Study On the Use of Oxidant Scrubbers for Elimination of Interferences Due to Nitrogen Dioxide in Analysis of Atmospheric Dimethylsulfide

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Neste trabalho foi avaliado o desempenho de filtros para dióxido de nitrogênio, buscando evitar perdas durante a amostragem de sulfetos orgânicos, provocadas por oxidantes atmosféricos. Diferentes compostos e misturas foram usadas para recobrir superfícies sólidas empregadas na preparação destes filtros. Um sistema automatizado de análise em fluxo foi utilizado para comparar a eficiência de retenção de dióxido de nitrogênio pelos filtros. Entre os materiais testados na preparação dos filtros, as melhores escolhas foram papel ou lã de vidro impregnados com a mistura de sulfato de ferro (II), ácido sulfúrico e ácido pirogalico e ainda os filtros feitos de papel impregnados com trietanolamina. Os resultados obtidos em laboratório com mistura de gás padrão de dimetilsulfeto e experimentos em campo confirmaram a qualidade dos filtros e indicaram que eles podem ser utilizados para evitar a oxidação de sulfetos orgânicos durante a sua amostragem.

In this work, oxidant scrubbers were evaluated for their ability to prevent sampling losses of dimethylsulfide caused by reactions with nitrogen dioxide. Various compounds and mixtures were used in the preparation of the oxidant scrubbers. An automatic flow analysis device was used to compare scrubbing efficiency for nitrogen dioxide. Among the scrubbers tested, the best were shown to be the one made with filter paper or glass wool coated with iron (II) sulfate, sulfuric acid and pyrogallic acid, and the one made from with paper coated with triethanolamine. The results obtained under laboratory conditions, using dimethylsulfide standard gas, and in field experiments confirmed that these scrubbers are suitable for the prevention of oxidation during sampling.

Keywords: dimethylsulfide, atmospheric trace gases, sampling technique, nitrogen dioxide scavenger.

Introduction

Sulfur gases and their reaction products play an important role in the chemistry of the global troposphere and in the biogeochemical sulfur cycle¹⁻⁵. Sulfur compounds are released into the atmosphere by various natural and anthropogenic sources. A substantial increase in anthropogenic sulfur emission has taken place in the last century. It is responsible for malodorous odor in sewage systems, in pulp and paper industry ⁶⁻⁸. In petrochemical processes sulfur compounds can be emitted and cause air pollution during oil prospecting, refining and when fuel is burned ^{7,9,10}. A few decades ago, biogenic H_2S was believed to be the dominant form in which gaseous sulfur was released to the atmosphere. The importance of H_2S emission has been continually revised ever since ^{3-5,11}. Current thought attributes an increasing role to dimethylsulfide (CH₃)₂S (DMS).

Although knowledge of the chemistry of sulfur compounds in the environment has much improved, there is

evidence to suggest that determination of reduced sulfur compounds may often be subject to significant inaccuracies 11 . The main source of uncertainty associated with these data occurs during the collection step. Frequently, the analysis of these compounds requires a cryogenic preconcentration step to enhance low atmospheric concentrations. The cotrapping of water has been a complex problem: the ice may limit sample volumes and water may be injected into columns 12 . However, the main problem is the destruction of sulfur compounds by oxidizing species $^{11-15}$. The species commonly present in the atmosphere, which may cause sampling losses, include ozone (O_3) , nitrogen dioxide (NO_2) and hydrogen peroxide (H_2O_2) .

Several types of oxidant scrubbers have been suggested to remove oxidants during cryogenic preconcentration. Braman and co-workers¹⁶ used glass pearl coated with Na₂CO₃, Barnard co-workers¹⁷ and Andreae and Raemdonck¹⁸ used packed tubes containing Na₂CO₃/ chromasorb as sulfur dioxide scrubbers, Bates and

Johnson¹⁹ used glass fiber filters coated with NaOH. A KOH filter scrubber presented rapid loss of efficiency compared to carbonate-based Anakrom scrubbers²⁰. Hofmann co-workers 12 used a Nafion drier in combination with cotton wadding as an oxidant scavenger. Davison and Allen¹³ investigated in-line scrubbers utilizing Na₂CO₃, FeSO₄.7H₂O, KI or KBr, either pure or coated onto a chromatographic support, which they found to be ineffective for oxidant removal in moderately polluted air. The KI scrubbers were, however, effective when used in clean remote marine air. Comparing the various oxidant scrubbers, Kittler co-workers 14 found the KI/glycerol/Vitrex filter to be superior to the scrubbers using KOH, NaOH, MnO₂ and Na₂CO₃. Other methods have included neutral aqueous potassium iodide²¹, ferrous sulfate²² and 100% cotton wadding in combination with a Nafion drier23. Although much work has been done on the analysis of organic sulfides, the nature of the oxidant interference remains largely unknown. The development and use of new scrubbers is an empirical process¹¹. So far there is no consensus about the use of oxidant scrubbers in the analysis of reduced sulfides.

The present work describes the development of oxidant scrubbers for organic sulfide analysis in air. The oxidant scrubbers were made out of solid supports coated with reactive species, to remove the oxidants before the collection of organic sulfide. Comparative data on the scrubbers was obtained by automated flow analysis. Dimethylsulfide, the commonest organic sulfide found in the air, and nitrogen dioxide, the most common oxidant found in the air, were used in the development of this work.

Experimental

Method of sampling and analysis of organic sulfides

The method used to sample and analyze organic sulfides is non-cryogenic and has already been reported 24,25 . Glass wool coated with sodium tetrachloropalladate (II) was shown to be very efficient for the collection of dimethylsulfide (DMS). The resulting complex ([Pd(DMS)_2Cl_2]) is only slightly soluble in water, but readily soluble in organic solvents. The solutions showed a well-defined absorption peak in the near UV region (e=17.05 x 10^3 l mol^-l cm^-l at 303 nm) and the absorbance values were proportional to the amount of DMS present. Gases such as $\rm H_2S$, CO, CS_2, COS, NH_3 and SO_2 did not cause interference because they neither react with sodium tetrachloropalladate (II) nor produce compounds soluble in the solvents used to extract the [Pd(DMS)_2Cl_2]_{(s)}. Oxidants such as nitrogen dioxide react with complexed DMS, interfering in subsequent DMS estimation.

The sodium tetrachloropalladate (II) solution was

prepared by dissolving $0.6\,\mathrm{mmol}\,\mathrm{PdCl_2}\,\mathrm{and}\,5.1\,\mathrm{mmol}\,\mathrm{NaCl}$ in $55\,\mathrm{mL}\,\mathrm{of}\,0.1\,\mathrm{mol}\,\mathrm{L}\,\mathrm{hydrochloric}\,\mathrm{acid}.$ This solution was used to coat $1.0\,\mathrm{g}\,\mathrm{of}\,\mathrm{glass}\,\mathrm{wool}.$ The coating procedure was that recommended in the previous paper²⁴.

Sampling and analysis followed the protocol: a) a sorbent tube containing substarte was connected to the stream of the DMS gas mixture; b) after collection of DMS, the packing substrate was transferred to an extraction tube; c) the contents of the extraction tube were shaken with toluene until dissolution of [Pd(DMS)₂Cl₂] was complete; d) the solution was transferred to a 5.0 mL volumetric flask and the volume was adjusted; e) a photometric measurement was made at 303 nm with a Varian-Cary 219 spectrophotometer, using quartz cuvettes of 10.0 mm optical path length.

Gas generation

The experimental arrangement is shown in Figure 1. The pumped air was purified in a column containing, in sequence, silica gel, soda lime and activated carbon. The air stream was maintained at a steady temperature by a coil and flowed into a glass chamber containing a permeation tube. The air flow was controlled at 430 mL min⁻¹ using a critical orifice. For the DMS standard gas mixture, a permeation tube emitting 519 ngDMS min⁻¹ at 30°C (VICI Metronics, Santa Clara, CA) was used. For the NO₂ standard gas mixture, the permeation tube described by Teckentrup and Klockow²⁶, filled with condensed NO₂ gas was used. The permeation rate of NO₂ was evaluated by the oxidation of an iodine solution, according to the procedure described by Littman and Benoliel²⁷. The permeation rate measured was $1.2 \mu g \, NO_2 \, min^{-1}$ at 27°C.

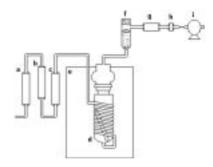


Figure 1. Schematic diagram of apparatus used to produce the gas mixture. a) silica gel; b) soda lime; c) activated carbon; d) permeation tube; e) thermostat bath; f) NO₂ scrubber; g) sorbent tube (Na₂PdCl₄); h) critical orifice; i) air pump.

Oxidant scrubber materials

Initially, solid substrates were prepared coated with substances known to react with nitrogen dioxide. Glycerol was used in some cases because of its humectant property, which should enhance the reactions with nitrogen dioxide. At first, 24 different compositions were tested. The supporting material was prepared from Whatman qualitative filter paper cut into small discs of 5mm diameter. The filter paper was soaked in solution, using 50mL of aqueous solution per 2.5 g. of filter paper. The filters were dried at 60°C in an stove with air circulation, producing the oxidant scrubber materials, which were stored in a desiccator in the dark.

Use of packed scrubbers

A glass tube with flow connections as in Figure 2 (1.5 cm internal diameter, 10 cm length) was filled with 3 g impregnated filter discs and closed with a Teflon stopper. The scrubber discs were retained in the tube with silanized wool glass. Kittler and co-workers¹⁴ proposed visual indication of the oxidation of iodide solution to measure the capacity of substrates for retention of oxidant. The blue/purple coloration of a starch-based iodine indicator was used to compare of several scrubbing systems. This visual comparison of the filters showed a discrepancy of up 24% among replicates. With the purpose of improving this evaluation standard, the measurement of the iodine formed by oxidation was adapted for automated flow analysis (Figure 3). The solution was prepared as suggested by Littman and Benoliel²⁷ and contained about 200g potassium iodide, 36g disodium phosphate dodecahydrate, and 14g potassium dihydrogen phosphate per liter of solution. The solution of iodine in 20% potassium iodide and starch exhibits a blue color with a characteristic absorption spectrum and an intense maximum at 477 nm. In this procedure, the scrubber was connected to the stream of air containing nitrogen dioxide. The air, after passing through the scrubber, readily oxidized the iodide to iodine by any nitrogen dioxide remaining. The absorbance of the iodine solution was continuously measured and recorded. The fast formation of iodine was a signal of scrubber breakthrough.

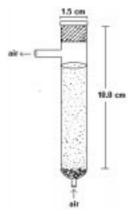


Figure 2. Construction of glass scrubber tube.

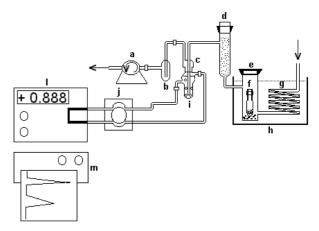


Figure 3. Schematic diagram of automated flow analysis for the measurement of iodine formation. a) air pump; b) security flask; c) bubble flask; d) NO₂ scrubber; e) permeation chamber; f) permeation tube; g) coil of glass; h) thermostat bath; i) iodide solution; j) peristaltic pump; l) spectrophotometer; m) chart recorder.

Results and Discussion

Interference of nitrogen dioxide

The reaction of nitrogen dioxide with the dimethylsulfide collected as $[Pd(DMS)_2Cl_2]_{(s)}$ was evaluated. The procedure involved two steps: first the $Na_2PdCl_{4(s)}$ sorbent tube was connected up to the stream of DMS gas mixture standard for 30 min; in the second step, air containing nitrogen dioxide was passed through the sorbent tube in different periods of time. The absorption spectra of the products formed are shown in Figure 4. The absorption spectra of compounds formed by reaction of nitrogen dioxide with the $[Pd(DMS)_2Cl_2]_{(s)}$ showed the transformation of this compound.

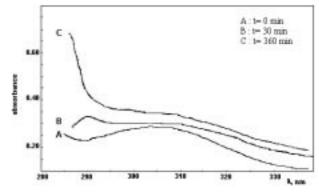


Figure 4. Absorption spectra of compounds formed from the reaction of nitrogen dioxide and $[Pd(DMS)_2Cl_2]$. The time that the air/NO₂ passed through the sorbent tube was: t=0 min (A); t=30 min (B) and, t=6 h (C).

Efficiency of oxidant scrubbers

In air sampling, suitable flow rate and duration of sampling

are two parameters which must be determined for a specific analytical method applied to the compound of interest. If, as here, a sampling time of 30 min at flow rate 430 mL min-1 is chosen for organic sulfide in air, then the oxidant scrubber should remain effective during this period. As a parameter for evaluating the efficiency of oxidant scrubbers for 30 min of sampling, the capacity of a mixture of NO₂ and air to oxidize the iodide was first measured. When the air containing nitrogen dioxide was passed directly through the iodide solution at 430 ml min-1, the absorbance of the solution increased linearly (Figure 5), and after 30 min, the signal measured was 0.330 (absorbance units), corresponding to the iodine formed by reaction of 36 µg of NO₂. In sequence, each one of the scrubbers was connected to the system upstream of the iodide solution and the absorbance of the solution measured. The oxidation of iodide depended on the quantity of NO₂ not retained by the scrubber. To evaluate the efficiency of each oxidant scrubber with respect to retention of NO₂ during a 30 min sampling interval, the following equation was used:

$$e = \frac{\left(0.330 - A_{30}\right)}{0.330} x100 \tag{1}$$

where e is the efficiency in percentage of an oxidant scrubber in retention of NO_2 after 30 min; 0.330 is the absorbance of the iodide solution after containing nitrogen dioxide was passed through the solution for 30 min; and A_{30} is the absorbance of the iodide solution after scrubbed air contained nitrogen dioxide had been passed through the solution for 30 min. Table 1 lists scrubber performance for the 24 different compositions of coating solution studied. The results showed a discrepancy of up to 5% among replicates.

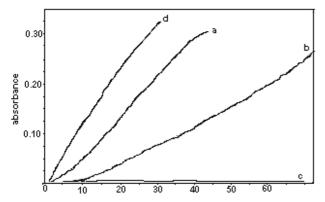


Figure 5. Absorbance of iodine formed. Air with NO₂ (36 μ g mL⁻¹) was scrubbed and passed through iodide solution; The oxidant scrubbers used were made with the following coating solutions: a) 10% (m/v) K₂CO₃; b) 5% (m/v) KI + 5% (v/V) glycerol (pH =7); c) 25% (m/v) ferrous ammonium sulfate + 1% (v/v) sulfuric acid + 5% (m/v) pyrogallic acid; d) air with NO₂ passed through iodide solution with no scrubber.

Table 1. Efficiency of oxidant scrubber in retention of 36 μ g NO₂ for 30 min. The glass tube was filled with different oxidant scrubber materials of coated supporting material (3 g). Scrubber capacity was evaluated by oxidation of iodide solution.

Coat Solution	Efficiency of oxidant
on filter paper (*)	scrubber after
	30 minutes %
10% K ₂ CO ₃	27
15% K ₂ CO ₃	50
25% K ₂ CO ₃	39
35% K ₂ CO ₃	42
$10\% \text{ K}_2\text{CO}_3 + 10\% \text{ glycerol}$	86
$15\% \text{ K}_2\text{CO}_3 + 10\% \text{ glycerol}$	78
$25\% \text{ K}_2\text{CO}_3 + 10\% \text{ glycerol}$	61
$35\% \text{ K}_2\text{CO}_3 + 10\% \text{ glycerol}$	69
10% KHCO ₃ + 10% glycerol	89
10% KOH + 10% glycerol	81
10% glycerol	49
10% sulfanilic acid	64
10% sulfanilic acid + 10% glycerol	89
10% sulfamic acid	36
10% sulfamic acid + 10% glycerol	72
10% pyridine + 10% H ₂ SO ₄ + 10% glyce	rol 33
10% urea + 10% H ₂ SO ₄	41
0,2M[Fe(II)EDTA] + 5% pyrogallic acid	>90
25% ferrous ammonium sulfate + 1% H ₂ S	SO ₄ +
5% pyrogallic acid	>90
5% KI + 5% glycerol (pH= 7)	52
25% diethanolamine	64
25% diethanolamine + 10% glycerol	68
25% triethanolamine	83
25% triethanolamine + 10% glycerol	86

(*)concentration of coated solution: % m/v if solute is solid or v/v for liquid

Effects of reduction of the amount of oxidant scrubber material

In order to design a scrubber that was more compact and easier to handle, the amounts of sorbent materials were reduced and the efficiency of scrubbers re-evaluated. Some sorbent were not tested because their preparation is more laborious. These experiments were carried out using a Teflon tube (0.3 cm internal diameter and 7.5 cm length) fitted with connectors on each end. The packing substrates, about 0.3 g, were held inside the Teflon tube by silanized glass wool. The results showed that the scrubbers maintained a good retention capacity for nitrogen dioxide (Table 2). These scrubbers were then evaluated during collection of DMS onto sampling tubes located downstream of the scrubbers.

Recovery of DMS using different oxidant scrubbers

It is important that the oxidant scrubbers should neither react with DMS, nor adsorb DMS present in the sample air stream. Tests were conducted to measure possible losses of analyte. The DMS standard gas was passed through the oxidant scrubber, and it was collected in the sampling tube over 30 min intervals. Table 3 shows the results of these experiments. The best coating solutions used were: a) 25% (m/v) ferrous ammonium sulfate, 1% (v/v) sulfuric acid and 5% pyrogallic acid; b) 25% triethanolamine. These solutions were also used to coat different supporting materials including: glass wool (fiber medium), silica gel (1-4 mm), and glass pearl (3 mm diameter). Scrubbers containing these materials were tested with the DMS standard gas using the same procedure. Only the solution of 25% (m/v) ferrous ammonium sulfate, 1% (v/v) sulfuric acid and 5% (v/v) pyrogallic acid coated on glass wool was shown to be inert towards DMS.

Table 2. Efficiency of oxidant scrubber in retention of 36 μ g NO₂ using the Teflon tube of 0.3 cm internal diameter and 7.5 cm length and with 0,3 g of coated support material.

Coat Solution on filter paper (*)	Efficiency of oxidant scrubber %
10% K ₂ CO ₃ + 10% glycerol	52
10% sulfanilic acid + 10% glycerol	52
10% sulfamic acid + 10% glycerol	69
25% ferrous ammonium sulfate +	
1% H ₂ SO ₄ + 5% pyrogallic acid	>90
25% diethanolamine	64
25% triethanolamine	85

Table 3. Interference of oxidant scrubbers in sampling of DMS. The DMS standard gas solution was passed through each scrubber collected for 30 min (15.6 μ g DMS).

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Scrubber	Relative DMS Recovery
10% K ₂ CO ₃ + 10% glycerol	71
10% sulfanilic acid + 10% glycerol	76
10% sulfamic acid + 10% glycerol	71
25% ferrous ammonium sulfate +	
1% H ₂ SO ₄ + 5% pyrogallic acid	97
25% diethanolamine	82
25% triethanolamine	96

Studies of preservation DMS collected in sampling tube

The stability of DMS collected as $[Pd(DMS)_2Cl_2]_{(s)}$ was assessed. DMS (15.6 µg) was initially collected in the sampling tube, after which the air/nitrogen dioxide mixture, scrubbed by passage through one of three devices, was passed through the sampling tube for 2 h (about 144 µg NO₂). The results show (Table 4) that the scrubbers employed can be used successfully for retention of NO₂, and hence preservation of DMS collected. It should be noted that the NO₂ concentration used in this experiment was many times higher than typical NO₂ concentrations found in the polluted ambient atmosphere.

Table 4. Losses of DMS collected. DMS (15.6 μg) was collected on the sorbent, then the scrubbed air/NO₂ mixture was passed through the Pd(DMS)₂Cl₁ for 2 h (14.4 mg NO₂).

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Scrubber Type (coating solution/substrate)	Relative DMS Recovery (%)
25% ferrous ammonium sulfate +	
1% H ₂ SO ₄ + 5% pyrogallic acid	
/ filter paper	98
25% ferrous ammonium sulfate +	
1% H ₂ SO ₄ + 5% pyrogallic acid	
/ glass wool	97
25% triethanolamine / filter paper	96

Test of oxidant scrubbers in field sampling

The analytical method reported here in was used to measure atmospheric DMS levels in the environment. The basic system for sampling comprised an air pump, a gasmeter, a sampling tube (glass wool coated with Na₂PdCl₄) and a scrubber. A dust filter made from silanized glass wool was placed in front of the filter pack. The oxidant scrubber made from the solution of 25% (m/v) ferrous ammonium sulfate, 1% (v/v) sulfuric acid and 5% (v/v) pyrogallic acid coated on to filter paper was used because it was easier to handle than the other filters. All samples were collected for 3 h because the levels of DMS were unknown. The sampling was conducted at the Chemistry Institute (Araraguara SP Brasil), July 1995. This site is an urban area and the trace gas compositions may have been affected by the presence of an automobile traffic highway within 500 m, and emissions from anaerobic wastewater treatment 2 Km distant. In the winter (June and July), periodic episodes of odor pollution have been noted at this site. Measurements carried out on polluted days resulted that the level of DMS varied from 3.8-12.4 µg m³. Figure 6 shows the absorption spectra of solutions resulting from field sampling with and without scrubbers. When the scrubber was used, the organic solution obtained from the sampling tube, showed a well-defined absorption peak, with the maximum absorbance at 303 nm characteristic of unoxidized [Pd(DMS)₂Cl₂]. On the other hand, when the filter pack was removed during sampling, the absorption spectrum of the solution obtained showed the existence of oxidized [Pd(DMS)₂Cl₂].

Conclusion

Direct analysis of DMS is generally impossible due to their low concentrations in air, which are frequently below the direct quantification limits of the current methods. Determination of atmospheric DMS employing preconcentration can result in very large sampling losses unless potential oxidants in the air sample are removed prior to the concentration step. Although much work has been done on the analy-

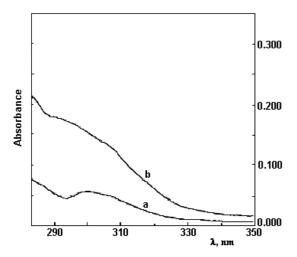


Figure 6. The absorption spectra of solution resulting from field sampling: a) with and b) without scrubber.

sis of DMS in air, the nature of the oxidant interference remains largely unknown. The gas phase reaction of DMS with atmospheric oxidants such as ozone or NO₂ is extremely slow^{28,29}. In the preconcentration procedure it is possible that the through surface sorption the concentration of both oxidant and sulfur compound are increased such that the reaction proceeds more rapidly than in the gas phase¹¹.

Oxidant scrubbers were evaluated for their ability to prevent sampling losses of DMS. The comparison of the filters showed a discrepancy of up to 5% among replicates. Scrubbers utilizing either (a) a mixture of 25% ferrous ammonium sulfate, 1% H₂SO₄ and 5% pyrogallic acid or (b) 25% triethanolamine possess efficient retention capacity for NO2. The scrubbers are easy to prepare and to store. The results showed that these scrubbers are suitable for the preservation of dimethylsulfide collected during atmospheric sampling. Furthermore, the technique employing such scrubbers, combined with the Na₂PdCl₄ sorbent used for preconcentration and analysis of DMS in ambient air, requires minimal sample handling and instrumentation. Results obtained under laboratory conditions and during the course of a small field experiment confirm that the use of a scrubber in combination with a Na₂PdCl₄ sorbent is suitable for measurements of DMS. Therefore, the described method offers a useful alternative for the determination of DMS in ambient air. In addition, use of the scrubbers described could probably be adapted to other analytical techniques of precocentration such as cryogenic trapping or adsoption on a solid support.

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References

- 1. Cullis, C. F.; Hirschler, M. M. *Atmos. Environ.* **1980**, *14*, 1263.
- 2. Moller, D. Atmos Environ 1984, 18, 29.
- 3. Kellogg, W. W; Candle, R. D.;.Allen, R. E; Lazrus; A. L; Martel, J. A *Science* **1972**, *175*, 587
- 4. Berresheeim, H. J. Geophys. Res. 1987, 92,13245
- 5. Cardoso A. A., ; Pitombo, L. R. M. *Quím. Nova* **1992**, 15, 219
- Cruse, H; Pomeroy, R. D. J. Amer. Waters Works Assoc. 1969, 61,677
- Finnerty, W. R. in *Bioconversion of Waste Materials* to *Industrial Product*; Martin, A. M. Ed; Elservier New York, 1991, p 475-500.
- 8. de Souza, T. L. C. JAPCA 1988, 38, 792
- 9. Kohl, A.; Riesenfeld, F. *Gas Purification*; 4th ed. Gulf Publishing Company, Texas, 1985.
- Taver, G. A.; Dasgupta, P. K. Atmos Environ 1995, 29, 1291
- 11. Saltzman, E. S.; Cooper, D. J. In *Biogenic Sulfur in the Environment;* Saltzman, E.S.; Cooper, D.J. Ed, Washington, ACS symposium serie, 1989, p. 330-335.
- 12. Hofmann, U.; Hofmann, R; Kesselmeier, J. *Atmos Environ* **1992**, *26*, 2445.
- 13. Davison, B. M; Allen A. G. *Atmos Environ* **1994**, 28. 1721.
- 14. Kittler, P.; Swan, H.; Ivey, J. *Atmos. Environ.* **1992**, 26, 2661.
- 15. Wardencki, W. J. Chromatogr. A 1998, 793, 1
- 16. Braman, R. S.; Ammons, J. M.; Bricker, J. L. *Anal. Chem.* **1978**, *50*, 992.
- 17. Barnard, W. R.; Andreae, M. O.; Watkins, W. E.; Bingemer, H.; Georgii, H. W. *J. Geophys. Res.* **1982**, 87,787.
- 18. Andreae, M. O.; Raemdonck, H. *Science* **1983**, 221,744.
- 19. Bates, T. S.; Johnson, J. E. EOS 1990, 71, 1254.
- 20. Saltzman, E.; Copper D. J. Atmos. Chem. 1988, 6, 191
- 21. Cooper, D. J.; Saltzman, E. S. *J. Atmos Chem.* **1991**, *12*, 153.
- 22. Watts, S. F. Environ. Technol. Lett. 1989, 10, 777.
- 23. Persson, C.; Leck, C. Anal. Chem. 1994, 66, 983
- 24. Pitombo, L. R. M.; Cardoso, A. A. *Intern. J. Environ. Anal. Chem.* **1994**, *54*, 221.
- 25. Cardoso, A. A. Tetracloropaladato (II) de Sódio

Sobre Sólidos, Sorventes Para Preconcentração de Compostos Reduzidos de Enxofre, Instituto de Química, USP **1991**, Ph.D. thesis.

26. Teckentrup, A.; Klockow, D. Anal. Chem. 1978, 50, 1728.

- 27. Littman, F. E.; Benoliel, R. W. Anal. Chem. 1953, 25, 1480.
- 28. Yin, F.; Grosjean; Seinfeld; J. H. *J. Geophys. Res.* **1986**, *91*, 14417
- 29. Grosjean; D. Environ. Sci. Technol. 1984, 18, 460.

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