RESEARCH ARTICLE



Climatic stability and contemporary human impacts affect the genetic diversity and conservation status of a tropical palm in the Atlantic Forest of Brazil

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Received: 10 May 2016 / Accepted: 23 December 2016 / Published online: 12 January 2017 © Springer Science+Business Media Dordrecht 2017

Abstract Understanding how historical and current environmental suitability and human impacts affect the genetic diversity on a large scale is essential to species management planning. However, most studies in conservation genetics are carried out at a local or regional scale and rarely on broad spatial scales such as an entire biome. We evaluated the relative contribution of historical and current environmental suitability, current landscape features and human impacts to explain genetic diversity, allelic richness and inbreeding variation among populations, using Euterpe edulis and the Atlantic forest as the model system. We fitted linear mixed models within a multiple competing hypotheses approach with model selection based on Akaike's Information Criteria. We showed that overall genetic diversity was lower in sites with absence of large seed dispersers and higher in sites with historically stable climate. Both seedling and adults showed to be negatively influenced by human impact factors; with adults mainly affected by the reduction of forest cover while seedlings by the loss of large seed dispersers. Thus, the current pattern

Electronic supplementary material The online version of this article (doi:10.1007/s10592-016-0921-7) contains supplementary material, which is available to authorized users.

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of genetic diversity in *E. edulis* is the result of historical instability during the mid-Holocene and recent anthropogenic impacts, mainly those that affect important ecological process such as seed dispersal. Thus, an efficient plan for species conservation must account for human impacts and environmental suitability and also assess the genetic diversity of seedlings and adults in fragmented landscapes.

Keywords Ecological niche modeling · Defaunation · Human impacts · Conservation genetics · *Euterpe edulis* · Atlantic Forest

Introduction

Understanding how past and current environmental suitability and human impact factors affect patterns of genetic diversity and biodiversity distribution is a challenge for both ecologists and biogeographers (Gaston 2000). The maintenance of genetic diversity in natural populations is essential for species management planning (Hughes et al. 2008), since species with low genetic diversity may be less likely to cope with environmental changes and therefore may become endangered or face extinction (Hoffmann and Willi 2008). Although genetic diversity has a fundamental role in species persistence and adaptation, few studies have reported the role of environmental suitability and human impacts in genetic diversity on a broad scale (Diniz-Filho et al. 2009; Ortego et al. 2012).

Several historical and contemporary factors may influence genetic diversity and inbreeding in natural populations. Historical factors related with climate and geological events may change the habitat suitability for species establishment or persistence and may therefore imprint a signal in the species genetic pool over time (Hewitt 2000).



The Quaternary climatic oscillation, for example, might have affected species genetic diversity due to the dynamics of population's expansion and contraction (Gugger et al. 2013). Population range contraction may decrease effective population size, leading to population bottlenecks and genetic drift, with the consequent loss of genetic diversity (Pauls et al. 2013). Loss of genetic diversity may also occur during population expansion into new habitats (Pauls et al. 2013) due to founder effects or recovery from population bottlenecks, since only a part of the original genetic diversity moves or persists into new habitats (Pauls et al. 2013). The colonization of new habitats during expansion can result in a high genetic admixture of immigrants arriving from different populations (Ortego et al. 2015), which could either increase genetic diversity due to heterosis or reduce genetic diversity if outbreeding depression occurs. Finally, sites with high environmental suitability and stable climate over time may preserve high genetic diversity in populations (Collevatti et al. 2014).

Contemporary factors that may affect genetic diversity include processes occurring at ecological scales and relate strongly with human activities (Miraldo et al. 2016). Global changes induced by human activities including climate change and conversion of land covers may affect species richness, abundance, and several key ecological processes (Fahrig 2003), such as pollination (González-Varo et al. 2009), seed dispersal (Uriarte et al. 2011) and animal movement (Eycott et al. 2012). Fragmentation and habitat loss are two factors identified as the main anthropogenic threats to genetic diversity (Lowe et al. 2005; DiBattista 2007; Aguilar et al. 2008). These anthropogenic threats may lead to a drastic reduction of population sizes, and consequently of effective population size (Frankham et al. 1999; Andersen et al. 2004). The probability of mating between closely related individuals may increase in small populations and, together with genetic drift, lead to a substantial reduction in genetic diversity (Breed et al. 2012). In these human-dominated scenarios, local extinction of medium- and large-body sized animals (i.e., defaunation; Dirzo and Miranda 1991) affects the composition and ecological functions of animal communities (Wright 2003). Seed dispersal is amongst the many ecological roles these animals provide to the community. Large-sized frugivores are important for long distance seed dispersal because they can move seeds hundreds of meters away from the mother plant. Therefore, these animals can promote gene flow and increase genetic diversity in plant populations (García et al. 2007; Jordano et al. 2007).

Although many studies focus separately on the effects of historical climate oscillation and human disturbances on genetic diversity, few evaluate the relative contribution of these historical and contemporary effects in the same system (Walter et al. 2012). Understanding how historical

events impact genetic diversity is important to predict whether populations are able or not to persist under future scenarios of environmental change (Pauls et al. 2013). In addition, evaluating the effects of contemporary factors like habitat fragmentation on genetic diversity helps identifying actions to prevent genetic erosion or to rescue genetic diversity of impacted populations. Therefore, quantifying the effects of historical and contemporary factors on species genetic pool may help design strategies for species and landscape conservation.

Large spatial and temporal scale studies are necessary to better understand how the historical and current factors shape genetic diversity across the distribution of natural populations and contribute to efficient species conservation planning (Huber et al. 2010). Large-scale studies can act as natural experiments to investigate historical climate effects on biodiversity because distinct sites may be differently affected by the oscillation in climate. Due to time lag effects, adult plants and seedlings may respond in a different way to human impacts (Van Geert et al. 2008). When estimated using microsatellite markers, the genetic diversity of adults of perennial plants often responds to past landscape conditions but not to recent habitat changes (Collevatti et al. 2001; Kramer et al. 2008). When evaluating ongoing processes, such as dispersal events culminating in gene flow, it is more appropriate to assess genetic variability in early recruitment stages (e.g., seed and seedlings in plants) rather than adults (Kramer et al. 2008).

The Atlantic forest biome has an extensive geographic range, presenting a wide variation in climate, topography and vegetation along its distribution (Ribeiro et al. 2011). It is also one of the most threatened ecosystems in South America, mainly due to anthropogenic disturbances (Rands et al. 2010; Tabarelli et al. 2010). Its current extension constitutes only 12% of its original 150 million hectares (Ribeiro et al. 2009). The heart of palm (Euterpe edulis, Arecaceae), a threatened Atlantic forest species, was once one of the dominant palms in this ecosystem (Henderson et al. 1997). The species is present in different vegetation types ranging from dense mountain rain forests to seasonally dry forests, where it is restricted to wet microhabitats. The edible meristem (heart of palm), obtained mainly by illegal exploitation, is used as source of raw material for food industries (Galetti and Fernandez 1998). Euterpe edulis is a monoic species with a predominantly outcrossed mating system with low amount of selfing (Gaiotto et al. 2003). Its flowers are mainly pollinated by small bees (Reis et al. 2000). The ripe fruits constitute an important food resource for animals, including more than 50 species of birds and mammals (Galetti et al. 2013). Several population genetics studies of E. edulis have been carried out at local or regional scales. Metadata combining the already published genetic diversity data may contribute to the



understanding of large-scale patterns (Lowe et al. 2005; DiBattista 2007) of *E. edulis*.

Here, we address the distribution of genetic diversity of E. edulis and evaluate whether past and current environmental variables and human-induced changes can explain variation among localities, using published data derived from microsatellite loci. First, we built ecological niche models for E. edulis in three different time periods (the Last Glacial Maximum, the mid-Holocene and the present-day) to understand how changes over time in habitat suitability for species establishment may have affected genetic diversity and inbreeding. Second, we evaluated the relative contribution of climatic habitat suitability in the past and in the present-day, together with current landscape features modified by human activities to explain the patterns in genetic diversity of E. edulis. We expect to find high genetic diversity in areas of high climatic stability and high habitat suitability. In addition, we expect to find stronger effects of human-modified environmental factors on seedlings than in adults, given that adults may have not had enough time to respond to the recent intensification of environmental changes caused by humans, mainly forest fragmentation and functional extinction of seed dispersers in the Atlantic Forest.

Methods

Literature search

We surveyed publications of population genetic studies on *E. edulis* using the following databases: Scielo (http://www.scielo.org), Web of Science (http://www.webofscience.bom), Scopus (http://www.scopus.com), and Google Scholar (http://www.scholar.google.com.br). The survey covered all publications available from 2001 to November 2015. We used the most frequent keywords in the area: "population genetics", "*E. edulis*", "microsatellites", "genetic diversity", "conservation genetics", and "heart of palm". We included only publications comprising populations from the northeast to southeast Brazilian Atlantic coast that used microsatellites as molecular markers (Fig. 1).

Genetic diversity response variables

Three genetic parameters were used as response variables in our analyses: the inbreeding coefficient (f); the genetic diversity, measured by expected heterozygosity (H_e) and allelic richness (AR). We chose these genetic parameters because they are present in most publications (Supplemental Information, Table A1) and are frequently used in

conservation genetics studies as surrogates for population evolutionary potential.

Predictive models

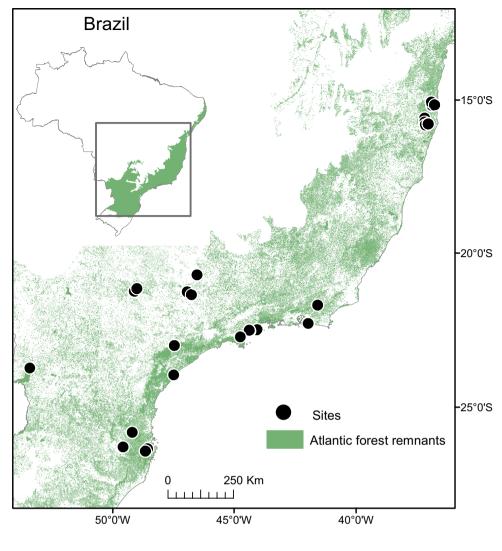
We used three groups of covariates to explain genetic diversity: (i) human-induced; (ii) climatic habitat suitability; and (iii) stability of climatic habitat suitability. Human-induced covariates were: (a) percentage of forest cover and (b) loss of large-body frugivores (i.e., defaunation). Climatic habitat suitability was measured through ecological niche modeling (ENM hereafter) for each of the time periods (the Last Glacial Maximum, the mid-Holocene and the present-day). We also measured the cumulative suitability as the sum of climatic habitat suitability over the three periods. The climatic stability was calculated as the difference between past and present climatic habitat suitability.

To quantify the covariates for modeling genetic diversity, we drew a buffer area with a 2 km radius around each of the 57 selected study sites from the surveyed literature. We chose this radius because the foraging distance of potential pollinators, such as *Plebleia droryana* and Trigona spinipes, can reach up to 540 and 840 m, respectively (Zurbuchen et al. 2010). Furthermore, seed dispersal distances by large-body bird species that are taxonomically related to the dispersers of E. edulis are likely to be shorter than 600 m (Holbrook 2011). We used the SOS Mata Atlântica map (2005, http://mapas.sosma.org.br/) and the ArcGIS 9.2 software (ENRI) to calculate the total area and the percentage of forest inside each 2 km-radius buffer area. Defaunation was defined as the absence or very low abundance of Ramphastidae (e.g., toucans and toucanets) and Cotingidae (e.g., bellbirds) species within the buffer area and was classified as a binary variable: defaunated (0) or non-defaunated (1). Species of these families are important long distance dispersers for E. edulis seeds (Holbrook 2011; Galetti et al. 2013). To assign the defaunation status of each site, we characterized the avian seed dispersal assemblage using the Wikiaves website (http://www.wikiaves.com.br) and available literature (Galetti et al. 2013; Morante-Filho et al. 2015).

To assess the climatic suitability of the study area we carried out ecological niche modeling of *E. edulis* combining species occurrences and climatic variables. A total of 229 occurrence records (Fig A1 in Supplementary material) were obtained from Species Link (http://www.splink.org.br), Jardim Botânico do Rio de Janeiro (http://www.jbrj.gov.br) and Biota Fapesp (http://www.biota.org.br/). All records were carefully checked for taxonomic reliability (i.e., synonymous and/or misidentification and duplicates). We carefully restricted our modeling area to encompass only Cerrado and Atlantic forest biomes (Fig. 2).



Fig. 1 Distribution of the 57 sampled sites with genetic information for *Euterpe edulis* across the Brazilian Atlantic forest biome in Brazil



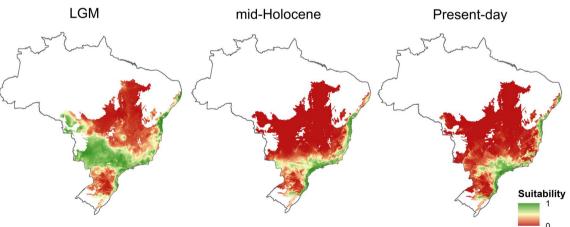


Fig. 2 Maps of mean climatic suitability across the ten ecological niche models (ENMs), five Atmosphere–Ocean General Circulation Models (AOGCMs) and three time periods (at the LGM, mid-Holocene and present, respectively) for *Euterpe edulis* in Brazil

The environmental space required for the ENMs was characterized by five bioclimatic variables: temperature seasonality, maximum temperature of the warmest month, precipitation of the wettest quarter, precipitation of the driest quarter, precipitation of the warmest quarter together with elevation and soil classes (http://www.embrapa.br).



Climatic variables were selected out of the 19 bioclimatic variables (Worldclim; http://worldclim.org; Hijmans et al. 2005) after performing a factor analysis with Varimax rotation to minimize collinearity among the variables when building the distribution models (For details on selection of variables see Supplementary material, Table A2). We derived distribution models using layers from the preindustrial period (representing current conditions; 0 ka), the mid-Holocene (6 ka) and the Last Glaciation Maxima (LGM, 21 ka). Current models were then projected onto three Atmosphere-Ocean General Circulation Models (AOGCMs; CCSM4, MIROC-ESM and MPI-ESM-P) obtained from Coupled Model Intercomparison Project Phase 5 (CMIP5; http://cmip-pcmdi.llnl.gov/cmip5/). We assumed soil and elevation as constraint variables (constant throughout time). All layers were used in a spatial resolution of 2.5 arc-min (ca. 5×5 km).

A total of 200 spatially independent localities were used for the models. ENM was carried out with biomod2 in R 3.1.2 (Thuiller et al. 2009). Six different algorithms were used (see Supplementary material for model parameterization): Artificial Neural Networks (ANN), Generalized Additive Models (GAM), Generalized Linear Models (GLM), Generalized Boosted Models (GBM), Random Forest (RF), and maximum entropy (MAXENT; Phillips and Dudík 2008). Usually ENM requires presence-absence data for model fitting and testing; however, there are not reliable absences available. Therefore, we chose to generate pseudo-absences randomly across the whole area (Ferrier et al. 2002), excluding cells with presences to ensure a complete sampling of the environmental space and minimum extrapolation when projecting to different historical period. To maximize model performance, pseudo-absences were weighted to ensure maintaining prevalence equal to 0.5 (Barbet-Massin et al. 2012). We used random data partitioning (80% training, 20% testing) and five repetitions for each model to assess model stability due to the absence of independent testing data. We assessed model accuracy through the calculation of the True Skill Statistic-TSS (Allouche et al. 2006) and the area under the receiver curve-AUC (Fielding and Bell 1997) (Table A3 in Supplementary material). Since results might differ among different algorithms, we performed an ensemble modeling (Araújo and New 2007) to calculate a consensus map that included only models with a TSS>0.7 and an AUC>0.75 and the resulting map was weighted by the individual model TSS value. Thus we generated a surface representing the suitability values for the occurrence of the species across our study area for both current and past climatic periods.

To access climatic stability we calculated the difference between the past (6 and 21 ka) and current (0 ka) suitability within the 2 km radius buffers for each site. Positive values indicate that current climatic suitability is lower than in the past (contraction population range) and negative values show that the current climatic suitability is higher than in the past (expansion population range). Zero value indicates that the site is climatically stable, which could either be associated with high environmental suitability or not. To account for the cumulative suitability over time, we also calculated the sum of suitability in the three different time periods.

Statistical analyses

We investigated the effects of the explanatory variables (i.e., defaunation status, percentage of forest cover, climatic habitat suitability in the present-day, cumulative climatic habitat suitability, stability from the mid-Holecene to present-day and stability from the LGM to present-day) on the three genetic parameters (i.e., f; H_{ρ} , AR). We fitted full models only for adults, only for seedlings and adults and seedlings combined for each genetic parameter using a linear mixed effects and the maximum-likelihood method using the R package lme4 (Bates et al. 2015), which is an adequate way to assess the influence of fixed effects given random structure. Defaunation status, percentage of forest cover, current climatic habitat suitability, cumulative climatic habitat suitability, climatic stability from the mid-Holocene to present-day and climatic stability from the LGM to present-day were considered as fixed effects. Ontogenetic stage (adults or seedlings) and "study" were included as random effects while sample size of each site was included as a weight parameter in all models. The "study" was included to take into account the difference in the set of microsatellites markers and in the number of populations and individuals sampled (Table A1). We assumed that these factors might influence the estimation of genetic parameters. The ontogenetic stage (adults and seedlings) was included as random effects only in the model that included all individuals. After fitting the full model, we fitted nested models with all possible combinations of the fixed effects.

Because the effect of historical and current factors on genetic variables tends to be complex and dependent on the interaction of diverse factors (Boscolo and Metzger 2011), we used a model selection with multiple competing hypotheses approach (Burnham and Anderson 2002). To select the best model, we used the Akaike Information Criteria (AIC), where the model with the lowest AICc (AIC corrected by sample size and number of parameters) (Burnham and Anderson 2002) was considered as the more plausible to explain observed patterns. Delta AICci (Δ AICci, where i represents each model) was calculated as the difference between AICc for the model i and the smallest observed AICc. We determined Akaike's weight



of evidence (wAICc) representing the relative contribution of model i to explain the observed pattern, given a set of competing models (Burnham and Anderson 2002). Models with Δ AICc<2 were considered as equally plausible to explain the observed pattern (Burnham and Anderson 2002). We also carried out a model averaging approach to compute the parameter average from all equivalent models (Δ AICc<2) and the significance of the estimated parameter values associated with each explanatory variable (Burnham and Anderson 2002). All analyses were performed using the *MuMIn* packages implemented in R version 3.0.2 (Barton 2016). We also analyzed if seedlings have on average lower levels of genetic diversity than adult populations.

To account for the presence of spatial autocorrelation, we ran linear mixed models with the restricted maximum-likelihood method with different spatial covariance structures (Gaussian, exponential and spherical) and compared them to a model without a spatial covariance structure. We ran these models by fitting all explanatory covariates as *fixed effects* and the study and ontogenetic stage as *random effects*.

Results

Literature search

We obtained eight publications, which accounted for 61 study sites. However, four sites were excluded from the analysis because they were outside of the Atlantic Forest biome; in the end, we used 57 sites (Fig. 1 and Supplemental Information Table A1). Most populations of E. edulis in the Atlantic forest biome showed high genetic diversity (Supplemental Information Table A1). H_e ranged from 0.57 to 0.86, and AR varied from 3.6 to 15.1. However, many populations showed high f value (Supplemental Information Table A1).

Climatic suitability and stability models

A total of 30 models were generated by the ENMs. From those, 83% were considered good under AUC criteria (Mean AUC, Table S3). The map of the modeled distribution under present conditions mostly agrees with the known current distribution for the species. The consensus map reveals suitable areas for the occurrence of the species across the coastal part of Brazil, from Pernambuco state in the northeast to Santa Catarina state in the south (Fig. 2). The region of higher climatic suitability coincides with the core area of Atlantic rain forests (Fig. 2) and shows the preference of *E. edulis* for hot and humid climates. Hindcasting to the LGM and the mid-Holocene showed a retraction of suitable areas through time, especially in areas

across central Brazil. The differences between past and current suitability values show a strong variation along the geographic distribution of the Atlantic Forest, with a reduction in habitat suitability in central Brazil during the mid-Holocene (i.e., retraction) and high stability and suitability since the LGM in areas along the coastal part of Brazil (Fig. 2; Table A1).

Effects of historical and current factors on genetic diversity

The results of the model selection indicated that models without a spatial covariance structure were considerably better for modeling H_e , AR and f than those with spatial covariance structures (Gaussian, exponential and spherical) (Supplementary material Table A4). Considering all individuals, variation in H_e and AR were best explained by the lack of large seed dispersers (Table 1). Defaunated sites presented lower H_e and AR than non-defaunated sites (Fig. 3a, b). The inbreeding coefficient (f) was best explained by climatic stability from the mid-Holocene to present-day (Table 1). The inbreeding coefficient was lower in areas with decreasing suitability values over time (habitat contraction) and was higher in areas under expansion (Fig. 3c).

We found that seedlings have on average lower levels of allelic richness than adult individuals (t=-2.49,p=0.01), however we did not find differences in H_a and f between ontogenetic stages (t=-0.03, p=0.97; t=-1.5, p=0.12, respectively). Considering only adults, variation of AR could not be explained by any of our competing models (Table 1). On the other hand, H_e was explained by climatic stability from the mid-Holocene to present-day (Table 1). Lower values of H_{ρ} were associated with populations in areas where climatic suitability decreased from the mid-Holocene to present-day (Fig. 3d). Nevertheless, the genetic diversity (H_e) remained high in areas under expansion or that were consistently suitable over time (Fig. 3d). Variation of f was best explained by the percentage of forest cover and by climatic stability from the mid-Holocene to present-day (Table 1). Populations in landscapes with higher percentage of forest cover showed lower f than populations in landscapes with lower forest cover (Fig. 3e). The model also shows that there is a decrease in the inbreeding coefficient when there is a retraction from suitable areas through time (positive climatic stability, Fig. 3f). Likewise, the inbreeding coefficient (f) increases in areas undergoing expansion over time (Fig. 3f). Considering only seedlings, variation in H_e was not explained by any of our competing models (Table 1); however, f was best explained by the climatic stability from the mid-Holocene to present-day (Table 1), as found for adults (Fig. 3g). Finally, defaunation



Table 1 Summary statistics for the model-averaging used to explain the genetic diversity pattern in populations of *Euterpe edulis* from 57 sites, distributed along the Brazilian Atlantic Rainforest

Parameter	H_e		f		AR	
	Model averaged (SE)	p-value	Model averaged (SE)	p-value	Model averaged (SE)	p-value
All individuals						
Defaunation*	0.29 (0.13)	0.03	_	_	0.24 (0.08)	0.00
% Forest cover	_	_	-0.13 (0.09)	0.18	-0.05 (0.05)	0.28
Current climatic suitability	-0.02 (0.23)	0.93	-0.12 (0.08)	0.17	-0.22 (0.11)	0.06
Climatic stability Holocene-present*	_	_	-0.29 (0.14)	0.04	_	_
Climatic stability LGM-present	0.15 (0.09)	0.10	-0.11 (0.11)	0.31	_	_
Cumulative suitability	0.22 (0.14)	0.12	-0.09 (0.08)	0.29	0.27 (0.16)	0.09
Adults						
Defaunation	0.48 (0.26)	0.09	_	_	0.28 (0.19)	0.17
% Forest cover*	_	_	-0.57 (0.14)	0.00	-0.16 (0.12)	0.21
Current climatic suitability	_	_	_	_	-0.36 (0.17)	0.06
Climatic stability Holocene-present*	-0.28 (0.13)	0.04	-0.33 (0.14)	0.03	_	_
Climatic stability LGM-present	_	_	_	_	_	_
Cumulative suitability	0.22 (0.12)	0.08	_	_	0.35 (0.20)	0.09
Seedlings						
Defaunation*	0.15 (0.18)	0.43	_	_	0.39 (0.11)	0.00
% Forest cover	0.09 (0.10)	0.39	_	_	-0.01 (0.06)	0.86
Current climatic suitability	0.11 (0.08)	0.23	-0.15 (0.09)	0.11	0.08 (0.13)	0.55
Climatic stability Holocene-present*	-0.19 (0.18)	0.31	-0.45 (0.17)	0.01	0.07 (0.11)	0.53
Climatic stability LGM-present	_	_	-0.22 (0.12)	0.08	-0.03 (0.07)	0.65
Cumulative suitability	0.11 (0.10)	0.29	-0.12 (0.10)	0.24	-0.01 (0.17)	0.92

Model averaged indicates the model-averaged estimates for fixed effects and SE the standard errors, p-value, p value for estimate coefficient values; H_{ϵ} expected heterozygozity; AR allelic richness; f inbreeding coefficient. Effects where p value < 0.05 are highlighted by stars (*) and in bold numbers

(Table 1) explained variation in AR. Non-defaunated sites presented higher AR than defaunated sites (Fig. 3h).

Discussion

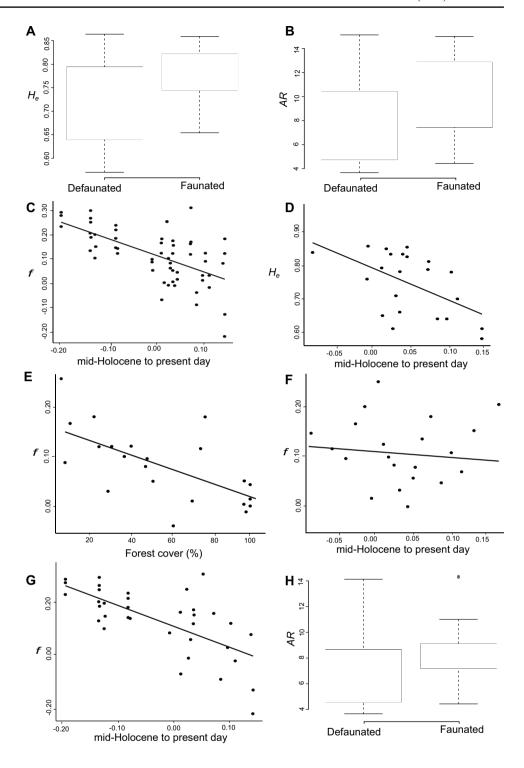
Our results show a retraction in the suitable areas for *E. edulis* through time and that the current pattern of *E. edulis* genetic diversity is the result of such historical population retraction and the current human impact. Combined effects of climatic habitat suitability, lack of large seed dispersers and habitat loss and fragmentation affected genetic diversity of heart of palm populations. Nevertheless, these observed patterns were not consistent across ontogenetic stages, as adults and seedlings are affected by different anthropogenic factors. The complex patterns found between different genetic parameters with historical environmental variables and human impacts indicate that it may be difficult to disentangle the processes involved in the origin and maintenance of genetic diversity.

The consensus map of climatic habitat suitability in the present-day (0 ka), mid-Holocene (6 ka BP) and LGM

(21 ka) revealed a contraction in climatically suitable areas for E. edulis from central sites towards the Brazilian Atlantic coast. Our ENM results showed that the potential distribution of E. edulis is currently narrower than it was in the mid-Holocene and in the LGM. The strong reduction in suitability during the mid-Holocene might have caused population extinction at least in part of the E. edulis distribution. Nevertheless, some areas, mainly in the Brazilian Atlantic coast, have been climatically stable through time and might have been important climatic refugia for E. edulis. In fact, we found that climatically stable sites can harbor a high amount of genetic diversity in heart of palm populations. Other studies also found that the Brazilian Atlantic coast may have been refuge for natural populations during the Quaternary climatic oscillation (Carnaval et al. 2009; Leite et al. 2016). Euterpe edulis populations in these refugia with stable or increasing suitability over time presented higher diversity compared to populations in areas with decreasing suitability since the mid-Holocene. Higher genetic diversity in populations under recent expansion probably reflects the admixture of immigrants arriving from different populations and also the maintenance of



Fig. 3 Relationship between genetic parameters and the main explanatory factors in 57 study sites of Euterpe edulis across the Atlantic forest in Brazil. **a** genetic diversity (H_a) and defaunation for all individuals, b allelic richness (AR) and defaunation for all individuals, c inbreeding coefficient (f) and climatic stability from mid-Holocene to present-day for all individuals, d genetic diversity (H_{ρ}) and climatic stability from mid-Holocene to present-day for adults, e inbreeding coefficient (f) and percentage of forest cover for adults, f inbreeding coefficient (f) and climatic stability from mid-Holocene to present-day for adults, g inbreeding coefficient (f) and climatic stability from mid-Holocene to present-day for seedlings, and h allelic richness (AR) and defaunation for seedlings



large effective population sizes (Ortego et al. 2015). On the other hand, range retractions due to low environmental suitability might lead to population bottlenecks, with loss of individuals and genetic diversity (Pauls et al. 2013).

Conversely, our results show that populations of both adults and seedlings in areas under recent expansion of climatic suitability have higher inbreeding coefficients. The high expected heterozygosity (H_e) despite high inbreeding

coefficients (f) observed in some populations may indicate assortative mating between *E. edulis* palms most likely due to short-distance pollination, spatial distribution of plants, fine-scale spatial genetic structure or flowering patterns (Ghazoul 2005). Another possible explanation is that selfing may occur more frequently to favor reproductive assurance, which may be advantageous in expanded populations (Arnaud-Haond et al. 2006; Pujol et al. 2009). Thus, we



suggest that populations in areas with expanded range are an admixture of individuals from different populations, yet with production of inbred descendants. It could be the result of increased reproductive assurance due to selfing and increased biparental inbreeding due to assortative mating.

On the other hand, we found low inbreeding coefficients in populations that have recently contracted. The reduction of habitat suitability may lead to a reduction of population size and density. Some authors argue that population density is critical when the distance between individuals leads to changes in the foraging behavior of pollinators (Ghazoul 2005; Côrtes et al. 2013). Low population density and higher distance among individuals may increase pollen dispersal distance due to high mobility of pollinators in search for resources (Côrtes et al. 2013). Low inbreeding coefficients may also be the result of loss of homozygous individuals due to lower survival success (i.e., inbreeding depression, Keller and Waller et al. 2002), mainly in low suitable habitats. Thus, we suggest that while contracting populations lose genetic diversity due to bottleneck effects, inbreeding remains low due to higher mobility of pollinators in low density populations or higher loss of homozygous individuals in unsuitable habitats.

Along with climatic suitability, anthropogenic factors may also affect the genetic diversity of E. edulis. Despite the high fragmentation and habitat loss E. edulis populations still present high genetic diversity within the Atlantic forest (Gaiotto et al. 2003; Conte et al. 2008; Carvalho et al. 2015). The high historical effective population size (Galetti et al. 2013) and wide historical climatic habitat suitability distribution may have led to high genetic diversity in currently small and isolated populations. However, we did find significant and negative effects of defaunation on the genetic diversity (H_a) and allelic richness of this species. The loss of large-body seed dispersers may result in the loss of very important ecological functions for plants that include the colonization of new habitats and connectivity between isolated populations (Pacheco and Simonetti 2000; Markl et al. 2012). Connectivity between populations is important for maintaining gene flow and, consequently, large effective population size and genetic diversity (Young et al. 1996). Moreover, habitat loss increased inbreeding levels in E. edulis. Habitat loss may affect the composition, abundance and behavior of pollinators communities, which in turn can limit pollination by increasing the probability of selfing and mating between closely related individuals. In addition, decrease in the amount of forest cover is commonly associated with higher fragmentation (Ribeiro et al. 2009), with the creation of smaller and more isolated habitat remnants. In this scenario, it is also possible that gene flow among patches within these fragmented landscapes was also negatively impacted, resulting in smaller effective population sizes and higher inbreeding.

Adults and seedlings can show distinct responses to anthropogenic disturbances (Van Geert et al. 2008). Due to time-lag effects it is expected that genetic diversity of adult perennial plants respond strongly to past landscape conditions rather than to recent habitat changes (Collevatti et al. 2001; Kramer et al. 2008). This is because changes in the composition and spatial configuration of landscapes occur faster than changes in the genetic diversity of arboreal plants, which can take many generations to respond genetically to human impacts (Landguth et al. 2010). Our results partially corroborate this hypothesis, as seedling and adults behave differently. Seedlings and adults inbreeding coefficients (f) were both influenced by climate variation from the mid-Holocene to present-day. Adults H_e was also influenced by the same historical condition. Both seedling and adults showed to be negatively influenced by human impacts; however adults were mainly affected by the reduction of forest cover while seedling by the loss of large seed dispersers.

Climatic models forecast an increase in mean global temperature due to anthropogenic factors (IPCC 2007). Species can cope with global climate changes if their populations have the capacity to adapt to new environmental conditions or to disperse and track suitable habitats (Dawson et al. 2011). Unfortunately, most natural habitats have been replaced by anthropogenic landscapes dominated by agricultural fields and urban settlements (Foley et al. 2005), which may reduce the capacity of individuals to disperse and find suitable habitat. The amount of genetic diversity is essential for species to cope with environmental changes (Jump et al. 2009). Although E. edulis still has high genetic diversity, we showed that the spatial distribution of this species, as well as its genetic composition, is affected by climate changes. Therefore, an efficient species conservation planning should include the conservation of climatically stable areas that also harbors large extensions of forest and diversified animal community to preserve species interactions and ecological functions such as pollination and seed dispersal.

Finally, our results suggest that the pattern of genetic diversity in *E. edulis* is the outcome of historical instability during the mid-Holocene and current anthropogenic factors, mainly those that affect important ecological processes such as seed dispersal. Current human impacts may enhance historical processes, as well as shape the genetic diversity of heart of palm. Thus, an efficient plan for species conservation is urgent and must account for human impacts and environmental suitability, as well as assess genetic diversity of seedlings and adults in fragmented landscapes.



Acknowledgements This work was supported by the competitive grants from CNPq (Project No. 445353/2014-7) and CAPES (PROCAD Project No. 88881.068425/2014-01) which we gratefully acknowledge. CSC received a CNPq grant (Project No. 401258/2012-2) and a scholarship from Fundação de Amparo à Pesquisa do Estado de São Paulo (FAPESP, No. 2014/01029-5). LBM receives a fellowship from CAPES Ciências sem Fronteira (Project CSF-PAJT/CAPES No. 88881.030318/2013-01). RGC and MCR have been continuously supported by grants and scholarships from CNPq and CAPES. MCR is funded by FAPESP (Project No. 2013/50421-2).

Author Contributions C.S.Carvalho conceptualized and performed the study, analyzed the data and wrote the paper. M.C.Ribeiro, R.G.Collevatti and L. Ballesteros-Mejia participated in the study design, analyzed the data and contributed to writing the paper. M.C.Côrtes assisted in the data analyses and contributed to writing the paper. A.S.Santos contributed to the study conception and to writing the paper.

References

- Aguilar R, Quesada M, Ashworth L et al (2008) Genetic consequences of habitat fragmentation in plant populations: susceptible signals in plant traits and methodological approaches. Mol Ecol 17:5177–5188. doi:10.1111/j.1365-294X.2008.03971.x
- Allouche O, Tsoar A, Kadmon R (2006) Assessing the accuracy of species distribution models: prevalence, kappa and the true skill statistic (TSS). J Appl Ecol 43:1223–1232. doi:10.1111/j.1365-2664.2006.01214.x
- Andersen LW, Fog K, Damgaard C (2004) Habitat fragmentation causes bottlenecks and inbreeding in the European tree frog (*Hyla arborea*). Proc Biol Sci 271:1293–1302. doi:10.1098/ rspb.2004.2720
- Araújo MB, New M (2007) Ensemble forecasting of species distributions. Trends Ecol Evol 22:42–47. doi:10.1016/j. tree.2006.09.010
- Arnaud-Haond S, Teixeira S, Massa SI et al (2006) Genetic structure at range edge: low diversity and high inbreeding in Southeast Asian mangrove (*Avicennia marina*) populations. Mol Ecol 15:3515–3525. doi:10.1111/j.1365-294X.2006.02997.x
- Barbet-Massin M, Jiguet F, Albert CH, Thuiller W (2012) Selecting pseudo-absences for species distribution models: how, where and how many? Methods Ecol Evol 3:327–338. doi:10.1111/j.2041-210X.2011.00172.x
- Barton, K (2016) MuMIn: multi-model inference, R package version 1.15.6.
- Bates D, Mächler M, Bolker B, Walker S (2015) Fitting linear mixedeffects models using lme4. J Stat Softw 67:1–48. doi:10.18637/ jss.v067.i01
- Boscolo D, Metzger JP (2011) Isolation determines patterns of species presence in highly fragmented landscapes. Ecography 34:1018–1029. doi:10.1111/j.1600-0587.2011.06763.x
- Breed MF, Gardner MG, Ottewell KM et al (2012) Shifts in reproductive assurance strategies and inbreeding costs associated with habitat fragmentation in Central American mahogany. Ecol Lett 15:444–452. doi:10.1111/j.1461-0248.2012.01752.x
- Burnham KKP, Anderson DRD (2002) Model selection and multimodel inference: a practical information-theoretic approach. Springer, New York
- Carnaval ACOQ, Hickerson MJ, Haddad CFB et al (2009) Stability predicts genetic diversity in the Brazilian Atlantic forest hotspot. Science 323:785–789. doi:10.1126/science.1166955

- Carvalho CS, Ribeiro MC, Côrtes MC et al (2015) Contemporary and historic factors influence differently genetic differentiation and diversity in a tropical palm. Heredity 115:216–224. doi:10.1038/hdy.2015.30
- Collevatti RG, Grattapaglia D, Hay JD (2001) Population genetic structure of the endangered tropical tree species *Caryocar brasiliense*, based on variability at microsatellite loci. Mol Ecol 10:349–356. doi:10.1046/j.1365-294X.2001.01226.x
- Collevatti RG, Lima-Ribeiro MS, Terribile LC et al (2014) Recovering species demographic history from multi-model inference: the case of a Neotropical savanna tree species. BMC Evol Biol 14:1–13. doi:10.1186/s12862-014-0213-0
- Conte R, dos Reis SM, Mantovani A, Vencovsky R (2008) Genetic structure and mating system of *Euterpe edulis* Mart. Populations: a comparative analysis using microsatellite and allozyme markers. J Hered 99:476–482. doi:10.1093/jhered/esn055
- Côrtes MC, Uriarte M, Lemes MR et al (2013) Low plant density enhances gene dispersal in the Amazonian understory herb *Heliconia acuminata*. Mol Ecol 22:5716–5729. doi:10.1111/mec.12495
- Dawson TP, Jackson ST, House JI et al (2011) Beyond predictions: biodiversity conservation in a changing climate. Science 332:53–58. doi:10.1126/science.1200303
- DiBattista JD (2007) Patterns of genetic variation in anthropogenically impacted populations. Conserv Genet 9:141–156. doi:10.1007/s10592-007-9317-z
- Diniz-Filho JAF, Nabout JC, Bini LM et al (2009) Niche modelling and landscape genetics of *Caryocar brasiliense* (Pequi tree: Caryocaraceae) in Brazilian Cerrado: an integrative approach for evaluating central-peripheral population patterns. Tree Genet Genomes 5:617–627. doi:10.1007/s11295-009-0214-0
- Dirzo R, Miranda A (1991) Altered patterns of herbivory and diversity in the forest understory: a case study of the possible consequences of contemporary defaunation. In: Price PW et al (eds) Plant-animal interactions: evolutionary ecology in tropical and temperate regions. Wiley, New York, pp 273–287
- Eycott AE, Stewart GB, Buyung-Ali LM et al (2012) A metaanalysis on the impact of different matrix structures on species movement rates. Landsc Ecol 27:1263–1278. doi:10.1007/ s10980-012-9781-9
- Fahrig L (2003) Effects of habitat fragmentation on biodiversity. Annu Rev Ecol Evol Syst 34:487–515. doi:10.1146/annurev.ecolsys.34.011802.132419
- Ferrier S, Watson G, Pearce J, Drielsma M (2002) Extended statistical approaches to modelling spatial pattern in biodiversity in northeast New South Wales. I. Species-level modelling. Biodivers Conserv 11:2275–2307. doi:10.1023/A:1021302930424
- Fielding AH, Bell JF (1997) A review of methods for the assessment of prediction errors in conservation presence/absence models. Environ Conserv 24:38–49. doi:10.1017/S0376892997000088
- Foley JA, DeFries R, Asner GP et al (2005) Global consequences of land use. Science 309:570–574. doi:10.1126/science.1111772
- Frankham R, Lees K, Montgomery ME et al (1999) Do population size bottlenecks reduce evolutionary potential? Anim Conserv 2:255–260. doi:10.1111/j.1469-1795.1999.tb00071.x
- Gaiotto FA, Grattapaglia D, Vencovsky R (2003) Genetic structure, mating system, and long-distance gene flow in heart of palm (*Euterpe edulis* Mart.). J Hered 94:399–406. doi:10.1093/jhered/esg087
- Galetti M, Fernandez JC (1998) Palm heart harvesting in the Brazilian Atlantic forest: changes in industry structure and the illegal trade. J Appl Ecol 35:294–301. doi:10.1046/j.1365-2664.1998.00295.x
- Galetti M, Guevara R, Côrtes MC et al (2013) Functional extinction of birds drives rapid evolutionary changes in seed size. Science 340:1086–1090. doi:10.1126/science.1233774



- García C, Jordano P, Godoy JA (2007) Contemporary pollen and seed dispersal in a *Prunus mahaleb* population: patterns in distance and direction. Mol Ecol 16:1947–1955. doi:10.1111/j.1365-294X.2006.03126.x
- Gaston KJ (2000) Global patterns in biodiversity. Nature 405:220–227. doi:10.1038/35012228
- Ghazoul J (2005) Pollen and seed dispersal among dispersed plants. Biol Rev Camb Philos Soc 80:413–443. doi:10.1017/S1464793105006731
- González-Varo JP, Arroyo J, Aparicio A (2009) Effects of fragmentation on pollinator assemblage, pollen limitation and seed production of Mediterranean myrtle (*Myrtus communis*). Biol Conserv 142:1058–1065. doi:10.1016/j.biocon.2009.01.017
- Gugger PF, Ikegami M, Sork VL (2013) Influence of late Quaternary climate change on present patterns of genetic variation in valley oak, *Quercus lobata*. Mol Ecol 22:3598–3612. doi:10.1111/ mec.12317
- Henderson A, Galeano-Garces G, Bernal R (1997) Field guide to the palms of the Americas. Princeton University Press, Princeton
- Hewitt G (2000) The genetic legacy of the Quaternary ice ages. Nature 405:907–913. doi:10.1038/35016000
- Hijmans RJ, Cameron SE, Parra JL et al (2005) Very high resolution interpolated climate surfaces for global land areas. Int J Climatol 25:1965–1978. doi:10.1002/joc.1276
- Hoffmann AA, Willi Y (2008) Detecting genetic responses to environmental change. Nat Rev Genet 9:421–432. doi:10.1038/nrg2339
- Holbrook KM (2011) Home range and movement patterns of toucans: implications for seed dispersal. Biotropica 43:357–364. doi:10.1111/j.1744-7429.2010.00710.x
- Huber PR, Greco SE, Thorne JH (2010) Spatial scale effects on conservation network design: trade-offs and omissions in regional versus local scale planning. Landsc Ecol 25:683–695. doi:10.1007/s10980-010-9447-4
- Hughes AR, Inouye BD, Johnson MTJ et al (2008) Ecological consequences of genetic diversity. Ecol Lett 11:609–623. doi:10.1111/j.1461-0248.2008.01179.x
- IPCC (2007) Climate change 2007: synthesis report. In: Core Writing Team, Pachauri RK, Reisinger A (eds) Contribution of working groups I, II and III to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change. IPCC, Geneva
- Jordano P, García C, Godoy JA, García-Castaño JL (2007) Differential contribution of frugivores to complex seed dispersal patterns. Proc Natl Acad Sci 104:3278–3282. doi:10.1073/pnas.0606793104
- Jump AS, Marchant R, Peñuelas J (2009) Environmental change and the option value of genetic diversity. Trends Plant Sci 14:51–58. doi:10.1016/j.tplants.2008.10.002
- Keller LF, Waller DM (2002) Inbreeding effects in wild populations. Trends Ecol Evol 17:230–241. doi:10.1016/S0169-5347(02)02489-8
- Kramer AT, Ison JL, Ashley MV, Howe HF (2008) The Paradox of forest fragmentation genetics. Conserv Biol 22:878–885. doi:10.1111/j.1523-1739.2008.00944.x
- Landguth EL, Cushman S a, Schwartz MK et al (2010) Quantifying the lag time to detect barriers in landscape genetics. Mol Ecol 19:4179–4191. doi:10.1111/j.1365-294X.2010.04808.x
- Leite YLR, Costa LP, Loss AC et al (2016) Neotropical forest expansion during the last glacial period challenges refuge hypothesis. Proc Natl Acad Sci 113:201513062. doi:10.1073/ pnas.1513062113
- Lowe AJ, Boshier D, Ward M et al (2005) Genetic resource impacts of habitat loss and degradation; reconciling empirical evidence and predicted theory for neotropical trees. Heredity 95:255–273. doi:10.1038/sj.hdy.6800725

- Markl JS, Schleuning M, Forget PM et al (2012) Meta-analysis of the effects of human disturbance on seed dispersal by animals. Conserv Biol 26:1072–1081. doi:10.1111/j.1523-1739.2012.01927.x
- Miraldo A, Li S, Borregaard MK et al (2016) An Anthropocene map of genetic diversity. Science 353:1532–1535. doi:10.1126/science.aaf4381
- Morante-Filho JC, Faria D, Mariano-Neto E, Rhodes J (2015) Birds in anthropogenic landscapes: the responses of ecological groups to forest loss in the Brazilian Atlantic forest. PLoS ONE 10:1–18. doi:10.1371/journal.pone.0128923
- Ortego J, Riordan EC, Gugger PF, Sork VL (2012) Influence of environmental heterogeneity on genetic diversity and structure in an endemic southern Californian oak. Mol Ecol 21:3210–3223. doi:10.1111/j.1365-294X.2012.05591.x
- Ortego J, Gugger PF, Sork VL (2015) Climatically stable landscapes predict patterns of genetic structure and admixture in the Californian canyon live oak. J Biogeogr 42:328–338. doi:10.1111/jbi.12419
- Pacheco LF, Simonetti JA (2000) Genetic structure of a mimosid tree deprived of it seed disperser, the spider monkey. Conserv Biol 14:1766–1775. doi:10.1046/j.1523-1739.2000.99182.x
- Pauls SU, Nowak C, Bálint M, Pfenninger M (2013) The impact of global climate change on genetic diversity within populations and species. Mol Ecol 22:925–946. doi:10.1111/mec.12152
- Phillips SJ, Dudík M (2008) Modeling of species distributions with Maxent: new extensions and a comprehensive evaluation. Ecography 31:161–175. doi:10.1111/j.0906-7590.2008.5203.x
- Pujol AB, Zhoua S, Vilasa JS, Pannella JR (2009) Reduced inbreeding depression after species range expansion. Proc Natl Acad Sci 106: 15379–15383. doi:10.1073/pnas.0902257106
- Rands MRW, Adams WM, Bennun L et al (2010) Biodiversity conservation: challenges beyond 2010. Science 329:1298–1303. doi:10.1126/science.1189138
- Reis MS, Fantini AC, Nodari RO, Reis A, Guerra MP, Mantovani A (2000) Management and conservation of natural populations in Atlantic rainforest: the case study of palm heart (*Euterpe edulis* Martius). Biotropica 32:894–902. doi:10.1111/j.1744-7429.2000.tb00627.x
- Ribeiro MC, Metzger JP, Martensen AC et al (2009) The Brazilian Atlantic forest: how much is left, and how is the remaining forest distributed? Implications for conservation. Biol Conserv 142:1141–1153. doi:10.1016/j.biocon.2009.02.021
- Ribeiro MC, Martensen AC, Metzger JP et al (2011) The Brazilian Atlantic forest: a shrinking biodiversity hotspot. In: Zachos FE, Habel JC (eds) Biodiversity hotspots. Springer, New York, pp 405–434
- Tabarelli M, Aguiar AV, Ribeiro MC et al (2010) Prospects for biodiversity conservation in the Atlantic Forest: Lessons from aging human-modified landscapes. Biol Conserv 143:2328–2340. doi:10.1016/j.biocon.2010.02.005
- Thuiller W, Lafourcade B, Engler R, Araújo MB (2009) BIOMOD—a platform for ensemble forecasting of species distributions. Ecography 32:369–373. doi:10.1111/j.1600-0587.2008.05742.x
- Uriarte M, Anciães M, da Silva MTB et al (2011) Disentangling the drivers of reduced long-distance seed dispersal by birds in an experimentally fragmented landscape. Ecology 92:924–937. doi:10.1890/10-0709.1
- Van Geert A, Van Rossum F, Triest L (2008) Genetic diversity in adult and seedling populations of *Primula vulgaris* in a fragmented agricultural landscape. Conserv Genet 9:845–853. doi:10.1007/s10592-007-9409-9
- Walter RP, Cena CJ, Morgan GE, Heath DD (2012) Historical and anthropogenic factors affecting the population genetic structure of ontario's inland lake populations of walleye (*Sander vitreus*). J Hered 103:831–841. doi:10.1093/jhered/ess066



Wright SJ (2003) The myriad consequences of hunting for vertebrates and plants in tropical forests. Perspect Plant Ecol Evol Syst 6:73–86. doi:10.1078/1433-8319-00043

Young A, Boyle T, Brown T (1996) The population genetic consequences of habitat fragmentation for plants. Trends Ecol Evol 11:413–418. doi:10.1016/0169-5347(96)10045-8

Zurbuchen A, Landert L, Klaiber J et al (2010) Maximum foraging ranges in solitary bees: only few individuals have the capability to cover long foraging distances. Biol Conserv 143:669–676. doi:10.1016/j.biocon.2009.12.003

