

## RESSALVA

Atendendo solicitação do autor ,  
o texto completo desta tese será  
disponibilizado somente a partir de  
17/12/2020.

**LETUSA MOMESSO MARQUES**

**IMPACTS OF NITROGEN APPLICATION ON FORAGE GRASSES TO MAIZE IN  
NO-TILLAGE SYSTEM**

**Botucatu**

**2019**

**LETUSA MOMESSO MARQUES**

**IMPACTS OF NITROGEN APPLICATION ON FORAGE GRASSES TO MAIZE IN  
NO-TILLAGE SYSTEM**

Thesis presented to Sao Paulo State University, College of Agricultural Sciences, to obtain Doctor of Philosophy degree in Agronomy (Agriculture).

Advisor: Prof. Dr. Carlos Alexandre Costa Crusciol

Co-advisor: Prof. Dr. Rogério Peres Soratto

**Botucatu**

**2019**

M357i

Marques, Letusa Momesso

Impacts of nitrogen application on forage grasses to maize in no-tillage system / Letusa Momesso Marques. -- Botucatu, 2019  
90 p.

Tese (doutorado) - Universidade Estadual Paulista (Unesp),  
Faculdade de Ciências Agrônômicas, Botucatu

Orientador: Carlos Alexandre Costa Crusciol

Coorientador: Rogério Peres Soratto

1. Brachiaria spp.. 2. Zea mays L.. 3. Crop residues. 4. Nitrogen recovery. 5. Tropical agriculture. I. Título.

Sistema de geração automática de fichas catalográficas da Unesp. Biblioteca da Faculdade de Ciências Agrônômicas, Botucatu. Dados fornecidos pelo autor(a).

Essa ficha não pode ser modificada.

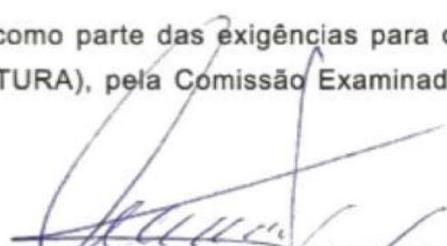
## CERTIFICADO DE APROVAÇÃO

TÍTULO DA TESE: IMPACTS OF NITROGEN APPLICATION ON FORAGE GRASSES TO MAIZE IN NO-TILLAGE SYSTEM

**AUTORA: LETUSA MOMESSO MARQUES**

**ORIENTADOR: CARLOS ALEXANDRE COSTA CRUSCIOL**

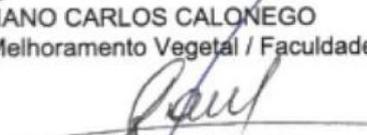
Aprovada como parte das exigências para obtenção do Título de Doutora em AGRONOMIA (AGRICULTURA), pela Comissão Examinadora:



Prof. Dr. CARLOS ALEXANDRE COSTA CRUSCIOL  
Produção e Melhoramento Vegetal / Faculdade de Ciências Agronômicas de Botucatu



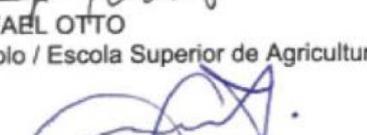
Prof. Dr. JULIANO CARLOS CALONEGO  
Produção e Melhoramento Vegetal / Faculdade de Ciências Agronômicas - UNESP - Câmpus de Botucatu



Pesquisador Dr. HEITOR CANTARELLA  
/ Instituto Agronômico de Campinas



Prof. Dr. RAFAEL OTTO  
Ciência do Solo / Escola Superior de Agricultura Luiz de Queiroz (ESALQ)



Pesquisador Dr. AILDSON PEREIRA DUARTE  
Centro de Grãos e Fibras / Instituto Agronômico de Campinas

Botucatu, 17 de junho de 2019

*To all those who work in science with  
hope their contributions and discoveries  
may change the world.*

## ACKNOWLEDGMENTS

First of all, I would like to thank Prof. Dr. *Carlos Alexandre Costa Crusciol* for having turned his eyes to my path, trusting in me and instigating me in my efforts to find answers. For being example as a person and researcher, and being always present, even in the distance during my exchange. It is an immense honor and pride to have you as my advisor. Thank you very much.

Thanks for my co-advisors, Prof. Dr. *Rogério Peres Soratto* (FCA/UNESP, Brazil), thank you for collaboration and incentive in the development of my research and my writing; and Dr. *Eiko Eurya Kuramae* (NIOO – KNAW, Netherlands), thank you for having me as a Ph.D student at NIOO, helping me and encouraging me to see my research from a different perspective - microbiology. Thanks for everything.

I thank all professors at Sao Paulo State University, College of Agricultural Sciences, for sharing their knowledge and giving me support during the development of my Ph.D. course, especially Prof. *Ciro Antonio Rosolem*, Prof. Dr. *Juliano Carlos Calonego* and Prof. Dr. *Edvaldo Aparecido Amaral da Silva*.

My special thanks for Dr. *Heitor Cantarella* for all assistance and for leaving open doors to help and deepen my research.

I am very thankful for Crusciol's research group through these years that somehow helped me on this journey: *Antonio Carlos Carmeis Filho, Ariani Garcia, Daniele Scudeletti, Cláudio Hideo Martins da Costa, Cleiton Alves, Gabriela Ferraz de Siqueira, Jayme Ferrari Neto, Jorge Martinelli, Luiz Jordão, Luiz Moretti, Murilo Campos, Nídia Raquel Costa, Marcelo Volf, Miriam Tarumoto and Pietro Micheri*. In particular, *Katiuça Sueko Tanaka* for always being present through my Ph.D course such as my co-worker and my friend; *Carlos Antonio Costa do Nascimento* for advising me and contributing with the data analyses. And also to all my friends and colleagues in Botucatu: *Beatriz Fabreti, Cassiano Puoli, Cristiano Pariz, Cristina Andreatti, Fernando Guidorizzi, Iago Cezar, Isabela Cocatto, João de Albuquerque, Lucas Canisares, Matheus Negrisoni, Michely Alves, Murilo de Souza, Natália Corniani and Tiara Guimaraes*.

I would like to thank Eiko's research group: *Afnan Suleiman, Lina Wong, Mattias Hollander, Marcio Leite, Maristela Calvente Moraes, Ohana Costa e Valéria Portela*. In special, thank you very much to the technician *Agaat Pijl* for teaching me a lot about molecular biology in the lab. I would also like to thank all my friends and colleagues in the Netherlands for supporting me in my exchange: *Adam Ossowicki, Anabele Aans, Anashein Diaz, Azkia Nurfikari, Dieke Boezen, Francisco Dini-Andreote, Grace Putra, Hu Jie, Isabela Fernandes, Je Seung, Jopie and Kees Wessels* (my Dutch family), *Marcelle Johnson, Mariana Avalos, Raul Masteling, Renan Pardal, Roos Keijzer, Stalin Sarango, Tatiana Moraes, Viviane Cordovez, and Wu Xiong*.

I am grateful for committee members, Dr. *Aildson Pereira Duarte*, Prof. Dr. *Carlos Alexandre Costa Crusciol*, Dr. *Heitor Cantarella*, Prof. Dr. *Juliano Carlos Calonego*, and Prof. Dr. *Rafael Otto*.

Thanks to Coordination for the Improvement of Higher Education Personnel (CAPES) for the scholarship (Grant: 88881.187743/2018-01), and the São Paulo Research Foundation (FAPESP) for funding part of the experiment in NUCLEUS project: a virtual joint center to deliver enhanced nitrogen use efficiency via an integrated soil-plant systems approach for the United Kingdom and Brazil (Grant 2015/50305-8).

I am very grateful for support and help granted by all members of Technical Section of Postgraduate and Department of Crop Science in São Paulo State University, College of Agricultural Sciences. Especially to *Amanda Bedette*, *Dorival Pires de Arruda*, *Edna Regina Prado*, *Eliane Gonçalves*, *Iara Brito*, *Taynan Ribeiro Moraes da Silva*, and *Valéria Cristina R. Giandoni*.

Furthermore, I would like to thank Fazenda Lageado, FCA – UNESP equip for their assistance in the field experiment, particularly *Casemiro Edson Alves* and *Ciro de Oliveira*.

Last but not the least, my special thanks to my family: *Paulo Donizete Marques*, *Edilene Francisca Momesso Marques*, *Sâmia Momesso Marques*, *Evaristo Luiz Momesso (in memorian)* and *Maria Natalina del Piccollo Momesso*. I thank you for having encouraged and helped me in all the moments of my life. Thank you for everything you have done and still do for me. Thank you for teaching me how to walk by myself so that can follow my own steps. For the education you gave me and for always being by my side. Thank you so much for all your love.

Finally, I would like to express my feeling of gratitude using the words of Gilbert K. Chesterton: "I would maintain that thanks are the highest form of thought, and that gratitude is happiness doubled by wonder".

Thank you all.

“The real voyage of discovery consists not in seeking new landscapes, but in having new eyes”.

Adapted from PROUST, M. Remembrance of Things Past: The Captive. 1923.

## ABSTRACT

The success of no-tillage system depends on the knowledge of the agricultural system as a whole. The use of grass *Urochloa* sp. as cover crop in agriculture results in slow organic material decomposition due to high biomass production and changes in soil microbe, in particular in biological processes related to nitrogen (N). Because N is a nutrient present in the main biochemical reactions in plants and microorganisms, N management requires special attention. Therefore, this research aimed to improve N-use efficiency from both agronomic and biological perspectives. The main objectives were to (i) assess the impact of N fertilizer and forage species on maize in the NT system, and (ii) determine the interactions between microbes x N x environmental factors. A field experiment was evaluated, in which palisade grass (*Urochloa brizantha*) and ruzigrass (*U. ruziziensis*) grown with four N management, included: (i) control zero-N (no N application), (ii) N applied on green cover crops at 35 days before maize seeding (35 DBS), (iii) N applied on cover crop residues at 1 day before maize seeding (1 DBS), and (iv) conventional method of N applied at sidedressing in maize growth), at a rate of 120 kg N ha<sup>-1</sup> as ammonium sulfate. The hypothesis of *Chapter 1* that N applied on alive cover crops or cover crop residues could replace N-sidedressing application (conventional method) for maize was confirmed when: (a) N was applied on palisade grass at 35 DBS or its residues at 1 DBS, and (b) N was applied on ruzigrass residues at 1 DBS. Due to results of first chapter, another experiment was conducted with the objective of assessing whether either the early N application on alive cover crops or on cover crop residues or the conventional method of N application contributed to the recovery of total-N and fertilizer <sup>15</sup>N by maize, by cover crop residues, and in the soil over growing season. Although the hypothesis that N applied on palisade grass to achieve high grain yields of maize was previously confirmed, the results *Chapter 2* showed that the best option is applying nitrogen fertilizer as the current fertilizer recommended method (40 kg N ha<sup>-1</sup> at maize seeding plus 120 kg N ha<sup>-1</sup> sidedressed in V<sub>6</sub> growth stage) for enhance grain yields of maize and N recovery from fertilizer.

**Keywords:** *Brachiaria*. *Zea mays* L.. Crop residues. <sup>15</sup>N. Nitrogen uptake efficiency. Tropical agriculture.

## RESUMO

O sucesso do sistema de plantio direto depende do conhecimento do sistema agrícola como um todo. O uso de gramíneas do gênero *Urochloa* como planta de cobertura resulta em lenta decomposição do material orgânico devido à alta produção de matéria seca e alterações nos microrganismos do solo, em particular nos processos biológicos relacionados ao nitrogênio (N). Como o N é um nutriente presente nas principais reações bioquímicas em plantas e microrganismos, o manejo deste nutriente requer atenção especial. Portanto, este trabalho de pesquisa teve como objetivo melhorar a eficiência do uso do manejo do N. O principal objetivo foi avaliar o impacto do adubo nitrogenado aplicado nas duas espécies de gramíneas ou nos seus resíduos para suprir a demanda e aumentar a produtividade de grãos do milho no sistema plantio direto. O experimento de campo foi conduzido durante três anos, no qual *Urochloa brizantha* e *U. ruziziensis* foram cultivadas com 4 manejos da adubação nitrogenada. Os manejos da adubação nitrogenada foram: (i) controle (zero aplicação de N), (ii) N aplicado 35 dias antes da semeadura do milho (35 DAS), (iii) N aplicado 1 dia antes da semeadura do milho (1 DAS), e (iv) método convencional (N aplicado em cobertura no crescimento do milho), com a dose de 120 kg ha<sup>-1</sup> de N da fonte sulfato de amônio. A hipótese no *Capítulo 1* de que o N aplicado nas plantas de cobertura ou nos resíduos destas plantas de cobertura poderiam ser substituir a aplicação de N em cobertura do atual método convencional para cultura do milho foi confirmada quando o N foi aplicado na *U. brizantha* aos 35 DAS ou em seus resíduos 1 DAS e quando o N foi aplicado nos resíduos da *U. ruziziensis* 1 DAS. Devido aos resultados observados no primeiro capítulo, o *Capítulo 2* objetivou avaliar se a aplicação antecipada de N (nas plantas de cobertura ou nos resíduos das plantas de cobertura) e a aplicação de N no método convencional contribui para o teor total de N e a recuperação do <sup>15</sup>N do fertilizante pelo milho, pelos resíduos das plantas de cobertura e no solo ao final da safra. Embora a hipótese de que a aplicação de N na *U. brizantha* tenha sido confirmada anteriormente para atingir altas produtividade de grãos de milho, os resultados do segundo capítulo mostraram que a aplicação do N deve ser realizada como recomendado no método convencional (40 kg ha<sup>-1</sup> na semeadura e 120 kg ha<sup>-1</sup> em cobertura) para, além de atingir altas produtividade, recuperar maior quantidade do fertilizante nitrogenado aplicado.

**Palavras-chave:** *Brachiaria*. Plantas de cobertura. Fertilizante <sup>15</sup>N. Eficiência do uso do nitrogênio. Sistema semeadura direta.

## LIST OF FIGURES

- Figure 1 – Daily rainfall (— bars), maximum (—lines) and minimum (—lines) temperatures observed in a meteorological station located near the field site at Botucatu, Sao Paulo State, Brazil, during periods in 2015-2016, 2016-2017, and 2017-2018 growing seasons..... 34
- Figure 2 – Experiment timeline of crop management and N application timing. Nitrogen application timing treatments in orange boxes are as follows: 35 DBS: 120 kg N ha<sup>-1</sup> broadcast over cover crop 35 days before maize seeding (5 days before cover crop termination); 1 DBS: 120 kg N ha<sup>-1</sup> broadcast over crop 1 day before maize seeding; Conventional method: 120 kg N ha<sup>-1</sup> sidedressed at V<sub>6</sub> growth stage of maize. The green box means application of 40 kg N ha<sup>-1</sup> at maize seeding to all N treatments..... 35
- Figure 3 – Exponential decomposition of biomass yield of *Urochloa brizantha* (a) and *Urochloa ruziziensis* (b), and amount of N in straw of *U. brizantha* (c) and *U. ruziziensis* (d) as affected by N application timing [(▲) 35 DBS: 120 kg N ha<sup>-1</sup> broadcast over grass cover crop 35 days before maize seeding plus 40 kg N ha<sup>-1</sup> in the maize seeding furrow, (◆) 1 DBS: 120 kg N ha<sup>-1</sup> broadcast over terminated cover crop 1 day before maize seeding plus 40 kg N ha<sup>-1</sup> in the maize seeding furrow, (■) conventional N application method: 40 kg N ha<sup>-1</sup> in the maize seeding furrow plus 120 kg N ha<sup>-1</sup> sidedressed in V<sub>6</sub> growth stage, and (●) control: no N application], depending on the days after cover crop termination. \*: Significant at 5% by F test. Vertical bars are indicative of the MSD value at 5% probability..... 41
- Figure 4 – Cover crop × N application timing interaction effect on the N concentration in leaves (a), plant height (b), number of grains per ears (c), 100-grain weight (d), grain yield (e), and N-use efficiency (f) of maize during three growing seasons. Nitrogen application timing treatments are as follows: 35 DBS: 120 kg N ha<sup>-1</sup> broadcast over cover crop 35 days before maize seeding plus 40 kg N ha<sup>-1</sup> in the maize seeding furrow; 1 DBS: 120 kg N ha<sup>-1</sup> broadcast over crop 1 day before maize seeding plus 40 kg N ha<sup>-1</sup> in the maize seeding furrow; Conventional method: 40 kg N ha<sup>-1</sup> in the maize seeding furrow plus 120 kg N ha<sup>-1</sup> sidedressed in V<sub>6</sub> growth stage; and Control: no N application. Different lowercase letters denote significant difference between cover corps and different uppercase letters denote significant difference among N application timing (LSD, *P* ≤ 0.05)..... 49
- Figure 5 – Pearson correlation between maize grain yield, maize dry matter, cover crop biomass at 0, 30, 60, 90 and 120 days after cover crops termination, ammonium in the soil at 0, 30 and 60 days after cover crops termination, and nitrate in the soil at 0, 30 and 60 days after cover crops termination. Pearson correlation coefficients of the blue ellipses are positive, and those of the red

ellipses are negative. The darker the color and the smaller the area of the ellipse, the greater the degree of correlation and the larger the absolute value of the Pearson correlation coefficient. The ellipses indicates significant differences of relationship between two variables ( $P > 0.05$ ). Variable names are on the horizontal and vertical. For the interpretation of the references to colors in this figure legend, the reader is referred to the bar..... 50

Figure 6 – Seasonal precipitation and average air temperature during study period in 2015-2016 growing seasons, and times of N application on cover crops 35 days before seeding (DBS) of maize, on cover crops residues 1 DBS of maize, and conventional method at sidedressing in V<sub>6</sub> growth stage of maize, and maize crop management..... 65

Figure 7 – Schematic representation of the microplot. The <sup>15</sup>N-fertilizer was applied in the central rows of the microplot, and the central and adjacent rows were sampling to <sup>15</sup>N calculations. The letters A and B represent the locations where soil sampling was performed in each microplot..... 67

Figure 8 – Cover crop × N application timing interaction effect on dry matter of cover crops (a and b) and N (c and d) at 0 (a and c) and 90 (b and d) days after termination (DAT) of cover crops. Nitrogen application timing treatments are as follows: 35 DBS: 120 kg N ha<sup>-1</sup> broadcast over cover crop 35 days before maize seeding plus 40 kg N ha<sup>-1</sup> at seeding; 1 DBS: 120 kg N ha<sup>-1</sup> broadcast over crop 1 day before maize seeding plus 40 kg N ha<sup>-1</sup> at seeding; Conventional method: 40 kg N ha<sup>-1</sup> at seeding furrow plus 120 kg N ha<sup>-1</sup> sidedressed at V<sub>6</sub> growth stage of maize; and Control: no N application. Different lowercase letters denote significant difference between cover corps and different uppercase letters denote significant difference between N application timing (LSD,  $P \leq 0.05$ )..... 72

Figure 9 – Cover crop × N application timing interaction effect on shoot dry matter (SDM) (a), N concentration in leaf (b), and grain yield (c) of maize. Nitrogen application timing treatments are as follows: 35 DBS: 120 kg N ha<sup>-1</sup> broadcast over cover crop 35 days before maize seeding plus 40 kg N ha<sup>-1</sup> in the maize seeding furrow; 1 DBS: 120 kg N ha<sup>-1</sup> broadcast over crop 1 day before maize seeding plus 40 kg N ha<sup>-1</sup> in the maize seeding furrow; Conventional method: 40 kg N ha<sup>-1</sup> in the maize seeding furrow plus 120 kg N ha<sup>-1</sup> sidedressed in V<sub>6</sub> growth stage; and Control: no N application. Different lowercase letters denote significant difference between cover corps and different uppercase letters denote significant difference between N application timing (LSD,  $P \leq 0.05$ )..... 74

Figure 10 – Cover crop × N application timing interaction effect on total-N content in plant (stover + plant) (a), N derived from fertilizer (NDF) in the stover (b) and grain (c), and percentage of fertilizer N on whole plant on plant N content ( $F_{index}$ ) (d). Nitrogen application timing treatments are as follows: 35 DBS: 120 kg N ha<sup>-1</sup> broadcast over cover crop 35 days before maize seeding plus 40 kg N ha<sup>-1</sup> in the maize seeding furrow; 1 DBS: 120 kg N ha<sup>-1</sup> broadcast over crop 1 day before maize seeding plus 40 kg N ha<sup>-1</sup> in the maize seeding furrow; and Conventional method: 40 kg N ha<sup>-1</sup> in the maize seeding furrow plus 120 kg N ha<sup>-1</sup> sidedressed in V<sub>6</sub> growth stage. Different lowercase letters denote significant difference between cover crops and different uppercase letters denote significant difference between N application timing (LSD,  $P \leq 0.05$ )..... 76

Figure 11 – Percentage of <sup>15</sup>N recovery (NRE, %) by soil 0-40 cm depth, straw of cover crops, stover and grain of maize as affected the cover crop and N application timing. Nitrogen application timing treatments are as follows: 35 DBS: 120 kg N ha<sup>-1</sup> broadcast over cover crop 35 days before maize seeding plus 40 kg N ha<sup>-1</sup> in the maize seeding furrow; 1 DBS: 120 kg N ha<sup>-1</sup> broadcast over crop 1 day before maize seeding plus 40 kg N ha<sup>-1</sup> in the maize seeding furrow; and Conventional method: 40 kg N ha<sup>-1</sup> in the maize seeding furrow plus 120 kg N ha<sup>-1</sup> sidedressed in V<sub>6</sub> growth stage. Different lowercase letters denote significant difference between cover crops and different uppercase letters denote significant difference between N application timing for same N fate (LSD,  $P \leq 0.05$ )..... 78

## LIST OF TABLES

Table 1 –	Chemical properties of field experiment soil (0-20 cm depth).....	35
Table 2 –	Dry matter yields and N accumulated in straw residues of palisade grass and ruzigrass under different N application timing in three growing seasons and sampling time (0, 30, 60, 90 and 120 days after desiccation).....	40
Table 3 –	Total N, ammonium (N-NH <sub>4</sub> <sup>+</sup> ) and nitrate (N-NO <sub>3</sub> <sup>-</sup> ) concentration in the soil (0-10 cm) at 0, 30, 60 and 90 days after cover crop termination (DAT) as affected by cover crops, N application timing and growing season.....	43
Table 4 –	Total N, ammonium (N-NH <sub>4</sub> <sup>+</sup> ) and nitrate (N-NO <sub>3</sub> <sup>-</sup> ) concentration in the soil (10-20 cm) at 0, 30, 60 and 90 days after cover crop termination (DAT) as affected by cover crops, N application timing and growing season.....	44
Table 5 –	Nutrient (N, P, K, Ca, Mg, and S) concentrations in the leaves of maize crop at 60 d after maize emergence as affected by the cover crop, N application timing, and growing season at Botucatu, Sao Paulo State, Brazil.....	45
Table 6 –	Shoot dry matter (DM) flowering, plant population, plant height, number of ears per plant, number of grains per ear, 100-grain weight, grain yield, and nitrogen use efficiency of maize crop as affected by cover crop, N application timing, and growing season, and analyses of variance at Botucatu, São Paulo, Brazil. Data reported are means of three harvest, except for the growing season average (GS).....	48
Table 7 –	Dry matter and N accumulated of cover crops at 0 and 120 days after desiccation as affected by cover crop and N application timing.....	71
Table 8 –	Shoot dry matter, N concentration of leaf at 60 days after maize emergence, grain yields of maize, and grain at maize harvest as affected by cover crop and N application timing.....	73
Table 9 –	Total-N contents in whole plant (stover + grain), distribution of N derived from fertilizer (NDFF) and <sup>15</sup> N recovery (NRE) in soil 0-40 cm depth, straw of cover crops, stover and grain of maize at harvest as affected by cover crop and N application timing.....	75

## SUMMARY

<b>GENERAL INTRODUCTION</b> .....	23
<b>CHAPTER 1 – CAN NITROGEN APPLICATION ON <i>Urochloa</i> COVER CROPS REPLACE THE STANDARD NITROGEN RECOMMENDATION METHOD FOR MAIZE?</b> .....	27
1.1 INTRODUCTION.....	27
1.2 MATERIAL AND METHODS.....	29
1.2.1 Field experimental characterization.....	29
1.2.2 Crop management.....	30
1.2.3 Sampling and analyses.....	34
1.2.4 Data statistical analyses.....	34
1.3 RESULTS.....	36
1.3.1 Decomposition kinetics of biomass yield and N accumulated of cover crops.....	36
1.3.2 N-NH <sub>4</sub> <sup>+</sup> , N-NO <sub>3</sub> <sup>-</sup> and total-N contents in the soil.....	38
1.3.3 Maize crop performance.....	42
1.3.4 Effects of biomass characteristics and n forms in the soil on grain yield of maize.....	47
1.4 DISCUSSION.....	47
1.5 CONCLUSIONS.....	53
REFERENCES.....	53
<b>CHAPTER 2 – WHAT IS THE BEST TIME FOR NITROGEN APPLICATION IN <i>Urochloa</i>-MAIZE CROP SYSTEM?</b> .....	58
2.1 INTRODUCTION.....	58
2.2 MATERIAL AND METHOD.....	61
2.2.1 Site description.....	61
2.2.2 Experimental design and treatments.....	62
2.2.3 Crop management and sampling.....	63
2.2.4 Isotopic labeled-N determination ( <sup>15</sup> N).....	65
2.2.5 Data statistical analyses.....	66
2.3 RESULTS.....	67
2.3.1 Cover crop and maize performance.....	67

2.3.2	Labeled nitrogen recovery.....	71
2.4	DISCUSSION.....	75
2.5	CONCLUSIONS.....	79
	REFERENCES.....	80
	<b>FINAL CONSIDERATIONS</b> .....	<b>85</b>
	<b>REFERENCES</b> .....	<b>87</b>

## GENERAL INTRODUCTION

Over time, the plant residues left on the soil surface in a no-tillage (NT) system gradually improve the physical, chemical, and biological characteristics of the soil (SILVA et al., 2014; TIRITAN et al., 2016; MORAES et al., 2019). Used frequently in tropical countries, this practice increases grain yields and improves environmental performance (DERPSCH et al., 2014; GUZMAN; GOLABI et al., 2017). In NT systems, food production costs are lower. It is easier to operate machines in fields, which improves soil sustainability, due to lower soil disturbances. However, the success of a no-tillage system depends on several important factors: growing crops in undisturbed soil, rotating crops and maintaining crop residues on the soil surface (DERPSCH et al., 2014; DUARTE et al., 2018).

Vegetal residues of cover crops are used in integrated livestock farming and grain food production systems (SILVA et al., 2015; MATEUS et al., 2016; MORAES et al., 2019; SCHUSTER et al., 2019). Use of these vegetal residues improves soil properties, inhibits the spread of diseases, reduces pests and weeds as well emissions of greenhouses gasses (FREITAS; LANDERS, 2014; MORAES et al., 2014; Mckenzie et al., 2016; SCHUSTER et al., 2019). The choice of the cover crop is very important and determines the success of the system (PARIZ et al., 2011). This is especially true in tropical soils due to the fact that plant characteristics impact biomass yield and the durability of soil coverage (LEITE et al., 2010; PAVINATO et al., 2017). Cover crop biomass contains nutrients extracted from deeper soil layers and plays an important role in nutrient cycling as a result of the release of nutrients during the decomposition process (CRUSCIOL; SORATTO, 2009; VERAS et al., 2016; ROSOLEM et al., 2017).

Legume and grass cover crops vary widely in their ability to cover the soil (CRUSCIOL et al., 2015; FAGERIA et al., 2016). Although legumes fix atmospheric

nitrogen, grasses are better at scavenging nutrients. Among the cover crops, grasses are most widely cultivated and used for livestock as well as agricultural activities by farmers (COSTA et al., 2017; PARIZ et al., 2017; CATUCHI et al., 2019). The most commonly used grasses are from the *Urochloa* genus. These grasses produce a large amount of biomass, which has high soil protection properties and nutrient-cycling efficiency (BORGHI et al., 2013; PACHECO et al., 2017; TANAKA et al., 2019). When cultivated as a cover crop, forage grass is managed as an annual crop in the system to produce biomass (BORGHI et al., 2013). The high dry matter production potential and the high C:N ratio of *Urochloa* result in slow decomposition and increase the possibility of cultivation in warmer regions, even in regions where other cover crops have accelerated rates of decomposition (TIMOSSI et al., 2007; ROSOLEM et al., 2017). Other characteristics, such as vigorous and deep root systems, favor water deficiency tolerance and absorption of nutrients in deeper soil layers, aiding nutrient cycling (CRUSCIOL; SORATTO, 2009; ALMEIDA et al., 2018). Thus, grasses perform well in drought conditions where most of the grain crops or other cover crops do not grow well (CASTRO et al., 2015; CRUSCIOL et al., 2015).

*Urochloa* species are less demanding when it comes to soil fertility. The roots tolerate aluminum toxicity and low P availability in the soil, which occurs often in acidic tropical soils (ARROYAVE et al., 2018). In addition, the roots of these grasses suppress soil nitrification, which is one of the key microbial processes (SUBBARAO et al., 2009, 2015). *Urochloa* species release brachialactone, a biological nitrification inhibitor (BNI), that blocks ammonia monooxygenase (AMO) and hydroxylamino oxidoreductase (HAO) ammonia oxidizing enzymatic pathways (SUBBARAO et al., 2007, 2009). In the complex soil-plant-atmosphere system, the major processes in the soil are nitrogen fixation, soil organic matter (SOM) mineralization, ammonification,

nitrification, and denitrification (WILCKE; LILIENFEIN, 2005; SUBBARAO et al., 2015; KUYPERS et al., 2018). Nitrogen plays a minor role in undisturbed temperate and tropical ecosystems, such as no-tillage systems, where nitrogen leakage is minimized and a large amount of nitrogen is retained in the soil (SUBBARAO et al., 2015). There are mechanisms of nitrogen conservation that involve short-circuiting mineralization, which microorganisms absorb nitrogen and return it to the soil, facilitating nitrogen accumulation in the soil when plants suppress nitrification and directly absorb organic nitrogen (SUBBARAO et al., 2015; KARWAT et al., 2017).

Nitrogen losses occur mainly through critical pathways of nitrification and denitrification (VAN GROENIGEN et al., 2015; ZHANG et al., 2015). *Urochloa* species cultivated in the no-tillage systems are expected to reduce nitrogen losses due to nitrate leaching and nitrous oxide emissions. One key role of tropical forage grasses has been suppressing leached nitrate and mitigating nitrous oxide emissions in pasture soils (BYRNES et al., 2017; KARWAT et al., 2017). However, there is a lack of information about forage grasses as cover crops in systems of grain food production. The nitrogen use efficiency (NUE) of fertilizer can be enhanced for grain crops when it is combined with nitrogen management on cover crops. Nevertheless, cover crop species affect the subsequent crops differently. This becomes important when investigating the influence of cover crops on the grass-maize system.

The organic nitrogen from the biomass of cover crops is degraded through mineralization and subsequently, nitrification in ammonium and nitrate in the soil (KUYPERS et al., 2018). The biomass of cover crops can drive shifts in soil microbial activity, resulting in mineral nitrogen. Since the plant roots take up nitrogen in the inorganic forms ammonium and nitrate (BOSCHIERO et al., 2018), the decomposition rates of nitrogen released in the soil determine synchrony/de-synchrony between soil

nitrogen mineralization and plant nitrogen demand (PERVEEN et al. 2014; ROSOLEM et al., 2017). There is a huge potential to reduce nitrogen fertilizer use and nitrogen loss in agricultural system composed of *Urochloa* (KARWAT et al. 2017; MOMESSO et al., 2019; ROCHA et al., 2019).

The current recommended method of applying nitrogen fertilizer to annual crops is to divide the application over two periods. The first application occurs when seeding a crop and the second occurs when the crop is in its growing stage in a manner called 'sidedressing'. However, an alternative method has been proposed which would actually supply the nitrogen fertilizer needed for annual crops, such as maize, during the cultivation of cover crops like *Urochloa*. The farmers would apply all the nitrogen either to cover crops or during pre-seeding of maize (BASSO; CERETTA, 2000; LARA CABEZAS et al., 2004, 2005; PÖTTKER; WIETHÖLTER, 2004), when the systems have high straw production (CERETTA et al., 2002). The early application of nitrogen could facilitate the main crop seeding, providing flexibility in the operational schedules of farmers. However, the application of all of the nitrogen on *Urochloa* cover crops does not supply the nitrogen to the subsequent maize crops due to the temporarily nitrogen immobilization. In systems composed of grasses grown in succession, microorganisms compete with plants for nitrogen in the soil during crop seeding due to increasing biological activity and consequent plant-microbe competition (MOMESSO et al., 2019).

An alternative and sustainable way to use nitrogen fertilizers could be to follow the initial recommended application of nitrogen fertilizer during maize seeding. However, instead of following the second half of the recommended application method, which says that nitrogen should be applied to growing crops (sidedressing), the nitrogen should actually be applied earlier to cover crops or on cover crop residues. Applying nitrogen directly to maize during seeding aims to reduce competition between

the maize crop and microbes for nitrogen immobilization in the soil. The application of nitrogen during maize seeding minimizes the competition between plants and microbes and thus avoids nitrogen immobilization. Applying nitrogen to the cover crops or cover crop residues may be a good substitute for sidedressing when growing forage species. The use of fertilized *Urochloa* can be effective to gradually provide nitrogen to subsequent maize crops during residue decomposition due to the great potential of *Urochloa* to produce biomass and tighten nitrogen cycling.

There has been a growing interest in manipulating plant growth in order to increase NUE and grain yields in tropical food production (BOWATTE et al., 2015). The nitrogen fertilizer used in grass systems enhances grain yields of annual crops by improving biomass production in agricultural systems. In a no tillage system, this biomass gradually releases nutrients to subsequent crops. Keeping this in mind, the current thesis starts by assessing how the forage grass and the maize crop are affected by the timing of nitrogen fertilizer application. In *Chapter 1*, the effects of the timing of nitrogen application on decomposition rates of cover crops was monitored for 3 years during a field experiment. In this experiment, biomass production, nitrogen released by cover crops and the availability of mineral nitrogen in the soil was evaluated in order to enhance grain yields of maize. In *Chapter 2*, the fate of nitrogen fertilizers  $[(^{15}\text{NH}_4)_2\text{SO}_4]$  applied at different times to maize and forage grasses was examined. This thesis presents potential strategies to optimize the sustainable use of nitrogen fertilizer in maize-forage systems and examines how grasses can supply nitrogen to subsequent maize crops in tropical soils during decomposition.

palisade grass at 35 DBS and on its residues at 1 DBS resulted in similar maize grain yields as the conventional method of N application. Thus, the use of palisade grass as a cover crop allows for early N application and is an alternative to the recommended sidedressing application. However, when the cover crop is ruzigrass, N fertilization must not be applied while the grass is still growing. Beyond crop nutrition, our results raise questions concerning the impact of the timing of N fertilization on effective N-fertilizer uptake by maize and the environmental consequences of N fertilization for microbial communities in agro-food systems.

## REFERENCES

AITA, C. *et al.* Consorciação de plantas de cobertura antecedendo o milho em plantio direto: I - Dinâmica do nitrogênio no solo. **Revista Brasileira de Ciência do Solo**, v. 28, n. 4, p. 739-749, 2004.

BANI, A. *et al.* The role of microbial community in the decomposition of leaf litter and deadwood. **Applied Soil Ecology**, v. 126, p. 75-84, 2018.

BORGHI, E. *et al.* Sorghum grain yield, forage biomass production and revenue as affected by intercropping time. **European Journal of Agronomy**, v. 51, p. 130-139, 2013.

BÜCHI, L. *et al.* Cover crops with reduced tillage. **Field Crops Research**, 107583, 2019.

BYRNES, R. C. *et al.* Biological nitrification inhibition by Brachiaria grasses mitigates soil nitrous oxide emissions from bovine urine patches. **Soil Biology and Biochemistry**, v. 107, p. 156–163, 2017.

CAMAROTTO, C. *et al.* Conservation agriculture and cover crop practices to regulate water, carbon and nitrogen cycles in the low-lying Venetian plain. **Catena**, v. 167, p. 236-249, 2018.

CANTARELLA, H. **Nitrogen**. Viçosa, Brazil, p. 375-470. 2007.

CANTARELLA, H. *et al.* **Recomendações de adubação e calagem para o Estado de São Paulo**. Campinas, Brazil, Inst. Agrônomo, p. 43–50, 1997.

CASTRO, G. S. A. *et al.* Management impacts on soil organic matter of tropical soils. **Vadose Zone Journal**, v. 14, p. 1-8, 2015.

CIAMPITTI, I. A.; VYN, T. J. A comprehensive study of plant density consequences on nitrogen uptake dynamics of maize plants from vegetative to reproductive stages. **Field Crops Research**, v. 121, p. 2–18, 2011.

COLEMAN, D. C. *et al.* **Decomposition and Nutrient Cycling**. In: Coleman, D.C., Callaham Jr., M.A., Crossley Jr., D.A. *Fundamentals of Soil Ecology*. 3<sup>rd</sup> Edition. 2018, 376p.

COSTA, C. H. M. *et al.* Nitrogen fertilization on palisadegrass: phytomass decomposition and nutrients release. **Pesquisa Agropecuaria Tropical**, v. 46, p. 159–168, 2016.

ECHER, F. R. *et al.* Initial Growth and nutrient absorption of cotton cultivated on congo grass residues. **Planta Daninha**, v. 30, p. 783–790, 2012.

EL-SHARKAWI, H. M. Effect of nitrogen sources on microbial biomass nitrogen under different soil types. **ISRN Soil Science**, p. 1–7, 2012.

FAGERIA, N. K.; BALIGAR, V. C. Enhancing nitrogen use efficiency in crop plants. **Advances in Agronomy**, v. 88, p. 97–185, 2005.

FELISMINO, M. F. *et al.* Meiotic stability in two valuable interspecific hybrids of *Brachiaria*. **Plant Breeding**, p. 402-408, 2012.

FIORETTO, A. *et al.* Lignin and cellulose degradation and nitrogen dynamics during decomposition of three leaf litter species in a Mediterranean ecosystem. **Soil Biology and Biochemistry**, v. 37, p. 1083-1091, 2005.

GATIBONI, L. C. *et al.* Microbial biomass and soil fauna during the decomposition of cover crops in no-tillage system. **Revista Brasileira de Ciência do Solo**, v. 35, n. 4, p. 1051-1057, 2011.

INSELSBACHER, E. *et al.* Early season dynamics of soil nitrogen fluxes in fertilized and unfertilized boreal forests. **Soil Biology and Biochemistry**, v. 74, p. 167–176, 2014.

KEENEY, D. R.; NELSON, D. W. **Nitrogen-Inorganic Forms**. Madison, WI: ASA SSSA, pp. 643-698, 1982.

KUZYAKOV, Y.; XU, X. Competition between roots and microorganisms for nitrogen: mechanisms and ecological relevance. **New Phytologist**, v. 198, n. 3, p. 656–669, 2013.

LIU, C.; SUN, X. A review of ecological stoichiometry: basic knowledge and advances. **References Module in Earth Systems and Environmental Sciences**, 2013.

MALAVOLTA, E. *et al.* **Avaliação do estado nutricional das plantas: princípios e aplicações**. Piracicaba: Potafos, 1997.

MARIANO, E. *et al.* Influence of nitrogen form supply on soil mineral nitrogen dynamics, nitrogen uptake, and productivity of sugarcane. **Agronomy Journal**, v. 107, p. 641–650, 2015.

MARSCHNER, H. **Mineral nutrition of higher plants**. London: Academic Press, 2012.

MOMESSO, L. *et al.* Impacts of nitrogen management on no-till maize production following forage cover crops. **Agronomy Journal**, v. 111, p. 639-649, 2019.

MORO, E. *et al.* Upland rice under no-tillage preceded by crops for soil cover and nitrogen fertilization. **Revista Brasileira de Ciencia do Solo**, v. 37, p. 1669-1677, 2013.

NUÑEZ, J. *et al.* Biological nitrification inhibition activity in a soil-grown biparental population of the forage grass, *Brachiaria humidicola*. **Plant Soil**, v. 426, p. 401–411, 2018.

PACHECO, L. P. *et al.* Biomass and nutrient cycling by cover crops in Brazilian cerrado in the State of Piauí. **Revista Caatinga**, v. 30, p. 13-23, 2017.

PACHECO, L. P. *et al.* Nutrient cycling by cover crops and yield of soybean and rice in no-tillage. **Pesquisa agropecuaria brasileira**, v. 48, n. 9, p. 1228-1236, 2013.

PACHECO, L. P. *et al.* Produção de fitomassa e acúmulo e liberação de nutrientes por plantas de cobertura na safrinha. **Pesquisa Agropecuaria Brasileira**, v. 46, p. 7-25, 2011.

PARIZ, C. M. *et al.* Straw decomposition of nitrogen-fertilized grasses intercropped with irrigated maize in an integrated crop-livestock system. **Revista Brasileira Ciencia Solo**, n. 35, p. 2029-2037, 2011.

PARK, B. S. *et al.* Arabidopsis nitrogen limitation adaptation regulates ORE1 homeostasis during senescence induced by nitrogen deficiency. **Nature Plants**, n. 4, p. 898-903, 2018.

ROCHA, K. F. *et al.* Fate of 15N fertilizer applied to maize in rotation with tropical forage grasses. **Field Crops Research**, v. 238, p. 35-44, 2019.

ROSOLEM, C. A. *et al.* Enhanced plant rooting and crop system management for improved N use efficiency. **Advances in Agronomy**, v. 146, p. 205–239, 2017.

ROSOLEM, C. A.; PACE, L.; CRUSCIOL, C. A. C. Nitrogen management in maize cover crop rotations. **Plant and Soil**, v. 264, p. 261-271, 2004.

SATTOLO, T. M. S. *et al.* Soil carbon and nitrogen dynamics as affected by land use change and successive nitrogen fertilization of sugarcane. **Agriculture, Ecosystems and Environment**, v. 247, p. 63-74, 2017.

SCHIMEL, J. P.; BENNETT, J. Nitrogen mineralization: challenges of a changing paradigm. **Ecology**, v. 85, p. 591–602, 2004.

SORATTO, R. P. Nitrogen early application and sources for common bean in succession to forage grasses in no-tillage system. **Tese de Livre-Docente**, Universidade Estadual Paulista “Júlio de Mesquita Filho” - Faculdade de Ciências Agrônômicas de Botucatu, 2011, 139p.

SOUZA, E. F. C. *et al.* Early growth of common bean cropped over ruzigrass residues. **Planta Daninha**, v. 32, p. 775–781, 2014.

SUBBARAO, G. *et al.* Genetic mitigation strategies to tackle agricultural GHG emissions: The case for biological nitrification inhibition technology. **Plant Science**, v. 262, p. 165–168, 2017.

SUBBARAO, G. V. *et al.* Evidence for biological nitrification inhibition in *Brachiaria* pastures. **Proceedings of the National Academy of Science of the United States of America**, v. 106, p. 17302–17307, 2009.

SUBBARAO, G. V. *et al.* Biological nitrification inhibition (BNI)—is it a widespread phenomenon? **Plant Soil**, v. 294, p. 5–18, 2007.

SUBBARAO, G. V. *et al.* Evidence for biological nitrification inhibition in *Brachiaria* pastures. **Proceedings of the National Academy of Sciences of the United States of America**, v. 106, n. 41, p. 17302–17307, 2009.

TANAKA, K. S. *et al.* Nutrients released by *Urochloa* cover crops prior to soybean. **Nutrient Cycling in Agroecosystems**, v. 113, p. 267-281, 2019.

THOMAS, R. J.; ASAKAWA, N. M. Decomposition of leaf litter from tropical forage grasses and legumes. **Soil Biology and Biochemistry**, v. 25, p. 1351-1361, 1993.

TURMEL, M. S. *et al.* Crop residue management and soil health: A systems analysis. **Agricultural Systems**, n. 134, p. 6-16, 2015.

USDA, United States Department of Agriculture. **Keys to soil taxonomy**. Twelfth Edition, p. 372, 2014.

USDA, United States Department of Agriculture. **USDA Agricultural Projections to 2028**, p. 108, 2019.

WANG, H. *et al.* Quality of fresh organic matter affects priming of soil organic matter and substrate utilization patterns of microbes. **Scientific Reports**, v. 5, n. 10102, 2015.

WORLDMETERS, **Wordometers 2019**. Available online at: <http://www.worldometers.info/worldpopulation/world-population-projections/> (Accessed February 20, 2019).

YANG, J.; UDVARDI, M. Senescence and nitrogen use efficiency in perennial grasses for forage and biofuel production. 2018. **Journal of Experimental Botany**, p. 855-865, 2017.

system to supply N demand and raise grain yield of maize. Besides that, the timing of N application in agricultural system is an important factor in increasing the maize grain yield and forage production. This study reported the important knowledge that higher grain yields of maize were achieved when N was applied on palisade grass and its residues or on ruzigrass residues, resulting in similar grain yield obtained in conventional method. However, the fate of N fertilizer in the system was not completely understood. Although the hypothesis of N applied on palisade grass to reach high grain yields of maize was confirmed, the application of nitrogen fertilizer must be applied as current recommended method (40 kg N ha<sup>-1</sup> at maize seeding plus 120 kg N ha<sup>-1</sup> sidedressed in V<sub>6</sub> growth stage) to enhance grain yields of maize and high N recovery from fertilizer. The conventional method is still better option to avoid loss of fertilizer. Additional studies should be conducted to better understand the changes that occur in soil microbiology, soil biochemistry, root composition and N losses.

## REFERENCES

- ALUWIHARE, Y. C. *et al.* Characterization and selection of phosphorus deficiency tolerant rice genotypes in Sri Lanka. **Rice Science**, v. 23, n. 4, p. 184-195, 2016.
- ALVARENGA, R. C. *et al.* Plantas de cobertura de solo para sistema plantio direto. **Informe Agropecuário**, Belo Horizonte, v. 22, p. 25-36, 2001.
- BINDRABAN, P. S. *et al.* Revisiting fertilisers and fertilisation strategies for improved nutrient uptake by plants. **Biology and fertility of soils**, v. 51, n. 8, p. 897-911, 2015.
- BYRNES, R. C. *et al.* Biological nitrification inhibition by Brachiaria grasses mitigates soil nitrous oxide emissions from bovine urine patches. **Soil Biology Biochemistry**, v. 107, p. 156–163, apr. 2017.
- CANTARELLA, H.; QUAGGIO, J. A.; FURLANI, A. C. M. **Recomendações de adubação e calagem para o Estado de São Paulo**. 2nd ed. Tech. Bull. Campinas: Instituto Agrônômico de Campinas, 1997.
- COÛTEAUX, M. M. *et al.* Litter decomposition, climate and litter quality. **Trends in Ecology & Evolution**, v. 10, n. 2, p. 63-66, feb. 1995.

CRUSCIOL, C. A. C. *et al.* Persistência de palhada e liberação de nutrientes do nabo forrageiro no plantio direto. **Pesquisa Agropecuária Brasileira**, v. 40, n. 2, p. 161–168, 2005.

FAGERIA, N. K.; BALIGAR, V. C. Enhancing nitrogen use efficiency in crop plants. **Advances in Agronomy**, n. 88, p. 97–185, 2005.

KARWAT, H. *et al.* Residual effect of BNI by *Brachiaria humidicola* pasture on nitrogen recovery and grain yield of subsequent maize. **Plant and Soil**, v. 420, n. 1-2, p. 389–406, 2017.

KEENEY, D. R.; NELSON, D. W. Nitrogen-Inorganic Forms. In: PAGE, A. L. **Methods of Soil Analysis**. Madison: ASA, SSSA, 1982. p. 643-698.

MALAVOLTA, E. *et al.* **Avaliação do estado nutricional das plantas: princípios e aplicações**. 2nd ed. Piracicaba: Potafos, 1997.

MARIANO, E. *et al.* Mineralisation and sorption of dissolved organic nitrogen compounds in litter and soil from sugarcane fields. **Soil Biology and Biochemistry**, v. 103, p. 522-532, dec. 2016.

MARSCHNER, H. **Mineral nutrition of higher plants**. 3<sup>rd</sup>. ed. London: Academic Press. 2012.

MEHRA, P. *et al.* Impact of carbonates on the mineralisation of surface soil organic carbon in response to shift in tillage practice. **Geoderma**, v. 339, n. 1, p. 94-105, apr. 2019.

MORRIS, T. F. *et al.* Strengths and limitations of nitrogen rate recommendations for corn and opportunities for improvement. **Agronomy Journal**, v. 110, n. 1, p. 1-37, jan. 2018.

OLIVEIRA, O. *et al.* Chemical and biological indicators of decline/degradation of *Brachiaria* pastures in the Brazilian Cerrado. **Agriculture, ecosystems and environment**, v. 103, n. 2, p. 289–300, jul. 2004.

PACHECO, L. P. *et al.* Nutrient cycling by cover crops and yield of soybean and rice in no-tillage. **Pesquisa Agropecuária Brasileira**, v. 48, n. 9, p. 1228-1236, 2013.

PACHECO, L. P. *et al.* Produção de fitomassa e acúmulo e liberação de nutrientes por plantas de cobertura na safrinha. **Pesquisa Agropecuária Brasileira**, v. 46, n. 1, p. 17-25, 2011.

RAO, I. Root distribution and production in native and introduced pastures in the South American savannas. Root demographics and their efficiencies in sustainable agriculture, grasslands and forest ecosystems. **Springer**, p.19–41, 1998.

SUBBARAO, G. V. *et al.* Suppression of soil nitrification by plants. **Plant Science**, v. 233, p. 155-164, apr. 2015.

THORBURN, P. J. *et al.* Modelling decomposition of sugar cane surface residues with APSIM–residue. **Field Crops Research**, v. 70, n. 3, p. 223–232, may. 2001.

USDA, United States Department of Agriculture. **Keys to soil taxonomy**. Twelfth Edition, 2014. 372 p.

USDA, United States Department of Agriculture. **USDA Agricultural Projections to 2028**. 2019. 108p.

YANG, J.; UDVARDI, M. Senescence and nitrogen use efficiency in perennial grasses for forage and biofuel production. **Journal of Experimental Botany**, v. 69, n. 4, p. 855-865, aug. 2017.

## FINAL CONSIDERATIONS

This study investigated during a period of three years (i.e. 2015-2018) the effects of grass cover crops and N management on grain yield, N-use efficiency and N recovery from fertilizer in maize. In no-tillage system, the N fertilizer applied on palisade grass (*Urochloa brizantha*) at 35 DBS or on its residues at 1 DBS, or on ruzigrass residues at 1 DBS resulted in similar grain yield of maize in conventional method of N application. However, since the earlier applications on cover crops had lower N recovery from fertilizer by maize grain, the application of nitrogen fertilizer must be applied as current recommended method (40 kg N ha<sup>-1</sup> at maize seeding plus 120 kg N ha<sup>-1</sup> sidedressed in V<sub>6</sub> growth stage) to enhance grain yields of maize and high N recovery from fertilizer.

## REFERENCES

- ALMEIDA, D. S. *et al.* *Urochloa ruziziensis* cover crop increases the cycling of soil inositol phosphates. **Biology and Fertility of Soils**, v. 54, n. 8, p. 935-947, nov. 2018.
- ARROYAVE, C. *et al.* A proteomic approach to the mechanisms underlying activation of aluminum resistance in roots of *Urochloa decumbens*. **Journal Inorganic Biochemistry**, v. 181, p. 145-151, apr. 2018.
- BASSO, C. J.; CERETTA, C. A. Manejo do nitrogênio no milho em sucessão a plantas de cobertura de solo sob plantio direto. **Revista Brasileira de Ciência do Solo**, v. 24, p. 905–915, 2000.
- BOWATTE, S. *et al.* W. Wide variation in nitrification activity in soil associated with different forage plant cultivars and genotypes. **Grass and Forage Science**, v. 71, p. 160-171, jun. 2015.
- BORGHI, E. *et al.* O. Intercropping time of corn and palisade grass and Guinea grass affecting grain yield and forage production. **Crop Science**, v. 53, n. 2, p. 629–636, mar. 2013.
- BOSCHIERO, B. N.; MARIANO, E.; TRIVELIN, P. C. O. “Preferential” ammonium uptake by sugarcane does not increase the <sup>15</sup>N recovery of fertilizer sources. **Plant Soil**, v. 429, n. 1-2, p. 253-269, aug. 2018.
- BYRNES, R. C. *et al.* Biological nitrification inhibition by *Brachiaria* grasses mitigates soil nitrous oxide emissions from bovine urine patches. **Soil Biology Biochemistry**, v. 107, p. 156–163, apr. 2017.
- CASTRO, G. S. A. *et al.* Management impacts on soil organic matter of tropical soils. **Vadose Zone Journal**, v. 14, p. 1–8, 2015.
- CATUCHI, T. A. *et al.* Nitrogen management of forage grasses for nutrition, seed production, and nutrients in residual straw. **Pesquisa Agropecuária Brasileira**, v. 54, apr. 2019.
- CERETTA, C. A. *et al.* Produção e decomposição de fitomassa de plantas invernais de cobertura de solo e milho, sob diferentes manejos da adubação nitrogenada. **Ciência Rural**, v. 32, n. 1, p. 49–54, 2002.
- COSTA, N. R. *et al.* M. Yield and nutritive value of the silage of corn intercropped with tropical perennial grasses. **Pesquisa Agropecuária Brasileira**, v. 52, p. 63-73, jan. 2017.
- CRUSCIOL, C. A.C. *et al.* Improving soil fertility and crop yield in a tropical region with palisadegrass cover crops. **Agronomy Journal**, v. 107, n. 6, p. 2271-2280, jun. 2015.

CRUSCIOL, C. A. C.; SORATTO, R. P. Nitrogen supply for cover crops and effects on peanut grown in succession under a no-till system. **Agronomy Journal**, v. 101, n. 1, p. 41-46, jan. 2009.

DERPSCH, R. *et al.* Why do we need to standardize no-tillage research? **Soil and Tillage Research**, v. 137, p. 16-22, apr. 2014.

DUARTE, A. P. *et al.* Concentração e exportação de nutrientes nos grãos de milho. **Informações Agronomicas**, v. 163, 2018.

FAGERIA, N. K. *et al.* Root growth, nutrient uptake and use efficiency by roots of tropical legume cover crops as influenced by phosphorus fertilization. **Journal of Plant Nutrition**, v. 39, n. 6, p. 781-792, feb. 2016.

FREITAS, P.L.; LANDERS, J. N. The transformation of agriculture in Brazil through development and adoption of zero tillage conservation agriculture. **International Soil and Water Conservation Research**, v. 2, n. 1, p. 35–46, mar. 2014.

GUZMAN, J.; GOLABI, M. H. Agroecosystem Net Primary Productivity and Carbon Footprint. In: AL-KAISI, M. M. **Soil Health and Intensification of Agroecosystems**. 1. ed. Academic Press, 2017. cap. 10, p. 215-229.

KARWAT, H. *et al.* Residual effect of BNI by *Brachiaria humidicola* pasture on nitrogen recovery and grain yield of subsequent maize. **Plant and Soil**, v. 420, n. 1-2, p. 389–406, nov. 2017.

KUYPERS, M. M. M.; MARCHANT, H. K.; KARTAL, B. The microbial nitrogen-cycling network. **Nature Reviews Microbiology**, v. 16, n. 5, p. 263-276, may. 2018.

LARA CABEZAS, W. A. R. *et al.* Influência da cultura antecessora e da adubação nitrogenada na produtividade de milho em sistema plantio direto e solo preparado. **Ciência Rural**, v. 34, n. 4, p. 1005–1013, 2004.

LARA CABEZAS, W. A. R. *et al.* Imobilização de nitrogênio da uréia e do sulfato de amônio aplicado em pré-semeadura ou cobertura na cultura de milho, no sistema plantio direto. **Revista Brasileira de Ciência do Solo**, v. 29, n. 2, p. 215–226, 2005.

LEITE, L. F. C. *et al.* Decomposição e liberação de nutrientes de resíduos vegetais depositados sobre Latossolo Amarelo no Cerrado Maranhense. **Revista Ciência Agrônômica**, v. 41, n. 1, p. 29–35, jan-mar. 2010.

MATEUS, G. P. *et al.* Sidedress nitrogen application rates to sorghum intercropped with tropical perennial grasses. **Agronomy Journal**, v. 108, n. 1, p. 433-447, jan. 2016.

MCKENZIE, S. V. *et al.* Integration of sheep grazing for cover crop termination into market gardens: Agronomic consequences of an ecologically based management strategy. **Renew Agriculture and Food Systems**, v. 32, n. 5, p. 389–402, sep. 2016.

MOMESSO, L. *et al.* Impacts of nitrogen management on no-till maize production following forage cover crops. **Agronomy Journal**, v. 111, n. 1, p. 639-649, mar. 2019.

MORAES, A. *et al.* Integrated crop-livestock systems as a solution facing the destruction of pampa and Cerrado biomes in South America by intensive monoculture systems. In: LEMAIRE, G. *et al.* **Agroecosystem Diversity**. Academic press, 2018. p. cap. 16, p. 257-273.

MORAES, M. T. *et al.* Soil load support capacity increases with time without soil mobilization as a result of age-hardening phenomenon. **Soil & Tillage Research**, v. 186, p. 128-134, mar. 2019.

MORAES, A. *et al.* Integrated crop-livestock systems in the Brazilian subtropics. **European Journal of Agronomy**, v. 57, p. 4-9, jul. 2014.

PARIZ, C. M. *et al.* Straw decomposition of nitrogen fertilized grasses intercropped with irrigated maize in an integrated crop-livestock system. **Revista Brasileira de Ciência do Solo**, v. 35, n. 6, p. 2029-2037, 2011.

PACHECO, L. P. *et al.* Biomass yield in production systems of soybean sown in succession to annual crops and cover crops. **Pesquisa Agropecuária Brasileira**, v. 52, n. 47, p. 582–591, 2017.

PARIZ, C. M. *et al.* Production, nutrient cycling and soil compaction to grazing of grass companion cropping with corn and soybean. **Nutrient Cycling in Agroecosystems**, v. 108, n. 1, p. 35-54, may. 2017.

PAVINATO, P. S. *et al.* Effects of cover crops and phosphorus sources on maize yield, phosphorus uptake, and phosphorus use efficiency. **Agronomy Journal**, v. 109, p. 1039-1047, may. 2017.

PERVEEN, N. *et al.* Priming effect and microbial diversity in ecosystem functioning and response to global change: A modeling approach using the symphony model. **Global Change Biology**, v. 20, p. 1174–1190, feb. 2014.

PÖTTKER, D.; WIETHÖLTER, S. Épocas e métodos de aplicação de nitrogênio em milho cultivado no sistema plantio direto. **Ciência Rural**, v. 34, n. 4, p. 1015–1020, jul-aug. 2004.

ROCHA, K. F. *et al.* Fate of <sup>15</sup>N fertilizer applied to maize in rotation with tropical forage grasses. **Field Crops Research**, v. 238, p. 35-44, may. 2019.

ROSOLEM, C. A. *et al.* Enhanced plant rooting and crop system management for improved N use efficiency. **Advances in Agronomy**, v. 146, p. 205–239, 2017.

SCHUSTER, M. Z. *et al.* Optimizing forage allowance for productivity and weed management in integrated crop-livestock systems. **Agronomy for Sustainable Development**, v. 39, n.18, apr. 2019.

SILVA, A. P. *et al.* Soil structure and its influence on microbial biomass in different soil and crop management systems. **Soil and Tillage Research**, v. 142, p. 42-53, sep. 2014.

SILVA, M. C. C. *et al.* Soil physical attributes and yield of winter common bean crop under a no-till system in the Brazilian cerrado. **Revista Caatinga**, v. 30, n. 1, jan-mar. 2017.

SUBBARAO, G. V. *et al.* Suppression of soil nitrification by plants. **Plant Science**, v. 233, p. 155-164, apr. 2015.

SUBBARAO, G. V. *et al.* Evidence for biological nitrification inhibition in *Brachiaria* pastures. **Proceedings of the National Academy of Sciences of the United States of America**. v. 106, n.41, p. 17302–17307, oct. 2009.

SUBBARAO, G. V. *et al.* NH<sub>4</sub><sup>+</sup> triggers the synthesis and release of biological nitrification inhibition compounds in *Brachiaria humidicola* roots. **Plant Soil**. v. 290, n. 1-2, p. 245-257, jan. 2007.

TANAKA, K. S. *et al.* Nutrients released by *Urochloa* cover crops prior to soybean. **Nutrient Cycling in Agroecosystems**, v. 113, n. 3, p. 267-281, apr. 2019.

TIMOSSI, P. C.; DURIGAN, J. C.; LEITE, G. J. Formação de palhada por braquiárias para adoção do sistema plantio direto. **Bragantia**, v. 66, n. 4, p. 617-622, 2007.

TIRITAN, C. S. *et al.* Tillage system and lime application in a tropical region: Soil chemical fertility and corn yield in succession to degraded pastures. **Soil & Tillage Research**, v. 155, p. 437-447, jan. 2016.

VAN GROENIGEN, J. W. *et al.* The soil N cycle: new insights and key challenges. **Soil**, n. 1, p. 235-256, 2015.

VERAS, M. S. *et al.* Cover Crops and Nitrogen Fertilization Effects on Nitrogen Soil Fractions under Corn Cultivation in a No-Tillage System. **Revista Brasileira de Ciência do Solo**, v. 40, apr. 2016.

WILCKE, W.; LILIENFEIN, J. Nutrient Leaching in Oxisols Under Native and Managed Vegetation in Brazil. **Soil Science Society of America Journal**, v. 69, n. 4, p. 1152-1161, jul. 2005.

ZHANG, J.; MÜLLER, C.; CAI, Z. Heterotrophic nitrification of organic N and its contribution to nitrous oxide emissions in soils. **Soil Biology and Biochemistry**, v. 84, p. 199-209, may. 2015.