

# Paleodistributions and Comparative Molecular Phylogeography of Leafcutter Ants (*Atta* spp.) Provide New Insight into the Origins of Amazonian Diversity

Scott E. Solomon<sup>1,2,3\*</sup>, Mauricio Bacci, Jr.<sup>3</sup>, Joaquim Martins, Jr.<sup>3</sup>, Giovanna Gonçalves Vinha<sup>3</sup>, Ulrich G. Mueller<sup>1</sup>

**1** Section of Integrative Biology, The University of Texas at Austin, Austin, Texas, United States of America, **2** Department of Entomology, Smithsonian Institution, Washington, D. C., United States of America, **3** Center for the Study of Social Insects, São Paulo State University, Rio Claro, São Paulo, Brazil

## Abstract

The evolutionary basis for high species diversity in tropical regions of the world remains unresolved. Much research has focused on the biogeography of speciation in the Amazon Basin, which harbors the greatest diversity of terrestrial life. The leading hypotheses on allopatric diversification of Amazonian taxa are the Pleistocene refugia, marine incursion, and riverine barrier hypotheses. Recent advances in the fields of phylogeography and species-distribution modeling permit a modern re-evaluation of these hypotheses. Our approach combines comparative, molecular phylogeographic analyses using mitochondrial DNA sequence data with paleodistribution modeling of species ranges at the last glacial maximum (LGM) to test these hypotheses for three co-distributed species of leafcutter ants (*Atta* spp.). The cumulative results of all tests reject every prediction of the riverine barrier hypothesis, but are unable to reject several predictions of the Pleistocene refugia and marine incursion hypotheses. Coalescent dating analyses suggest that population structure formed recently (Pleistocene-Pliocene), but are unable to reject the possibility that Miocene events may be responsible for structuring populations in two of the three species examined. The available data therefore suggest that either marine incursions in the Miocene or climate changes during the Pleistocene—or both—have shaped the population structure of the three species examined. Our results also reconceptualize the traditional Pleistocene refugia hypothesis, and offer a novel framework for future research into the area.

**Citation:** Solomon SE, Bacci M Jr, Martins J Jr, Vinha GG, Mueller UG (2008) Paleodistributions and Comparative Molecular Phylogeography of Leafcutter Ants (*Atta* spp.) Provide New Insight into the Origins of Amazonian Diversity. PLoS ONE 3(7): e2738. doi:10.1371/journal.pone.0002738

**Editor:** Peter M. Bennett, University of Kent, United Kingdom

**Received:** January 10, 2008; **Accepted:** June 10, 2008; **Published:** July 23, 2008

**Copyright:** © 2008 Solomon et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

**Funding:** S.E.S. was funded by a DDIG grant from the National Science Foundation (DEB 0407772), the IGERT program in computational phylogenetics at The University of Texas at Austin, graduate fellowships from the Section of Integrative Biology at The University of Texas at Austin, and a grant from the Amazon Conservation Association and the NSF International Research Fellowship Program (IRFP #07012333). M.B.J. was funded by FAPESP (06/00185-7), CAPES (Aux-UT-165/2005) and CNPq (310826/2006-3 and 479990/2006-9). J.M.J. was funded by CAPES (Brazil). G.G.V. was funded by FAPESP (05/54250-1). Additional funding was provided by NSF IRCEB Grant DEB-0110073 to U.G.M.

**Competing Interests:** The authors have declared that no competing interests exist.

\* E-mail: solomons@si.edu

## Introduction

Tropical regions around the world are well known for their rich diversity of life. Yet, the reasons why the tropics harbor more species than temperate and arctic regions remain unclear [1,2,3,4]. The Amazon Basin has been of particular interest in this matter, as it harbors perhaps the world's greatest terrestrial biodiversity [5,6,7,8]. As is true for the study of speciation in general [9], much focus has been placed on the biogeography of processes generating diversity in the Amazon Basin, specifically on how allopatry can be achieved in a landscape without obvious geographic barriers (although the presence of now invisible barriers, such as ancient "arches" has been suggested [10,11,12,13]). Although a plethora of hypotheses have been suggested, three stand out as the most widely discussed. These are the Pleistocene refugia hypothesis, the marine incursion hypothesis, and the riverine barrier hypothesis.

The Pleistocene refugia hypothesis has been responsible for generating the most interest in the field [12,13,14], but has also become the most heavily criticized [15]. First proposed by Haffer in 1969 [16], this hypothesis suggests that historical climate changes,

specifically during periods of glacial maxima, restricted the distribution of wet forests in Amazonia. Under this model, species that inhabited these forests (birds in Haffer's original hypothesis but later expanded to include all terrestrial species [12]) would likewise have become more isolated, resulting in the possibility for allopatric speciation. Haffer [16] proposed the presence of several Pleistocene forest refugia along the periphery of the Amazon Basin, reasoning that these mountainous regions would have enough surface relief to remain moist, even during periods of widespread aridity, by generating local precipitation [17].

Although some studies [18,19,20,21,22] have found support for the predictions of the Pleistocene refugia hypothesis (see Table 1 for a list of predictions), most have not [23,24,25,26,27]. Furthermore, the refugia hypothesis has been criticized because (1) geological and paleoclimatic data do not generally support the conclusion that wet forests were highly fragmented during the Pleistocene [15,28,29,30]; (2) the locations and size of forest refugia, if they did exist, would be different for each species because of different environmental tolerances [12,14]; (3) some areas that have been proposed as refugia because they appear to

contain greater species diversity can be explained as artifacts of sampling biases [31]; and (4) the ages of many extant Amazonian species pre-date the Pleistocene, suggesting they were generated by earlier mechanisms [12,14,32]. These criticisms have led some researchers to call for the complete dismissal of the Pleistocene refugia hypothesis on the grounds that it has been sufficiently discredited [15].

The marine incursion hypothesis stems from evidence that tectonic events combined with elevated sea levels, most recently during the mid-Miocene (approximately 10–15 mya), flooded much of the Amazon Basin in salty or brackish water [33,34,35,36,37,38]. Such incursions would have restricted all terrestrial organisms inhabiting the Amazon region to become isolated in areas of higher elevation, namely near the Andes to the west, the Guiana Shield to the north, and the Brazilian Shield to the south. Under this model, the resulting isolation would permit allopatric divergence of these populations. Support for the marine incursion hypothesis has so far been found in woodcreepers [23] and freshwater fish [39].

The riverine barrier hypothesis can be traced to early observations on vertebrate distributions by Alfred Russell Wallace [40]. This hypothesis suggests that tropical rivers serve as barriers to gene flow for terrestrial organisms. These rivers, which are wide and numerous in Amazonia, may promote divergence of populations restricted to either side [14,41,42,43,44,45]. This hypothesis has received mixed support. On the one hand, major Amazonian rivers do seem to restrict dispersal of passerine birds [46], small primates [47], lizards [48,49], frogs [50] and Riodinid butterflies [51]. However, extensive molecular and morphological work on small mammals and frogs along the Juruá River, a tributary of the Amazon, failed to detect a significant river barrier effect [10,41,43,44].

Two recent developments have allowed new insights into the predictions made by these hypotheses (see Table 1). First, advances in molecular techniques have not only increased the amount of data available for analysis, they also permit a more quantitative evaluation of species and population histories, which are essential for testing competing hypotheses on tropical diversification [14]. Although molecular reconstructions of the biogeography of past speciation events seems promising, the dynamic nature of species' geographic ranges makes these inferences somewhat tenuous [52]. An alternative approach is to examine the current population structure of widespread species. Such phylogeographic analyses

can provide insight into the processes responsible for generating allopatry by giving not only a snapshot of the current population structure, but also a window into the past through the reconstruction of gene trees and historical demography [14,53,54].

The second recent development combines reconstructions of paleoclimates with a flurry of novel techniques for modeling species distributions under current as well as past (or future) climate conditions. Such “paleodistribution” analyses provide a means of independently assessing the extent to which past climate has influenced species' geographic ranges [55,56,57,58], thereby avoiding assumptions about the presence and location of putative forest refugia and thus bypassing several of the major criticisms of the Pleistocene refugia hypothesis.

Several recent studies have demonstrated the utility of combining molecular phylogeography and paleodistribution reconstruction in a complimentary fashion to test a priori biogeographic hypotheses [55,59,60,61]. However, paleoclimate data for the Amazon basin are not nearly as complete as for other regions, such as the Australian Wet Tropics [62], so such an approach has not yet been utilized for Amazonian species. Furthermore, the few studies that have used a molecular phylogeographic approach to test these supposedly universal hypotheses have primarily focused on vertebrate taxa [10,23,27,43,44,45,63], which represent only a small proportion of the total diversity of the Amazonian region [5,6,7,64].

We used three co-distributed species of leafcutter ants in the genus *Atta* (Formicidae: Attini) to test the Pleistocene refugia, marine incursion, and riverine barrier hypotheses using a combination of paleodistribution modeling and comparative molecular phylogeography. Leafcutter ants are widespread throughout the Neotropics [65,66]. They are generalist herbivores, cutting fresh vegetation as a food source for their mutualistic fungal gardens [67,68]. Due to their tendency to forage on crops and ornamental plants [69], leafcutter ants are considered to be major agricultural pests, and have been described as the dominant herbivores of the Neotropics [66,70]. They also play a key ecological role in nutrient cycling as they bring organic material deep into their subterranean nests [71,72].

Three leafcutter ant species, *A. cephalotes*, *A. sexdens*, and *A. laevigata*, are ideal for testing the hypotheses in question because (1) they co-occur throughout much of the Amazon Basin, as well as in adjacent areas [65,73], (2) they diversified within the relevant time frame for the hypotheses in question [74], (3) the three species

**Table 1.** Summary of the predictions of each hypothesis and overview of the methods used to test them (\*Diversification any time subsequent to the formation of the Amazon river (5–12 mya) would be consistent with the riverine barrier hypothesis, therefore only diversification prior to 12 mya would falsify this prediction; we chose not to use this as a test of the riverine barrier hypothesis because it is nearly impossible to reject for these species, which originated no more than 14 mya [81].).

Predictions	Pleistocene refugia	Marine incursion	Riverine barrier	Methods used
Reciprocal monophyly of populations:	in different refugia	in Eastern base of Andes, Brazilian Shield, and/or Guiana Shield	on opposite banks of Amazon River	Parametric bootstrap, Bayesian hypothesis tests
Basal populations are located:	in refugia	in Eastern base of Andes, Brazilian Shield, and/or Guiana Shield	N/A	ML and Bayesian gene tree reconstruction
Derived populations are located:	outside refugia	in Amazonian lowlands	N/A	ML and Bayesian gene tree reconstruction
Barrier to gene flow:	areas between refugia	Amazonian lowlands	Amazon River	AMOVA, Mantel tests
Population history includes:	bottlenecks and expansion	bottlenecks and expansion	N/A	Mismatch distributions, Tajima's D
Population structure formed:	during Pleistocene (10 kya–1.8 mya)	during Miocene (10–15 mya)	N/A*	IM

doi:10.1371/journal.pone.0002738.t001

differ in their environmental tolerances [75,76], permitting an evaluation of how historical climatic changes have differentially influenced each, and (4) they can be easily collected due to their enormous colony sizes [66,77].

We used these three species as independent tests of the predictions of each hypothesis (summarized in Table 1). Furthermore, we hypothesized that, since these species have similar distributions, dispersal abilities, and life histories [65,73,75,76,78,79,80], the riverine barrier hypothesis and marine incursion hypothesis should both apply equally to all three species. However, because the three species chosen in this study display a continuum of tolerance to aridity, such that *A. cephalotes* is the least tolerant of aridity, *A. laevigata* the most tolerant, and *A. sexdens* intermediate between the two [76], we hypothesized that each species would respond differently to historical climate change during the Pleistocene. Specifically, we predicted that increasing aridity during the Pleistocene, reaching a climax at the last glacial maximum (LGM; approximately 21,000 ybp) would have most restricted the distribution of the least arid-tolerant species, *Atta cephalotes*, while expanding the range of the most arid-tolerant, *Atta laevigata*, with *A. sexdens* affected to an intermediate extent. To test these predictions, we used a rigorous statistical framework combining paleodistribution modeling with gene tree reconstructions, population genetic analyses, historical demographic analyses, and coalescent dating analyses.

## Results

Maps comparing the potential geographic range of each species under current conditions and during the last glacial maximum (LGM), approximately 21 kya, are shown in Figure 1. For current conditions, the area under the receiver operating characteristic curve (AUC) was 0.996, 0.983, and 0.986 for *A. cephalotes*, *A. laevigata*, and *A. sexdens*, respectively. Furthermore, out of the ten different thresholds (see Methods) used to obtain a binary (i.e. presence/absence) prediction, all ten were significantly better than random models for all three species. The cumulative probability thresholds (chosen such that they minimized the commission (false positive) rate for current conditions; see Methods) for *A. cephalotes*, *A. sexdens*, and *A. laevigata* were 1, 5, and 5, respectively.

The projected distribution of each species at the LGM is shown in panels D–F of Figure 1. Putative refugial areas, used for subsequent hypothesis testing, were defined as contiguous areas (i.e. solid green in Figure 1D–F) projected to have been suitable habitat for a given species at the LGM (areas within colored circles in Figure 1D–F). Areas predicted to have been suitable at the LGM, but for which no samples were obtained, were logically excluded for the purposes of hypothesis testing. For *A. cephalotes*, the potential LGM range spanned most of the Amazon Basin, with a contiguous population throughout the Guiana Shield (Figure 1D). This range is somewhat reduced from the estimated current potential distribution of the species (Figure 1A.). Other areas with high probability of occurrence during the LGM include the Atlantic Coastal Forests of Brazil, lower Central America and the Chocó region of South America west of the Andes, and upper Central America into central Mexico (the latter two regions are separated by an area, corresponding to modern day Honduras, predicted to have only very small patches of suitable habitat and was therefore not considered a refugium for hypothesis testing). For *A. sexdens*, the paleodistribution model predicts a more fragmented potential distribution during the LGM (Figure 1E). The largest block of inhabitable range was in the southwestern Amazon Basin, from approximately just west of Manaus to the southwestern edge of the Peruvian Andes. Other blocks of

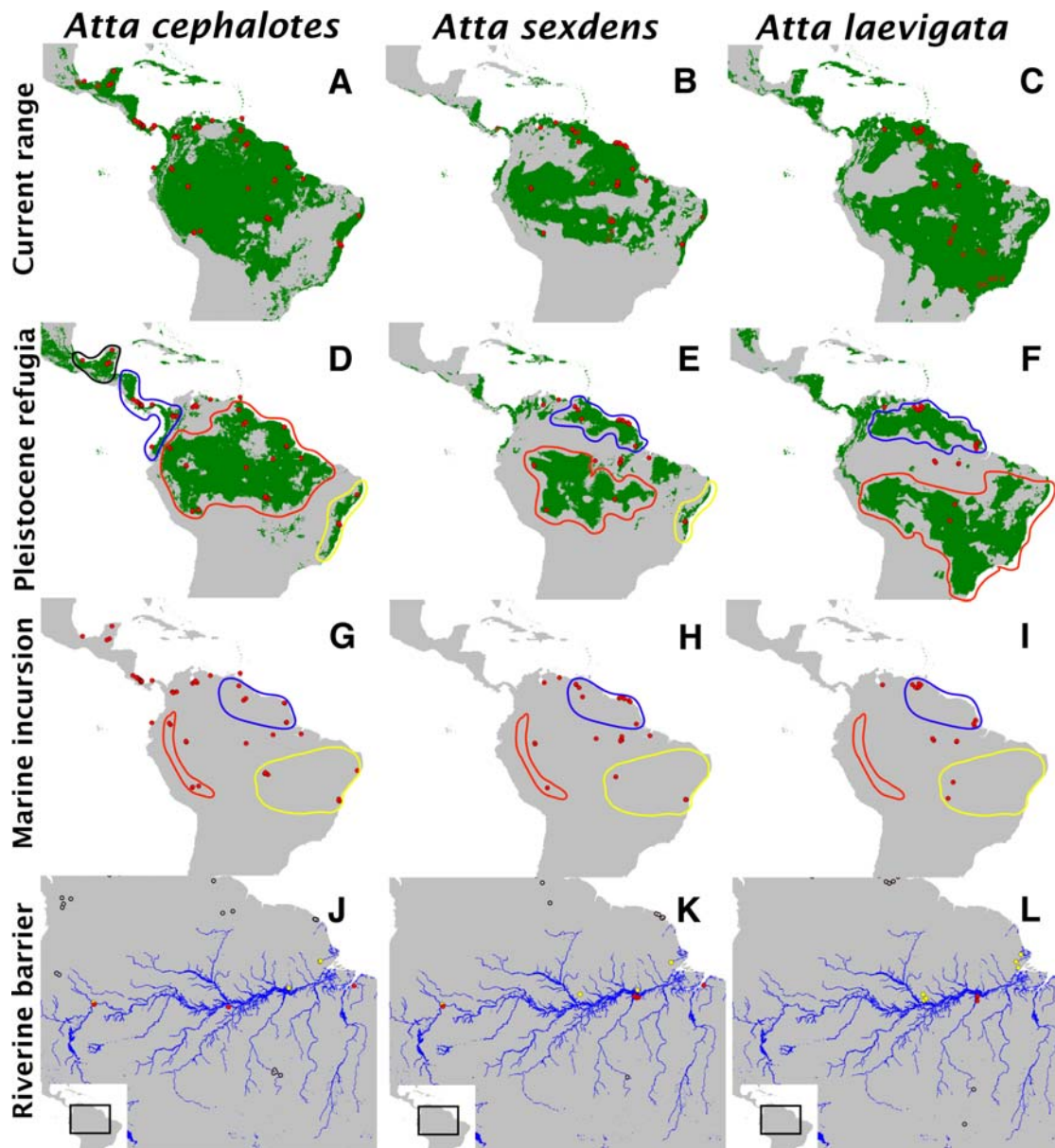
inhabitable areas during the LGM for *A. sexdens* include the Guiana Shield, the Atlantic Coastal Forests of Brazil, an area south of the mouth of the Amazon River roughly between Belem and São Luis, northwestern Colombia/eastern Panama, and Nicaragua. For *A. laevigata*, the model predicted the presence of a large area of unsuitable habitat spanning much of the Amazon Basin (Figure 1F). The remaining areas of suitable habitat occur to the north and south of the Amazon Basin, and are themselves somewhat fragmented.

The topologies of mitochondrial gene trees are shown in Figures 2–4. With one exception, these topologies were not consistent with reciprocal monophyly of the populations predicted by any of the three hypotheses, as determined by parametric bootstrap and Bayesian hypothesis tests (Table S2). The exception was the gene tree for *A. sexdens*, in which the populations predicted by the Pleistocene refugia hypothesis were reciprocally monophyletic (parametric bootstrap  $p=0.15$ ; Bayesian posterior probability = 0.843). However, the gene trees for *A. cephalotes* and *A. laevigata* did have the relevant basal and derived populations as predicted by both the Pleistocene refugia and marine incursion hypotheses. The gene tree for *A. sexdens* is split at the base into two reciprocally monophyletic clades that correspond to geographically distinct populations, such that no statement could be made about which populations are basal versus derived.

Population genetic analyses (AMOVA and Mantel tests) failed to find any evidence that the lower Amazon River has served as a barrier to gene flow for any of the three species (Tables S3–S4). For the Pleistocene refugia and marine incursion hypotheses, analyses of molecular variance (AMOVA) rejected the predicted barrier in all cases (Table S3) except for the barrier predicted by the Pleistocene refugia hypothesis for *A. cephalotes* (40.19% of variance explained by the refugia dictated by paleoclimate reconstructions;  $p=0.00098$ ). In contrast, partial Mantel tests (Table S4) could not reject the barrier predicted by the Pleistocene refugia or marine incursion hypotheses for any of the three species (*A. cephalotes*: marine incursion  $r=-0.149$ ,  $p=0.00003$ ; Pleistocene refugia  $r=0.076$ ,  $p=0.00589$ ; *A. sexdens*: marine incursion  $r=-0.396$ ,  $p=0.00138$ ; Pleistocene refugia  $r=0.251$ ,  $p=0.00009$ ; *A. laevigata*: marine incursion/Pleistocene refugia  $r=0.472043$ ,  $p=0.0073$ ).

Evidence for population bottlenecks and subsequent expansions was mixed in the two tests used (Table S5). For the purposes of discussion, an inference of population expansion was only made in the three instances in which both goodness-of-fit measures used to evaluate mismatch distributions, as well as Tajima's  $D$  statistic, were all consistent with population expansion (*A. cephalotes*, Pleistocene refugia: Atlantic Coast population [SSD = 0.0200829,  $p=0.299$ ; Harpending's Raggedness Index = 0.08930211,  $p=0.3$ ; Tajima's  $D=-1.65893$ ,  $p=0.033$ ]; *A. sexdens*, marine incursion: Brazilian Shield population [SSD = 0.20368588,  $p=0.137$ ; Harpending's Raggedness Index = 0.47,  $p=0.191$ ; Tajima's  $D=-1.21852$ ,  $p=0.026$ ]; *A. laevigata*, marine incursion/Pleistocene refugia: Guiana Shield population [SSD = 0.01959799,  $p=0.181$ ; Harpending's Raggedness Index = 0.10577614,  $p=0.212$ ; Tajima's  $D=-2.31554$ ,  $p=0$ ]). In all other instances, at least one statistic was inconsistent with population expansion, or there were insufficient data.

Coalescent dating analyses that estimated the oldest measurable split ( $T_{div}$ ) between extant populations for each species are shown in Figure 5. The mode, upper, and lower 95% confidence intervals of  $T_{div}$  are given in Table S6. In all three species, the posterior distribution of  $T_{div}$  has a peak within the Pleistocene, and a long tail that extends into the Pliocene and/or Miocene. The long tail results in a rather wide 95% confidence interval, and is partially



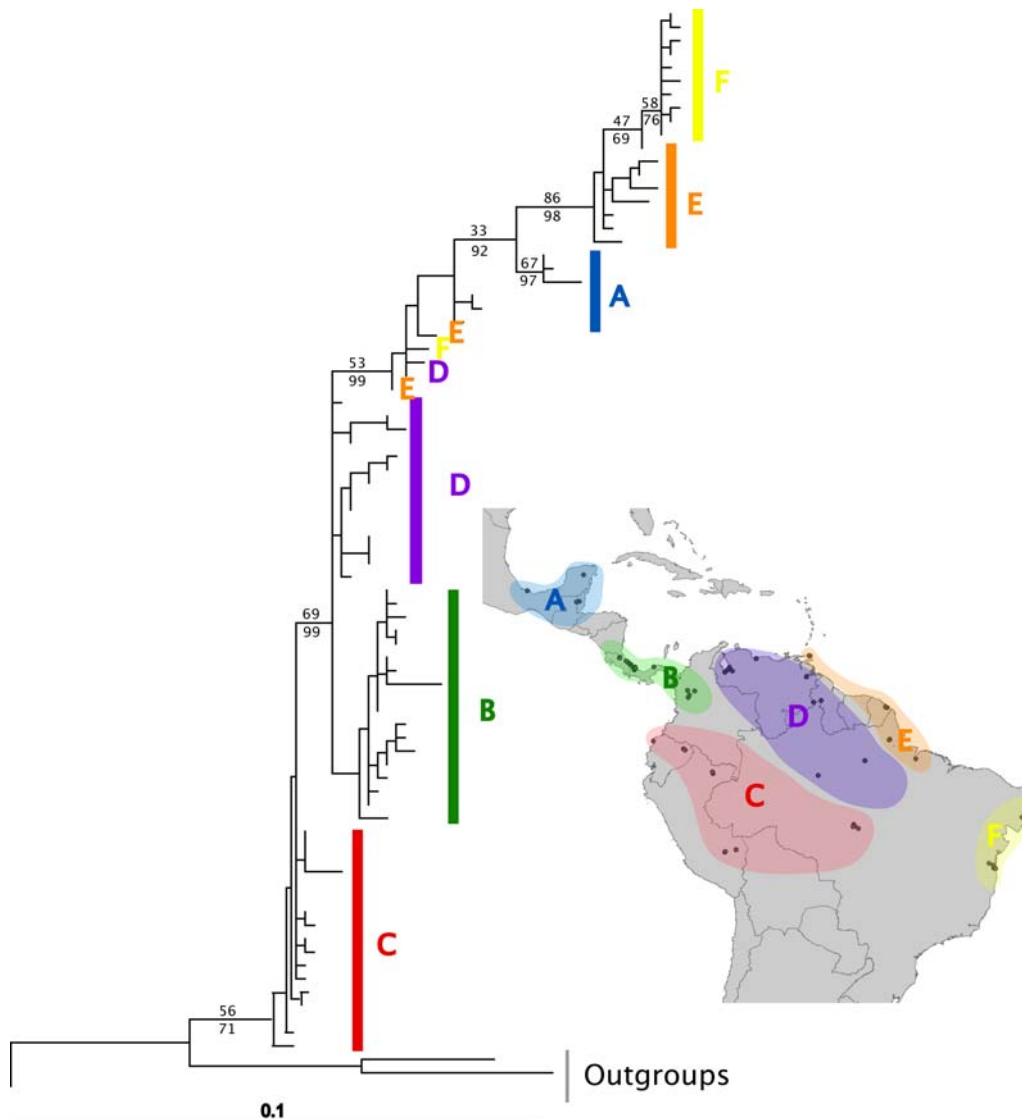
**Figure 1. Overview of populations sampled and groupings used in hypothesis tests (left to right: *Atta cephalotes*, *Atta sexdens*, *Atta laevigata*).** A–C: Results of maxent binary distribution models for the three species under current conditions. Areas predicted to be suitable for each species under current climate conditions are shaded in green. Populations used in this study are shown with red circles; populations for which molecular data were obtained are indicated by filled circles, while populations used only for distribution modeling are indicated by open circles. D–F: Paleodistributions of the three species at the LGM (21 kya) estimated by projecting the maxent model for current conditions onto climate layers from the LGM. Red circles indicate populations used in molecular analyses; Regions outlined with colored lines show population groupings used to test the Pleistocene refugia hypothesis. G–I: Population groupings used to test the marine incursion hypothesis are circled with colored lines (red = Andes, blue = Guiana Shield, yellow = Brazilian Shield); populations for which molecular data were obtained are indicated by filled circles. J–L: Populations used to test the riverine barrier hypothesis are shown with yellow or red circles, indicating populations located north or south of the Amazon river, respectively. Populations for which molecular data were obtained but are located away from the Amazon river (and are therefore not considered in tests of this hypothesis) are shown with empty, black circles.  
doi:10.1371/journal.pone.0002738.g001

due to the high value ( $t_{\max} = 133$ , corresponding to 14 mya) used as an upper bound for the time since divergence in all three species. This value was chosen based on the results of dating analyses for the tribe Attini, in which the crown group of leafcutter ants were estimated to have originated between 8 and 14 mya [81]. The value for  $t_{\max}$  used in this study is thus somewhat conservative and likely extended the 95% confidence interval farther than would a lower value; however, given the data

currently available, it would not be justified to use a lower value for  $t_{\max}$ .

The 95% confidence interval for population divergence in *Atta cephalotes* extends from the mid-Pleistocene (819 kya) to the lower Pliocene (4.893 mya), but does not include the Miocene (Figure 5). It therefore appears that the population structure currently present in *A. cephalotes* formed too recently to be explained by marine incursions during the Miocene. For the other two species,





**Figure 2. Maximum likelihood gene tree for *Atta cephalotes*.** Support values are 100 ML Bootstrap (top) and Bayesian posterior probabilities (bottom). Outgroup sequences used for rooting were from *A. columbica*, *A. texana*, and *A. mexicana*. Uppercase letters correspond to regions shown on map.

doi:10.1371/journal.pone.0002738.g002

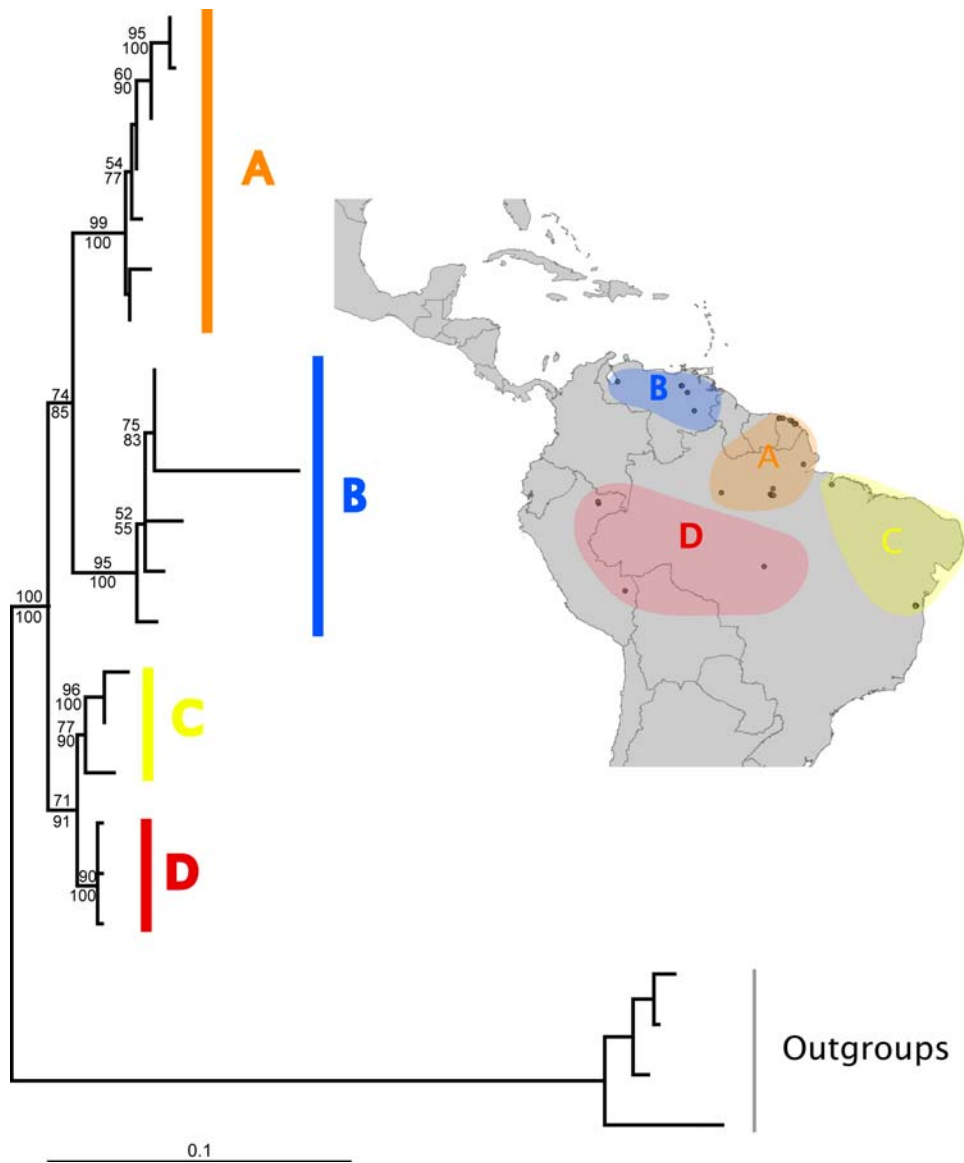
however, the upper 95% confidence limit (13.279 mya and 12.817 mya in *A. sexdens* and *A. laevigata*, respectively) extends into the Miocene, including the period between 10 and 15 mya when marine incursions into the Amazon Basin are thought to have achieved their greatest extent [38]. The wider confidence interval in these two species may also be due to the smaller sample sizes for *A. sexdens* (N = 46) and *A. laevigata* (N = 30) compared with *A. cephalotes* (N = 118).

## Discussion

Combining the results of the paleodistribution models with the molecular phylogeographic analyses (Table 2), the accumulated data rejected every prediction of the riverine barrier model for all three species examined. The results of AMOVA and Mantel tests for the presence of barriers to gene flow, as well as the topologies of mitochondrial gene trees (in which closely related haplotypes are found on opposite river banks), suggest that gene flow regularly

occurs across the lower Amazon River in all three species. Although the exact dispersal abilities of *Atta* species are not known, typical flight distances for mated queens are thought to be less than 2 km (Mueller, pers. obs.), with a maximum range of no more than 50 km [82]. The main channel of the lower Amazon river (e.g. near the city of Santerém) is between 1 and 3 km in width, although the seasonal floodplain can be 20 to 40 km wide in the wet season [83]. The floodplain width is probably more relevant as a dispersal barrier to leafcutter ants since they do not survive in seasonally inundated soils (Solomon, pers. obs.). Although the potential barrier effects of other major rivers in the Amazon Basin were not tested in this analysis, the lack of a significant effect of the lower Amazon River suggests that smaller rivers are unlikely to structure populations of leafcutter ants.

In contrast, discriminating between the Pleistocene refugia and marine incursion hypotheses was more difficult. This difficulty is due in part to the similar predictions that these hypotheses make (Table 1), since some areas reconstructed as refugia are also areas



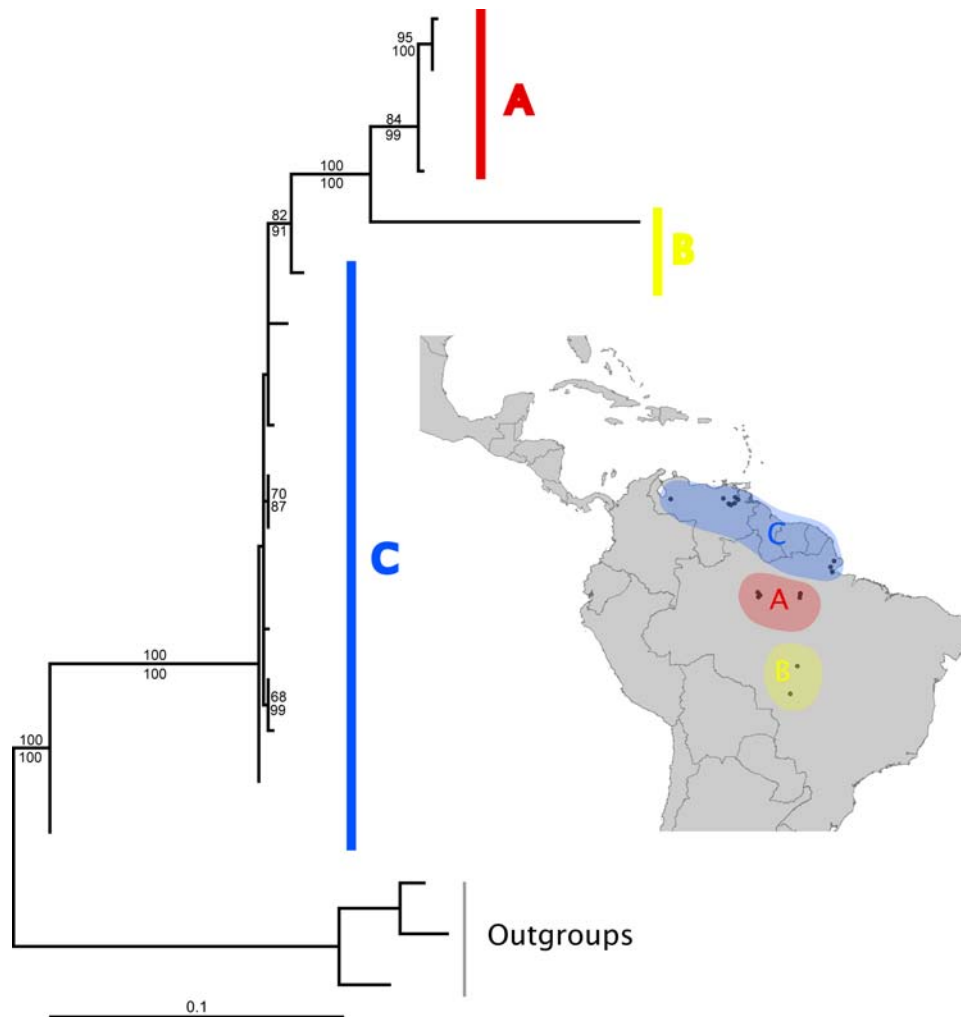
**Figure 3. Maximum likelihood gene tree for *Atta sexdens*.** Support values are 100 ML Bootstrap (top) and Bayesian posterior probabilities (bottom). Outgroup sequences used for rooting were from *A. laevigata*. Uppercase letters correspond to regions shown on map. doi:10.1371/journal.pone.0002738.g003

that would have avoided flooding during marine incursions (Figure 1) [39,84,85]. However, by reconstructing the paleodistribution of each species independently, our approach to testing the Pleistocene refugia hypothesis avoids this issue (in part; see below) since (1) we do not make the assumption that only areas of high surface relief served as refugia, and (2) the areas reconstructed as refugia are different for each species whereas the areas that avoided marine incursions are the same for each species. Nevertheless, the reconstructed paleodistribution for *Atta laevigata* at the LGM (Figure 1F) coincides with the areas unaffected by marine incursions (i.e. the Brazilian Shield and the Guiana Shield), so the predictions of these two hypotheses were largely identical for this species.

Paleodistribution modeling of species ranges during the LGM also addresses one of the major criticisms of the Pleistocene refugia hypothesis, namely that the locations and size of putative forest refugia are likely to be different for every species considered [12,14]. The results of paleodistribution models in the current

study strengthen this argument, since each of the congeneric species examined is predicted to have responded differently to environmental conditions at the LGM (Figure 1). Interestingly, the paleoclimate model used in this study predicts that conditions supporting wet forests persisted throughout much of the Amazon Basin during the LGM, as is suggested by an increasing amount of fossil pollen and other geological information [15,29]. However, this reconstruction of Pleistocene climate conditions contradicts claims by proponents of the refugia model that wet forest only existed along the margins of the Amazon Basin during the LGM [13,16,17,86].

The molecular data provided mixed support for the predictions of the Pleistocene refugia and the marine incursion hypotheses (Table 2). Reciprocal monophyly of the relevant populations was only found in one instance: the gene tree of *Atta sexdens* as predicted by the Pleistocene refugia hypothesis. However, failure to detect reciprocal monophyly does not necessarily indicate that the predictions of a given hypothesis have been invalidated, since



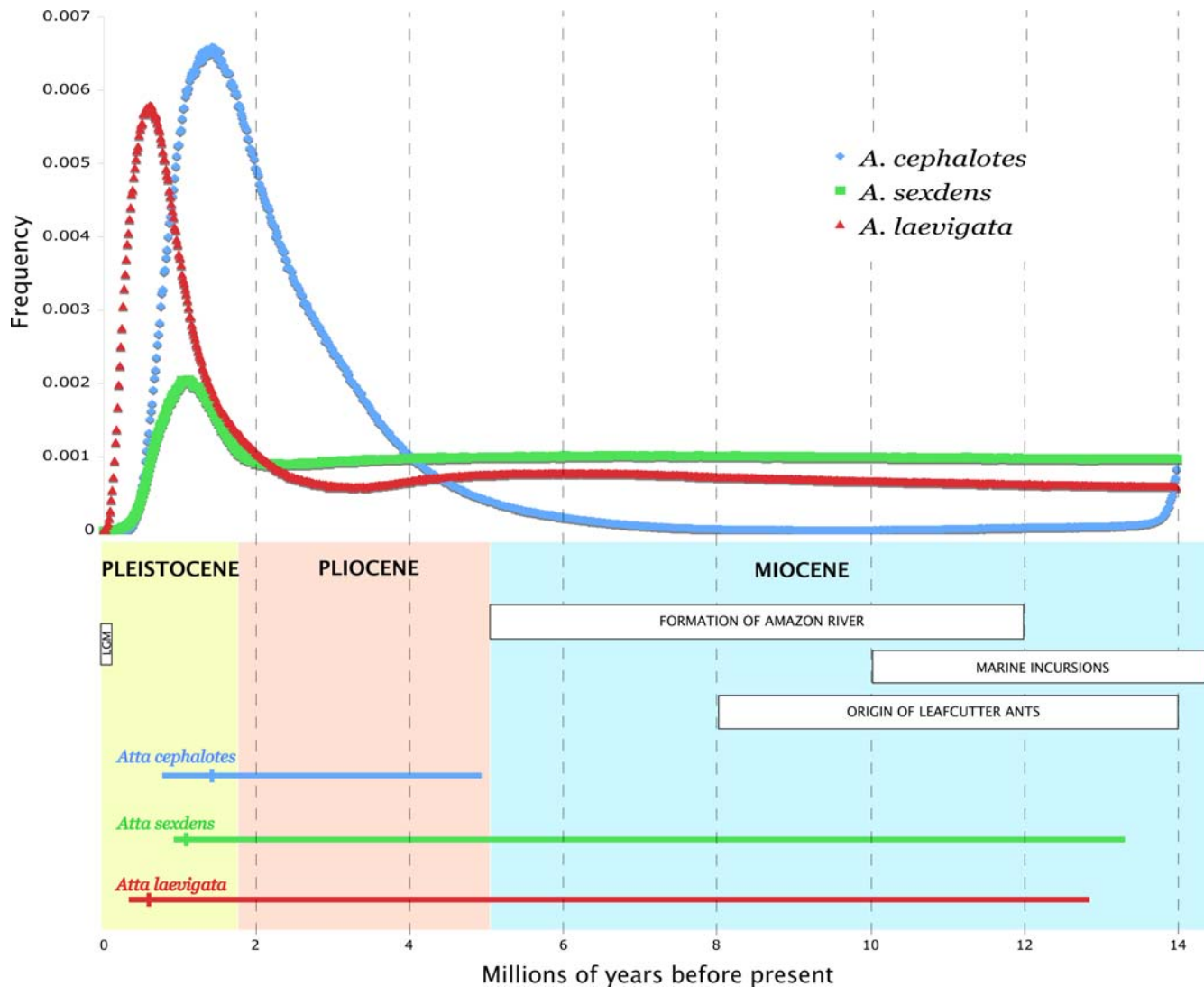
**Figure 4. Maximum likelihood gene tree for *Atta laevigata*.** Support values are 100 ML bootstrap replicates (top) and Bayesian posterior probabilities (bottom). Outgroup sequences used for rooting were from *A. sexdens*. Uppercase letters correspond to regions shown on map. doi:10.1371/journal.pone.0002738.g004

incomplete lineage sorting is expected to also produce paraphyletic and polyphyletic gene trees as populations are split by barriers to gene flow [87]. Thus, although detecting reciprocal monophyly provides strong support for the predicted genealogical history of a species, failure to detect it does not necessarily indicate that the relevant populations are not diverging in the expected manner, especially if the suspected barrier promoting divergence appeared recently. Visual inspection of the gene tree topologies (Figures 2–4) offers an alternative way of interpreting a species genealogical history that is less sensitive to the effects of incomplete lineage sorting. Such an approach shows that the populations expected to be more basal and/or more derived did in fact occupy the positions predicted for *Atta cephalotes* by the Pleistocene refugia hypothesis, but not the marine incursion hypothesis. The gene tree for *Atta laevigata* shows the predicted positions for both these hypotheses (which make identical predictions, as explained above). However, because of the reciprocal monophyly of the relevant populations of *Atta sexdens*, the gene tree could not determine which populations were more basal or derived for this species.

The Pleistocene refugia and marine incursion hypotheses predict that areas that were historically unsuitable for a species to occur (due to inappropriate climatic conditions for the former, flooded areas for the latter) formed barriers to gene flow. The two

methods used to test for the presence of these barriers (AMOVA and Mantel tests) did not always provide congruent results (Table 2, and Tables S3–S4). In fact, the Mantel tests failed to reject the barrier of interest in all six instances, while the AMOVA found no evidence for the barrier of interest in the case of the marine incursion hypothesis for *Atta cephalotes*, both the marine incursion and Pleistocene refugia hypotheses for *Atta sexdens*, and both hypotheses (which, again, make identical predictions) for *Atta laevigata*. It would therefore appear that the AMOVA is a more sensitive way of testing for the presence of gene flow barriers, although we are not aware of any studies that directly compare the discriminatory power of these two commonly used tests.

The two methods used to test for population bottlenecks and subsequent expansions, as predicted by both the Pleistocene refugia and marine incursion hypotheses, also occasionally gave conflicting results (Table 2). We interpreted the results in a conservative manner, such that population bottlenecks and expansions were only inferred in instances in which the results were unanimously consistent with such a demographic history. This was the case in three instances: one population (“Atlantic Coast”) of *A. cephalotes* predicted by the Pleistocene refugia hypothesis, one population (“Brazilian Shield”) of *A. sexdens* predicted by the marine incursion hypothesis, and one population



**Figure 5. Timeline of diversification in Amazonian *Atta* species.** *Top:* Posterior distributions of  $T_{div}$ , the time since the oldest population division for each species as reconstructed for each species using the program IM. *Bottom:* The 95% confidence limits for diversification in each species are represented by horizontal bars, with a vertical bar indicating the best estimate for  $T_{div}$ , assuming a mutation rate of 9.5 substitutions per site per million years and a generation time of 4 years.  
doi:10.1371/journal.pone.0002738.g005

(“Guiana Shield”) of *A. laevigata* predicted by both hypotheses. The results of these tests therefore do not provide much assistance in discriminating between the Pleistocene refugia and marine incursion hypotheses.

Despite the largely similar predictions about geographic population structure made by the Pleistocene refugia and marine incursion hypotheses, these hypotheses operate on vastly different temporal scales. On the one hand, marine incursions are thought to have taken places during the Miocene [33,34,37,39,84], (approximately 10–15 mya), whereas conditions thought to promote refugia existed, generally speaking, during the Pleistocene (1.8 mya–10 kya) and reached a climax during the last glacial maximum (21 kya) [13,16,17,18,88]. The results of our coalescent dating analyses indicate that the population structure observed today in all three species formed between 371,000 and 13.279 million years ago, a time period spanning the Pleistocene, Pliocene, and late Miocene. The 95% confidence interval for population divergence in *A. cephalotes* does not include the Miocene, suggesting that marine

incursions are unlikely to have been responsible for forming the population structure observed today in this species. However, despite a trend toward diversification during the Pleistocene in all three species (Figure 5), marine incursions could not be ruled out as the source of population divergence in *A. sexdens* or *A. laevigata*. More precise dating of the origin of each of these species will require a species-level phylogenetic analysis of the leafcutter ants as well as their closest living relatives, *Trachymyrmex*, which have fossils that can provide calibration points [81].

Although our results are not able to differentiate between many of the predictions of the Pleistocene refugia and marine incursion hypotheses, except possibly for *A. cephalotes*, it is important to recognize, as some other authors have also noted [12,23,25,50], that these hypotheses are not necessarily mutually exclusive. Marine incursions in the Miocene could have been followed by isolation into refugia during the Pleistocene, with species present at both times being affected by both. It is therefore possible that our inability to differentiate between these two “competing” hypotheses may be



**Table 2.** Overview of results.

Species	Prediction	Test	Riverine barrier	Pleistocene refugia	Marine incursion
<i>A. cephalotes</i>	Reciprocal monophyly of relevant populations	Parametric bootstrap	1/1	1/1	1/1
		Bayesian hypothesis tests	1/1	1/1	1/1
	Relevant basal and derived populations	ML and Bayesian trees	N/A	0/1	1/1
	Evidence for predicted barrier to gene flow	AMOVA	1/1	0/1	1/1
		Mantel Tests	1/1	0/1	0/1
	History of population expansions	Mismatch Distributions	N/A	1/8	0/6
		Tajima's <i>D</i>	N/A	2/4	3/3
	Appropriate age of oldest population division	IM	N/A	0/1	1/1
<i>A. sexdens</i>	Reciprocal monophyly of relevant populations	Parametric bootstrap	1/1	0/1	1/1
		Bayesian hypothesis tests	1/1	0/1	1/1
	Relevant basal and derived populations	ML and Bayesian trees	N/A	*	*
	Evidence for predicted barrier to gene flow	AMOVA	1/1	1/1	1/1
		Mantel Tests	1/1	0/1	0/1
	History of population expansions	Mismatch Distributions	N/A	3/6	1/4
		Tajima's <i>D</i>	N/A	3/3	1/2
	Appropriate date for oldest population division	IM	N/A	0/1	0/1
<i>A. laevigata</i> **	Reciprocal monophyly of relevant populations	Parametric bootstrap	1/1	1/1 **	1/1 **
		Bayesian hypothesis tests	1/1	1/1 **	1/1 **
	Relevant basal and derived populations	ML and Bayesian trees	N/A	0/1 **	0/1 **
	Evidence for predicted barrier to gene flow	AMOVA	1/1	1/1 **	1/1 **
		Mantel Tests	1/1	0/1 **	0/1 **
	History of population expansions	Mismatch Distributions	N/A	0/2 **	0/2 **
		Tajima's <i>D</i>	N/A	1/2 **	1/2 **
	Appropriate date for oldest population division	IM	N/A	0/1 **	0/1 **

The number of instances (statistical tests per species or per population) in which the relevant prediction could be rejected are indicated followed after a slash by the total number of instances (e.g. 1/2 means that one out of the two tests rejected the prediction); predictions which are not applicable are indicated by "N/A" ("the gene tree for *A. sexdens* could not resolve which populations were basal or derived; \*\*the predictions for the Pleistocene refugia and marine incursion models are identical for *A. laevigata*).

doi:10.1371/journal.pone.0002738.t002

more a function of their inherent compatibility than their mutual exclusivity.

It should also be noted that, despite some support for the Pleistocene refugia hypothesis, our results do not support its traditional formulation, namely that isolated pockets of wet forest at the periphery of the Amazon Basin were the refugia responsible for diversification in all species [13,17]. Instead, the results of our paleodistribution reconstructions, and to a large extent the genetic data, suggest that the species restricted to wet forest, *A. cephalotes*, was the most widespread at the LGM, while the species most closely associated with open habitat, *A. laevigata*, was the most fragmented. This result is exactly opposite that predicted by the traditional Pleistocene refugia hypothesis, and suggests a more general model for the role of Pleistocene climate change in generating diversity in the Amazon region. Instead of restricting the role that allopatry has played only to inhabitants of wet, lowland forests, it seems likely that inhabitants of all Amazonian habitats should be subject to distributional shifts that could generate population structure.

Indeed, the emerging picture of Amazonia during the Pleistocene, based on data from fossil pollen [15,28,29,30], simulations of paleoclimate, paleohabitat, and species' paleodistributions [58,89], and, increasingly, by genetic data from Amazonian species [23] all point toward a similar scenario: temperatures, precipitation and carbon dioxide levels were all lower than today, but forests

nevertheless remained widespread, and therefore species restricted to forest habitats were not dissected in the way envisioned by Haffer and colleagues [13,16,17,86]. Nevertheless, our results suggest that these climate changes, perhaps acting on top of effects from earlier events such as marine incursions, may have been sufficient to drive diversification in some Amazonian species.

Our results suggest that the role that climate change has played in the diversification of Amazonian species should be revisited, but that other mechanisms that may act in concert should also be considered. That climate change in general is linked to diversification processes is also suggested by a number of recent studies that span various taxa, time periods and geographic regions [63,90,91,92,93,94,95]. The relationship between climate change and diversity is of particular interest for predicting the biotic effects of future climate change [96,97,98,99,100]. Combining paleodistribution modeling with comparative, molecular phylogeography across a diversity of taxa is likely to provide a productive framework for future research into this area.

## Materials and Methods

### Collection of samples and molecular analyses

Samples for molecular analysis were obtained from 118 *Atta cephalotes* colonies, 46 *Atta sexdens* colonies, and 30 *Atta laevigata*

colonies, spanning the known geographic range of each species (Table S7). Sampling locations were chosen to allow testing of the hypotheses in question and to maximize coverage within each species' geographic range. Individual worker ants were collected at nests or along foraging trails and preserved in 95% ethanol during transport to The University of Texas at Austin (for samples collected outside Brazil) or São Paulo State University (UNESP), Rio Claro, SP, Brazil (for samples collected in Brazil), where they were stored at 4°C. The location of all samples was recorded using a handheld GPS unit (Garmin eTrex).

Two disjunct sections of mitochondrial DNA, encompassing part of the Cytochrome Oxidase I (COI) and tRNA-Leucine (tRNA<sub>Leu</sub>) genes, as well as the entire intergenic spacer between COI and tRNA<sub>Leu</sub> were sequenced for all samples. The sequences were concatenated into a single alignment for each species that varied in length from 635 base pairs in *A. cephalotes* to 701 base pairs in *A. sexdens* and *A. laevigata*. Several nuclear pseudogenes were accidentally amplified and sequenced for *A. cephalotes* (described in [101]) and were not used in subsequent analyses; all sequences included in the final alignments for each species appeared to be functional, mitochondrial loci, as no premature stop codons or frameshift mutations were detected. Additional sequences for outgroup taxa (*Atta columbica*, *Atta mexicana*, and *Atta texana*; see Table S7), used for phylogenetic analyses of *A. cephalotes* were obtained from specimens available in the Mueller Lab at The University of Texas at Austin. Sequence information for all samples was deposited in GenBank (Accession Numbers EU847821-EU848214).

Total genomic DNA was extracted from one individual per colony using either the DNeasy Blood and Tissue Kit (QIAGEN) or the AccuPrep Genomic DNA Extraction Kit (Bioneer, Inc.). Several sets of mtDNA primers (Table S1 and References S1) were used to amplify two sections of the cytochrome oxidase I (COI) gene, as well as an intergenic spacer, and a portion of the tRNA-Leucine gene. PCR reactions contained 1 ul each of genomic DNA (approximately 10 ng), 1X reaction buffer, dNTPs, and MgCl<sub>2</sub>, 0.04 ul of Taq polymerase, and 5.96 ul of water for a total reaction volume of 10 ul. Average PCR conditions were as follows, with slight modifications depending on the annealing temperatures of individual primer pairs: Initial denaturation at 95°C for 3 minutes was followed by 35 cycles of 95°C for 5 seconds, and an annealing temperature that increased by 0.5°C for each successive round of amplification, beginning at 45°C, for 20 seconds each round, with a final elongation step of 68°C for 15 seconds. PCR products were analyzed by running 3 ul of the product on a 1.5% agarose gel and subsequently visualized with ethidium bromide staining. For samples that successfully amplified, the remaining 7 ul of PCR product were purified by polyethylene glycol (PEG) precipitation, using a 1:1 PCR product/20% PEG mixture which was incubated for 15 min at 37°C followed by a 10-min centrifugation at 2,688×g and two washes with 80% ethanol.

Cycle sequencing reactions were performed for both forward and reverse sequences using the ABI BigDye Terminator Kit (version 3.1). Sephadex column purification was used to clean the cycle-sequencing product, which was then analyzed on a PRISM 3100 genetic analyzer (Applied Biosystems). Forward and reverse sequences were assembled into individual contigs using SeqMan II v.5.05 (DNASTAR) and alignments between sequences were created initially using Clustal X [102] and then adjusted manually in MacClade v. 4.06 [103].

### Paleodistribution modeling

Estimates of the current and historical potential geographic ranges of each species were made using Maxent version 2.3 [104].

Maxent uses presence-only species occurrence records (i.e. latitudes & longitudes of known species sightings) and environmental data (i.e. GIS layers) as input. In general, the maxent approach seeks to estimate an unknown ("target") distribution using incomplete information about the target distribution and a given set of constraints. For modeling species potential geographical ranges, the occurrence data are considered to be the incomplete sample of a larger, unknown geographical distribution, and the environmental data are used as constraints [104,105]. A recent comparison of methods for niche-based modeling of species potential ranges under current conditions identified Maxent as among the best approaches available in terms of predictive performance [56].

For each species, a model was constructed for the current potential range using known collection localities (see below) and current climate conditions; the model was then projected onto a reconstruction of climate layers for the LGM to obtain a potential geographic range of each species at the LGM. Localities used as known presence records for each species of leafcutter ant (Table S7) came primarily from observations by the authors. Additional localities were obtained from A. Himler, N. Gerardo, C. Currie, A. Little, A. Mikheyev, and S. Villamarin. Geographic coordinates for each locality were obtained using a handheld GPS unit (Garmin eTrex). Museum specimens, although abundant for many species of *Atta*, were generally not used in these analyses because they often do not contain detailed geographic coordinates indicating where the collection was made.

For current environmental conditions, twenty bioclimatic layers for the entire New World were obtained from the WorldClim dataset (<http://www.worldclim.org>; version 1.4), each with a resolution of approximately 10 km. The methods used to generate these layers are described in Hijmans et al. [108]. The "auto features" option was selected in Maxent for all analyses. In addition, the following settings were used for the full training runs for each species: 500 maximum iterations, a convergence threshold of 1.0E-5, "minimize memory use," and a regularization multiplier equal to 1.0 [104].

Two approaches were used to determine whether the predictions for current conditions generated by Maxent were better than random predictions. First, the area under the receiver Operating Characteristic curve (AUC), a commonly used measurement for comparison of model performance [56], was calculated for each species. The AUC varies from 0 to 1, with greater scores indicating better discrimination ability; an AUC greater than 0.5 indicates that the model discriminates better than random [56].

Second, a separate analysis was conducted by randomly splitting the localities into two sets: training and testing. The training set (75% of localities for *A. cephalotes*, 90% for *A. laevigata* and *A. sexdens*) was used to build the model while the testing set was used to test the predictive ability of that model. The number of localities used for testing versus training was dependent on how many sites were available for each species. To test the predictive ability of the model, Maxent's cumulative prediction was converted to a binary (i.e. presence vs. absence) prediction. Ten different thresholds automatically generated by Maxent were used for this conversion and the extrinsic omission rate (the fraction of test localities that are outside the area in which the species is predicted to occur) was tested against the null hypothesis that it is no better than a random prediction (of equal area) using a one-tailed binomial test [104]. The same settings were used as for the full training runs, except that all of the available samples were used to build the model.

Estimates of the potential geographic range of each species during the last glacial maximum (LGM, approximately 21 kya)

were made by projecting the model generated under current environmental conditions onto a reconstruction of the same environmental variables at the LGM (see [57] for an explanation of how these layers were generated). A binary (presence vs. absence) prediction for the LGM was necessary for hypothesis tests (see below). To obtain a binary prediction, threshold values were chosen that minimized the commission (false positive) rate for current conditions, based on absence data obtained from recent surveys (S. Solomon, unpublished). The cumulative probability thresholds chosen for *A. cephalotes*, *A. sexdens*, and *A. laevigata* were 1, 5, and 5, respectively. The results of the paleodistribution models were used in subsequent analyses to provide a priori population groupings for all tests of the refugia hypothesis in the following way: areas that were predicted to provide contiguous blocks of suitable habitat during the LGM (using the binary prediction) were grouped together as a single population (Figure 1: C, F, I); areas that were predicted not to be suitable were ignored for the purposes of hypothesis testing (see below).

### Gene tree topology tests

Each hypothesis makes a specific prediction about the genealogical relationships between populations across the geographic range of each species (see Table 1). Specifically, given enough time, isolated populations that have diverged evolutionarily are expected to become reciprocally monophyletic [14,87]. The relationships predicted by a strict interpretation of each hypothesis, assuming complete lineage sorting, were converted into backbone constraint topologies as follows. For the riverine barrier hypothesis, populations occurring on either bank (i.e. north and south) of the Amazon River should be reciprocally monophyletic. For the marine incursion hypothesis, populations near the eastern base of the Andes, on the Brazilian Shield, and on the Guyana Shield should be reciprocally monophyletic. For the refugia hypothesis, populations that were predicted by the paleodistribution models to persist during the last glacial maximum (Figure 1, middle rows) should be reciprocally monophyletic.

To determine whether these predictions were met, mitochondrial DNA gene trees were estimated, using unique haplotypes, with maximum likelihood and Bayesian inference techniques. Maximum likelihood searches were performed with a beta version of GARLI [109] that allows backbone constraints (version 0.952 Beta), with default settings and parameters estimated according to the model of evolution selected using the Akaike Information Criterion (AIC) as implemented in ModelTest [110]. The best tree consistent with the constraint topology for each hypothesis was then found using identical settings. In order to assess whether the null hypothesis represented by the constraint trees could be rejected, the difference between the log-likelihood values of the best constrained and best unconstrained trees was used as a test statistic, with statistical significance assessed through simulation (parametric bootstrap or SOWH test [111,112]). One hundred simulated datasets were generated using Seq-gen [113], with parameters estimated by PAUP\* [114] from the best constrained tree under each constraint. Constrained and unconstrained searches were performed in GARLI on the simulated data using identical settings as for the empirical data. The distribution of differences between constrained and unconstrained searches on the simulated data was used to assess the significance of the test statistic; the  $p$  value was equal to the number of simulated datasets (out of 100 replicates) with a difference in log-likelihood scores between constrained and unconstrained searches greater than the empirical difference. The null hypothesis (i.e. constraint topology) was rejected when  $p$  values were less than 0.05.

Bayesian searches were conducted in MrBayes version 3.1.2 [115]. Four separate runs were conducted, each with four incrementally heated chains and uninformative, default priors; convergence and optimal burn-in were assessed as described in [116] using the program MrConverge (A. Lemmon, in prep.). After discarding burn-in, the posterior samples of tree topologies for each run were combined in PAUP\*; the combined posterior sample was then filtered with the constraint tree for each hypothesis. The proportion of trees retained by the filter was the Bayesian posterior probability of that hypothesis.

### Population Genetic Structure

To determine whether populations are structured as predicted by each of the hypotheses in question (Table 1), two types of population-genetic analyses were performed, using all ingroup haplotypes for each. Analysis of molecular variance (AMOVA) was used, as implemented in Arlequin 3.11 [117], to calculate the percentage of variance explained by *a priori* population groupings in a hierarchical framework [118]. The population structure was defined for each species/hypothesis, as for the constraint trees in phylogenetic analyses. Tamura and Nei distances with an alpha shape parameter were used to compute the pairwise distance matrix for all AMOVA calculations, as this is the most complex model of sequence evolution currently available in Arlequin [117]. Transitions and transversions were given equal weight, while deletions (i.e. gaps) were ignored. Statistical significance of variance components was assessed using the permutation procedures described in the Arlequin user's manual (<http://cmpg.unibe.ch/software/arlequin3/arlequin31.pdf>).

To further test for the presence of barriers to gene flow, as predicted for each hypothesis, simple and partial Mantel tests [119,120] were conducted on the following matrices. First, the pairwise maximum likelihood genetic distance between individuals (as defined *a priori* for each species/hypothesis) was calculated in PAUP, using the model of sequence evolution selected by the AIC in ModelTest [110]. Second, the pairwise geographic distance (in kilometers) was calculated using the program Range (<http://earthquake.usgs.gov/research/software/#Range>). Third, the presence or absence of a potential barrier between two individuals was coded as a binary character and converted to a pairwise barrier matrix. If the straight-line distance between two individuals crossed the barrier of interest (e.g. the Amazon River in the case of the riverine barrier hypothesis), then the barrier was coded as present; if not, the barrier was coded as absent.

For each hypothesis/species, simple Mantel tests assessed the correlation between pairwise genetic distance matrices and the pairwise barrier matrix. Furthermore, isolation by distance was tested for by a simple Mantel test of the pairwise genetic distance and pairwise geographic distance. If both of the above tests were statistically significant, a partial Mantel test was conducted to determine whether the genetic distance between individuals was correlated with the presence of a potential barrier when the effects of geographic distance are removed. All Mantel tests were conducted with the program *zt* [121] and used 10,000 permutations to assess statistical significance.

### Demographic Analyses

Two types of analyses were performed using Arlequin 3.11 [117] to test the predictions of both the refugia and marine incursion hypotheses that populations restricted to an isolated region should show signs of population bottlenecks and subsequent population expansion (Table 1). Tajima's  $D$  statistic [122] which is expected to be negative for populations that have experienced recent population growth [123], was calculated for each

population grouping (Figure 1) for each hypothesis. Significance was tested, as described in the Arlequin manual (<http://cmpg.unibe.ch/software/arlequin3/arlequin31.pdf>) by simulating random samples under a model of population equilibrium, where the  $p$  value is equal to the number of simulated values less than or equal to the observed value of  $D$ .

Second, pairwise nucleotide mismatch distributions were calculated for each population. A population that is at equilibrium is expected to have a multi-model mismatch distribution due to the stochastic shape of its gene tree, whereas populations that have experienced recent growth should have a unimodal mismatch distribution resulting from a star-like gene tree [124,125]. A model of stepwise population expansion was estimated using a generalized least-square approach [126], and its validity was tested as follows: The sum of squared deviations (SSD) between the observed and the simulated (i.e. expected) mismatch distributions was used as a test statistic; 1000 bootstrap simulations of the data were performed, and the SSD was calculated for each; the null hypothesis of population expansion was rejected when fewer than 5% of the simulated SSD values were greater than the observed SSD. To further test whether the observed mismatch distributions deviated from the null expectations characteristic of an expanding population, Harpending's Raggedness Index [127] was calculated. This index has greater values for distributions that are multimodal, as expected for stationary (i.e. non-expanding) populations. Significance for Harpending's Raggedness Index was assessed through bootstrap simulation as described for the SSD.

### Coalescent dating of population divergence

The refugia and marine incursion hypotheses make similar predictions about how populations should be structured (see Table 1). However, these two hypotheses make predictions on vastly different temporal scales. On the one hand, the Pleistocene refugia model predicts that current population structure formed during or subsequent to the Pleistocene, 10,000 to 1.8 million years ago. In contrast, the population structure predicted by the marine incursion hypothesis should date to the Miocene, approximately 10–15 million years ago.

To discriminate between these alternative scenarios, a coalescent dating approach was used. The results of the phylogenetic analyses for each species were used to determine where the most basal split occurred between all sampled populations. The approximate date of this split, in years before present (ybp), was estimated using the isolation-with-migration model developed by Nielsen and Wakeley [128] as implemented in the program IM [129]. This program simultaneously approximates the divergence time ( $t$ ) between two populations that share a common ancestor, the migration rates ( $m_1$  and  $m_2$ ) between these populations, the proportion of the ancestral population that founded each of the resulting populations ( $s$  and  $1-s$ ) and a measure of genetic diversity for the ancestral ( $\theta_{A_0}$ ) as well as both resulting populations ( $\theta_{A_1}$ ,  $\theta_{A_2}$ ) in a Bayesian framework using a Markov chain Monte Carlo method. The program assumes that the diverging populations are not exchanging migrants with any other populations [128].

Preliminary analyses were conducted on each population pair to assess mixing of the chains, as well as to determine appropriate priors for the parameters that were not of interest (i.e. all but  $t$ ; see Table S6 for a list of the priors used for each species). The upper limit for the prior distribution of  $t$ ,  $t_{\max}$ , was determined based on recent estimates for the origin of the genus *Atta* [81]; the oldest possible date recovered by that study for the origin of the crown group of leafcutter ants, 14 mya (Schultz and Brady 2008 Table S3), was used as  $t_{\max}$  for all three species in our study. All searches

used the HKY model of sequence evolution (currently the most appropriate model available in IM for mtDNA evolution), a generation time of 4 years (based on life history data from Autuori [130] and observations by the authors) and uninformative priors. After the first 100,000 steps, which were discarded as burnin, searches proceeded until the following criteria were satisfied: (1) the minimum ESS was at least 100, (2) no trends were observable in plots of parameter values throughout the course of the run, and (3) the results from at least 3 independent runs using the same data and prior values converged on similar posterior distributions.

The estimates for  $t$  were converted into time in years since divergence ( $T_{\text{div}}$ ) using the equation,  $T_{\text{div}} = t * u$ , where  $u$  is the mutation rate in substitutions per site per year. The mutation rate for COI was estimated based on unpublished sequence data for the same gene from species spanning the tribe Attini and from divergence times within the Attini, as reconstructed by Schultz and Brady [81]; the resulting value of 9.5 substitutions per site per million years is consistent with an estimate of the average mutation rate for COI in a recent survey across the arthropods [131].

### Supporting Information

**Table S1** Mitochondrial DNA primers used for amplification and sequencing of ants in the present study.

Found at: doi:10.1371/journal.pone.0002738.s001 (0.05 MB DOC)

**Table S2** Results of gene tree topology tests. For the parametric bootstrap analyses,  $p$  values less than 0.05 indicate rejection of the null hypothesis (i.e. the constraint tree). Bpp is the Bayesian posterior probability of a given constraint topology (\*The predictions of the Pleistocene refugia and marine incursion hypotheses are identical for *A. laevigata*).

Found at: doi:10.1371/journal.pone.0002738.s002 (0.07 MB DOC)

**Table S3** Results of Analyses of Molecular Variance (AMOVA). For each hypothesis, population structure was defined as predicted by each hypothesis (see text). The percentage of variance explained by each hierarchical grouping is shown, with an asterisk indicating statistical significance as assessed by permutation. The "among regions" grouping is the grouping of interest for the purposes of hypothesis testing in this study (Negative percentages and percentages greater than 100 should be interpreted as not significantly different than zero and 100, respectively).

Found at: doi:10.1371/journal.pone.0002738.s003 (0.07 MB DOC)

**Table S4** Results of simple and partial Mantel tests of matrix correlation. For each hypothesis, the correlation between corrected, pairwise genetic distance between individuals and the presence or absence of the barrier of interest was tested using a simple Mantel test (Gen Dist×Barrier). The correlation between genetic and geographic distances (Gen Dist×Geog Dist) was assessed to test for isolation by distance. If a significant correlation was found between both matrix comparisons, a partial Mantel test was conducted on all three matrices to determine whether the presence of the barrier of interest was significantly correlated with genetic distance once the effects of geographic distance are factored out (Partial). All tests used 10,000 permutations to assess statistical significance.

Found at: doi:10.1371/journal.pone.0002738.s004 (0.06 MB DOC)

**Table S5** Results of demographic analyses. Pairwise nucleotide mismatch distributions and Tajima's (1989)  $D$  tests were used to

test for historical population expansion for populations defined a priori for each hypothesis.

Found at: doi:10.1371/journal.pone.0002738.s005 (0.08 MB DOC)

**Table S6** Summary of coalescent dating analyses using the program IM. Left panel: priors used for estimating Tdiv, the time since earliest population divergence for each species. Right panel: the posterior estimate for Tdiv, as well as the lower (95Lo) and upper (95Hi) 95% confidence limits for each species.

Found at: doi:10.1371/journal.pone.0002738.s006 (0.04 MB DOC)

**Table S7** List of all samples used, their geographic locations, and GenBank Accession numbers for samples used in molecular analyses. (BR = Brazil; BZ = Belize; CO = Colombia; CR = Costa Rica; EC = Ecuador; FG = French Guiana; GT = Guatemala; GU = Guyana; MX = Mexico; PA = Panama; PU = Peru; TR = Trinidad; US = United States; VZ = Venezuela)

Found at: doi:10.1371/journal.pone.0002738.s007 (0.44 MB DOC)

## References S1

Found at: doi:10.1371/journal.pone.0002738.s008 (0.02 MB DOC)

## Acknowledgments

The authors wish to thank the following people and institutions for providing access to material without which this study could not have been

completed: J. Lattke (Museo del Instituto de Zoología Agrícola 'Francisco Fernández Yépez', Maracay, Venezuela), K. McGuire (University of Michigan), A. Ortiz Reyes (Universidad Nacional de Colombia, Medellín, Colombia), S. Sanchez-Peña (Universidad Autónoma Agraria Antonio Narro, Saltillo, Coahuila, Mexico), José Santisteban (Universidad Nacional Agraria La Molina, Lima, Peru); Autoridad Nacional del Ambiente, Panama (Permit numbers SE/AH 037-02 and SE/A-29-02); Ministerio del Ambiente y Energía, Costa Rica (Permit No. 15365). Fieldwork was conducted with assistance from the Organization for Tropical Studies (Costa Rica), the Smithsonian Tropical Research Institute (Panama), the Amazon Conservation Association (Peru), and the following individuals: R.M.M. Adams, A.G.D. Bieber, C. Currie, S.A. da Silva, O.P.G. Darrault, R. Horth, I.R. Leal, A. Mikheyev, C. Rabeling, E. Rodriguez, J. Scott, A. Silva, and others. M. Cooper, S. Haferkamp, H. Luong, A. Mikheyev, S. Narasimhan, J. Scott, and D. Seval provided assistance with molecular analyses. J. Brown, S. Hedtke, L. Pomara, S. Ron, and D. Zwickl, and the computational phylogenetics discussion group at UT-Austin, provided invaluable support with data analysis. Helpful comments on the manuscript were provided by J. Brown, L. Gilbert, S. Hedtke, M. Kirkpatrick, R. Linder, A. Mikheyev, C. Rabeling, S. Ron, K. Young and two anonymous reviewers.

## Author Contributions

Conceived and designed the experiments: SES. Performed the experiments: SES JMJ GGV. Analyzed the data: SES. Contributed reagents/materials/analysis tools: MJB UGM. Wrote the paper: SES MJB UGM.

## References

- Hillebrand H (2004) On the generality of the latitudinal diversity gradient. *American Naturalist* 163: 192–211.
- Weir JT, Schluter D (2007) The latitudinal gradient in recent speciation and extinction rates of birds and mammals. *Science* 315: 1574–1576.
- Wright S, Keeling J, Gillman L (2006) The road from Santa Rosalia: A faster tempo of evolution in tropical climates. *Proceedings of the National Academy of Sciences of the United States of America* 103: 7718–7722.
- Rohde K (1992) Latitudinal gradients in species-diversity - the search for the primary cause. *Oikos* 65: 514–527.
- Wilson EO (1987) The arboreal ant fauna of Peruvian Amazon forests - a 1st assessment. *Biotropica* 19: 245–251.
- Lewinsohn TM, Freitas AVL, Prado PI (2005) Conservation of terrestrial invertebrates and their habitats in Brazil. *Conservation Biology* 19: 640–645.
- Wilson EO (1999) *The Diversity of Life*. New York: W. W. Norton and Company.
- Gentry AH (1988) Tree species richness of upper Amazonian forests. *Proceedings of the National Academy of Sciences of the United States of America* 85: 156–159.
- Coyne JA, Orr HA (2004) *Speciation*. Sunderland, MA: Sinauer Associates, Inc.
- Lougheed SC, Gascon C, Jones DA, Bogart JP, Boag PT (1999) Ridges and rivers: a test of competing hypotheses of Amazonian diversification using a dart-poison frog (*Epidobates femoralis*). *Proceedings of the Royal Society of London Series B-Biological Sciences* 266: 1829–1835.
- Elmer KR, Davila JA, Lougheed SC (2007) Cryptic diversity and deep divergence in an upper Amazonian leaf litter frog, *Eleutherodactylus ockendeni*. *BMC Evolutionary Biology* 7.
- Bush MB (1994) Amazonian speciation - a necessarily complex model. *Journal of Biogeography* 21: 5–17.
- Haffer J (1997) Alternative models of vertebrate speciation in Amazonia: An overview. *Biodiversity and Conservation* 6: 451–476.
- Moritz C, Patton JL, Schneider CJ, Smith TB (2000) Diversification of rainforest faunas: An integrated molecular approach. *Annual Review of Ecology and Systematics* 31: 533–563.
- Colinvaux PA, Irion G, Rasanen ME, Bush MB, de Mello J (2001) A paradigm to be discarded: Geological and paleoecological data falsify the HAFFER & PRANCE refuge hypothesis of Amazonian speciation. *Amazoniana-Limnologia Et Oecologia Regionalis Systemae Fluminis Amazonas* 16: 609–646.
- Haffer J (1969) Speciation in Amazonian forest birds. *Science* 165: 131–137.
- Haffer J, Prance GT (2001) Climatic forcing of evolution in Amazonia during the Cenozoic: On the refuge theory of biotic differentiation. *Amazoniana-Limnologia Et Oecologia Regionalis Systemae Fluminis Amazonas* 16: 579–605.
- Brown KS, Sheppard FRS, Turner JRG (1974) Quaternary refugia in tropical America: evidence from race formation in *Heliconius* butterflies. *Proceedings of the Royal Society of London, Series B* 187: 369–378.
- Brown KS (1982) Paleogeology and regional patterns of evolution in Neotropical forest butterflies. In: Prance GT, ed (1982) *Biological diversification in the tropics*. New York: Columbia University Press. pp 255–308.
- Mayr E, Ohara RJ (1986) The biogeographic evidence supporting the Pleistocene forest refuge hypothesis. *Evolution* 40: 55–67.
- Fjelds J (1994) Geographical patterns for relict and young species of birds in Africa and South-America and implications for conservation priorities. *Biodiversity and Conservation* 3: 207–226.
- Brower AVZ (1996) Parallel race formation and the evolution of mimicry in *Heliconius* butterflies: A phylogenetic hypothesis from mitochondrial DNA sequences. *Evolution* 50: 195–221.
- Aleixo A (2004) Historical diversification of a Terra-firme forest bird superspecies: A phylogeographic perspective on the role of different hypotheses of Amazonian diversification. *Evolution* 58: 1303–1317.
- Aleixo A (2006) Historical diversification of floodplain forest specialist species in the Amazon: a case study with two species of the avian genus *Xiphorhynchus* (Aves: Dendrocolaptidae). *Biological Journal of the Linnean Society* 89: 383–395.
- Cheviron ZA, Hackett SJ, Capparella AP (2005) Complex evolutionary history of a Neotropical lowland forest bird (*Lepidothrix coronata*) and its implications for historical hypotheses of the origin of Neotropical avian diversity. *Molecular Phylogenetics and Evolution* 36: 338–357.
- Dick CW, Roubik DW, Gruber KF, Bermingham E (2004) Long-distance gene flow and cross-Andean dispersal of lowland rainforest bees (Apidae: Euglossini) revealed by comparative mitochondrial DNA phylogeography. *Molecular Ecology* 13: 3775–3785.
- Smith TB, Schneider CJ, Holder K (2001) Refugial isolation versus ecological gradients. *Genetica* 112: 383–398.
- Colinvaux PA, DeOliveira PE, Moreno JE, Miller MC, Bush MB (1996) A long pollen record from lowland Amazonia: Forest and cooling in glacial times. *Science* 274: 85–88.
- Colinvaux PA, De Oliveira PE, Bush MB (2000) Amazonian and Neotropical plant communities on glacial time-scales: The failure of the aridity and refuge hypotheses. *Quaternary Science Reviews* 19: 141–169.
- Colinvaux PA, De Oliveira PE (2001) Amazon plant diversity and climate through the Cenozoic. *Palaeogeography Palaeoclimatology Palaeoecology* 166: 51–63.
- Nelson BW, Ferreira CAC, Dasilva MF, Kawasaki ML (1990) Endemism centers, refugia and botanical collection density in Brazilian Amazonia. *Nature* 345: 714–716.
- Wilf P, Cuneo NR, Johnson KR, Hicks JF, Wing SL, et al. (2003) High plant diversity in Eocene South America: Evidence from Patagonia. *Science* 300: 122–125.
- Hovikoski J, Rasanen M, Gingras M, Roddaz M, Brusset S, et al. (2005) Miocene semidiurnal tidal rhythmites in Madre de Dios, Peru. *Geology* 33: 177–180.



34. Hovikoski J, Rasanen M, Gingras M, Lopez S, Romero L, et al. (2007) Palaeogeographical implications of the Miocene Quendeque Formation (Bolivia) and tidally-influenced strata in southwestern Amazonia. *Palaeogeography Palaeoclimatology Palaeoecology* 243: 23–41.
35. Latrubesse EM, da Silva SAF, Cozzuol M, Absy ML (2007) Late Miocene continental sedimentation in southwestern Amazonia and its regional significance: Biotic and geological evidence. *Journal of South American Earth Sciences* 23: 61–80.
36. Rasanen ME, Linna AM, Santos JCR, Negri FR (1995) Late Miocene tidal deposits in the Amazonian foreland basin. *Science* 269: 386–390.
37. Vonhof HB, Wesselingh FP, Kaandorp RJG, Davies GR, van Hinte JE, et al. (2003) Paleogeography of Miocene Western Amazonia: Isotopic composition of molluscan shells constrains the influence of marine incursions. *Geological Society of America Bulletin* 115: 983–993.
38. Webb SD (1995) Biological implications of the middle Miocene Amazon seaway. *Science* 269: 361–362.
39. Lovejoy NR, Albert JS, Crampton WGR (2006) Miocene marine incursions and marine/freshwater transpositions: Evidence from Neotropical fishes. *Journal of South American Earth Sciences* 21: 5–13.
40. Wallace AR (1852) On the monkeys of the Amazon. *Proceedings of the Zoological Society of London* 20: 107–110.
41. Gascon C, Malcolm JR, Patton JL, da Silva MNF, Bogart JP, et al. (2000) Riverine barriers and the geographic distribution of Amazonian species. *Proceedings of the National Academy of Sciences of the United States of America* 97: 13672–13677.
42. Matocq MD, Patton JL, da Silva MNF (2000) Population genetic structure of two ecologically distinct Amazonian spiny rats: Separating history and current ecology. *Evolution* 54: 1423–1432.
43. Patton JL, Dasilva MNF, Malcolm JR (1994) Gene genealogy and differentiation among arboreal spiny rats (Rodentia, Echimyidae) of the Amazon basin - a test of the riverine barrier hypothesis. *Evolution* 48: 1314–1323.
44. Patton JL, Da Silva MNF, Malcolm JR (2000) Mammals of the Rio Jurua and the evolutionary and ecological diversification of Amazonia. *Bulletin of the American Museum of Natural History*. pp 1–306.
45. Peres CA, Patton JL, da Silva MNF (1996) Riverine barriers and gene flow in Amazonian saddle-back tamarins. *Folia Primatologica* 67: 113–124.
46. Hayes FE, Sewlal JAN (2004) The Amazon River as a dispersal barrier to passerine birds: effects of river width, habitat and taxonomy. *Journal of Biogeography* 31: 1809–1818.
47. Hershkovitz P (1977) *Living New World monkeys (Platyrrhini) with an introduction to primates*. Chicago, IL: University of Chicago Press.
48. Avila-Pires TCS (1995) Lizards of Brazilian Amazonia (Reptilia: Squamata). *Zoologische Verhandlungen* 299: 1–706.
49. Pellegrino KCM, Rodrigues MI, Waite AN, Morando M, Yassuda YY, et al. (2005) Phylogeography and species limits in the *Gymnodactylus darwini* complex (Gekkonidae, Squamata): genetic structure coincides with river systems in the Brazilian Atlantic Forest. *Biological Journal of the Linnean Society* 85: 13–26.
50. Funk WC, Caldwell JP, Peden CE, Padial JM, De la Riva I, et al. (2007) Tests of biogeographic hypotheses for diversification in the Amazonian forest frog, *Physalaemus petersi*. *Molecular Phylogenetics and Evolution* 44: 825–837.
51. Hall JPW, Harvey DJ (2002) The phylogeography of Amazonia revisited: New evidence from riodinid butterflies. *Evolution* 56: 1489–1497.
52. Losos JB, Glor RE (2003) Phylogenetic comparative methods and the geography of speciation. *Trends in Ecology & Evolution* 18: 220–227.
53. Avise JC, Arnold J, Ball RM, Bermingham E, Lamb T, et al. (1987) Intraspecific phylogeography - the mitochondrial-DNA bridge between population-genetics and systematics. *Annual Review of Ecology and Systematics* 18: 489–522.
54. Knowles LL (2004) The burgeoning field of statistical phylogeography. *Journal of Evolutionary Biology* 17: 1–10.
55. Hugall A, Moritz C, Moussalli A, Stanisic J (2002) Reconciling paleodistribution models and comparative phylogeography in the Wet Tropics rainforest land snail *Gnatosiphia bellendenkerensis* (Brazier 1875). *Proceedings of the National Academy of Sciences of the United States of America* 99: 6112–6117.
56. Elith J, Graham CH, Anderson RP, Dudik M, Ferrier S, et al. (2006) Novel methods improve prediction of species' distributions from occurrence data. *Ecography* 29: 129–151.
57. Ruegg KC, Hijmans RJ, Moritz C (2006) Climate change and the origin of migratory pathways in the Swainson's thrush, *Catharus ustulatus*. *Journal of Biogeography* 33: 1172–1182.
58. Bonaccorso E, Koch I, Peterson AT (2006) Pleistocene fragmentation of Amazon species' ranges. *Diversity and Distributions* 12: 157–164.
59. Richards CL, Carstens BC, Lacey Knowles L (2007) Distribution modeling and statistical phylogeography: an integrative framework for generating and testing alternative biogeographical hypotheses. *Journal of Biogeography* 34: 1833–1845.
60. Carstens B, Richards C (2007) Integrating coalescent and ecological niche modeling in comparative phylogeography. *Evolution* 61: 1439–1454.
61. Carnaval AC, Moritz C Historical climate modeling predicts patterns of current biodiversity in the Brazilian Atlantic forest. *Journal of Biogeography Online* Early: 28-March 2008.
62. Bermingham E, Dick CW, Moritz C, eds (2005) *Tropical rainforests: past, present, and future*. Chicago, IL: The University of Chicago Press.
63. Kosciński A, Handford P, Tubaro PL, Sharp S, Loughheed SC ( ) Pleistocene climatic cycling and diversification of the Andean treefrog, *Hypsiboas andinus*. *Molecular Ecology* 17: 2012–2025.
64. Erwin TL (1991) How many species are there - revisited. *Conservation Biology* 5: 330–333.
65. Weber NA (1972) *Gardening Ants, The Attines*. Philadelphia, PA: The American Philosophical Society.
66. Hölldobler B, Wilson EO (1990) *The ants*. Cambridge, MA: Harvard University Press.
67. Mueller UG, Schultz TR, Currie CR, Adams RMM, Malloch D (2001) The origin of the attine ant-fungus mutualism. *Quarterly Review of Biology* 76: 169–197.
68. Mueller UG, Gerardo NM, Aanen DK, Six DL, Schultz TR, et al. (2005) The evolution of agriculture in insects. *Annual Review of Ecology Evolution and Systematics* 36: 563–595.
69. Cherrett JM, Peregrine DJ (1976) A review of the status of leaf-cutting ants and their control. *Proceedings of the Association of Applied Biologists* 84: 124–128.
70. Wirth R, Herz H, Ryel RJ, Beyschlag W, Hölldobler B (2003) *Herbivory of Leaf-Cutting Ants: A Case Study on Atta colombica in the Tropical Rainforest of Panama*. Berlin, Heidelberg, New York: Springer Verlag.
71. Moutinho P, Nepstad DC, Davidson EA (2003) Influence of leaf-cutting ant nests on secondary forest growth and soil properties in Amazonia. *Ecology* 84: 1265–1276.
72. Garretson M, Stetzel JF, Halpern BS, Hearn DJ, Lucey BT, et al. (1998) Diversity and abundance of understory plants on active and abandoned nests of leaf-cutting ants (*Atta cephalotes*) in a Costa Rican rain forest. *Journal of Tropical Ecology* 14: 17–26.
73. Gonçalves CR (1967) As formigas cortadeiras da Amazônia, dos generos "Atta" Fabr. e "Acromyrmex" Mayr (Hym., Formicidae). *Atas do Simpósio sobre a Biota Amazônica 5 (Zoologia)*. pp 181–202.
74. Schultz TR, Brady SG (2008 (in press)) Major evolutionary transitions in ant agriculture. *Proceedings of the National Academy of Sciences of the USA*.
75. Gonçalves CR (1960) *Distribuição, biologia e ecologia das saúvas*. *Divulgação Agronômica* 12: 2–10.
76. Weber NA (1959) Ecological relations of three *Atta* species in Panama. *Ecology* 50: 141–147.
77. Moreira AA, Forti LC, Andrade APP, Boaretto MAC, Lopes JFS (2004) Nest architecture of *Atta laevigata* (F. Smith, 1858) (Hymenoptera : Formicidae). *Studies on Neotropical Fauna and Environment* 39: 109–116.
78. Cherrett JM (1986) The biology pest status and control of leaf-cutting ants. *Russell, G E*. pp 1–38.
79. Cherrett JM (1986) The economic importance and control of leaf-cutting ants. In: *Vinson SB, ed (1986) Economic impact and control of social insects*. New York: Praeger. pp 165–192.
80. Cherrett JM (1986) History of leaf cutting ant problem. *Lofgren, C S and R K Vander Meer*. pp 10–17.
81. Schultz TR, Brady SG (2008) Major evolutionary transitions in ant agriculture. *Proceedings of the National Academy of Sciences of the USA Early Edition (March 24)*.
82. Moser JC (1967) Mating activities of *Atta texana* (Hymenoptera, Formicidae). *Insectes Sociaux* 14: 295–312.
83. Goulding M, Barthem R, Ferreira E (2003) *The Smithsonian Atlas of the Amazon*. Washington, DC: Smithsonian Books.
84. Lovejoy NR, Bermingham E, Martin AP (1998) Marine incursion into South America. *Nature* 396: 421–422.
85. Nores M (1999) An alternative hypothesis for the origin of Amazonian bird diversity. *Journal of Biogeography* 26: 475–485.
86. Simpson BB, Haffer J (1978) Speciation patterns in Amazonian forest biota. *Annual Review of Ecology and Systematics* 9: 497–518.
87. Avise JC (2000) *Phylogeography: the history and formation of species*. Cambridge, MA: Harvard University Press.
88. Bush MB, Silman MR (2004) Observations on Late Pleistocene cooling and precipitation in the lowland Neotropics. *Journal of Quaternary Science* 19: 677–684.
89. Mayle FE, Beerling DJ, Gosling WD, Bush MB (2004) Responses of Amazonian ecosystems to climatic and atmospheric carbon dioxide changes since the last glacial maximum. *Philosophical Transactions of the Royal Society of London Series B-Biological Sciences* 359: 499–514.
90. Hardman M, Hardman LM (2008) The relative importance of body size and paleoclimatic change as explanatory variables influencing lineage diversification rate: An evolutionary analysis of bullhead catfishes (Siluriformes : Ictaluridae). *Systematic Biology* 57: 116–130.
91. Gamble T, Simons AM, Colli GR, Vitt IJ (2008) Tertiary climate change and the diversification of the Amazonian gecko genus *Gonatodes* (Sphaerodactylidae, Squamata). *Molecular Phylogenetics and Evolution* 46: 269–277.
92. Jansson R, Davies TJ (2008) Global variation in diversification rates of flowering plants: energy vs. climate change. *Ecology Letters* 11: 173–183.
93. Mayhew PJ, Jenkins GB, Benton TG (2008) A long-term association between global temperature and biodiversity, origination and extinction in the fossil record. *Proceedings of the Royal Society B-Biological Sciences* 275: 47–53.
94. Carnaval AC, Bates JM (2007) Amphibian DNA shows marked genetic structure and tracks Pleistocene climate change in northeastern Brazil. *Evolution* 61: 2942–2957.

95. Tolley KA, Chase BM, Forest F (2008) Speciation and radiations track climate transitions since the Miocene Climatic Optimum: a case study of southern African chameleons. *Journal of Biogeography Online Early*: 28-March 2008.
96. Parmesan C (1996) Climate and species' range. *Nature* 382: 765–766.
97. Parmesan C, Yohe G (2003) A globally coherent fingerprint of climate change impacts across natural systems. *Nature* 421: 37–42.
98. Parmesan C (2006) Ecological and evolutionary responses to recent climate change. *Annual Review of Ecology Evolution and Systematics* 37: 637–669.
99. Pounds JA, Fogden MPL, Campbell JH (1999) Biological response to climate change on a tropical mountain. *Nature* 398: 611–615.
100. Wilson RJ, Gutierrez D, Gutierrez J, Monserrat VJ (2007) An elevational shift in butterfly species richness and composition accompanying recent climate change. *Global Change Biology* 13: 1873–1887.
101. Martins Jr J, Solomon SE, Mikheyev AS, Mueller UG, Ortiz A, Bacci Jr M (2007) Nuclear mitochondrial-like sequences in ants: evidence from *Atta cephalotes* (Formicidae: Attini). *Insect Molecular Biology* 16: 777–784.
102. Thompson JD, Plewniak F, Poch O (1999) A comprehensive comparison of multiple sequence alignment programs. *Nucleic Acids Research* 27: 2682–2690.
103. Maddison DR, Maddison WP (2000) *MacClade 4: analysis of phylogeny and character evolution*. 4.0 ed. Sunderland, MA: Sinauer Associates, Inc.
104. Phillips SJ, Anderson RP, Schapire RE (2006) Maximum entropy modeling of species geographic distributions. *Ecological Modeling* 190: 231–259.
105. Dudik M, Phillips SJ, Schapire RE (2004) Performance guarantees for regularized maximum entropy density estimation. *Learning Theory, Proceedings*. Berlin: Springer-Verlag Berlin. pp 472–486.
106. Anderson RP (2003) Real vs. artefactual absences in species distributions: tests for *Oryzomys albigularis* (Rodentia: Muridae) in Venezuela. *Journal of Biogeography* 30: 591–605.
107. Vasconcelos HL, Cherratt JM (1995) Changes in leaf-cutting ant populations (Formicidae, Attini) after the clearing of mature forest in Brazilian Amazonia. *Studies on Neotropical Fauna and Environment* 30: 107–113.
108. Hijmans RJ, Cameron SE, Parra JL, Jones PG, Jarvis A (2005) Very high resolution interpolated climate surfaces for global land areas. *International Journal of Climatology* 25: 1965–1978.
109. Zwickl DJ (2006) Genetic algorithm approaches for the phylogenetic analysis of large biological sequence datasets under the maximum likelihood criterion: The University of Texas at Austin.
110. Posada D, Crandall KA (1998) Model selection and model averaging in phylogenetics: advantages of the AIC and Bayesian approaches over likelihood ratio tests. *Bioinformatics* 14: 817–818.
111. Hillis DM, Mable BK, Moritz C (1996) Applications of molecular systematics: the state of the field and a look to the future. In: Hillis DM, Moritz C, Mable BK, eds (1996) *Molecular Systematics*. 2nd ed. Sunderland, MA: Sinauer Associates, Inc. pp 515–543.
112. Goldman N, Anderson JP, Rodrigo AG (2000) Likelihood-based tests of topologies in phylogenetics. *Systematic Biology* 49: 652–670.
113. Rambaut A, Grassly NC (1997) Seq-Gen: An application for the Monte Carlo simulation of DNA sequence evolution along phylogenetic trees. *Computer Applications in the Biosciences* 13: 235–238.
114. Swofford DL (2002) *PAUP\*: Phylogenetic Analysis Using Parsimony (and Other Methods)* 4.0 Beta. 4 ed. Sunderland, MA: Sinauer Associates, Inc.
115. Ronquist F, Huelsenbeck JP (2003) MrBayes 3: Bayesian phylogenetic inference under mixed models. *Bioinformatics* 19: 1572–1574.
116. Brown JM, Lemmon AR (2007) The importance of data partitioning and the utility of Bayes factors in Bayesian phylogenetics. *Systematic Biology* 56: 643–655.
117. Excoffier L, Laval G, Schneider S (2005) Arlequin ver. 3.0: An integrated software package for population genetics data analysis. *Evolutionary Bioinformatics Online* 14: 47–50.
118. Excoffier L, Smouse PE, Quattro JM (1992) Analysis of molecular variance inferred from metric distances among DNA haplotypes - application to human mitochondrial-DNA restriction data. *Genetics* 131: 479–491.
119. Mantel N (1967) The detection of disease clustering and a generalized regression approach. *Cancer Research* 27: 209–220.
120. Smouse PE, Long JC, Sokal RR (1986) Multiple-regression and correlation extensions of the Mantel test of matrix correspondence. *Systematic Zoology* 35: 627–632.
121. Bonnet E, Van de Peer Y (2002) zt: a software tool for simple and partial Mantel tests. *Journal of Statistical software* 7: 1–12.
122. Tajima F (1989) The effect of change in population-size on DNA polymorphism. *Genetics* 123: 597–601.
123. Rogers AR (1995) Genetic-evidence for a Pleistocene population explosion. *Evolution* 49: 608–615.
124. Slatkin M, Hudson RR (1991) Pairwise comparisons of mitochondrial-DNA sequences in stable and exponentially growing populations. *Genetics* 129: 555–562.
125. Rogers AR, Harpending H (1992) Population-growth makes waves in the distribution of pairwise genetic-differences. *Molecular Biology and Evolution* 9: 552–569.
126. Schneider S, Excoffier L (1999) Estimation of past demographic parameters from the distribution of pairwise differences when the mutation rates vary among sites: Application to human mitochondrial DNA. *Genetics* 152: 1079–1089.
127. Harpending HC (1994) Signature of ancient population-growth in a low-resolution mitochondrial-DNA mismatch distribution. *Human Biology* 66: 591–600.
128. Nielsen R, Wakeley J (2001) Distinguishing migration from isolation: A Markov chain Monte Carlo approach. *Genetics* 158: 885–896.
129. Hey J (2005) On the number of New World founders: A population genetic portrait of the peopling of the Americas. *PloS Biology* 3: 965–975.
130. Autuori M (1947) Contribuição para o conhecimento da Saúva (*Atta* spp.) IV. O saúveiro depois da primeira revoada (*Atta sexdens rubropilosa* Forel, 1908). *Arquivos do Instituto de Biologia de São Paulo* 187: 39–70.
131. Quek SP, Davies SJ, Itino T, Pierce NE (2004) Codiversification in an ant-plant mutualism: Stem texture and the evolution of host use in *Crematogaster* (Formicidae: Myrmicinae) inhabitants of *Macaranga* (Euphorbiaceae). *Evolution* 58: 554–570.
132. Simon C, Frati F, Beckenbach A, Crespi B, Liu H, et al. (1994) Evolution, weighting, and phylogenetic utility of mitochondrial gene-sequences and a compilation of conserved polymerase chain-reaction primers. *Annals of the Entomological Society of America* 87: 651–701.
133. Wetterer JK, Schultz TR, Meier R (1998) Phylogeny of fungus-growing ants (Tribe Attini) based on mtDNA sequence and morphology. *Molecular Phylogenetics and Evolution* 9: 42–47.

Copyright of PLoS ONE is the property of Public Library of Science and its content may not be copied or emailed to multiple sites or posted to a listserv without the copyright holder's express written permission. However, users may print, download, or email articles for individual use.