Influence of estrous cycle and gestation on cardiovascular system of bitches


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ABSTRACT

The purpose of this study was to evaluate the consequences of the estrous cycle and gestation on the cardiovascular system of bitches evaluated through echocardiographic assessment. Fifteen healthy pregnant bitches aged 2 to 6 years and weighing 8.8 ± 2.5 kg were included in the study. Heart rate and blood pressure were clinically evaluated. M-mode, spectral, and tissue Doppler-echocardiography assessments were performed during the anestrous and estrous stages, every 15 days during pregnancy (15, 30, 45, and 60 days), and 45 days postpartum. The data for evaluated variables were submitted for statistical analysis. Heart rate (P = 0.286) and systolic blood pressure (P = 0.686) were not changed as a result of pregnancy or estrus, while ejection fraction (P = 0.0016) and fractional shortening (P = 0.00004) changed. The variables transmural flow, isovolumetric relaxation time, aortic and pulmonary flow, and some variables related to myocardium movement obtained via tissue Doppler assessment varied (P < 0.05) among the reproductive periods studied. In conclusion, gestation resulted in changes in echocardiography variables, and consequently, on maternal hemodynamics, with myocardial contractility increasing during pregnancy, resulting in greater systolic efficiency and ventricular compliance.

1. Introduction

During gestation, the formation of a uteroplacental vascular bed promotes an increase in the compliance of blood vessels (Nautrup, 1998; Di Salvo et al., 2006; Blanco et al., 2009), with a consequent reduction in peripheral vascular resistance (PVR) and cardiac output (CO). These changes, associated with adaptations in the renin-angiotensin-aldosterone system, are responsible for the gestational plasma volume that culminates in volume overload. The heart adapts to these changed circulatory demands by becoming hypertrophic. Stroke volume increases, maximizing the supply of nutrients to tissues (Gilson et al., 1992; Blanco et al., 2011) so that the requirements for fetal development are met (Blanco et al., 2008; Egbbali et al., 2005).

Characterization of the cardiovascular system adaptations in pregnant bitches has been an object of studies in veterinary medicine, revealing the importance of these variables to myocardial contractility (systolic function) and cardiac output (Blanco et al.,...
pregnancy (15, 30, 45, and 60 days), and 45 days postpartum. To determine the phases of the estrous cycle, cytological evaluation
through echocardiographic examination.

2. Material and methods

This study was approved by institutional Animal Ethics and Welfare Committee (protocol nº 023950/14). Fifteen healthy bitches
of different breeds, 2 to 6 years and weighing 8.8 ± 2.5 kg were used and evaluated while in anestrus, estrus, every 15 days during
pregnancy (15, 30, 45, and 60 days), and 45 days postpartum. To determine the phases of the estrous cycle, cytological evaluation
was conducted (Feldman and Nelson 2003).

Gestational diagnosis was made at approximately 15 days of gestation by gestational vesicle detection, considering the first
day from the natural mating as Day 0. An Acuson S2000® (Siemens®, Germany) with a multi-frequency linear transducer (9.0 MHz) was
used for these observations. The gestational stage was confirmed according to the method described by Feliciano et al. (2015).

2.1. Heart rate systolic blood pressure

Heart rate was measured by thoracic auscultation and systolic blood pressure (SBP; mmHg) was measured in triplicate using the
Doppler method (Dv10 PLX/Microem®). The bitches were maintained in right-side recumbency, an appropriate cuff (size 40 · 50% of
limb circumference) was placed around the left thoracic limb above the carpus joint, and the Doppler sensor was positioned over the
medial metacarpal artery (Mucha, 2001).

2.2. Echocardiography

The M-Mode, Doppler, and tissue Doppler echocardiography, were performed with the bitches in lateral recumbency using the
ultrasonic device Acuson S2000. Measurements were taken according to Boon, (1998) over three consecutive estrous cycles and were
analyzed by one trained expert.

The interventricular septum thickness in diastole (IVSr; mm) and in systole (IVSs; mm), left ventricular end diastolic (LVDd; mm)
and end systolic (LVDs; mm) diameter, left ventricular free wall thickness in diastole (LVFWd; mm) and in systole (LVFWs; mm),
ejection (EF), and shortening (FS) fractions, were measured in M-mode. These measurements were taken from the short axis of the left
ventricle in the cordal plane, with the cursor aligned perpendicular to the interventricular septum and equidistant from the papillary
muscles. Calculations of the systolic function were obtained using the Teichholz technique.

On a short axis, measurement of the relationship between the diameters of the left atrium and the aortic diameter (LA:Ao) were
assessed using cardiac base imaging. In a plane directed to the cardiac base, the peak velocity of the pulmonary valve (PVVmáx) was
measured, with the sampling volume positioned next to the pulmonary leaflets on the face opposite the pulmonary artery.

The five-chamber apical image was obtained through the left parasternal window. The cursor was aligned parallel to the aortic
flow, with the sampling volume positioned next to the aortic leaflets on the face opposite to the aorta. From this, the peak velocity of
the aortic valve (AVVmax) was measured.

The pulsed Doppler mitral flow velocity curves were used for analysis of diastolic function, as was the study of the mitral (septal
and lateral) flow in an apical four-chamber image by tissue Doppler assessment (Fig. 1). To obtain the mitral flow, the volume sample
pulsed Doppler was placed at the height of the commissural edges of the cusps with the mitral valve being open. The filter and gain
were adjusted to the minimum possible values that were compatible with obtaining an adequate image of the flow spectrum. With
this curve, peak velocities were obtained of the E wave (E), corresponding to the early peak velocity of the trans-mitral flow, and of
the A wave (A), corresponding to the late peak velocity of the trans-mitral flow; thus obtaining the ratio of E:A.

For measurement of the isovolumetric relaxation time (IVRT), the cursor of the pulsed Doppler was positioned equidistant be-
tween the outflow of the left ventricle and the trans-mitral flow, determining the time from the end of the trans-aortic flow to the
beginning of rapid ventricular filling (E). The tissue Doppler evaluation was performed with a volume sample positioned on the septal
(s) and lateral (lat) margins of the mitral annulus, allowing measurement of the peak velocity of the left myocardial wall during
systole (S). Early mitral annular diastolic movement - the rapid filling phase (E'), late mitral annular diastolic movement - the atrial
contraction phase (A'), and the E:A’ and E:E’ ratios were calculated. These were considered for the analysis of velocity measurements
for each margin (septal and lateral).
2.3. Statistical analysis

The R software was used for statistical analysis. Experimental design constituted blocks (bitches) randomized with subdivided parcel times (periods). Residual normality and homoscedasticity of variances were previously tested. Measures were compared between periods by repeated ANOVA, if results were significant, means were compared by Tukey test and polynomial contrasts (linear, quadratic and cubic). For variables where there was not a normal distribution, the Friedman test and Dunns post-test were applied. The level of significance was set at $P = 0.05$.

3. Results

The mean gestational period was 61.0 ± 2.5 days, and the litter sizes ranged from 4 to 6 healthy pups. The bitches did not have any gestational or fetal delivery complications. The mean heart rate was 124.9 ± 26.6 beats per minute and SBP 127.5 ± 15.1 mm Hg and there were no differences in the values for these variables during the estrous cycle, gestation or postpartum ($P = 0.2860$ and $P = 0.6860$, respectively), and values were consistent with values previously reported for this species during gestation (Fig. 2).

For IVSd, IVSs, LVDd, LVDs, LVFWd, LVFWs, and LA:Ao, there was no variations ($P > 0.05$) in the values for these variables during the periods evaluated (Table 1). The values for measurements of the left ventricular chamber for all bitches were within the reference standard range when average body weights of 8.8 ± 2.5 kg were considered.

The EF increased by 15% and FS by 20% from the 30th day of gestation, with the peak value occurring at 45th and 60th day of gestation, respectively ($P < 0.001$). When the values during gestation were compared with those during estrus and anestrus, the effects on these variables can be explained by linear regression models ($R^2 = 0.65$ and $R^2 = 0.68$, respectively). After 45 days from the day of parturition, only the FS returned to the values recorded in anestrus (Fig. 2).

When the values for the variables E, E:IVRT and E:A, E:E’ (lat) were assessed there were no differences in the variations ($P > 0.05$) during the reproductive periods evaluated. The data for variables A, E:A, IVRT, PVVmáx, AVVmáx, E’ (s) and E’ (lat), A’ (s) and A’ (lat), S’ (s) and S’ (lat), and E:E’ (s), however, varied when values during the various reproductive periods were assessed ($P < 0.05$; Table 2).

The A wave of the transmitral flow gradually increased until the 45th day of gestation, when this value was 80% greater than the value when bitches were in anestrus. Subsequently, values decreased to those that were similar to those during estrus, however, remained greater during postpartum period than when bitches were in anestrus., This effect was explained by a quadratic regression model ($R^2 = 0.83$; Fig. 2).

$$EF = 62.5 + 0.16 \times \text{gestational day}$$

$$FS = 31.9 + 0.14 \times \text{gestational day}$$

When the values for the variables E, E:IVRT and E:A, E:E’ (lat) were assessed there were no differences in the variations ($P > 0.05$) during the gestation period evaluated. The data for variables A, E:A, IVRT, PVVmáx, AVVmáx, E’ (s) and E’ (lat), A’ (s) and A’ (lat), S’ (s) and S’ (lat), and E:E’ (s), however, varied when values during the various reproductive periods were assessed ($P < 0.05$; Table 2).

The A wave of the transmitral flow gradually increased until the 45th day of gestation, when this value was 80% greater than the value when bitches were in anestrus. Subsequently, values decreased to those that were similar to those during estrus, however, remained greater during postpartum period than when bitches were in anestrus., This effect was explained by a quadratic regression model ($R^2 = 0.83$; Fig. 2).

$$A = 0.7 - 0.0009 \times \text{gestational day} + 0.00002 \times \text{gestational day}^2$$

The E:A ratio decreased by 50% during gestation to the 45th day of gestation when compared to the ratio during anestrus. There was a subsequent increase in the ratio anestrus during postpartum period to those that were present during anestrus with this effect being explained by a cubic regression model ($R^2 = 0.78$; $P = 0.001$; Fig. 2).

$$E: A = 1.6 + 0.02 \times \text{gestational day} - 0.001 \times \text{gestational day}^2 + 0.00001 \times \text{gestational day}^3$$

The IVRT was greater during anestrus, when compared to the measurements during estrus and the postpartum period. The IVRT increased during the first 15 days of gestation (10%) and there was a subsequent decrease in values to where these were less than those during anestrus at the 45th and 60th days of gestation (15%). This effect was explained by a cubic regression model ($R^2 = 0.38$;
Fig. 2. Graphical representation of: A1) heart rate (HR); and B1) systolic blood pressure (SBP). A2) ejection fraction (EF); and B2) fractional shortening (FS). A3) late peak velocity of the trans-mitral flow (A); and B3) ratio of the early peak velocity of the trans-mitral flow: A (E: A). Evaluated in bitches during anestrus (AZ), estrus (EZ), pregnancy days (PD) 15, 30, 45 and 60, and 45 days postpartum (PP). Similar letters are indicative of a lack of differences ($P > 0.05$) and the red line indicates the regression model that estimates the changes in these variables during gestation.
Similar letters are indicative of a lack of difference (P > 0.05).

Table 1
Mean values and standard deviation of B-mode and M-mode variables during anestrus (AZ), estrus (EZ), pregnancy days (PD) 15.30.45.60 days, and postpartum (45 days PP) in bitches

<table>
<thead>
<tr>
<th>Variables</th>
<th>Anestrus</th>
<th>Estrus</th>
<th>15 (PD)</th>
<th>30 (PD)</th>
<th>45 (PD)</th>
<th>60 (PD)</th>
<th>45 (PP)</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>IVSs (mm)</td>
<td>9.2 ± 1.9</td>
<td>11.2 ± 2.0</td>
<td>10.7 ± 2.3</td>
<td>10.5 ± 1.6</td>
<td>11.1 ± 1.9</td>
<td>9.9 ± 1.8</td>
<td>10.1 ± 1.7</td>
<td>0.073</td>
</tr>
<tr>
<td>IVSd (mm)</td>
<td>6.9 ± 0.8</td>
<td>8.2 ± 0.7</td>
<td>7.9 ± 1.1</td>
<td>8 ± 1.4</td>
<td>8.5 ± 1.6</td>
<td>7.8 ± 2.1</td>
<td>7.7 ± 1.5</td>
<td>0.365</td>
</tr>
<tr>
<td>LVDs (mm)</td>
<td>14.3 ± 1.6</td>
<td>10.7 ± 2.3</td>
<td>16.6 ± 3.2</td>
<td>17.1 ± 2.6</td>
<td>17.1 ± 3.9</td>
<td>15.3 ± 3.2</td>
<td>15.3 ± 3.1</td>
<td>0.131</td>
</tr>
<tr>
<td>LVDd (mm)</td>
<td>21.9 ± 3.6</td>
<td>25.5 ± 2.8</td>
<td>25.1 ± 4.1</td>
<td>26.54 ± 4.1</td>
<td>27.1 ± 4.6</td>
<td>26 ± 3.8</td>
<td>25.1 ± 5.8</td>
<td>0.118</td>
</tr>
<tr>
<td>LVFWs (mm)</td>
<td>9.9 ± 1.8</td>
<td>10.6 ± 2.3</td>
<td>11.0 ± 2.5</td>
<td>10.5 ± 2.3</td>
<td>10.8 ± 1.8</td>
<td>9.8 ± 1.5</td>
<td>10.5 ± 1.8</td>
<td>0.590</td>
</tr>
<tr>
<td>LVFWd (mm)</td>
<td>7.8 ± 1.5</td>
<td>7.5 ± 1.3</td>
<td>8.1 ± 2.0</td>
<td>7.6 ± 1.7</td>
<td>7.8 ± 1.8</td>
<td>6.6 ± 1.1</td>
<td>7.4 ± 1.4</td>
<td>0.222</td>
</tr>
<tr>
<td>EF (%)</td>
<td>65.3 ± 7.8b</td>
<td>64.2 ± 10.1b</td>
<td>63.7 ± 7.8b</td>
<td>66.5 ± 4.6b</td>
<td>70.2 ± 7.7ab</td>
<td>72.7 ± 6.2a</td>
<td>72.1 ± 5.6a</td>
<td>0.001</td>
</tr>
<tr>
<td>FS (%)</td>
<td>34.1 ± 6.3sc</td>
<td>34.3 ± 7.8c</td>
<td>32.8 ± 5.5c</td>
<td>35.4 ± 3.6bc</td>
<td>38.6 ± 6.0abc</td>
<td>40.3 ± 4.9a</td>
<td>39.9 ± 5ab</td>
<td>0.001</td>
</tr>
<tr>
<td>LA: Ao (mm)</td>
<td>1.3 ± 0.2</td>
<td>1.3 ± 0.3</td>
<td>1.3 ± 0.2</td>
<td>1.3 ± 0.1</td>
<td>1.3 ± 0.2</td>
<td>1.3 ± 0.2</td>
<td>1.3 ± 0.2</td>
<td>0.941</td>
</tr>
</tbody>
</table>

IVSs, interventricular septum in systole; IVSd, interventricular septum in diastole; LVDs, left ventricular end systolic diameter; LVDd, left ventricular end diastolic diameter; LVFWs, left ventricular free wall in systole; LVFWd, left ventricular free wall in diastole; EF, ejection fraction; FS, shortening fraction; LA: Ao left atrium: aortic root ratio. Similar letters are indicative of a lack of difference (P > 0.05).

Table 2
Values (mean ± SD or * median ± IQR) obtained by spectral and tissue Doppler during anestrus, estrus, pregnancy days (PD) 15, 30, 45, 60 days, and after delivery (45 Days PP) in bitches

<table>
<thead>
<tr>
<th>Variables</th>
<th>Anestrus</th>
<th>Estrus</th>
<th>15 (PD)</th>
<th>30 (PD)</th>
<th>45 (PD)</th>
<th>60 (PD)</th>
<th>45 (PP)</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>E (m/s)*</td>
<td>0.8 ± 0.2</td>
<td>0.9 ± 0.1</td>
<td>0.8 ± 0.2</td>
<td>0.8 ± 0.1</td>
<td>0.9 ± 0.2</td>
<td>1 ± 0.3</td>
<td>0.92 ± 0.2</td>
<td>0.137</td>
</tr>
<tr>
<td>A (m/s)</td>
<td>0.4 ± 0.04c</td>
<td>0.5 ± 0.1bc</td>
<td>0.6 ± 0.2ab</td>
<td>0.6 ± 0.3ab</td>
<td>0.8 ± 0.2a</td>
<td>0.7 ± 0.2ab</td>
<td>0.6 ± 0.1ab</td>
<td>0.001</td>
</tr>
<tr>
<td>E:A (m/s)*</td>
<td>2.1 ± 0.4b</td>
<td>1.7 ± 0.4b</td>
<td>1.5 ± 0.3a</td>
<td>1.4 ± 0.6b</td>
<td>1.3 ± 0.7c</td>
<td>1.4 ± 0.8b</td>
<td>1.6 ± 0.4ab</td>
<td>0.001</td>
</tr>
<tr>
<td>IVRT (ms)*</td>
<td>64.0 ± 10.5a</td>
<td>61.1 ± 11.8ab</td>
<td>71.2 ± 12.8ab</td>
<td>67.2 ± 13.5ab</td>
<td>57.2 ± 10.7c</td>
<td>55 ± 8.7c</td>
<td>61 ± 12.7ac</td>
<td>0.010</td>
</tr>
<tr>
<td>E:E' (s)</td>
<td>0.013 ± 0.01</td>
<td>0.01 ± 0.01</td>
<td>0.01 ± 0.01</td>
<td>0.01 ± 0.01</td>
<td>0.01 ± 0.01</td>
<td>0.01 ± 0.02</td>
<td>0.01 ± 0.001</td>
<td>0.309</td>
</tr>
<tr>
<td>PVVmáx</td>
<td>0.94 ± 0.21c</td>
<td>0.9 ± 0.3bc</td>
<td>0.9 ± 0.2d</td>
<td>1 ± 0.2d</td>
<td>1.1 ± 0.2a</td>
<td>1.1 ± 0.1ab</td>
<td>1.1 ± 0.2d</td>
<td>0.001</td>
</tr>
<tr>
<td>AVVmáx</td>
<td>1.2 ± 0.3a</td>
<td>1.2 ± 0.1ab</td>
<td>1.1 ± 0.3b</td>
<td>1.1 ± 0.2ab</td>
<td>1.3 ± 0.2ab</td>
<td>1.3 ± 0.4a</td>
<td>1.3 ± 0.4ab</td>
<td>0.004</td>
</tr>
</tbody>
</table>

E', early mitral annular diastolic movement; A', late mitral annular diastolic movement; E:A, ratio of peak velocity of the trans-mitral flow in early (E) and late (A); E:E', ratio of mitral annular diastolic velocity in early (E) and late (A); E:E, ratio of early peak velocity of the trans-mitral flow and early mitral annular diastolic movement; IVRT, isovolumetric relaxation time; E:IVRT*, ratio of early peak velocity of the trans-mitral flow and isovolumetric relaxation time; PVVmáx, peak velocity of the pulmonary valve; AVVmáx, peak velocity of the aortic valve; S', peak velocity of the left myocardial wall measured during systole (P < 0.05).

A, late peak velocity of the trans-mitral flow; A', late mitral annular diastolic movement; E, early peak velocity of the trans-mitral flow; E', early mitral annular diastolic movement; A', late mitral annular diastolic movement; E:A, ratio of peak velocity of the trans-mitral flow in early (E) and late (A); E:E', ratio of mitral annular diastolic velocity in early (E) and late (A); E:E, ratio of early peak velocity of the trans-mitral flow and early mitral annular diastolic movement; IVRT, isovolumetric relaxation time; E: IVRT*, ratio of early peak velocity of the trans-mitral flow and isovolumetric relaxation time; PVVmáx, peak velocity of the pulmonary valve; AVVmáx, peak velocity of the aortic valve; S', peak velocity of the left myocardial wall measured during systole (P < 0.05).

Similar letters are indicative of a lack of difference (P > 0.05).

P = 0.01; Fig. 3).

IVRT = 62.3 + 1.8 × gestational day – 0.08 × gestational day² + 0.0008 × gestational day³

The PVVmáx was similar during anestrus, estrus, and the postpartum period. During gestation, the PVVmáx increased to the 45th day (17%) and decreased slightly until the 60th day of gestation. This effect was explained by a cubic regression model (R² = 0.83; P = 0.001; Fig. 4).

PVVmáx = 0.95–0.01 × gestational day + 0.0006 × gestational day²–0.000006 × gestational day³

The AVVmáx was similar during anestrus, estrus, postpartum, and gestation, however, there was a slight increase on the 60th when compared to the 15th day of gestation (Fig 4). The E' (lat) and E' (s) were similar during anestrus, estrus, and the postpartum period (P > 0.05). The E' (lat) decreased gradually until the 30th day of gestation, and increased between the 45th and 60th day of gestation. This effect was explained by a cubic regression model (R² = 0.68; P = 0.036; Fig. 5).

E'lat = 0.12–0.0013 × gestational day + 0.0006 × gestational day²–0.000001 × gestational day³

The E' (s) increased gradually during gestation peaking on the 60th day of gestation (17%). This effect was explained by a linear
The A’ (s) was less (P = 0.031) during estrus than anestrus, and similar (P > 0.05) during the other reproductive phases that variables were assessed. The A’ (lat) was similar during estrus, anestrus, and the postpartum period, however, values increased (P = 0.003) to the 45th day of gestation (Fig. 5). The S’ (lat) was less during anestrus than the postpartum period, increasing significantly to the 45th day of gestation (30%), and subsequently decreasing gradually until the postpartum period. This effect was explained by a cubic regression model (R² = 0.70; P = 0.001; Fig. 5).

\[ E' = 0.1 + 0.0004 \times \text{gestational day} \]

The A’ (s) was less (P = 0.031) during estrus than anestrus, and similar (P > 0.05) during the other reproductive phases that variables were assessed. The A’ (lat) was similar during estrus, anestrus, and the postpartum period, however, values increased (P = 0.003) to the 45th day of gestation (Fig. 5). The S’ (lat) was less during anestrus than the postpartum period, increasing significantly to the 45th day of gestation (30%), and subsequently decreasing gradually until the postpartum period. This effect was explained by a cubic regression model (R² = 0.70; P = 0.001; Fig. 5).

\[ S' (\text{lat}) = 0.11 - 0.002 \times \text{gestational day} + 0.00007 \times \text{gestational day}^2 - 0.000001 \times \text{gestational day}^3 \]

The S’ (s) was similar during anestrus, estrus, and the postpartum period, and there was a decrease (P = 0.012) to the 15th and 30th day of gestation when compared to values obtained during anestrus followed by an increase that resulted in values at 45 days of gestation similar to that found in anestrus. (Fig. 5).

The E:E’ (s) was similar during anestrus, estrus, and the postpartum period. During gestation, the E:E’ (s) gradually decreased (P = 0.035) and the least values (12%) were observed on the 45th day of gestation (Fig. 6).

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**Fig. 3.** Graphical representation of isovolumetric relaxation time (IVRT) evaluated in bitches during anestrus (AZ), estrus (EZ), pregnancy days 15, 30, 45 and 60, and 45 days postpartum (PP). Similar letters are indicative of a lack of differences (P > 0.05) and the red line indicates the regression model that estimates the changes in values of these variables during gestation.

**Fig. 4.** Graphical representation of: A) peak velocity of the pulmonary valve (PVVmáx); B) peak velocity of the aortic valve (AVVmáx); evaluated in bitches during anestrus (AZ), estrus (EZ), pregnancy days 15, 30, 45 and 60, and 45 days postpartum (PP). Similar letters are indicative of a lack of differences (P > 0.05) and the red line indicates the regression model that estimates the values for these variables during gestation.
4. Discussion

The main cardiovascular adaptations during the gestational process in healthy bitches occurred in the last third of gestation, when compared with values during anestrus, estrus and the postpartum period. These changes probably occurred due to a more effective systolic performance and to an increase in ventricular compliance. These results are consistent with findings in previous studies (Abbott, 2010; Blanco et al., 2011; Blanco et al., 2012; Souza et al., 2017).

Heart Rate did not vary and these findings are consistent with those of Souza et al. (2017), where vagal tone was found to
downregulate the heart rate in response to an increase in the hemodynamic demand. This response occurs so the heart rate does not increase to harmful rates during gestation and cause damage to the cardiovascular system.

Variables relating to the thickness and diameter of the cardiac chambers were not different when compared among the reproductive stages evaluated in the present study, these findings are consistent with the results of Souza et al. (2017). In contrast in women was reported an physiological hypertrophy of the eccentric type as consequence of hemodynamic overload, when the myocardium adapts by Frank-Starling mechanism (Eghbali et al., 2005; Fok et al., 2006; Patey et al., 2017). Same results were related in previous studies in bitches (Abbott, 2010; Blanco et al., 2012) but without an significant increase in the thickness of the myocardial wall or karyometry analysis which allows them to reach this conclusion (Liberatori Filho et al., 1998). It is therefore inferred that, the absence of myocardial remodeling evidence in bitches is probably due to the shorter gestation length, in which structural changes of the myocardium do not reach the magnitude of those for humans.

The velocities of the pulmonary (PVVmáx) and aortic (AVVmáx) flows were greater during gestation as evidenced by those between the 45th and 60th days of gestation. These findings are consistent with an improvement in systolic function at the end of gestation that was previously reported by Abbott (2010). Another indicator of the improvement in the systolic function during this period was the increase in wave S' obtained using the tissue Doppler exam from the 30th to 60th days of gestation. This type of adaptation during the first third of gestation has been reported in women (Bamfo et al., 2007) and reflects contractility increases of the left ventricle which was probably influenced by preload augmentation.

Hemodynamic changes during gestation also affect left ventricular compliance. Structural realignment of myocardial tissue during this phase principally that of the collagen fibers may influence diastolic function. Although the trans-mitral flow analysis results indicated there was a decrease in the E:A ratio in the last third of gestation, the peak velocity of the A and A' waves increased during this same period due to a great influence of atrial contraction, which increases ventricular filling (Fok et al., 2006). Additionally, these waves also increased at the 45th day of gestation relating an increase in the diastolic filling rate and greater ventricular compliance.

The E:E' ratio, which is a marker of left ventricular diastolic pressure (Wu et al., 2008) was less between the 45th and 60th days of gestation, when ventricular relaxation became inadequate for compensation. During this time period, the atrial pressure increased, and the E wave and E:E' ratio increased (Boon, 2011). In the present study, therefore, the ventricle was more compliant, as verified by E' wave increase and E:E' ratio decrease. The E wave did not change gradually during gestation, but was greater on the 45th day of gestation, resulting in a lesser ratio during this period with a lesser ventricular pressure being present probably due to a more compliant ventricle.

The values for the IVRT were greater on the 15th of gestation, and this may be a result of a decrease in the ventricular relaxation (Boon, 2011), not necessarily a lesser diastolic filling, but a ventricle that was less compliant. The lesser values at the 45th and 60th days of gestation related an increase in ventricular compliance during the last third of gestation as result of associations with values for other variables.

In conclusion, the healthy gestational process in bitches results in changes in maternal hemodynamics as validated in the present study by associated changes in echocardiography variables when compared with to the other reproductive phases evaluated. During gestation, the myocardial contractility increases resulting in improved systolic efficiency and increased ventricular compliance. By understanding the normal physiological changes that occurred in the present study, it becomes possible to investigate issues involving the pathophysiology of heart disease and its repercussions on gestation.
Conflict of Interest

The authors declare that they have no competing interests.

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