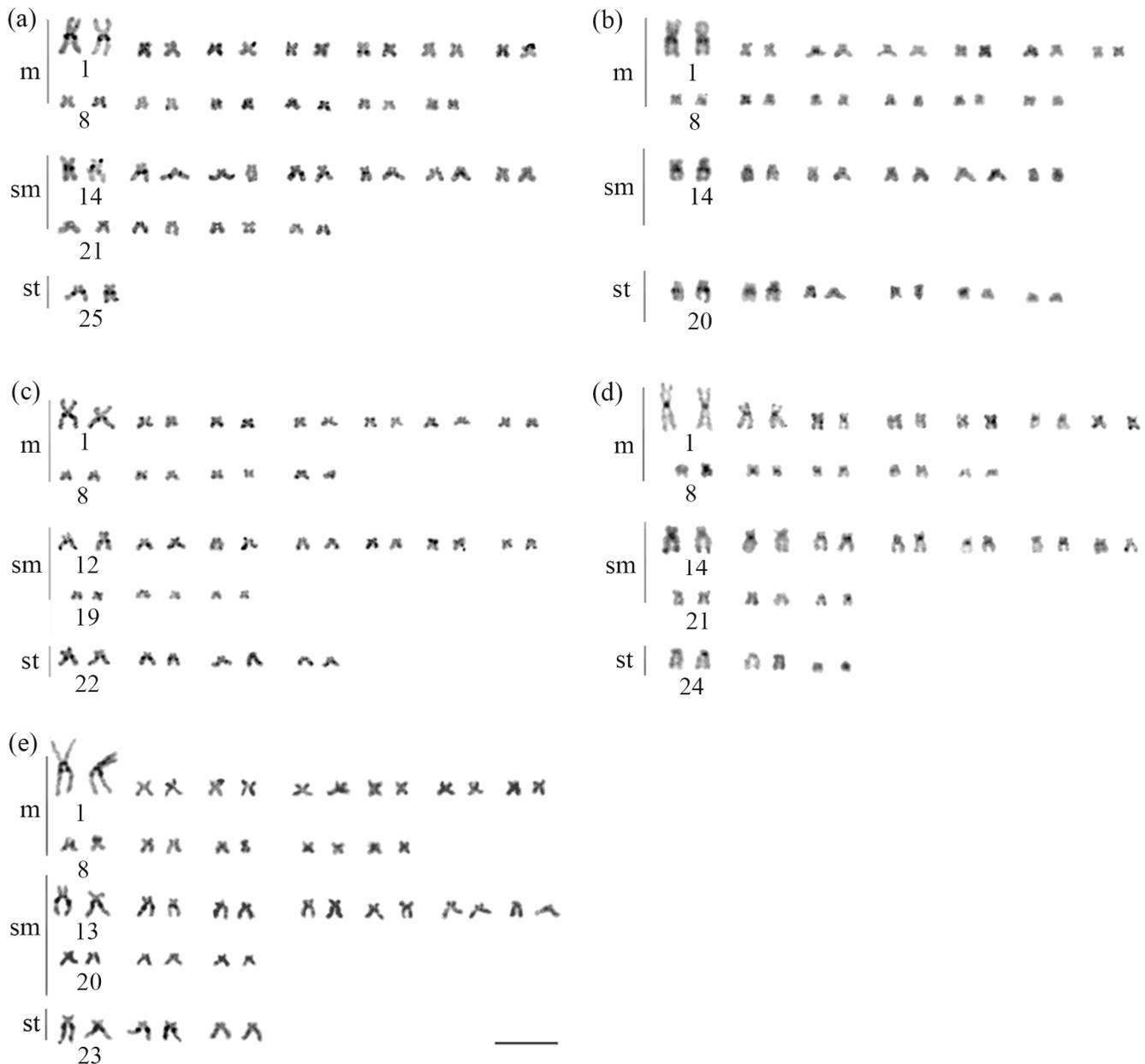
composition was  $\pi_T = 0.30$ ,  $\pi_C = 0.27$ ,  $\pi_A = 0.25$ , and  $\pi_G = 0.18$ . The estimated average transition/transversion rate was 9. The 733 bp-long fragment of the *MYH6* gene was obtained for most samples, except *Brycon atrocaudatus* and *Brycon petrosus*; those sequences were considered missing data in the multiple alignments. The *MYH6* fragment yielded 53 variable sites, 30 of them were parsimony informative, and the average nucleotide composition was  $\pi_T = 0.24$ ,  $\pi_C = 0.22$ ,  $\pi_A = 0.30$ , and  $\pi_G = 0.24$ . The estimated average transition/transversion rate was 2.

The *S72* 356 bp-long fragment was obtained for most samples, except for *Brycon atrocaudatus*, which was included as missing data in the multiple alignment. The *S72* fragment yielded 172 variable sites, 77 of them were parsimony informative, and the average nucleotide composition was  $\pi_T = 0.34$ ,  $\pi_C = 0.19$ ,  $\pi_A = 0.26$ , and  $\pi_G = 0.21$ . The estimated average transition/transversion rate was 11. Twelve heterozygote sites were observed among the 32 specimens and the nucleotide code followed the International Union of Pure and Applied Chemistry



**Fig 3. Distribution of active NORs in the last cell interphase in *Brycon* species that occur in the basins of eastern Brazil.** *Brycon devillei* (a), *B. insignis* (b), *B. ferox* (c), *B. opalinus* (d), and *B. vermelha* (e). The bar represents 10  $\mu$ m.

doi:10.1371/journal.pone.0137843.g003



**Fig 4. Heterochromatic patterns of *Brycon* species that occur in the basins of eastern Brazil.** *Brycon devillei* (a), *B. insignis* (b), *B. opalinus* (c), *B. ferox* (d), and *B. vermelha* (e). The bar represents 10  $\mu$ m.

doi:10.1371/journal.pone.0137843.g004

(IUPAC). The final matrix with DNA sequences concatenated resulted in 2,109 bp fragment deposited in TreeBase ([www.treebase.org](http://www.treebase.org)) under number XXXX.

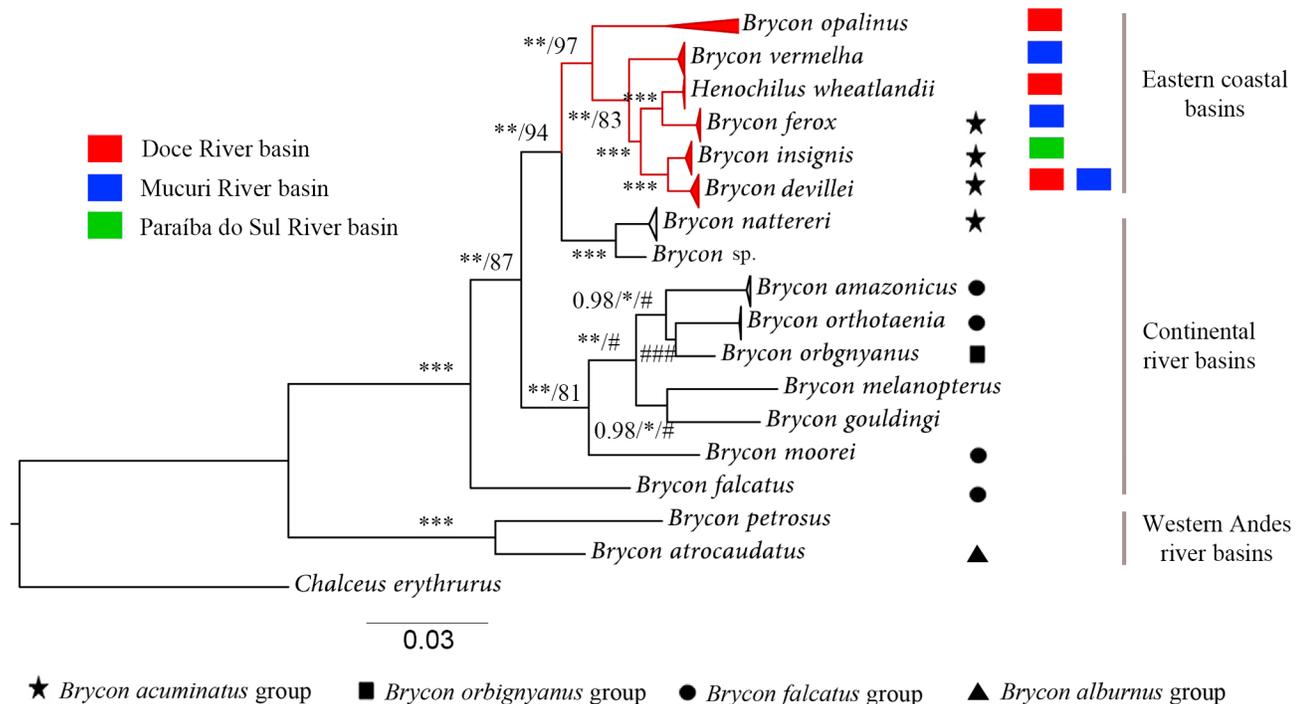
The most suitable partitions indicated by Partition Finder was composed of five subsets: (1) molecular model K80+I+G for *16S* gene partition, *COI*\_1<sup>st</sup> codon position and *MYH6*\_3<sup>rd</sup> codon position; (2) molecular model F81 for *COI*\_2<sup>nd</sup> codon position; (3) molecular model GTR+G for *COI*\_3<sup>rd</sup> codon position; (4) molecular model F81+I for *MYH6*\_1<sup>st</sup> and *MYH6*\_2<sup>nd</sup> codon position, and (5) HKY+G for *S72* partition.

**Table 2. Bryconin species, karyotypic formulae, Ag-NORs and heterochromatic patterns.**

Species	Karyotypic formulae	Ag-NORs	Heterochromatic patterns
<i>Brycon devillei</i>	26m+22sm+2st	On terminal region of the long arm of the first pair of subtelocentric chromosomes.	Pericentromeric block on the long arm of the first pair of the metacentric chromosomes.
<i>Brycon ferox</i>	26m+12sm+12st	On terminal region of the long arm of the first pair of subtelocentric chromosomes.	Pericentromeric block on the long arm of the first pair of the metacentric chromosomes.
Species	Karyotypic formulae	Ag-NORs	Heterochromatic patterns
<i>Brycon insignis</i>	22m+20sm+2st	On terminal region of the long arm of the first pair of subtelocentric chromosomes.	Pericentromeric block on the long arm of the first pair of the metacentric chromosomes.
<i>Brycon opalinus</i>	24m+20sm+6st	On terminal region of the long arm of the first pair of subtelocentric chromosomes.	Pericentromeric block on the long arm of the first pair of the metacentric chromosomes.
<i>Brycon vermelha</i>	24m+20sm+6st	On terminal region of the long arm of the first pair of subtelocentric chromosomes.	Pericentromeric block on the long arm of the first pair of the metacentric chromosomes and centromeric block was also observed on the first chromosome pair.

doi:10.1371/journal.pone.0137843.t002

The phylogenetic hypothesis obtained with concatenated genes with Bayesian inference (MrBayes), maximum likelihood (RAxML) and maximum parsimony (PAUP) recovered two groups to the east and to the west of the Andean relief. Within the eastern Andean group, one sub-group was formed by the coastal species *B.insignis*, *B. devillei*, *B. ferox*, *H. wheatlandii*, *B. vermelha*, and *B. opalinus* (Fig 5).



**Fig 5. Hypothesis of phylogenetic relationships among some species of the subfamily Bryconinae.** The numbers and symbols at nodes indicate the posterior probability (first) and bootstraps (second and third) obtained by Bayesian, maximum likelihood and maximum parsimony approaches, respectively. \* denotes posterior probability = 1 and bootstrap values = 100. # denotes posterior probability lower than 0.9 and bootstrap lower than 80%. Symbols indicate three cis-Andean and one trans-Andean morphological groups proposed by Howes (1982). The “*B. guatemalensis*” species group were not included in the analyses. Painted squares denote the coastal basins of eastern Brazil where samples were collected. Bars represent the molecular distance.

doi:10.1371/journal.pone.0137843.g005

## Discussion

Karyotypes composed of  $2n = 50$  metacentric, submetacentric, and subtelocentric chromosomes characterize the Bryconinae [13, 22, 23, 25–27, 51] and *Salminus* species [27, 52, 53]. The close phylogenetic relationship between *Brycon* spp. and *Salminus* spp. led some researchers to propose the Family Bryconidae [12]. Despite the stable diploid number, the differences in the chromosomal formulae among *Brycon* spp. from southeastern Brazil indicate a preponderance of pericentric and possible paracentric inversions in the karyotypic evolution of this group. The stable karyotypical macrostructure and the basal position of *B. opalinus* and *B. vermelha* suggest that their karyotypic formula of  $24m+20sm+6st$  is a plesiomorphy in southeastern Brazilian bryconins. Pericentric inversions occurred in the ancestor of *H. wheatlandii* and *B. ferox* resulting in the chromosome formula of  $26m+12sm+12st$  shared by these two species. Finally, the dichotomy between *B. devillei* and *B. insignis* involved the retention of 26 metacentrics in *B. devillei* and alteration in the number of submetacentrics and subtelocentrics, whereas the karyotype of *B. insignis* involved changes in the number of metacentrics, submetacentrics, and subtelocentrics. Pericentric inversions are relevant in the speciation of several groups of animals, including fish [19], insects [54, 55], and reptiles [56], and may represent potential isolation factors among among sympatric fishes with similar reproductive habits.

The presence of NORs in the terminal regions of the long arms of the first subtelocentric pair of chromosomes in the southeastern *Brycon* spp. is similar to that previously reported in *H. wheatlandii* [13], corroborating a second pattern that diverged from other Neotropical Bryconinae such as *B. cephalus*, *B. orthotaenia*, *B. hilarii*, and *B. orbignyanus*, which have a pair of NORs on the second submetacentric chromosome pair [22, 23, 26]. Although Almeida-Toledo *et al.* [22] and Margarido and Galetti Jr. [23] indicate that *B. insignis* is characterized by NORs in a pair of submetacentric chromosomes, the *B. insignis* sample included in the present study showed sites of NORs in its first pair of subtelocentric chromosomes. This difference suggests the existence of polymorphism in this species, because both samples were collected in the Paraíba do Sul River Basin. One possible exception for the presence of NORs in the subtelocentric chromosomes in the continental species is *B. amazonicus* [25]. Although Mariguela *et al.* [25] indicate that this species has its NORs in a subtelocentric chromosome pair, these authors included the NOR-bearing chromosome in the submetacentric chromosome group. Therefore, the presence of NORs in a subtelocentric chromosome pair is restricted to Bryconinae spp. from the coastal basins of southeastern Brazil.

Another unique feature of the eastern coastal bryconine is the distribution pattern of heterochromatic blocks in the chromosomes. All *Brycon* species present in the southeastern Brazilian coastal watersheds, including *H. wheatlandii*, lack equilocal heterochromatic blocks in their first chromosome pair, whereas *Brycon* species occurring the continental watersheds have heterochromatic equilocal blocks [23, 25, 27]. Indeed, the presence of equilocal heterochromatic blocks in *B. amazonicus*, *B. hilarii*, *B. orthotaenia*, and *Salminus hilarii* may indicate a plesiomorphic feature within the Bryconidae [12, 25, 27]. Consequently, the loss of equilocality is a synapomorphy that characterizes the southeastern coastal bryconins, whereas the presence of heterochromatic block in the telomeric region of the first pair is an autapomorphy in *H. wheatlandii* [13].

As previously indicated, the variation in heterochromatine patterns among species of *Brycon* were used by Margarido and Galetti Jr. [23] to propose the existence of two species groups within this taxon. The first group includes the species *B. cephalus*, *B. hilarii*, and *B. orbignyanus* "that revealed telomeric bands in **some** metacentric chromosomes," whereas the second group was characterized by having "**predominantly** centromeric and pericentromeric positive C band, **mainly** in submetacentric chromosomes". This second group is represented by *B. orthotaenia*,

*B. falcatus*, and *B. insignis*. A putative third pattern of heterochromatin was proposed for *H. wheatlandii*, which was characterized by telomeric marks on the first metacentric chromosome pair and predominantly pericentromeric markings in the subtelocentric chromosomes [13]. Silva *et al.* [13] hypothesized that bryconine occurring in the coastal basins of eastern Brazil might share the third pattern, which is supported by the results obtained in the present study.

To date, the cytogenetic data of Neotropical fishes have shown some major trends: karyotypic stability in Anostomidae [57, 58], Prochilodontidae [58, 59–61], and Curimatidae [57, 62], and the existence of species complexes in the genera *Astyanax* [63] and *Hoplias* [64]. However, the cytogenetic pattern observed in bryconine from Brazilian southeastern coastal basins represents the first case in which synapomorphies of chromosomal banding show monophyly of well-defined morphological species.

A well-supported clade formed by bryconine from eastern Brazilian coastal river basins was recovered in the molecular analyses. This topology differs from the one proposed by Abe *et al.* [14] where *Brycon nattereri* appears as more related to the eastern coastal basins bryconine than *Brycon opalinus*. Species sampling, such as the inclusion of *B. devillei* and the use of genes with faster substitution rates may have influenced this result. Long branches and low bootstrap values using maximum likelihood and maximum parsimony analyses were shown on the ancestral node shared by *B. nattereri* and other coastal bryconine clade (without *B. opalinus*) on the concatenated tree of Abe and colleagues (Fig 3 from Abe *et al.* [14]). Low levels of phylogenetic support among eastern coastal bryconines are more evident on Abe and colleagues analysis (Fig 4 from Abe *et al.* [14]) and *B. nattereri* and *B. opalinus* sister species status may be the result of long-branch attraction between them. Likewise, our analysis showed a more resolved phylogenetic relationship among all eastern coastal basin bryconine species, including *B. insignis*, *B. ferox*, *B. vermelha* and *H. wheatlandii*, which is unresolved in Abe *et al.* [14].

The presence of one well-defined clade composed exclusively by species present in the eastern coastal river basins, seems to be the result of the long independent paleohydrological history of these watersheds. The coastal basins of eastern Brazil are isolated from the continental watersheds by Serra da Mantiqueira and Serra do Espinhaço, which represent the boundaries of the distribution ranges of various fish species [8, 9]. It has been reported that the patterns of cytogenetic and molecular variation in the predator *Hoplias malabaricus* are consistent with the paleohydrological history of the coastal basins of eastern Brazil [10], although this species also shows evidence of geodispersal between coastal and other continental basins [16]. The pattern of *Brycon* distribution in the coastal basins partially fits the subdivision of the areas of endemism among the Paraíba do Sul and Doce Rivers proposed by Carvalho [65]. Furthermore, Menezes [66] proposed that the Atlantic coastal drainages can be divided into three regions of endemism: the Northern region of the Doce River to the mouth of the Jequitinhonha River; a Central region from the Cubatão River to the Itabapoana River to the north, characterized by the occurrence of *Oligosarcus hepsetus*; and a Southern region, characterized by the presence of *Oligosarcus jenynsii* and *Oligosarcus robustus*. In addition, Abell *et al.* [67] also classified drainages into six ecoregions based on the compositions of species in different taxonomic levels: Northeast Atlantic, Paraíba do Sul, Fluminense, Ribeira de Iguape, Southeastern Atlantic, Tramandai-Mampituba, and Laguna dos Patos. However, Buckup [68] indicated that the limits and relations of endemism along the Brazilian Shield are not yet well defined.

The phylogeographic analyses carried out in the present study based on the cytogenetic and molecular data also shed light on the taxonomic status of some Bryconidae. According to the phylogenetic hypothesis, *H. wheatlandii* appears to be closely related to *B. ferox* from the Mucuri River, which corroborates the evidence that *Brycon* is a paraphyletic group [11, 12, 14]. The high degree of morphological divergence seen in *H. wheatlandii* relative to its sister species has been considered as an example of adaptive radiation [69], which in this case seems to be

unrelated to the extinction of intermediate forms. The position of *B. devillei* and *B. insignis* as reciprocally monophyletic and allopatric species is reinforced by the high degree of cytogenetic divergence between them.

Specimens of *B. devillei* from the Doce and the Mucuri rivers shared the same cytogenetic and molecular characters, indicating that these populations belong to the same species. This is especially informative for the taxonomy of this species. Although Lima *et al.* [1] state that the precise type locality of *B. devillei* is unknown, the Mucuri River drains into the Atlantic Ocean in the southern tip of the Bahia State, which is consistent with the original Castelnau's description [1]. The close relatedness of the Doce and Mucuri populations can be explained by the recently paleohydrology of these watersheds. As a result of eustatic sea level variations during the periods of glacial maximum, the existing drainage of the Doce and Mucuri rivers probably converged in paleo channels that are currently represented by isobaths of 60 m and surrounded by 30-m isobaths [70]. These Pleistocene paleochannels are delimited to the south by the mouth of the Doce River and to the north by Abrolhos Formation, and explain the patterns of distribution and phylogeography of the freshwater fish fauna [10] unrelated to Bryconidae.

The results of the present study demonstrate that the Bryconinae from the coastal basins of southeastern Brazil form a phylogenetic and phylogeographic unit. The cytogenetic characteristics that differentiate these fish from their continental congeners are: presence of a pair of subtelocentric chromosomes bearing NORs (vs. submetacentric) and the absence of equilocal heterochromatic blocks in their first chromosome pair; this monophyly is strongly supported by molecular data. Our results partially corroborate the "*Brycon acuminatus*" group proposed by Howes in 1982 [2], keeping *B. devillei*, *B. ferox*, and *B. insignis*; and including *B. opalinus* (which was a member of Owen's "falcatus" group), *B. vermelha* (described by Lima and Castro [4]), and *H. weatlandii* (formerly considered unrelated to *Brycon* [11]) whereas it excludes *B. nattereri*.

## Supporting Information

**S1 Fig. Chromosome spread from Ag-NOR banding protocols presented in this work.** *Brycon devillei* (a); *Brycon ferox* (b); *Brycon insignis* (c); *Brycon opalinus* (d), and *Brycon vermelha* (e).  
(TIF)

## Acknowledgments

This work was supported by the Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES), Conselho Nacional de Pesquisa (CNPq), and Fundação de Amparo à Pesquisa do Estado de Minas Gerais (FAPEMIG). We thank Flávio César Thadeo de Lima for specimen identification, the Piabanha Project for donating *Brycon insignis*, Luiz R. Malabarba for donating tissue samples, and the students of the Laboratory of Molecular Systematics of the Universidade Federal de Viçosa (UFV) for field and laboratory assistance.

## Author Contributions

Conceived and designed the experiments: JAD. Performed the experiments: NMT PCS US. Analyzed the data: JAD NMT PCS US. Contributed reagents/materials/analysis tools: JAD CO. Wrote the paper: JAD NMT PCS US JCZ.

## References

1. Lima FCT (2003) Subfamily Bryconinae. In: Malabarba LR, Reis RE, Vari RP, Lucena ZMS, Lucena CAS, editors. *Phylogeny and Classification of Neotropical Fishes*. Porte Alegre: Edipucrs. Pp. 174–181.
2. Howes GJ (1982) Review of the genus *Brycon* (Teleostei: Characoidei). *Bull Br Mus Nat Hist (Zool)* 43: 1–47.
3. Botero-Botero A, Ramírez-Castro H (2011) Ecología trófica de la Sabaleta *Brycon henni* (Pisces: Characidae) en el río Portugal de Piedras, Alto Cauca. *Rev MVZ Córdoba* 16: 2349–2355.
4. Lima FCT, Castro RMC (2000) *Brycon vermelha*, a new species of characid fish from the Rio Mucuri, a coastal river of eastern Brazil (Ostariophysi: Characiformes). *Ichthyol Explor Freshw* 11: 55–62.
5. Weitzman SH, Menezes NA & Weitzman MJ (1988) Phylogenetic biogeography of the *Glandulocaudini* (Teleostei: Characiformes, Characidae) with comments on the distribution of other freshwater fishes in eastern and southeastern Brazil. Proceedings of a Workshop on Neotropical Distribution Patterns. In: PE Vanzolini, WR Heyer, editor. *Academia Brasileira de Ciências*.
6. Bizerril CRSF (1994) Análise taxonômica e biogeográfica da ictiofauna de água doce do leste do Brasil. *Acta Biol. Leopold.* 16: 51–80.
7. Buckup PA (2011) The Eastern Brazilian Shield. In: Albert JS & Reis Historical RE, editor. *Biogeography of Neotropical Freshwater Fishes*. University of California Press, Berkeley, EUA.
8. Ribeiro AC (2006) Tectonic history and the biogeography of the fresh water fishes from the coastal drainages of eastern Brazil: an example of faunal evolution associated with a divergent continental margin. *Neotrop Ichthyol* 4: 225–246.
9. Ingenito LFS & Buckup PA (2007) The Serra da Mantiqueira as a biogeographic barrier for fishes, southeastern Brazil. *J Biogeogr* 34: 1173–1182.
10. Pereira TL, Santos U, Schaefer CE, Souza GO, Paiva SR, Malabarba LR, Schmidt EE & Dergam JA (2012) Dispersal and vicariance of *Hoplias malabaricus* (Bloch, 1794) (Teleostei, Erythrinidae) populations of the Brazilian continental margin. *J Biogeogr* 40: 905–914.
11. Hilsdorf S, Oliveira C, Lima FCT, Matsumoto CK (2008) A phylogenetic analysis of *Brycon* and *Hemochilus* (Characiformes, Characidae, Bryconinae) based on the mitochondrial gene 16S rRNA. *Genet Mol Biol* 31: 366–371.
12. Oliveira C, Avelino GS, Abe KT, Mariguela TC, Benine RC, Ortí G, Vari RP, Corrêa e Castro RM (2011) Phylogenetic relationships within the speciose family Characidae (Teleostei: Ostariophysi: Characiformes) based on multilocus analysis and extensive in group sampling. *BMC Evol Biol* 11: 275. doi: [10.1186/1471-2148-11-275](https://doi.org/10.1186/1471-2148-11-275) PMID: [21943181](https://pubmed.ncbi.nlm.nih.gov/21943181/)
13. Silva PC, Santos U, Travenzoli NM, Zanuncio JC, Cioffi MB, Dergam JA (2012) The unique karyotype of *Hemochilus wheatlandii*, a critically endangered fish living in a fast-developing region in Minas Gerais State, Brazil. *PLoS ONE* 7: e42278. doi: [10.1371/journal.pone.0042278](https://doi.org/10.1371/journal.pone.0042278) PMID: [22848754](https://pubmed.ncbi.nlm.nih.gov/22848754/)
14. Abe KT, Mariguela TC, Avelino GS, Foresti F, Oliveira C (2014) Systematic and historical biogeography of the Bryconidae (Ostariophysi: Characiformes) suggesting a new rearrangement of its genera and an old origin of Mesoamerican ichthyofauna. *BMC Evol Biol* 14: 152. doi: [10.1186/1471-2148-14-152](https://doi.org/10.1186/1471-2148-14-152) PMID: [25005252](https://pubmed.ncbi.nlm.nih.gov/25005252/)
15. Santos U, Volcker CM, Belei FA, Cioffi MB, Bertollo LAC, Paiva SR & Dergam JA (2009) Molecular and karyotypic phylogeography in the Neotropical *Hoplias malabaricus* (Erythrinidae) fish in eastern Brazil. *J Fish Biol* 75: 2326–2343. doi: [10.1111/j.1095-8649.2009.02489.x](https://doi.org/10.1111/j.1095-8649.2009.02489.x) PMID: [20738690](https://pubmed.ncbi.nlm.nih.gov/20738690/)
16. Jacobina UP, Affonso PRAM, Carneiro PLS, Dergam JA (2009) Biogeography and comparative cytogenetics between two populations of *Hoplias malabaricus* (Bloch, 1794) (Ostariophysi: Erythrinidae) from coastal basins in the State of Bahia, Brazil. *Neotrop Ichthyol* 7: 617–622.
17. Howell WM, Black DA (1980) Controlled silver—staining of nucleolus organizer regions with a protective colloidal developer: a 1-step method. *Experientia* 36: 1014–1015. PMID: [6160049](https://pubmed.ncbi.nlm.nih.gov/6160049/)
18. Sumner AT (1972) A simple technique for demonstrating centromeric heterochromatin. *Exp Cell Res* 75: 304–306. PMID: [4117921](https://pubmed.ncbi.nlm.nih.gov/4117921/)
19. Kavalco KF, Pazza R, Bertollo LAC, Moreira-Filho O (2005) Molecular cytogenetics of *Oligosarcus hepsetus* (Teleostei, Characiformes) from two Brazilian locations. *Genetica* 124: 85–91. PMID: [16011006](https://pubmed.ncbi.nlm.nih.gov/16011006/)
20. Artoni RF, Vicari MR, Almeida MC, Moreira-Filho O, Bertollo LAC (2009) Karyotype diversity and fish conservation of southern field from South Brazil. *Re Fish Biol Fisheries* 19: 393–401
21. de Barros LC, Santos U, Cioffi MB, Dergam JA (2015) Evolutionary Divergence Among *Oligosarcus* spp. (Ostariophysi, Characidae) from the São Francisco and Doce River Basins: *Oligosarcus solitarius* Menezes, 1987 Shows the Highest Rates of Chromosomal Evolution in the Neotropical Region. *Zebrafish* 12: 102–110. doi: [10.1089/zeb.2014.1030](https://doi.org/10.1089/zeb.2014.1030) PMID: [25602472](https://pubmed.ncbi.nlm.nih.gov/25602472/)

22. Almeida-Toledo LF, Bigoni AP, Bernardino G, Foresti F, Toledo-Filho SA (1996) Karyotype and NOR conservatism with heterochromatin reorganization in Neotropical Bryconids. *Caryologia* 49: 35–43.
23. Margarido VP, Galetti PM Jr. (1996) Chromosome studies in fish of the genus *Brycon* (Characiformes, Characidae, Bryconinae). *Cytobios* 85: 219–228.
24. López DD, Palacio GV, Cortes TR, Angel MO (2008) Caracterización citogenética del pez neotropical *Brycon henni* (Pisces: Characidae). *Rev Biol Trop* 56: 1619–1628.
25. Mariguela TC, Nirchio M, Ron E, Gaviria JI, Foresti F, Oliveira C (2010) Cytogenetic characterization of *Brycon amazonicus* (SpixetAgassiz, 1829) (Teleostei: Characidae) from Caicaradel Orinoco, Venezuela. *Comp Cytogenet* 4: 185–193.
26. Wasko AP, Galetti PM Jr. (2000) Mapping 18s ribosomal genes in fish of the genus *Brycon* (Characidae) by fluorescence in situ hybridization (FISH). *Genet Mol Biol* 23: 135–138.
27. Margarido VP, Galetti PM Jr (1999) Heterochromatin patterns and karyotype relationships within and between the genera *Brycon* and *Salminus* (Pisces, Characidae). *Genet Mol Biol* 22: 357–361.
28. Molina WF, Galetti PM Jr. (2004) Karyotypic changes associated to the dispersive potential on Pomacentridae (Pisces, Perciformes). *J Exp Mar Biol Ecol* 309: 109–119.
29. Garcia C, MoREIRA-FILHO O (2005) Cytogenetical analyses in three fish species of the genus *Pimelodus* (Siluriformes: Pimelodidae) from Rio São Francisco: considerations about the karyotypical evolution in the genus. *Neotrop Ichthyol* 3: 285–289.
30. Wasko AP, Martins C, Wright JM., Galetti PM Jr (2001) Molecular organization of 5S rDNA in fishes of the genus *Brycon*. *Genome* 44: 893–902. PMID: [11681614](#)
31. Lucena CAS, Calegari BB, Pereira EHL, Dallegrave E (2013) O uso de óleo de cravo na eutanásia de peixes. *Bol Soc Bras Ictiol* 105: 20–24.
32. Bertollo LAC, Takahashi C, Moreira-Filho O (1978) Cytotaxonomic considerations on *Hoplias lacerdae* (Pisces, Erythrinidae). *Braz J Genet* 1: 103–120.
33. Levan A, Fredga K, Sandberg AA (1964) Nomenclature for centromeric position on chromosomes. *Hereditas* 1: 201–220.
34. Boyce TM, Zwick ME, Aquadro CF (1989) Mitochondrial DNA in the bark weevils: size, structure and heteroplasmy. *Genetics* 123: 825–836. PMID: [2612897](#)
35. Ward RD, Zemlak TS, Innes BH, Last PR, Hebert PDN (2005) DNA barcoding Australia's fish species. *Philos T Roy Soc B* 360: 1847–1857.
36. Ivanova NV, Zemlak TS, Hanner RH, Hebert PDN (2007) Universal primer cocktails for fish DNA barcoding. *Mol Ecol Notes* 7:544–548.
37. Palumbi SR (1996) Nucleic acids II: the polymerase chain reaction. In: Hillis D, Moritz C, Mable B, editor. *Molecular Systematics*. Sunderland: USA.
38. Li CG, Orti G, Zhang & Lu G (2007) A practical approach to phylogenomics: The phylogeny of ray-finned fish (Actinopterygii) as a case study. *BMC Evol Biol* 7: 44. PMID: [17374158](#)
39. Cooke GM and Beheregaray LB (2007) Extremely high variability in the S72 intron of the Amazonian cardinal tetra (*Paracheirodon axelrodi*). *J Fish Biol* 71: 132–140.
40. Higgins D, Thompson J, Gibson T, Thompson JD, Higgins DG, Gibson TJ (1994) CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice. *Nucleic Acids Res* 22: 4673–4680. PMID: [7984417](#)
41. Edgar RC (2004) MUSCLE: a multiple sequence alignment method with reduced time and space complexity. *BMC Bioinformatics* 5:113. PMID: [15318951](#)
42. Tamura KPD, Peterson NSG, Nei M & Kumar S (2011) MEGA5: Molecular Evolutionary Genetics Analysis using maximum likelihood, evolutionary distance, and maximum parsimony Methods. *Mol Biol Evol* 28: 2731–2739. doi: [10.1093/molbev/msr121](#) PMID: [21546353](#)
43. Lanfear R, Calcott B, Ho SYW, Guindon S (2012) PartitionFinder: Combined Selection of Partitioning Schemes and Substitution Models for Phylogenetic Analyses. *Mol Biol Evol* 29: 1695–1701. doi: [10.1093/molbev/mss020](#) PMID: [22319168](#)
44. Ronquist F, Teslenko M., Van der Mark P, Ayres DL, Darling A, Höhna S, Larget B, Liu L, Suchard MA, Huelsenbeck JP (2012) MrBayes 3.2: efficient Bayesian phylogenetic inference and model choice across a large model space. *Syst Biol* 61: 539–542. doi: [10.1093/sysbio/sys029](#) PMID: [22357727](#)
45. Rambaut A & Drummond AJ (2009) Tracer v1.5.0 <http://beast.bio.ed.ac.uk/Tracer>.
46. Stamatakis A (2014) "RAxML Version 8: A tool for Phylogenetic Analysis and Post-Analysis of Large Phylogenies". In *Bioinformatics*, open access link: <http://bioinformatics.oxfordjournals.org/content/early/2014/01/21/bioinformatics.btu033.abstract?keytype=ref&ijkey=VTEggUJYCDcf0kP>.

47. Miller MA, Pfeiffer W, Schwartz T (2010) "Creating the CIPRES Science Gateway for inference of large phylogenetic trees" in Proceedings of the Gateway Computing Environments Workshop (GCE), 14 Nov. 2010, New Orleans, LA pp 1–8.
48. Swofford DL (2003) PAUP\*: phylogenetic analysis using parsimony (\*and other methods). Version 4. Sunderland, Massachusetts: Sinauer Associates
49. Felsenstein J (1985) Confidence limits on phylogenies: an approach using the bootstrap. *Evolution* 39: 783–791.
50. Zander RH (2004) Minimal Values for Reliability of Bootstrap and Jackknife Proportions, Decay Index, and Bayesian Posterior Probability. *Phylo Informatics* 2:1–13.
51. Parada S, Arias JA, Cruz PE (2003) Caracterización cariotípica del yamu (*Brycon siebenthalae*). *Rev Orinoquia* 7: 42–46.
52. Castro RMC, Vari RP, Vieira F, Oliveira C (2004) A phylogenetic analysis and redescription of the genus *Henochilus* (Characiformes, Characidae). *Copeia* 3: 496–506.
53. Souza IL, Santos-Silva LK, Venere Paulo César, Moreira-Filho O (2008) Molecular cytogenetics of *Salmminus* fish (Characiformes) based on 5S and 18S rRNA genes hybridization, fluorochrome staining and C-banding. *Micron* 39: 1036–1041. PMID: [17988879](#)
54. Noor MAF, Grams KL, Bertucci LA, Reiland J (2001) Chromosomal inversions and the reproductive isolation of species. *PNAS* 98:21.
55. White BJ, Cheng C, Sangaré D, Lobo NF, Collins FH, Besansky NJ (2009) The population genomics of trans-specific inversion polymorphisms in *Anopheles gambiae*. *Genetics* 183: 275–288. doi: [10.1534/genetics.109.105817](#) PMID: [19581444](#)
56. Sessions SK (2008) Evolutionary cytogenetics in salamanders. *Chromosome Res* 16:183–201. doi: [10.1007/s10577-007-1205-3](#) PMID: [18293112](#)
57. Carvalho ML, Oliveira C, Navarrete ML, Froehlich O, Foresti F (2002) Nuclear DNA content determination in Characiformes fish (Teleostei, Ostariophysi) from the Neotropical region. *Genet Mol Biol* 25: 49–55.
58. Cioffi MB, Kejnovský E, Marquioni V, Poltronieri J, Molina WF, Diniz D, Bertollo LAC (2012) The key role of repeated DNAs in sex chromosome evolution in two fish species with ZW sex chromosome system. *Mol Cytogenet* 5: 28. doi: [10.1186/1755-8166-5-28](#) PMID: [22658074](#)
59. Oliveira C, Nirchio M, Granado A, Levy S (2003) Karyotypic characterization of *Prochilodus mariae*, *Semaprochilodus kneri* and *S. laticeps* (Teleostei: Prochilodontidae) from Caicara del Orinoco, Venezuela. *Neotrop Ichthyol* 1: 47–52.
60. Vicari MR, Almeida MC, Bertollo LAC, Moreira-Filho O, Artoni RF (2006). Cytogenetic analysis and chromosomal characteristics of the polymorphic 18S rDNA in the fish *Prochilodus lineatus* (Characiformes, Prochilodontidae). *Genet Mol Biol* 29: 621–625.
61. Voltolin TA, Senhorini JA, Oliveira C, Foresti F, Bortolozzi J, Porto-Foresti Fabio (2010) B-chromosome frequency stability in *Prochilodus lineatus* (Characiformes, Prochilodontidae). *Genetica* 138: 281–284. doi: [10.1007/s10709-009-9420-9](#) PMID: [19882308](#)
62. De Rosa LVS, Foresti F, Martins C, Oliveira C, Wasko AP (2007) Cytogenetic analyses of two Curimatidae species (Pisces, Characiformes) from the Paranapanema and Tietê Rivers. *Braz J Biol* 67: 333–338. PMID: [17876445](#)
63. Artoni RF, Shibatta AO, Gross MC, Schneider CH, Almeida MC, Vicari MR, Bertollo LAC (2006) *Astyanax* aff. *Fasciatus* Cuvier, 1819 (Teleostei, Characidae): evidences of a species complex in the upper rio Tibagi basin (Paraná, Brazil). *Neotrop Ichthyol* 4: 1997–2002.
64. Bertollo LAC, Born GG, Dergam JA, Fenocchio AS, Moreira-Filho O (2000) A biodiversity approach in the Neotropical Erythrinidae fish, *Hoplias malabaricus*. Karyotypic survey, geographic distribution of cytotypes and cytotaxonomic considerations. *Chromosome Res* 8: 603–613. PMID: [11117356](#)
65. Carvalho TP (2007) Distributional patterns of freshwater fishes in coastal Atlantic drainages of eastern Brazil: a preliminary study applying parsimony analysis of endemism. *Darwiniana* 45: 65–67.
66. Menezes NA (1988) Implications of the distribution patterns of the species of *Oligosarcus* (Teleostei, Characidae) from central and southern South America. Proceedings of a Workshop on Neotropical Distribution Patterns (ed. by W.R. Heyer and P.E. Vanzolini), pp. 295–304. Academia Brasileira de Ciências, Rio de Janeiro, Brazil.
67. Abell R, Thieme ML, Revenga C, et al. (2008) Freshwater ecoregions of the world: a new map of biogeographic units for freshwater biodiversity conservation. *Bio Science* 58: 403–414.
68. Buckup PA (2011) The eastern Brazilian Shield. Historical biogeography of Neotropical freshwater fishes (ed. by Albert J.S. and Reis R.E.), pp. 203–210. University of California Press, Berkeley, CA.

69. Albert JS, Petry P, Reis RE (2011) Major Biogeographic and Phylogenetic Patterns, (ed. by Albert J.S. and Reis R.E.) pp. 45–46. University of California Press, Berkeley, CA.
70. Kowsmann RO & Costa MPA (1979) Sedimentação quaternária da margem continental brasileira e das áreas oceânicas adjacentes. Série do Projeto REMAC. Rio de Janeiro, PETROBRÁS/CENPES/DINTEP. Série do Projeto REMAC, v.8, 55p.