Comparison of Crystallinity between Natural Hydroxyapatite and Synthetic cp-Ti /HA Coatings

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1. Introduction

The diversity of applications of biomaterials, the different chemical composition and the need for appropriate morphological forms for its use, requires interdisciplinary research work that considers aspects such as: surface treatment, synthesis routes, processing techniques (in various forms), degradability as well as local and systemic biocompatibility. In a similar manner, the development of bioceramics and prosthesis also require interdisciplinary research work1.

In this context, the different areas of research and development can contribute significantly to increase the number of applications of bioceramics, by developing novel and more efficient biomaterials, by elucidating mechanisms that govern bone regeneration and by determining physical and chemical characteristics of different metal surfaces on which in-vitro nucleation of calcium phosphate can be stimulated. Creation of a rough surface or a porous film on an implant can reduce the long periods required for interfacial bonding between implant and bone. It can also increase the surface area available for the juxtaposition bone-implant and enhance osseointegration2. This means that the normal remodeled bone is in contact with the implant surface with no interposition of connective tissue, but bone cells3.

Medicine and dentistry have studied calcium phosphate–based ceramics for over thirty years. Interest in a particular phase of calcium phosphates, the hydroxyapatite (HA), \( \text{Ca}_10(\text{PO}_4)_6(\text{OH})_2 \), appeared because of its great similarity with the main component present in the mineral phase of bones. The high bioactivity (strong ceramic/bone tissue interaction) and biocompatibility of HA increased its acceptance as an implant4. Coating of metallic implants, such as commercially pure titanium (cp-Ti) and its alloys with HA is one of its main applications. The basic idea is to combine the advantageous mechanical properties of the metals with the excellent biocompatibility and bioactivity of the HA.

Among the various methods used to obtain HA coatings, the physical techniques5-7, produce high quality coatings and high density films. The main disadvantages of these techniques are the use of sophisticated and expensive equipment and reduced adhesion, because of a lack of chemical bonding, between the metal and the apatite coating. These are the main reasons for studying novel techniques, most of which are chemical methods4,8,9.

The biomimetic4,10,11 method is one of the most promising techniques for the production of biomaterials in body environment conditions. This method consists in immersing the substrate to be coated in a synthetic solution (SBF - Simulated body fluid), with chemical composition and pH similar to that of blood plasma and temperature similar to that of the human body. This technique permits complex forms, porous and temperature sensitive materials, such as polymers, to be coated. Further, the HA obtained using this method is similar to the biological one.

2. Materials and Methods

2.1. Treatment of cp-Ti with NaOH solution

The cp-Ti substrate was cut into small bars 0.5 × 0.5 × 0.3 cm and polished with a 150 mesh sandpaper. The substrates were washed in an ultrasonic cleaner with alcohol, acetone and deionized water for 10 minutes. Immediately afterwards, the substrates were etched in 5M NaOH solution for 24 hours at 60 °C and heat treated at 600 °C12.

2.2. Apatite coating on cp-Ti

After the treatment with NaOH solution, the substrates were immersed in a solution of sodium silicate for 24 hours at 37 °C. Subsequently, the substrate was treated with a SBF solution, for slow and organized nucleation of apatite on the substrate to be coated. Immediately after, the substrates were immersed for six days at 37 °C in a solution 1.5 SBF for the growth of apatite. Table 1 shows the ionic concentrations of the blood plasma and all the solutions used in this process.
2.3. Heat treatment of the apatite coating

After coating the substrates, the samples were heat treated at 400, 500, 600, 700 and 800 °C, without atmospheric control and at heating rate of 5 °C/min.

2.4. Characterization of the apatite coating

After the heat treatment, all the coatings were characterized using scanning electron microscopy (SEM), infrared spectroscopy (FTIR) and X-ray diffraction analysis (XRD).

3. Results and Discussion

Figure 1 shows the scanning electron micrographs of the cp-Ti surface after alkaline treatment. As a thin passive layer of titanium dioxide (TiO₂) forms spontaneously on the cp-Ti substrate, it was treated with 5M NaOH to produce a more favorable and active surface on the passive layer of TiO₂, to induce the precipitation of apatite.

During the alkaline treatment, the passive TiO₂ surface can react in the following manner:

\[
\begin{align*}
\text{TiO}_2 + \text{OH}^- & \rightarrow \text{HTiO}_3^- \\
\text{Ti} + 3\text{OH}^- & \rightarrow \text{Ti(}OH)_3^+ + 4e^-
\end{align*}
\]

The formation of insufficient Ti-OH groups, as a consequence of the amphoteric nature of TiO₂, inhibits the formation of enough nuclei on the cp-Ti to produce a dense and uniform layer of apatite. Therefore, the immersion of these substrates in a solution of a salt which has a tendency to increase the –OH bridges and not displace the equilibrium of the media, will increase the active sites on the cp-Ti surface. In this investigation a solution of sodium silicate was used to increase apatite nucleation, through the formation of Si–OH groups, which is accelerated when immersed in a SBF solution for 24 hours (Table 1). When the substrates were immersed in a SBF solution that is 1.5 times more concentrated than the original one, (Table 1) the apatite nuclei grew spontaneously by consumption of calcium and phosphate ions of the solution, to form a uniform apatite layer.

The composition of the apatite coating on the hydrogel surface was determined by XRD (Figure 3). It can be observed that for heat treatments at 400, 500 and 600 °C, the X-ray diffractograms of the coatings exhibit three wide bands at approximately 2θ = 26°, 20 = 29° and 20 = 32°. These bands are characteristic of the large

\[
\begin{align*}
\text{Ti(OH)}_3^+ + e^- & \rightarrow \text{TiO}_2\text{H}_2\text{O} + \frac{1}{2}\text{H}_2(g) \\
\text{Ti(OH)}_3^+ + \text{OH}^- & \rightarrow \text{Ti(OH)}_4^- \\
\text{TiO}_2\text{H}_2\text{O} + \text{OH}^- & \rightarrow \text{HTiO}_3^- \text{H}_2\text{O}
\end{align*}
\]

The negatively charged species combine with the alkaline ions in the aqueous solution, to form an alkaline layer of titane hydrogel. During the heat treatment, this hydrogel layer is dehydrated and stabilized as a crystalline sodium titane layer, Figure 2.

Figure 2. XDR pattern of sodium titanate on cp-Ti after treatment with NaOH and heat treatment at 600 °C.

Table 1. Ionic concentrations of the solutions used for the apatite coating (mmol.dm⁻³).

<table>
<thead>
<tr>
<th></th>
<th>Na⁺</th>
<th>K⁺</th>
<th>Ca²⁺</th>
<th>Mg²⁺</th>
<th>HCO₃⁻</th>
<th>Cl⁻</th>
<th>HPO₄²⁻</th>
<th>SO₄²⁻</th>
<th>SiO₃²⁻</th>
</tr>
</thead>
<tbody>
<tr>
<td>Blood</td>
<td>142.0</td>
<td>5.0</td>
<td>2.5</td>
<td>1.5</td>
<td>27.0</td>
<td>103.0</td>
<td>1.0</td>
<td>0.5</td>
<td>-</td>
</tr>
<tr>
<td>Plasma</td>
<td>142.0</td>
<td>5.0</td>
<td>2.5</td>
<td>1.5</td>
<td>4.2</td>
<td>148.0</td>
<td>1.0</td>
<td>0.5</td>
<td>-</td>
</tr>
<tr>
<td>SBF</td>
<td>1.5 SBF</td>
<td>213.0</td>
<td>7.5</td>
<td>3.8</td>
<td>6.3</td>
<td>223.0</td>
<td>1.5</td>
<td>0.75</td>
<td>-</td>
</tr>
<tr>
<td>Na₂SiO₄</td>
<td>2.0</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>1.0</td>
<td>-</td>
<td>-</td>
<td>3.6</td>
<td>-</td>
</tr>
</tbody>
</table>
number of peaks associated with apatite phases, indicating the presence of phases with low crystallinity, similar to that of biological apatite. The well defined peaks at 2θ = 27.5° and 2θ = 36° are due to the Ti substrate.

The low crystallinity of the phases in the coating leads to instabilities when implanted. According to literature16, amorphous calcium phosphates and mostly, the tetracalcium phosphate and tricalcium phosphate phases are more soluble than HA, leading to rapid disintegration of the coating and loss of fixation of the implant, known as reabsorption.

Figure 3d-3e shows the increase in crystallinity of the apatite coating with heat treatment at 700 and 800 °C. It is possible to identify different types of apatites that precipitated on the cp-Ti substrate. Besides the crystalline HA phase, some peaks of octacalcium phosphate (OCP), Ca8H2(PO4)6.5H2O and hydrated magnesium phosphate, Mg3(PO4)2.H2O, were observed.

The OCP phase is considered to be the important one as it participates in bone formation and also because it is a precursor of the HA phase. According to Feng14, to obtain calcium phosphate coatings on Ti surfaces, the solution must be above the equilibrium value for OCP saturation. Once the OCP growth velocity exceeds its hydrolysis value, it is possible to form a mixture of OCP and HA phases19.

By varying the heat treatment temperature, it was possible to obtain a HA with crystalline structure similar to that of biological HA, as shown in Figure 4. This reveals the high efficiency of the biomimetic method and the influence of heat treatment on the coatings obtained by this method.

The scanning electron micrographs of the apatite coatings on cp-Ti show a deposit with uniform morphology, composed of spherical particles with size in the range 3-5 μm, and some agglomerates formed by fusion of some particles during the heat treatments. It can be observed that with increase in heat treatment temperature, the morphology of the coatings are similar and free of cracks. After heat treatments at 700 and 800 °C, a new phase, characterized as Mg3(PO4)2.H2O appeared, and this was also detected by XRD. The EDS analysis confirmed the appearance of magnesium as shown in Figure 5.

The crystallinity of the HA coating on cp-Ti surfaces and the phases of the phosphate precipitates during immersion were analyzed by infra-red (IR) spectroscopy.

Figure 6a, 6b shows strong bands attributable to stretching and deformation of the O-H group of water, and can be observed at 3300 and 1660 cm⁻¹, respectively. Three bands related to the CO3²⁻ group at 1457, 1417 and 877 cm⁻¹, characterizing a carbonated hydroxyapatite were also observed. Because of the appearance of these bands it can be concluded that the PO4³⁻ ions are substituted by the carbonate ions in the HA structure according to Ca8,3₁,7(PO4)₄,₃(CO3)₃,₇(HPO4)₀,₇(OH)₀,₃,₇, where ₀ means a vacancy. The band at 1100 cm⁻¹ is attributed to P-O stretching, whereas the bands at 560 and 473 cm⁻¹ are associated to deformation vibration of the P-O in the PO4 group. This indicates that the phosphate ions have an ideal tetrahedral structure.

Figure 6c-6e show the spectra of the HA coating on cp-Ti after heat treatment at 600, 700 and 800 °C. Absence of the band associated to the stretching mode of the O-H group of water is worth noting. The crystalline hydroxyapatite has two bands, at 3570 and 630 cm⁻¹, as observed in the figure. The P-OH bands, characteristic of the HPO4²⁻ group, were observed at 960 and 870 cm⁻¹, confirming the presence of the octacalcium phosphate.

4. Conclusions

A coating with a uniform, homogeneous and porous apatite layer was obtained using a slow nucleation process. The major phase was apatite with small quantities of other phases, and the octacalcium phosphate phase was the HA precursor. Upon heat treatment at temperatures between 400 and 600 °C, the phases exhibited low crystallinity, and this increased at temperatures above 700 °C. To heat treat the coatings, 600 °C was considered the ideal temperature to efficiently attain the HA phase with low crystallinity, similar to that of biological HA.

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Figure 5. SEM micrographs of phosphate coatings on cp-Ti after a) 400; b) 500; c) 600; d) 700; and e) 800 °C. (f) Morphology and EDX analysis of phosphate precipitate.

Figure 6. FTIR transmission spectra of phosphate coating on cp-Ti after heat treatment at: a) 400; b) 500; c) 600; d) 700; and e) 800 °C.

References


