

RESSALVA

Atendendo solicitação do(a) autor(a), o texto completo deste trabalho será disponibilizado somente a partir de 20/02/2019.

UNIVERSIDADE ESTADUAL PAULISTA “JÚLIO DE MESQUITA FILHO”

Instituto de Biociências de Botucatu

Departamento de Física e Biofísica

UNIVERSITAT DE BARCELONA

Facultat de Farmàcia

“Structural studies of PLA₂-like toxins and development of
the structure solution method SEQUENCE SLIDER”

RAFAEL JUNQUEIRA BORGES

Botucatu – SP
January / 2017

UNIVERSIDADE ESTADUAL PAULISTA “JÚLIO DE MESQUITA FILHO”

Instituto de Biociências de Botucatu

Departamento de Física e Biofísica

UNIVERSITAT DE BARCELONA

Facultat de Farmàcia

CONSEJO SUPERIOR DE INVESTIGACIONES CIENTÍFICAS

Instituto de Biología Molecular de Barcelona

Departamento de Biología Estructural

PARC CIENTIFIC DE BARCELONA

Crystallographic Methods Laboratory

PROGRAMA DE PÓS-GRADUAÇÃO EM CIÊNCIAS BIOLÓGICAS (GENÉTICA)

PROGRAMA DE DOCTORAT EM BIOTECNOLOGIA

“Structural studies of PLA₂-like toxins and development of
the structure solution method SEQUENCE SLIDER”

Doctoral thesis presented by Rafael Junqueira Borges, Bachelor degree in
Biomedical Sciences, Ms.C. in Genetics, for the degree of Doctor (Ph.D.) from the
University of State of Sao Paulo and the University of Barcelona.

Thesis director: Dr. Marcos Roberto de Mattos Fontes, UNESP Prof.

Thesis co-director: Dr. Isabel Usón Finkenzeller, ICREA Prof.

Botucatu – SP
January / 2017

FICHA CATALOGRÁFICA ELABORADA PELA SEÇÃO TÊC. AQUIS. TRATAMENTO DA INFORM.
DIVISÃO TÉCNICA DE BIBLIOTECA E DOCUMENTAÇÃO - CÂMPUS DE BOTUCATU - UNESP
BIBLIOTECÁRIA RESPONSÁVEL: ROSANGELA APARECIDA LOBO-CRB 8/7500

Borges, Rafael Junqueira.

Structural studies of PLA₂-like toxins and development of the structure solution method SEQUENCE SLIDER / Rafael Junqueira Borges. - Botucatu, 2017

Tese (doutorado) - Universidade Estadual Paulista "Júlio de Mesquita Filho", Instituto de Biociências de Botucatu
Orientador: Marcos Roberto de Mattos Fontes
Coorientador: Isabel Usón Finkenseller
Capes: 20901003

1. Cobra venenosa - Veneno. 2. Fosfolipases. 3. Cristalografia. 4. Biologia molecular.

Palavras-chave: Crystallography; Methods development; PLA₂-like proteins; Phasing; Snake venom.

To all those who fight to eradicate human ignorance and suffering.

Acknowledgements

I express my sincere acknowledgements to the institutions and people that contributed for the development of my thesis.

Dr. Marcos Fontes, for the incentive and teachings since my early undergraduate research and for the given background that allowed me to adventure over new scientific fields and laboratories

Dr. Isabel Usón, for the innumerable scientific discussions personally and via video-conference, for the patience, persistence and dedication to introduce me to phasing and programming, and for trusting my work.

Dr. Ney Lemke, for guiding me over mathematics, normal model analysis and scientific ideas with metaphoric examples.

Dr. Mario Oliveira, for the aid collecting SAXS experiments and interpreting results.

Dr. George Sheldrick, for the hospitality receiving me in his laboratory in University of Goettingen and for being always available to answer any complicated question from chemistry to applied crystallography with an astonishing simplicity.

Dr. Randy Read, who received me in his laboratory in University of Cambridge, for the impressive scientific clear thoughts and for the assertive suggestions over my ideas.

Henrique Barcellos Campanelli, for the help in the experiments and for the opportunity to allow me to teach what I have acquired over my scientific career.

Our collaborators: Dr. Bruno Lomonte, Dr. Maria Laura Fernandez, Dr. Andreimar Soares, Dr. Luis Ponce-Soto, Dr. Sergio Marangoni, Dr. Salomón Huacahuire-Veja, for the various supplied snake venom proteins and inhibitors, and Dr. Ehmke Pohl, Dr. Isabel dos Santos, Edson José Comparetti, for the supplied crystallographic datasets.

LNLS-MX1&2 scientific instrumentation team, Dr. Ana Carolina de Mattos Zeri, Me. Alexandre Lo Bianco Dos Santos and Luciano Braga Candido, for the help setting the X-ray beamline to better meet our needs.

LN BIO mass spectrometry technicians, Dra. Bianca Alves Pauletti and Romênia Domingues, for the effort to aid sample preparation and results interpretation.

FAPESP, for the financial support during a 4-year project (Process number 2013/00873-4 and 2014/11182-5).

My colleagues in the Structural Molecular Laboratory and the Crystallographic Methods Laboratory, in special to Claudia and Massimo, for the enriching discussion and for pushing me to my best.

My colleagues in the laboratories I visited, for the effort to fasten my adaptation and for extending my scientific visit into a cultural exchange and discovery.

All my friends over the cities I lived during this sandwich PhD, Botucatu, Barcelona, Goettingen and Cambridge, for the unique experiences, relaxing moments and amazing trips.

My parents, brother and family, for the endless love, care and support.

“To raise new questions, new possibilities, to regard old problems from a new angle, requires creative imagination and marks real advance in science.”

Albert Einstein

ABSTRACT

Phospholipases A₂ (PLA₂s) are one of the main component of *Bothrops* genus snakes and one of the main responsible for muscular necrosis in envenomation, a consequence not neutralized by administration of antiophidian serum. These proteins are myotoxic by disrupting membrane phospholipids by a catalytic mechanism dependent of calcium or by perturbing membrane integrity by an independent of calcium mechanism not fully known. Usually, snake toxins are purified directly from the natural source, extracted venom, and purity is a challenge due to the co-existence of various isoforms. The objectives of this thesis were to understand the non-catalytic membrane perturbation mechanism by studying the snake myotoxins and to propose a new crystallographic method to deal with impure samples, called SEQUENCE SLIDER. We performed structural X-ray crystallography studies, complemented with other biophysical techniques, such as small-angle X-ray scattering, with three bothropic myotoxins in *apo* state and complexed to natural products and inhibitors. We proposed local and global measurements to characterize and relate these states to toxin function. With the method SEQUENCE SLIDER, we aided elucidation of structures with partial sequence known by evaluating different side chain against real space correlation coefficient calculated from diffraction data. Furthermore, we developed SEQUENCE SLIDER to increase the scope of the crystallographic phasing program ARCIMBOLDO to lower resolution than the usual 2 Å. In such cases, different hypotheses of sequences of the partial traces of polyalanine are evaluated simultaneously and pushed through autotracing until the structure is phased. We improved the myotoxic mechanism comprehension and developed a structure solution method for challenging crystallographic structures.

Keywords: Structural molecular biology; Phospholipases A₂-like; Snake venom; phasing; ARCIMBOLDO.

RESUMO

As fosfolipases A₂ (PLA₂s) são um dos maiores constituintes protéicos do veneno botrópico e um dos responsáveis pela necrose muscular, consequência esta não eficazmente neutralizada pela administração do soro antiofídico. Estas proteínas são tóxicas através do rompimento ou perturbação da membrana celular em um mecanismo catalítico dependente de cálcio e outro independente, sendo este último não totalmente elucidado. Usualmente, estas toxinas são obtidas diretamente do veneno das serpentes, sendo sua purificação um desafio pela coexistência de diferentes isoformas. O objetivo desta tese foi compreender o mecanismo miotóxico independente de cálcio através de estudos estruturais e propor nova metodologia que trate de dados cristalográficos de toxinas provenientes de amostras impuras, chamada SEQUENCE SLIDER. Para tanto, cristalografia e outras técnicas biofísicas, como espalhamento de raios X a baixo ângulo, serão utilizados para estudar três miotoxinas ofídicas em estado nativo e complexado com produtos naturais e inibidores. Nós propusemos medidas locais e globais para caracterizar e relacionar a estrutura dessas toxinas a função. Com o SEQUENCE SLIDER, pudemos elucidar as estruturas de toxinas inéditas cuja sequência era parcialmente conhecida. Esta nova metodologia proposta consiste em avaliar diferentes cadeias laterais contra o coeficiente de correlação em espaço real calculado a partir dos dados cristalográficos. Em paralelo, desenvolvemos o SEQUENCE SLIDER no âmbito do método cristalográfico *ab initio* ARCIMBOLDO com objetivo de aumentar seu escopo a dados com resolução mais baixa que 2 Å. Nestes casos, diferentes hipóteses de sequências para as soluções parciais de polialanina são avaliadas mutualmente e enviadas a autotraçamento até que grande parte da estrutura seja faseada. Pudemos melhorar a compreensão do mecanismo miotóxico e desenvolver um programa que poderá auxiliar resolução de estruturas cristalográficas desafiadoras.

Palavras chave: Biologia molecular estrutural; Fosfolipases A₂ homólogas; Veneno de serpente; faseamento; ARCIMBOLDO.

RESUM

Les fosfolipases A₂ (PLA₂s) de les serps del gènere *Bothrops* són un dels principals responsables de la necrosi muscular en l'enverinament, una conseqüència no neutralitzada per l'administració de sèrum antiofídic. Aquestes proteïnes són miotòxiques pel seu efecte disruptiu sobre la membrana fosfolipídica, bé per un mecanisme catalític dependent de calci o bé per un mecanisme no dependent de calci poc descrit a la bibliografia. En general, les toxines de serp es purifiquen directament de la seva font natural, el verí extret, i la purificació és un repte a causa de la coexistència de diverses isoformes. Els objectius d'aquesta tesi van ser entendre el mecanisme no catalític de pertorbació de la membrana mitjançant l'estudi de miotoxines de serp i proposar un nou mètode cristal·logràfic per fer front a les mostres impures, anomenat SEQUENCE SLIDER. Es van realitzar estudis estructurals de cristal·lografia de raigs X, complementats amb altres tècniques biofísiques, com SAXS (dispersió de raigs X d'angle petit), amb tres miotoxines en estat apo i acomplexades amb productes naturals i inhibidors. Hem proposat mesuraments locals i globals per caracteritzar i relacionar aquests estats amb la funció de la toxina. Amb el mètode SEQUENCE SLIDER, hem facilitat l'elucidació de les estructures amb seqüència parcial coneguda mitjançant l'avaluació de diferents cadenes laterals contra el coeficient de correlació de l'espai real calculat a partir de dades de difracció. D'altra banda, hem desenvolupat SEQUENCE SLIDER per augmentar l'abast del programa ARCIMBOLDO amb resolució més baixa que l'habitual de 2 Å. En aquests casos s'avaluen simultàniament diferents hipòtesis de seqüències de les traces parcials de polialanina i es proven mitjançant l'autotraçat fins que l'estructura sigui resolta. Hem millorat la comprensió del mecanisme miotòxic i hem desenvolupat un mètode de solució estructural per a estructures cristal·logràfiques desafiantes.

Paraules clau: Biologia molecular estructural; Fosfolipases A₂-like; Verí de serp; Phasing; ARCIMBOLDO.

INDEX

| | |
|--|----|
| LIST OF FIGURES..... | 12 |
| LIST OF TABLES | 15 |
| LIST OF ABBREVIATIONS AND ACRONYMS..... | 17 |
| 1 INTRODUCTION..... | 19 |
| THESIS OUTLOOK..... | 20 |
| 2 STRUCTURAL STUDIES OF SNAKE VENOM PLA ₂ S AND PLA ₂ -LIKE PROTEINS | 22 |
| 2.1 Study of venom | 23 |
| 2.2 Snake venom accidents in the World and Brazil..... | 23 |
| 2.3 Snake venom PLA ₂ and its catalytic mechanism | 24 |
| 2.4 Snake venom PLA ₂ -like membrane perturbation mechanism..... | 25 |
| 2.5 Myotoxicity of some bothropic PLA ₂ S and PLA ₂ -like proteins independent of calcium..... | 31 |
| 2.6 Objective | 31 |
| 2.7 Material and Methods..... | 32 |
| 2.8 Comparison of available snake venom PLA ₂ -like proteins structures | 36 |
| 2.8.1 Quaternary structure possibilities..... | 36 |
| 2.8.2 Global measurement: orientation and translations between almost identical objects | 40 |
| 2.8.3 Tertiary structure variability: Local measurement using C α and C β distances..... | 46 |
| 2.8.4 PLA ₂ -like toxins hydrophobic channel characterization and accessibility | 48 |
| 2.8.5 Results and discussion of global and local measurement..... | 52 |
| 2.8.6 Evaluation of flexibility of PLA ₂ -like protein in active and inactive states..... | 54 |
| 2.8.7 New steps in the myotoxic mechanism of PLA ₂ -like toxins..... | 56 |
| 2.8.8 Structural studies of BthTX-I complexed with zinc..... | 57 |
| 2.8.8.1 Zinc interaction site and its relationship to inhibition..... | 58 |
| 2.8.9 Structural studies of BthTX-I complexed with MMV | 60 |
| 2.8.10 Concluding remarks | 70 |
| 2.9 Structural studies of snake venom basic D49-PLA ₂ S independent of calcium..... | 71 |

| | | |
|-----------|---|-----|
| 2.9.1 | Small-angle X-ray scattering studies of BthTX-II | 71 |
| 2.9.2 | Crystallographic studies of basic PLA ₂ s | 75 |
| 2.9.2.1 | Monomeric BthTX-II complexed to fatty acids | 75 |
| 2.9.2.1 | Apo BthTX-II..... | 78 |
| 2.9.2.3 | Apo PrTX-III..... | 80 |
| 2.9.3 | Discussion | 80 |
| 2.9.3 | Conclusion..... | 84 |
| 3 | SEQUENCE SLIDER..... | 86 |
| 3.1 | Crystallography | 87 |
| 3.2 | The phase problem and structure elucidation..... | 88 |
| 3.3 | Crystallography, ab initio, and ARCIMBOLDO | 91 |
| 3.4 | Structure elucidation of crystal containing multiple isoforms | 95 |
| 3.5 | Objective | 97 |
| 3.6 | Material and Methods..... | 97 |
| 3.7 | SEQUENCE SLIDER into venoms | 100 |
| 3.7.1 | Algorithm description | 100 |
| 3.7.2 | Known test case..... | 102 |
| 3.7.2.1 | Synthetic AtxA..... | 103 |
| 3.7.2.1.1 | Full search | 103 |
| 3.7.2.1.2 | Restricted search | 103 |
| 3.7.2.2 | Natural AtxC | 104 |
| 3.7.2.2.1 | Full search | 104 |
| 3.7.2.2.2 | Restricted search | 104 |
| 3.7.2.3 | Comparison of AtxA and C datasets | 104 |
| 3.7.3 | Unknown structures..... | 108 |
| 3.7.3.1 | BbTX-III..... | 108 |
| 3.7.3.1.1 | Preliminary SEQSLIDER run: CONSTRUCTIVE and FULL SEARCH mode | 111 |

| | | |
|-----------|---|-----|
| 3.7.3.1.2 | Introducing mass spectrometry results as restriction..... | 113 |
| 3.7.3.1.3 | Discussion of restricted evaluation of residues | 113 |
| 3.7.3.2 | Mt-I | 115 |
| 3.7.3.2.1 | Full search mode | 116 |
| 3.7.3.2.2 | Results and discussion of restricted search mode..... | 116 |
| 3.8 | SEQSLIDER in ARCIMBOLDO scope | 122 |
| 3.8.1 | SEQUENCE SLIDER program..... | 123 |
| 3.8.2 | MltC | 128 |
| 3.8.3 | FrmR E64H de Salmonella enterica..... | 135 |
| 3.9 | Conclusion..... | 139 |
| 4 | REFERENCES..... | 141 |
| 5 | SUPPLEMENTARY MATERIAL | 152 |
| 6 | ATTACHMENTS | 168 |
| 7 | APPENDIX | 170 |

LIST OF FIGURES

| | |
|---|----|
| Figure 2.1 Structures and interaction site of snake venom PLA ₂ (A) and PLA ₂ -like proteins (B). | 29 |
| Figure 2.2 Drawing of the PLA ₂ -like monomer and dimers. | 30 |
| Figure 2.3 Compact dimer of PLA ₂ -like proteins and dos Santos, Soares and Fontes (2009) angle proposition..... | 30 |
| Figure 2.4 - Superposition in cartoon of two basic Asp49-PLA ₂ s, BthTX-II and PrTX-III, and an acid PLA ₂ , DacuTX. | 31 |
| Figure 2.5 - Representation of the aeronautical (Tait-Bryan) angles of movement in an airplane and its similarity to our proposed PLA ₂ -like protein angles description. | 41 |
| Figure 2.6 - Graph of the RMSF in Å by residues of available bothropic PLA ₂ -like crystallographic structures. | 42 |
| Figure 2.7 - Cartoon representation of asymmetry in C-termini of BthTX-I/Zn canonical (in orange) and non-canonical monomer (in brown). | 47 |
| Figure 2.8 - Comparison of ligands interacting to PLA ₂ -like protein H48 (in A) and to MDiS (in B). | 49 |
| Figure 2.9 - Tunnels analysis for bothropic PLA ₂ -like crystal structures in the inactive (A and B) and active states (C). | 50 |
| Figure 2.10 - The hydrophobic channel accessibility with tunnel volume calculation, the distance of MDiS residues and monomer-monomer angle for all bothropic available PLA ₂ -like toxins. | 53 |
| Figure 2.11 - Angles between monomers along the range of -6 to 1.5 Å of NM09 of Inac and along the range of 1.5 and -6 Å of NM08 of Act..... | 56 |
| Figure 2.12 - Structural transitions of the C-termini from the BthTX-I that occurs during hydrophobic molecule entrance..... | 57 |
| Figure 2.13 - BthTX-I and zinc ions interacting regions and coordination distances (black dashes)... .. | 60 |
| Figure 2.14 - Molecular structure of MMV (12-methoxy-4-methyl-voachalotine)..... | 62 |
| Figure 2.15 - Effect of MMV in BthTX-I quaternary and secondary structure. | 62 |
| Figure 2.16 - Complex of BthTX-I with MMV in sticks. | 65 |
| Figure 2.17 - Structural comparison between BthTX-I/MMV with other PLA ₂ -like protein structures. | 66 |
| Figure 2.18 - BthTX-I different dimers in cartoon representation. | 69 |
| Figure 2.19 - Experimental SAXS curves of BthTX-II in 50 mM HEPES pH 7.4..... | 72 |
| Figure 2.20 - Experimental SAXS curves of BthTX-II in low concentration (1 mg/mL) and in 50 mM sodium citrate pH 5.2. | 73 |

| | |
|---|-----|
| Figure 2.21 - Experimental SAXS curves of BthTX-II in high concentration (5 mg/mL) and in 50 mM sodium citrate pH 5.2. | 73 |
| Figure 2.22 - Experimental SAXS curves of BthTX-II in 50 mM HEPES pH 7.4 and theoretical curve of monomeric BthTX-II structure. | 74 |
| Figure 2.23 - Crystal of basic and bothropic PLA ₂ s..... | 77 |
| Figure 2.24 - Ligands found in the BthTX-II crystallographic structures..... | 79 |
| Figure 2.25 - Structural comparison of cartoon representation of monomers of BthTX-II/STE with canonical calcium binding loop and apo BthTX-II and PrTX-III with distorted calcium binding loop. | 82 |
| Supplementary Figure 2.26 - Normal Mode (NM) 7 to 10 analysis of structures in intermediate (in A) and active state (in B) and their monomer-monomer angles (ψ , θ , and ϕ)..... | 157 |
| | |
| Figure 3.1 - Portrait of a human composed of assembly of vegetables (A) and “portrait” of a protein composed of assembly of small fragments on the right (B)..... | 92 |
| Figure 3.2 - Unknown structures solved with ARCIMBOLDO versions. | 94 |
| Figure 3.3 - Program flow of SEQSLIDER applied to venoms. Colored background indicates the use of external programs. | 100 |
| Figure 3.4 - Amino acids with similar atomic composition. | 108 |
| Figure 3.5 - Sticks representation of residues differences in Atx isoforms A (purple) and C (dark yellow) and respective polder maps. | 108 |
| Figure 3.6 - Sequence similarity of BbTX-III against PLA ₂ s. | 109 |
| Figure 3.7 - R _{factors} and identity along first round of SEQSLIDER full search and constructive mode with BbTX-III dataset starting from a polyala model. | 112 |
| Figure 3.8 – Residue 1 and 12 and their polder map (3.5 σ) correspondent to Asn and Iso, respectively, different than mass spectrometry results. | 114 |
| Figure 3.9 - Sequence similarity of Mt-I against PLA ₂ s. | 115 |
| Figure 3.10 – Sticks representation of divergent residues RSCC/MS of Mt-I with their polder maps (3.5 σ). | 118 |
| Figure 3.11 – Sticks representation of 109Y of Mt-I and its hydrogen network with its polder maps (3.5 σ). | 119 |
| Figure 3.12 – Cartoon representation of Mt-I with uncertain side chain in sticks and circled in dashes. | 119 |
| Figure 3.13 - Divergent RSCC between chain A/B of Mt-I and their respective polder maps (3.5 σ). | 122 |

| | |
|---|-----|
| Figure 3.14 - Program flow of SEQSLIDER. Colored background indicates the use of external programs..... | 124 |
| Figure 3.15 - Structure of a coil coiled after ARCIMBOLDO phasing with electron density with 1.5 σ | 124 |
| Figure 3.16 - Alignments of predicted secondary structure elements of known sequence (in A) against structure fragments from chain in model (in B)..... | 127 |
| Figure 3.17 - Cartoon representation of MltE (in red PDB id: 2Y8P) and of MltC (transparent in green PDB id: 4C5F)..... | 128 |
| Figure 3.18 - Cartoon representation of main chain expansions of MltC prior and after SEQSLIDER runs..... | 131 |
| Figure 3.19 - Post mortem graph of SHELXE of different stages of MltC phasing by SEQSLIDER. 0 is prior to SEQSLIDER launch, 1 and 2 are the first and second SEQSLIDER runs, respectively.... | 132 |
| Figure 3.20 - Evaluation of the hypotheses of chain E and its plots of R, R_{free} , CC_{mc} , CC_{sc} | 132 |
| Figure 3.21 - Evaluation of the hypotheses of evaluated chains in SEQSLIDER cycle 1 by FOM and identity..... | 133 |
| Figure 3.22 - Evaluation of the hypotheses of evaluated chain in SEQSLIDER cycle 2 by FOM and identity..... | 133 |
| Figure 3.23 - The different fragments of the missing N-terminal domain of MltC. | 135 |
| Figure 3.24 - Evaluation of the hypotheses of all chains in SEQSLIDER by FOM and identity (%). 136 | |
| Figure 3.25 – Graphs of the improvement of Real Space Correlation Coefficient (RSCC) and $R_{factors}$ with increase of number of residues assigned by SEQSLIDER run. | 138 |
| Figure 3.26 – Alignment of the best hypothesis of each SEQSLIDER evaluation by chain with complete sequence in top. | 139 |
| Supplementary Figure 3.27 - Graph of local variability of similar sequences to Atx..... | 158 |
| Supplementary Figure 3.28 - Graph of local variability of similar sequences to BbTX-III..... | 158 |
| Supplementary Figure 3.29 - Graph of local variability of similar sequences to Mt-I..... | 159 |

LIST OF TABLES

| | |
|---|-----|
| Table 2.1 - Dimeric evidence of the PLA ₂ -like toxins whose crystallographic structures were elucidated. | 37 |
| Table 2.2 - Summary of PLA ₂ -like crystallographic models and their dissociation calculations. | 38 |
| Table 2.3 - Summary of local and global measurements of PLA ₂ -like crystallographic models..... | 39 |
| Table 2.4 - Data collection statistics of the X-ray diffraction of the crystals of BthTX-I/MMV..... | 64 |
| Table 2.5 - Composition of complexes within crystal contacts of BthTX-I/MMV structure | 68 |
| Table 2.6 - SAXS measurement of BthTX-II in citrate and HEPES buffer and theoretical scattering of different BthTX-II quaternary assemblies..... | 74 |
| Table 2.7 - Experimental data of dynamic light scattering (DLS) of BthTX-II in buffer of sodium citrate and HEPES..... | 74 |
| Table 2.8 - Crystallization setup of BthTX-II and PrTX-III crystals..... | 75 |
| Table 2.9 - X-ray data collection and refinement of BthTX-II/STE and apo BthTX-II and PrTX-III.. | 76 |
| Table 2.10 - BthTX-II/STE and apo BthTX-II and PrTX-III comparison in RMSD (C α). | 82 |
| Supplementary Table 2.11 - Summary of PLA ₂ -like crystallographic models and their dissociation calculations..... | 153 |
| Supplementary Table 2.12 - Summary of region binding of ligands of available bothropic PLA ₂ -like crystallographic models..... | 154 |
| Supplementary Table 2.13 - List of residues that composes the inner and outer of the BthTX-I/Zn dimer cavity and their chemical properties calculated by MOLE 2.0. | 155 |
| Supplementary Table 2.14 - List of residues that composes the tunnels and the chemical properties of these tunnels calculated by MOLE 2.0..... | 155 |
| Supplementary Table 2.15 - BthTX-I/MMV comparison with other bothropic PLA ₂ -like toxin structures in RMSD (C α). | 156 |
| Table 3.1 - Overall data statistics of crystallographic models Atx..... | 102 |
| Table 3.2 - Summary of full and restricted SEQSLIDER searches of Atx datasets..... | 105 |
| Table 3.3 - Summary of evaluated residues in 3G8G and 3G8H of SEQSLIDER full search run whose true residue did not score best..... | 106 |

| | |
|--|-----|
| Table 3.4 - X-ray data collection and refinement with best obtained model of BbTX-III and MT-I | 110 |
| Table 3.5 - Correlation coefficient of PLA ₂ and BbTX-II residues whose mass spectrometry matched..... | 112 |
| Table 3.6 - SEQSLIDER summary of BbTX-III run with RSCC phylogenetic restriction of residues having divergent results with mass spectrometry results | 114 |
| Table 3.7 - SEQSLIDER summary of Mt-I run with RSCC residues diverging mass spectrometry match or phylogenetic analysis..... | 118 |
| Table 3.8 - SEQSLIDER summary of Mt-I run with ambiguous RSCC possibilities | 119 |
| Table 3.9 - SEQSLIDER summary of Mt-I run of residues undetermined..... | 120 |
| Table 3.10 - SEQSLIDER summary of Mt-I residues whose RSCC of chains diverge..... | 121 |
| Table 3.11 - SEQSLIDER scoring function of the local alignment | 127 |
| Table 3.12 - Different strategies to improve initial phases from MltC partial solution and SHELXE autotracing..... | 129 |
| Table 3.13 - Secondary structure chain composition, number of hypotheses generated by SEQLISDER and maximum observed identity..... | 132 |
| Table 3.14 - SEQSLIDER overview on FrmR E64H dataset..... | 136 |
| Supplementary Table 3.15 - Correlation coefficient of residues whose scored matched mass spectrometry results of BbTX-III | 160 |
| Supplementary Table 3.16 - SEQSLIDER summary of BbTX-III run with phylogenetic restriction of residues having more than one mass spectrometry match | 161 |
| Supplementary Table 3.17 - SEQSLIDER summary of BbTX-III run with phylogenetic restriction of unknown residues..... | 161 |
| Supplementary Table 3.18 - SEQSLIDER summary of BbTX-III residues whose electron density was poor | 162 |
| Supplementary Table 3.19 - Correlation coefficient of residues with clear distinction of Mt-I | 163 |
| Supplementary Table 3.20 - SEQSLIDER run on Mt-I and summary of convergence results | 165 |

LIST OF ABBREVIATIONS AND ACRONYMS

| | |
|-------------------|--|
| ASU | Asymmetric unit |
| Atx | Ammodytoxin |
| B1MMV | Structure BthTX-I complexed to MMV |
| BPB | 4-bromophenacyl bromide |
| BthTX-I | First protein identified from <i>Bothrops jararacussu</i> venom |
| BthTX-II | Second protein identified from <i>Bothrops jararacussu</i> venom |
| CC | Correlation Coefficient |
| CC7 | 7 Ca centroid coordinates |
| CC _i | initial CC |
| CC _f | final CC |
| CC _{mc} | RSCC of main chain atoms |
| CCS _a | RSCC calculated by <i>EDSTATS</i> |
| CC _{sc} | RSCC of side chain atoms |
| CD | Circular Dichroism |
| dMDiS | distance between C β of MDiS residues 121, and 125 |
| iFace | interface-binding surface |
| LNLS | Laboratório Nacional de Luz Síncrotron |
| MDiS | Membrane-Disruption Site comprehended by L121, and F125 |
| MDoS | Membrane-Docking Site comprehended by K115, R118, and K20 |
| MMV | 12-methoxy-4-methyl-voachalotine |
| MR | Molecular Replacement |
| M _w | Molecular weight |
| NM | Normal Mode |
| NSLS-I | National Synchrotron Light Source I |
| PDB | Protein Databank |
| PEG | polyethylene glycol |
| PCC | RSCC calculated by <i>PHENIX.POLDER</i> |
| PLA _{2s} | Phospholipases A2 |
| PrTX-III | Third protein identified from <i>Bothrops pirajai</i> venom |
| R _g | radius of gyration |
| RSCC | Real Space Correlation Coefficient |
| SAXS | Small Angle X-ray Scattering |
| SEQSLIDER | SEQUENCE SLIDER |
| R _h | Hydrodynamic radius |
| RMSD | root-mean-square deviation |
| RMSF | root-mean-square fluctuation |
| SS | Secondary Structure |
| wMPE _i | initial weighted mean phase error |
| wMPE _f | final weighted mean phase error |

| Amino acid | 3-letter abbreviation | 1-letter abbreviation |
|---------------|-----------------------|-----------------------|
| Alanine | Ala | A |
| Arginine | Arg | R |
| Asparagine | Asn | N |
| Aspartic acid | Asp | D |
| Cysteine | Cys | C |
| Glutamic acid | Glu | E |
| Glutamine | Gln | Q |
| Glycine | Gly | G |
| Histidine | His | H |
| Isoleucine | Ile | I |
| Leucine | Leu | L |
| Lysine | Lys | K |
| Methionine | Met | M |
| Phenylalanine | Phe | F |
| Proline | Pro | P |
| Serine | Ser | S |
| Threonine | Thr | T |
| Tryptophan | Trp | W |
| Tyrosine | Tyr | Y |
| Valine | Val | V |

1 INTRODUCTION

THESIS OUTLOOK

This thesis focuses on the study of the mechanism of action of the phospholipases A₂ (PLA₂S) and PLA₂-like toxins from the snakes of *Bothrops* genus and on the development of the structure solution method called SEQUENCE SLIDER (SEQSLIDER).

The first chapter starts with a general introduction on the importance to study snake venom and its constituents, on ophidic accidents epidemiology and on the toxic effect of PLA₂ and PLA₂-like proteins that are still not neutralized but current antivenom. The structural information available is correlated with what is known on their toxic mechanism as a background for the discussion of our results. The methodologies that we proposed are shown as a result of this thesis and are not included in the Material and Methods.

We present results and discussions on PLA₂-like proteins in **section 2.8**. We start proposing three complementary methodologies that better characterize the more than 20 available structures. First, measuring the geometric orientation between the two identical monomers of the dimeric PLA₂-like proteins structures using Euler Angles. Second, we measure the local variability of the monomers. And third, we calculate hydrophobic channel accessibility. We discuss the results of these three methodologies finding different toxin states related to how they damage membranes. The hypothesis raised in previous sections is tested with Normal Mode Analysis. Essential to the proposition of the previous analysis, we present the results of the complexes of BthTX-I, purified from *Bothrops jararacussu* venom, with the inhibitors zinc and MMV.

We present in **section 2.9** the experimental results of two basic PLA₂ that may resemble PLA₂-like proteins, BthTX-II and PrTX-III from *Bothrops jararacussu* and *Bothrops pirajai*, respectively. SAXS, DLS and crystallography are used to better characterize their structures. Their possible mechanism of action is discussed relating to the literature information.

Chapter 3 focuses on the second objective of this project, the proposition of a method that aids solution of crystallographic challenging datasets. A brief history of crystallography and its current challenges are described. The environment in which SEQSLIDER originated is outlined in the ARCIMBOLDO scope and in the complexity composition of venoms.

We describe SEQSLIDER focused in venoms in **section 3.7**. We describe the algorithm and how it evaluates different side chain using real space correlation coefficient. First, we test the algorithm on high resolution known structures from both natural and synthetic source. Second, we elucidate two previously unknown myotoxins with SEQSLIDER.

We describe SEQSLIDER focused in phasing within the ARCIMBOLDO scope in **section 3.8**. Again, the algorithm is first described, a known structure is elucidated, MltC, followed by study of an unknown case, FrmR E64H.

In the appendix, we attach the published articles related to this thesis. The proposition of a comprehensive myotoxic mechanism (FERNANDES et al., 2013), a review of the available structures of PLA₂-like proteins (FERNANDES et al., 2014) and the functional and structural study of BthTX-I with zinc (BORGES et al., 2017) are all related to **section 2.8**. At last, the application ARCIMBOLDO_LITE in a large set of structures (SAMMITO et al., 2015) is related to **section 3.3**.

This thesis present two symbiotic objectives, the understanding of toxins mechanism of action and the proposition of methodologies that aid solution of structures of medical importance. The structural description of PLA₂s and PLA₂-like proteins allowed the proposition of additional steps in these toxins mechanism of action. Three different bioinformatics tools are employed to evaluate both local and global structural features that may also be used to characterize structural movement from Molecular Dynamics models and other family of proteins. As a side chain evaluator, SEQSLIDER may aid structure solution of protein with unknown fold within ARCIMBOLDO scope or within sequence uncertainties.

4 REFERENCES

REFERENCES

- ADAMS, P. D. et al. PHENIX: a comprehensive Python-based system for macromolecular structure solution. *Acta Crystallographica Section D Biological Crystallography*, v. 66, n. 2, p. 213–221, 22 jan. 2010.
- ALEXANDROV, V. et al. Normal modes for predicting protein motions: a comprehensive database assessment and associated Web tool. *Protein Science: A Publication of the Protein Society*, v. 14, n. 3, p. 633–643, mar. 2005.
- ALTSCHUL, S. F. et al. Basic local alignment search tool. *Journal of Molecular Biology*, v. 215, n. 3, p. 403–410, 5 out. 1990.
- AMBROSIO, A. L. B. et al. A molecular mechanism for Lys49-phospholipase A2 activity based on ligand-induced conformational change. *The Journal of biological chemistry*, v. 280, n. 8, p. 7326–7335, 25 fev. 2005.
- ANDRIÃO-ESCARSO, S. H. et al. Myotoxic phospholipases A2 in Bothrops snake venoms: Effect of chemical modifications on the enzymatic and pharmacological properties of bothropstoxins from Bothrops jararacussu. *Biochimie*, v. 82, n. 8, p. 755–763, Agosto 2000.
- ARCIMBOLDO, G. **The Greengrocer**. Free media repository. Disponível em: <https://upload.wikimedia.org/wikipedia/commons/4/49/Arcimboldo_Vegetables.jpg>. Acesso em: 5 out. 2016.
- ARNI, R. K. et al. Structure of a calcium-independent phospholipase-like myotoxic protein from Bothrops asper venom. *Acta crystallographica. Section D, Biological crystallography*, v. 51, n. Pt 3, p. 311–317, 1 maio 1995.
- ARTOLA-RECOLONS, C. et al. Structure and cell wall cleavage by modular lytic transglycosylase MltC of Escherichia coli. *ACS chemical biology*, v. 9, n. 9, p. 2058–2066, 19 set. 2014.
- BAHNSON, B. J. Structure, function and interfacial allostereism in phospholipase A2: insight from the anion-assisted dimer. *Archives of biochemistry and biophysics*, v. 433, n. 1, p. 96–106, 1 jan. 2005.
- BATINA, M. DE F. et al. Inhibition of the lethal and myotoxic activities of Crotalus durissus terrificus venom by Tabernaemontana catharinensis: identification of one of the active components. *Planta medica*, v. 66, n. 5, p. 424–428, jun. 2000.
- BATTYE, T. G. G. et al. iMOSFLM: a new graphical interface for diffraction-image processing with MOSFLM. *Acta Crystallographica Section D Biological Crystallography*, v. 67, n. 4, p. 271–281, 18 mar. 2011.
- BERG, O. G. et al. Interfacial Enzymology: The Secreted Phospholipase A2-Paradigm. *Chemical Reviews*, v. 101, n. 9, p. 2613–2654, 1 set. 2001.
- BERNSTEIN, F. C. et al. The Protein Data Bank: a computer-based archival file for macromolecular structures. *Journal of Molecular Biology*, v. 112, n. 3, p. 535–542, 25 maio 1977.
- BORGES, R. J. **Estudos estruturais com fosfolipases A2 homólogas de veneno botrópicos em presença de íons com importância funcional**. Botucatu: Universidade Estadual Paulista, Instituto de Biociências de Botucatu, 2012.
- BORGES, R. J. et al. Functional and structural studies of a Phospholipase A2-like protein complexed to zinc ions: Insights on its myotoxicity and inhibition mechanism. *Biochimica et Biophysica Acta (BBA) - General Subjects*, v. 1861, n. 1, p. 3199–3209, jan. 2017.
- BRASIL. Ministério da Saúde. Secretaria de Vigilância em Saúde. Departamento de Vigilância Epidemiológica. In: **Doenças Infeciosas e Parasitárias: Guia de Bolso**. 7. ed. Brasília, DF: Ministério da Saúde, 2008. p. 28–31.
- BRASIL. Ministério da Saúde. Secretaria de Vigilância em Saúde. Departamento de Vigilância Epidemiológica. **Casos de acidentes por serpentes. Brasil, Grandes Regiões e Unidades Federadas 2000 a 2015**. [s.l.: s.n.]. Disponível em: <<http://portalsaude.saude.gov.br/images/pdf/2016/janeiro/20/1-Casos-Ofidismo-2000-2015.pdf>>.
- BRASIL. Ministério da Saúde. Secretaria de Vigilância em Saúde. Departamento de Vigilância Epidemiológica. **Óbitos por serpentes. Brasil, Grandes Regiões e Unidades Federadas 2000 a 2015**. [s.l.:

s.n.]. Disponível em: <<http://portalsaude.saude.gov.br/images/pdf/2016/janeiro/20/3-Obitos-Ofidismo-2000-2015.pdf>>.

BRICOGNE, G. et al. **BUSTER**. Cambridge, United Kingdom: Global Phasing Ltd., 2011.

BROOKS, B. R. et al. CHARMM: the biomolecular simulation program. **Journal of computational chemistry**, v. 30, n. 10, p. 1545–1614, 30 jul. 2009.

BULTRÓN, E.; GUTIÉRREZ, J. M.; THELESTAM, M. Effects of *Bothrops asper* (terciopelo) myotoxin III, a basic phospholipase A2, on liposomes and mouse gastrocnemius muscle. **Toxicon: official journal of the International Society on Toxinology**, v. 31, n. 2, p. 217–222, fev. 1993.

CALVETE, J. J. et al. Venoms, venomics, antivenomics. **FEBS letters**, v. 583, n. 11, p. 1736–1743, 5 jun. 2009.

CALVETE, J. J. et al. Snake venomics of the Central American rattlesnake *Crotalus simus* and the South American *Crotalus durissus* complex points to neurotoxicity as an adaptive pedomorphic trend along *Crotalus* dispersal in South America. **Journal of proteome research**, v. 9, n. 1, p. 528–544, jan. 2010.

CALVETE, J. J.; JUÁREZ, P.; SANZ, L. Snake venomics. Strategy and applications. **Journal of mass spectrometry: JMS**, v. 42, n. 11, p. 1405–1414, nov. 2007.

CARDOSO, F. F. **Estudos estruturais e funcionais da interação entre derivados do ácido cinâmico e fosfolipase A2 homóloga do veneno de *Bothrops jararacussu***. Botucatu: Universidade Estadual Paulista, Instituto de Biociências de Botucatu, 2016.

CARVALHO, C. DE. **Ação do extrato bruto da *Tabernaemontana catharinensis* e de seu alcalóide isolado sobre as atividades de fosfolipases A2 ofídicas em preparação neuromuscular de camundongos**. Botucatu: Universidade Estadual Paulista, Instituto de Biociências de Botucatu, 7 out. 2011.

CASEWELL, N. R. et al. Complex cocktails: the evolutionary novelty of venoms. **Trends in Ecology & Evolution**, v. 28, n. 4, p. 219–229, abr. 2013.

CHANG, C. C.; LEE, C. Y. ISOLATION OF NEUROTOXINS FROM THE VENOM OF *BUNGARUS MULTICINCTUS* AND THEIR MODES OF NEUROMUSCULAR BLOCKING ACTION. **Archives Internationales De Pharmacodynamie Et De Thérapie**, v. 144, p. 241–257, 1 jul. 1963.

CHIOATO, L. et al. Mapping of the structural determinants of artificial and biological membrane damaging activities of a Lys49 phospholipase A2 by scanning alanine mutagenesis. **Biochimica et biophysica acta**, v. 1768, n. 5, p. 1247–1257, maio 2007.

CHOVANCOVA, E. et al. CAVER 3.0: a tool for the analysis of transport pathways in dynamic protein structures. **PLoS computational biology**, v. 8, n. 10, p. e1002708, 2012.

CORRÊA, L. C. et al. Crystal structure of a myotoxic Asp49-phospholipase A2 with low catalytic activity: Insights into Ca²⁺-independent catalytic mechanism. **Biochimica et biophysica acta**, v. 1784, n. 4, p. 591–599, abr. 2008.

COSTA, T. R. et al. Myotoxic phospholipases A2 isolated from *Bothrops brazili* snake venom and synthetic peptides derived from their C-terminal region: Cytotoxic effect on microorganism and tumor cells. **Peptides**, v. 29, n. 10, p. 1645–1656, out. 2008.

CROWFOOT, D. et al. X-ray crystallographic investigation of the structure of penicillin. In: **Chemistry of Penicillin**. Clarke H, Johnson T, Robinson JR ed. Princeton, NJ: University Press, 1949. p. 310–367.

DAVIS, I. W. et al. MolProbity: all-atom contacts and structure validation for proteins and nucleic acids. **Nucleic acids research**, v. 35, n. Web Server issue, p. W375–383, jul. 2007.

DE AZEVEDO, W. F. et al. Crystal structure of piratoxin-I: a calcium-independent, myotoxic phospholipase A2-homologue from *Bothrops pirajai* venom. **Toxicon: Official Journal of the International Society on Toxinology**, v. 36, n. 10, p. 1395–1406, out. 1998.

DE OLIVEIRA, A. H. et al. A pH-induced dissociation of the dimeric form of a lysine 49-phospholipase A2 abolishes Ca²⁺-independent membrane damaging activity. **Biochemistry**, v. 40, n. 23, p. 6912–6920, 12 jun. 2001a.

- DE OLIVEIRA, A. H. et al. The effect of resonance energy homotransfer on the intrinsic tryptophan fluorescence emission of the bothropstoxin-I dimer. **Biochemical and biophysical research communications**, v. 284, n. 4, p. 1011–1015, 22 jun. 2001b.
- DE OLIVEIRA, A. H. C.; FERREIRA, T. L.; WARD, R. J. Reduced pH induces an inactive non-native conformation of the monomeric bothropstoxin-I (Lys49-PLA2). **Toxicon: official journal of the International Society on Toxinology**, v. 54, n. 3, p. 373–378, 1 set. 2009.
- DE OLIVEIRA, M. et al. Antagonism of myotoxic and paralyzing activities of bothropstoxin-I by suramin. **Toxicon: Official Journal of the International Society on Toxinology**, v. 42, n. 4, p. 373–379, 15 set. 2003.
- DELATORRE, P. et al. Crystal structure of Bn IV in complex with myristic acid: a Lys49 myotoxic phospholipase A₂ from Bothrops neuwiedi venom. **Biochimie**, v. 93, n. 3, p. 513–518, mar. 2011.
- DHAWAN, D. K.; CHADHA, V. D. Zinc: a promising agent in dietary chemoprevention of cancer. **The Indian Journal of Medical Research**, v. 132, p. 676–682, dez. 2010.
- DÍAZ, C. et al. The effect of myotoxins isolated from Bothrops snake venoms on multilamellar liposomes: relationship to phospholipase A₂, anticoagulant and myotoxic activities. **Biochimica et biophysica acta**, v. 1070, n. 2, p. 455–460, 9 dez. 1991.
- DÍAZ, C. et al. Cleavage of the NH₂-terminal octapeptide of Bothrops asper myotoxic lysine-49 phospholipase A₂ reduces its membrane-destabilizing effect. **Archives of biochemistry and biophysics**, v. 312, n. 2, p. 336–339, 1 ago. 1994.
- DÍAZ, C. et al. Modulation of the susceptibility of human erythrocytes to snake venom myotoxic phospholipases A(2): role of negatively charged phospholipids as potential membrane binding sites. **Archives of biochemistry and biophysics**, v. 391, n. 1, p. 56–64, 1 jul. 2001.
- DIEBEL, J. **Representing Attitude: Euler Angles, Unit Quaternions, and Rotation Vectors** Stanford, California 94301–9010, , 20 out. 2006. Disponível em: <https://www.astro.rug.nl/software/kapteyn/_downloads/attitude.pdf>
- DOLEY, R.; KINI, R. M. Protein complexes in snake venom. **Cellular and molecular life sciences: CMLS**, v. 66, n. 17, p. 2851–2871, set. 2009.
- DOS SANTOS, J. I. et al. Structural and functional studies of a bothropic myotoxin complexed to rosmarinic acid: new insights into Lys49-PLA₂ inhibition. **PloS one**, v. 6, n. 12, p. e28521, 2011a.
- DOS SANTOS, J. I. et al. Structural, functional, and bioinformatics studies reveal a new snake venom homologue phospholipase A₂ class. **Proteins**, v. 79, n. 1, p. 61–78, jan. 2011b.
- DOS SANTOS, J. I.; SOARES, A. M.; FONTES, M. R. M. Comparative structural studies on Lys49-phospholipases A(2) from Bothrops genus reveal their myotoxic site. **Journal of structural biology**, v. 167, n. 2, p. 106–116, ago. 2009.
- EALICK, S. E. Now we're cooking: new successes for shake-and-bake. **Structure (London, England: 1993)**, v. 5, n. 4, p. 469–472, 15 abr. 1997.
- EMSLEY, P. et al. Features and development of Coot. **Acta Crystallographica Section D Biological Crystallography**, v. 66, n. 4, p. 486–501, 24 mar. 2010.
- EVANS, P.; MCCOY, A. An introduction to molecular replacement. **Acta Crystallographica Section D Biological Crystallography**, v. 64, n. 1, p. 1–10, 1 jan. 2008.
- EVANS, P. R. An introduction to data reduction: space-group determination, scaling and intensity statistics. **Acta Crystallographica Section D Biological Crystallography**, v. 67, n. 4, p. 282–292, 1 abr. 2011.
- EVRARD, G. X. et al. Assessment of automatic ligand building in ARP/wARP. **Acta Crystallographica. Section D, Biological Crystallography**, v. 63, n. Pt 1, p. 108–117, jan. 2007.
- FEIGIN, L. A.; SVERGUN, D. I. **Structure Analysis by Small-Angle X-Ray and Neutron Scattering**. Softcover reprint of the original 1st ed. 1987 edition ed. [s.l.] Springer, 2013.
- FENTON, A. W. et al. Arthropod venom citrate inhibits phospholipase A₂. **Toxicon: Official Journal of the International Society on Toxinology**, v. 33, n. 6, p. 763–770, jun. 1995.

- FERNANDES, C. A. H. et al. Comparison between apo and complexed structures of bothropstoxin-I reveals the role of Lys122 and Ca(2+)-binding loop region for the catalytically inactive Lys49-PLA(2)s. **Journal of structural biology**, v. 171, n. 1, p. 31–43, jul. 2010.
- FERNANDES, C. A. H. et al. Crystallization and preliminary X-ray diffraction analysis of three myotoxic phospholipases A2 from *Bothrops brazili* venom. **Acta Crystallographica. Section F, Structural Biology and Crystallization Communications**, v. 68, n. Pt 8, p. 935–938, 1 ago. 2012.
- FERNANDES, C. A. H. et al. Structural bases for a complete myotoxic mechanism: Crystal structures of two non-catalytic phospholipases A2-like from *Bothrops brazili* venom. **Biochimica et biophysica acta**, v. 1834, n. 12, p. 2772–2781, dez. 2013.
- FERNANDES, C. A. H. et al. A structure-based proposal for a comprehensive myotoxic mechanism of phospholipase A2-like proteins from viperid snake venoms. **Biochimica Et Biophysica Acta**, 29 set. 2014.
- FERNANDES, C. A. H. et al. Structural Basis for the Inhibition of a Phospholipase A2-Like Toxin by Caffeic and Aristolochic Acids. **PLoS One**, v. 10, n. 7, p. e0133370, 2015.
- FERREIRA, S. H.; ROCHA E SILVA, M. Potentiation of bradykinin by dimercaptopropanol (bal) and other inhibitors of its destroying enzyme in plasma. **Biochemical Pharmacology**, v. 11, p. 1123–1128, dez. 1962.
- FRANCIS, B. et al. Myotoxin II from *Bothrops asper* (Terciopelo) venom is a lysine-49 phospholipase A2. **Archives of biochemistry and biophysics**, v. 284, n. 2, p. 352–359, 1 fev. 1991.
- FRANCIS, B.; SEEBART, C.; KAISER, I. I. Citrate is an endogenous inhibitor of snake venom enzymes by metal-ion chelation. **Toxicon: Official Journal of the International Society on Toxinology**, v. 30, n. 10, p. 1239–1246, out. 1992.
- FUGLEBAKK, E.; ECHAVE, J.; REUTER, N. Measuring and comparing structural fluctuation patterns in large protein datasets. **Bioinformatics (Oxford, England)**, v. 28, n. 19, p. 2431–2440, 1 out. 2012.
- GRATTAN, B. J.; FREAKE, H. C. Zinc and cancer: implications for LIV-1 in breast cancer. **Nutrients**, v. 4, n. 7, p. 648–675, jul. 2012.
- GUTIÉRREZ, J. M. et al. Snake venomomics and antivenomics: Proteomic tools in the design and control of antivenoms for the treatment of snakebite envenoming. **Journal of Proteomics, Venomics**. v. 72, n. 2, p. 165–182, 6 mar. 2009.
- GUTIÉRREZ, J. M.; THEAKSTON, R. D. G.; WARRELL, D. A. Confronting the Neglected Problem of Snake Bite Envenoming: The Need for a Global Partnership. **PLoS Medicine**, v. 3, n. 6, jun. 2006.
- HELUANY, N. F. et al. Effects induced by bothropstoxin, a component from *Bothrops jararacussu* snake venom, on mouse and chick muscle preparations. **Toxicon: official journal of the International Society on Toxinology**, v. 30, n. 10, p. 1203–1210, out. 1992.
- HENDRICKSON, W. A. Evolution of diffraction methods for solving crystal structures. **Acta Crystallographica Section A Foundations of Crystallography**, v. 69, n. 1, p. 51–59, 1 jan. 2013.
- HODGKIN, D. C. et al. The Structure of Vitamin BFormula I. An Outline of the Crystallographic Investigation of Vitamin BFormula. **Proceedings of the Royal Society A: Mathematical, Physical and Engineering Sciences**, v. 242, n. 1229, p. 228–263, 29 out. 1957.
- HUANCAHUIRE-VEGA, S. et al. Structural and functional characterization of brazilitoxins II and III (BbTX-II and -III), two myotoxins from the venom of *Bothrops brazili* snake. **Toxicon: Official Journal of the International Society on Toxinology**, v. 54, n. 6, p. 818–827, nov. 2009.
- HÜNEFELD, F. L. **Die Chemismus in der tierescher Organisation**. Leipzig: FA Brockhouse, 1840.
- JANOTA, A. et al. Improving the precision and speed of Euler angles computation from low-cost rotation sensor data. **Sensors (Basel, Switzerland)**, v. 15, n. 3, p. 7016–7039, 2015.
- JASKOLSKI, M.; DAUTER, Z.; WLODAWER, A. A brief history of macromolecular crystallography, illustrated by a family tree and its Nobel fruits. **The FEBS journal**, v. 281, n. 18, p. 3985–4009, set. 2014.
- KABSCH, W. A solution for the best rotation to relate two sets of vectors. **Acta Crystallographica Section A**, v. 32, n. 5, p. 922–923, 1 set. 1976.

- KABSCH, W. XDS. **Acta crystallographica. Section D, Biological crystallography**, v. 66, n. Pt 2, p. 125–132, fev. 2010.
- KARPLUS, P. A.; DIEDERICHS, K. Linking crystallographic model and data quality. **Science (New York, N.Y.)**, v. 336, n. 6084, p. 1030–1033, 25 maio 2012.
- KASTURIRATNE, A. et al. The global burden of snakebite: a literature analysis and modelling based on regional estimates of envenoming and deaths. **PLoS medicine**, v. 5, n. 11, p. e218, 4 nov. 2008.
- KENDREW, J. C. et al. A three-dimensional model of the myoglobin molecule obtained by x-ray analysis. **Nature**, v. 181, n. 4610, p. 662–666, 8 mar. 1958.
- KINI, R. M. Excitement ahead: structure, function and mechanism of snake venom phospholipase A2 enzymes. **Toxicon: official journal of the International Society on Toxinology**, v. 42, n. 8, p. 827–840, 15 dez. 2003.
- KLEYWEGT, G. J.; JONES, T. A. Homo Crystallographicus—Quo Vadis? **Structure**, v. 10, n. 4, p. 465–472, abr. 2002.
- KONAREV, P. V. et al. *PRIMUS*: a Windows PC-based system for small-angle scattering data analysis. **Journal of Applied Crystallography**, v. 36, n. 5, p. 1277–1282, 1 out. 2003.
- KONAREV, P. V. et al. ATLAS 2.1, a program package for small-angle scattering data analysis. **Journal of Applied Crystallography**, v. 39, n. 2, p. 277–286, 12 mar. 2006.
- KRISSINEL, E.; HENRICK, K. Inference of macromolecular assemblies from crystalline state. **Journal of molecular biology**, v. 372, n. 3, p. 774–797, 21 set. 2007.
- KRIVOV, G. G.; SHAPOVALOV, M. V.; DUNBRACK, R. L. Improved prediction of protein side-chain conformations with SCWRL4. **Proteins**, v. 77, n. 4, p. 778–795, dez. 2009.
- LEE, W. H. et al. Structural basis for low catalytic activity in Lys49 phospholipases A2—a hypothesis: the crystal structure of piratoxin II complexed to fatty acid. **Biochemistry**, v. 40, n. 1, p. 28–36, 9 jan. 2001.
- LIEBSCHNER, D. et al. Polder maps: improving OMIT maps by excluding bulk solvent. **Acta Crystallographica Section D Biological Crystallography**, n. (accepted), 2017.
- LIN, Y. et al. Docking phospholipase A2 on membranes using electrostatic potential-modulated spin relaxation magnetic resonance. **Science (New York, N.Y.)**, v. 279, n. 5358, p. 1925–1929, 20 mar. 1998.
- LITTLE, P. J. et al. Zinc and cardiovascular disease. **Nutrition (Burbank, Los Angeles County, Calif.)**, v. 26, n. 11–12, p. 1050–1057, dez. 2010.
- LIZANO, S.; LAMBEAU, G.; LAZDUNSKI, M. Cloning and cDNA sequence analysis of Lys(49) and Asp(49) basic phospholipase A(2) myotoxin isoforms from *Bothrops asper*. **The International Journal of Biochemistry & Cell Biology**, v. 33, n. 2, p. 127–132, fev. 2001.
- LOMONTE, B. et al. The phospholipase A2 homologues of snake venoms: biological activities and their possible adaptive roles. **Protein and peptide letters**, v. 16, n. 8, p. 860–876, 2009.
- LOMONTE, B. Identification of linear B-cell epitopes on myotoxin II, a Lys49 phospholipase A₂ homologue from *Bothrops asper* snake venom. **Toxicon: official journal of the International Society on Toxinology**, v. 60, n. 5, p. 782–790, out. 2012.
- LOMONTE, B.; ANGULO, Y.; CALDERÓN, L. An overview of lysine-49 phospholipase A2 myotoxins from crotalid snake venoms and their structural determinants of myotoxic action. **Toxicon: official journal of the International Society on Toxinology**, v. 42, n. 8, p. 885–901, 15 dez. 2003.
- LOMONTE, B.; GUTIÉRREZ, J. M. A new muscle damaging toxin, myotoxin II, from the venom of the snake *Bothrops asper* (terciopelo). **Toxicon: official journal of the International Society on Toxinology**, v. 27, n. 7, p. 725–733, 1989.
- LOMONTE, B.; GUTIÉRREZ, J. M. Phospholipases A2 From Viperidae Snake Venoms: How do They Induce Skeletal Muscle Damage? **Acta Chim. Slov.**, v. 58, p. 647–658, 2011.
- MAGRO, A. J. et al. Crystal structures of BnSP-7 and BnSP-6, two Lys49-phospholipases A(2): quaternary structure and inhibition mechanism insights. **Biochemical and biophysical research communications**, v. 311, n. 3, p. 713–720, 21 nov. 2003.

- MARAGANORE, J. M. et al. A new class of phospholipases A2 with lysine in place of aspartate 49. Functional consequences for calcium and substrate binding. **The Journal of biological chemistry**, v. 259, n. 22, p. 13839–13843, 25 nov. 1984.
- MARCHI-SALVADOR, D. P. et al. Crystal structure of a phospholipase A(2) homolog complexed with p-bromophenacyl bromide reveals important structural changes associated with the inhibition of myotoxic activity. **Biochimica et biophysica acta**, v. 1794, n. 11, p. 1583–1590, nov. 2009.
- MCCOY, A. J. et al. Likelihood-enhanced fast translation functions. **Acta Crystallographica. Section D, Biological Crystallography**, v. 61, n. Pt 4, p. 458–464, abr. 2005.
- MCCOY, A. J. et al. Phaser crystallographic software. **Journal of Applied Crystallography**, v. 40, n. 4, p. 658–674, 13 jul. 2007.
- MCGUFFIN, L. J.; BRYSON, K.; JONES, D. T. The PSIPRED protein structure prediction server. **Bioinformatics (Oxford, England)**, v. 16, n. 4, p. 404–405, abr. 2000.
- MCPHERSON, A. **Introduction to macromolecular crystallography**. 2nd ed ed. Hoboken, N.J: Wiley-Blackwell, 2009.
- MILLÁN, C.; SAMMITO, M.; USÓN, I. Macromolecular ab initio phasing enforcing secondary and tertiary structure. **IUCrJ**, v. 2, n. Pt 1, p. 95–105, 1 jan. 2015.
- MILLER, R. et al. On the application of the minimal principle to solve unknown structures. **Science (New York, N.Y.)**, v. 259, n. 5100, p. 1430–1433, 5 mar. 1993.
- MONTECUCCO, C.; GUTIÉRREZ, J. M.; LOMONTE, B. Cellular pathology induced by snake venom phospholipase A2 myotoxins and neurotoxins: common aspects of their mechanisms of action. **Cellular and molecular life sciences: CMLS**, v. 65, n. 18, p. 2897–2912, set. 2008.
- MORA-OBANDO, D. et al. Role of enzymatic activity in muscle damage and cytotoxicity induced by Bothrops asper Asp49 phospholipase A2 myotoxins: are there additional effector mechanisms involved? **PeerJ**, v. 2, p. e569, 2014.
- MURAKAMI, M. T. et al. Inhibition of myotoxic activity of Bothrops asper myotoxin II by the anti-trypanosomal drug suramin. **Journal of molecular biology**, v. 350, n. 3, p. 416–426, 15 jul. 2005.
- MURAKAMI, M. T. et al. Interfacial surface charge and free accessibility to the PLA2-active site-like region are essential requirements for the activity of Lys49 PLA2 homologues. **Toxicon: official journal of the International Society on Toxinology**, v. 49, n. 3, p. 378–387, 1 mar. 2007.
- MURAKAMI, M. T. et al. Biochemical and structural investigations of Bothropstoxin-II, a myotoxic Asp49 phospholipase A2 from Bothrops jararacussu venom. **Protein and peptide letters**, v. 15, n. 9, p. 1002–1008, 2008.
- MURSHUDOV, G. N. et al. REFMAC5 for the refinement of macromolecular crystal structures. **Acta Crystallographica. Section D, Biological Crystallography**, v. 67, n. Pt 4, p. 355–367, abr. 2011.
- NÚÑEZ, V. et al. Structural and functional characterization of myotoxin I, a Lys49 phospholipase A2 homologue from the venom of the snake Bothrops atrox. **Toxicon: Official Journal of the International Society on Toxinology**, v. 44, n. 1, p. 91–101, jul. 2004.
- OSMAN, D. et al. The Effectors and Sensory Sites of Formaldehyde-responsive Regulator FrmR and Metal-sensing Variant. **The Journal of Biological Chemistry**, v. 291, n. 37, p. 19502–19516, 9 set. 2016.
- OTWINOWSKI, Z.; MINOR, W. Processing of X-ray diffraction data collected in oscillation mode. In: [s.l.] Elsevier, 1997. v. 276p. 307–326.
- PAPADOPOULOS, J. S.; AGARWALA, R. COBALT: constraint-based alignment tool for multiple protein sequences. **Bioinformatics (Oxford, England)**, v. 23, n. 9, p. 1073–1079, 1 maio 2007.
- PATTERSON, A. L. A Fourier Series Method for the Determination of the Components of Interatomic Distances in Crystals. **Physical Review**, v. 46, n. 5, p. 372–376, 1 set. 1934.
- PEDERSEN, J. Z. et al. Autocatalytic acylation of phospholipase-like myotoxins. **Biochemistry**, v. 34, n. 14, p. 4670–4675, 11 abr. 1995.

- PEREIRA, P. S. et al. Chemical constituents from *Tabernaemontana catharinensis* root bark: a brief NMR review of indole alkaloids and in vitro cytotoxicity. **Química Nova**, v. 31, n. 1, p. 20–24, 2008.
- PERUTZ, M. F. et al. Structure of haemoglobin: a three-dimensional Fourier synthesis at 5.5-Å resolution, obtained by X-ray analysis. **Nature**, v. 185, n. 4711, p. 416–422, 13 fev. 1960.
- PETTERSEN, E. F. et al. UCSF Chimera—a visualization system for exploratory research and analysis. **Journal of Computational Chemistry**, v. 25, n. 13, p. 1605–1612, out. 2004.
- PONCE-SOTO, L. A. et al. Structural and functional properties of BaTX, a new Lys49 phospholipase A2 homologue isolated from the venom of the snake *Bothrops alternatus*. **Biochimica Et Biophysica Acta**, v. 1770, n. 4, p. 585–593, abr. 2007.
- PRASAD, A. S. et al. Zinc in cancer prevention. **Nutrition and Cancer**, v. 61, n. 6, p. 879–887, 2009.
- PRASAD, A. S. Discovery of human zinc deficiency: 50 years later. **Journal of trace elements in medicine and biology: organ of the Society for Minerals and Trace Elements (GMS)**, v. 26, n. 2–3, p. 66–69, jun. 2012.
- PRAŽNIKAR, J. et al. Averaged kick maps: less noise, more signal... and probably less bias. **Acta Crystallographica. Section D, Biological Crystallography**, v. 65, n. Pt 9, p. 921–931, set. 2009.
- RCSB PDB. **Graph of Yearly Growth of Total Structures**. Disponível em: <<http://www.rcsb.org/pdb/statistics/contentGrowthChart.do?content=total&seqid=100>>. Acesso em: 13 nov. 2016.
- RENETSEDER, R. et al. A comparison of the crystal structures of phospholipase A2 from bovine pancreas and *Crotalus atrox* venom. **The Journal of biological chemistry**, v. 260, n. 21, p. 11627–11634, 25 set. 1985.
- RIGDEN, D. J. et al. The structure of the D49 phospholipase A2 piratoxin III from *Bothrops pirajai* reveals unprecedented structural displacement of the calcium-binding loop: possible relationship to cooperative substrate binding. **Acta crystallographica. Section D, Biological crystallography**, v. 59, n. Pt 2, p. 255–262, fev. 2003.
- ROCHA E SILVA, M.; BERALDO, W. T.; ROSENFELD, G. Bradykinin, a hypotensive and smooth muscle stimulating factor released from plasma globulin by snake venoms and by trypsin. **The American Journal of Physiology**, v. 156, n. 2, p. 261–273, fev. 1949.
- RODRIGUES, V. M. et al. Geographic variations in the composition of myotoxins from *Bothrops neuwiedi* snake venoms: biochemical characterization and biological activity. **Comparative biochemistry and physiology. Part A, Molecular & integrative physiology**, v. 121, n. 3, p. 215–222, nov. 1998.
- RODRIGUES-SIMIONI, L. et al. No role for enzymatic activity or dantrolene-sensitive Ca²⁺ stores in the muscular effects of bothropstoxin, a Lys49 phospholipase A2 myotoxin. **Toxicon: official journal of the International Society on Toxinology**, v. 33, n. 11, p. 1479–1489, nov. 1995.
- RODRÍGUEZ, D. et al. Practical structure solution with ARCIMBOLDO. **Acta crystallographica. Section D, Biological crystallography**, v. 68, n. Pt 4, p. 336–343, abr. 2012.
- RODRÍGUEZ, D. D. et al. Crystallographic ab initio protein structure solution below atomic resolution. **Nature methods**, v. 6, n. 9, p. 651–653, set. 2009.
- ROSSMANN, M. G.; BLOW, D. M. The detection of sub-units within the crystallographic asymmetric unit. **Acta Crystallographica**, v. 15, n. 1, p. 24–31, 1 jan. 1962.
- RUSSO, E. Special Report: The birth of biotechnology. **Nature**, v. 421, n. 6921, p. 456–457, 23 jan. 2003.
- SALVADOR, G. H. M. et al. Structural and phylogenetic studies with MjTX-I reveal a multi-oligomeric toxin—a novel feature in Lys49-PLA2s protein class. **PLoS one**, v. 8, n. 4, p. e60610, 2013a.
- SALVADOR, G. H. M. et al. Structural and functional studies with mytoxin II from *Bothrops moojeni* reveal remarkable similarities and differences compared to other catalytically inactive phospholipases A₂-like. **Toxicon: official journal of the International Society on Toxinology**, v. 72, p. 52–63, set. 2013b.
- SALVADOR, G. H. M. et al. Structural and functional evidence for membrane docking and disruption sites on phospholipase A2-like proteins revealed by complexation with the inhibitor suramin. **Acta Crystallographica. Section D, Biological Crystallography**, v. 71, n. Pt 10, p. 2066–2078, out. 2015.

- SALVADOR, G. H. M. et al. Crystal structure of a phospholipase A2 from Bothrops asper venom: Insights into a new putative “myotoxic cluster”. **Biochimie**, v. 133, p. 95–102, fev. 2017.
- SAMMITO, M. et al. Exploiting tertiary structure through local folds for crystallographic phasing. **Nature methods**, v. 10, n. 11, p. 1099–1101, nov. 2013.
- SAMMITO, M. **Enforcing secondary and tertiary structure for crystallographic phasing**. Barcelona: Universitat de Barcelona, Facultat de Farmàcia, 2015.
- SAMMITO, M. et al. ARCIMBOLDO_LITE: single-workstation implementation and use. **Acta Crystallographica. Section D, Biological Crystallography**, v. 71, n. Pt 9, p. 1921–1930, 1 set. 2015.
- SÁNCHEZ, E. E.; RODRÍGUEZ-ACOSTA, A. Inhibitors of snake venoms and development of new therapeutics. **Immunopharmacology and immunotoxicology**, v. 30, n. 4, p. 647–678, 2008.
- SAUL, F. A. et al. Comparative structural studies of two natural isoforms of ammodytoxin, phospholipases A2 from Vipera ammodytes ammodytes which differ in neurotoxicity and anticoagulant activity. **Journal of Structural Biology**, v. 169, n. 3, p. 360–369, mar. 2010.
- SCHALOSKE, R. H.; DENNIS, E. A. The phospholipase A2 superfamily and its group numbering system. **Biochimica et biophysica acta**, v. 1761, n. 11, p. 1246–1259, nov. 2006.
- SCOTT, D. L. et al. Interfacial catalysis: the mechanism of phospholipase A2. **Science (New York, N.Y.)**, v. 250, n. 4987, p. 1541–1546, 14 dez. 1990a.
- SCOTT, D. L. et al. Crystal structure of bee-venom phospholipase A2 in a complex with a transition-state analogue. **Science (New York, N.Y.)**, v. 250, n. 4987, p. 1563–1566, 14 dez. 1990b.
- SEHNAL, D. et al. MOLE 2.0: advanced approach for analysis of biomacromolecular channels. **Journal of Cheminformatics**, v. 5, n. 1, p. 39, 2013.
- SHELDRIK, G. M. Experimental phasing with SHELXC/D/E: combining chain tracing with density modification. **Acta Crystallographica Section D Biological Crystallography**, v. 66, n. 4, p. 479–485, 24 mar. 2010.
- SHELDRIK, G. M. et al. Ab initio phasing. **International Tables for Crystallography**, v. F, n. ch. 16.1, p. 413–432, 2012.
- SOARES, A. M. et al. Structural and functional characterization of BnSP-7, a Lys49 myotoxic phospholipase A(2) homologue from Bothrops neuwiedi pauloensis venom. **Archives of biochemistry and biophysics**, v. 378, n. 2, p. 201–209, 15 jun. 2000.
- SOARES, A. M. et al. Dissociation of enzymatic and pharmacological properties of piratoxins-I and -III, two myotoxic phospholipases A2 from Bothrops pirajai snake venom. **Archives of biochemistry and biophysics**, v. 387, n. 2, p. 188–196, 15 mar. 2001.
- SOARES, A. M. et al. Mn(2+) ions reduce the enzymatic and pharmacological activities of bothropstoxin-I, a myotoxic Lys49 phospholipase A(2) homologue from Bothrops jararacussu snake venom. **The international journal of biochemistry & cell biology**, v. 34, n. 6, p. 668–677, jun. 2002.
- SOARES, A. M. et al. Medicinal plants with inhibitory properties against snake venoms. **Current medicinal chemistry**, v. 12, n. 22, p. 2625–2641, 2005.
- SOARES, A. M.; GIGLIO, J. R. Chemical modifications of phospholipases A2 from snake venoms: effects on catalytic and pharmacological properties. **Toxicon: official journal of the International Society on Toxinology**, v. 42, n. 8, p. 855–868, 15 dez. 2003.
- STORONI, L. C.; MCCOY, A. J.; READ, R. J. Likelihood-enhanced fast rotation functions. **Acta Crystallographica. Section D, Biological Crystallography**, v. 60, n. Pt 3, p. 432–438, mar. 2004.
- SVERGUN, D.; BARBERATO, C.; KOCH, M. H. J. *CRY SOL* – a Program to Evaluate X-ray Solution Scattering of Biological Macromolecules from Atomic Coordinates. **Journal of Applied Crystallography**, v. 28, n. 6, p. 768–773, 1 dez. 1995.
- SVERGUN, D. I. Determination of the regularization parameter in indirect-transform methods using perceptual criteria. **Journal of Applied Crystallography**, v. 25, n. 4, p. 495–503, 1 ago. 1992.

- TAYLOR, G. The phase problem. **Acta Crystallographica. Section D, Biological Crystallography**, v. 59, n. Pt 11, p. 1881–1890, nov. 2003.
- TERWILLIGER, T. C. et al. Iterative model building, structure refinement and density modification with the PHENIX AutoBuild wizard. **Acta Crystallographica. Section D, Biological Crystallography**, v. 64, n. Pt 1, p. 61–69, jan. 2008a.
- TERWILLIGER, T. C. et al. Iterative-build OMIT maps: map improvement by iterative model building and refinement without model bias. **Acta Crystallographica. Section D, Biological Crystallography**, v. 64, n. Pt 5, p. 515–524, maio 2008b.
- THOMAS, A. et al. Tertiary and quaternary conformational changes in aspartate transcarbamylase: a normal mode study. **Proteins**, v. 34, n. 1, p. 96–112, 1 jan. 1999.
- THORN, A.; SHELDRIK, G. M. Extending molecular-replacement solutions with SHELXE. **Acta crystallographica. Section D, Biological crystallography**, v. 69, n. Pt 11, p. 2251–2256, nov. 2013.
- TICLI, F. K. et al. Rosmarinic acid, a new snake venom phospholipase A2 inhibitor from *Cordia verbenacea* (Boraginaceae): antiserum action potentiation and molecular interaction. **Toxicon: official journal of the International Society on Toxinology**, v. 46, n. 3, p. 318–327, 1 set. 2005.
- TOYAMA, M. H. et al. A quick procedure for the isolation of dimeric piratoxins-I and II, two myotoxins from *Bothrops pirajai* snake venom. N-terminal sequencing. **Biochemistry and Molecular Biology International**, v. 37, n. 6, p. 1047–1055, dez. 1995.
- TOYAMA, M. H. et al. Purification and Amino Acid Sequence of MP-III 4R D49 Phospholipase A2 from *Bothrops pirajai* Snake Venom, a Toxin with Moderate PLA2 and Anticoagulant Activities and High Myotoxic Activity. **Journal of Protein Chemistry**, v. 18, n. 3, p. 371–378, 1 abr. 1999.
- UTKIN, Y. N. Animal venom studies: Current benefits and future developments. **World Journal of Biological Chemistry**, v. 6, n. 2, p. 28–33, 26 maio 2015.
- VERHEIJ, H. M. et al. Methylation of histidine-48 in pancreatic phospholipase A2. Role of histidine and calcium ion in the catalytic mechanism. **Biochemistry**, v. 19, n. 4, p. 743–750, 19 fev. 1980.
- VERONESE, E. L. G. et al. Inhibition of the myotoxic activity of *Bothrops jararacussu* venom and its two major myotoxins, BthTX-I and BthTX-II, by the aqueous extract of *Tabernaemontana catharinensis* A. DC. (Apocynaceae). **Phytomedicine: international journal of phytotherapy and phytopharmacology**, v. 12, n. 1–2, p. 123–130, jan. 2005.
- VONRHEIN, C. et al. Data processing and analysis with the autoPROC toolbox. **Acta Crystallographica. Section D, Biological Crystallography**, v. 67, n. Pt 4, p. 293–302, abr. 2011.
- WANG, B. C. Resolution of phase ambiguity in macromolecular crystallography. **Methods in Enzymology**, v. 115, p. 90–112, 1985.
- WARRELL, D. A. Snake bite. **Lancet**, v. 375, n. 9708, p. 77–88, 2 jan. 2010.
- WATANABE, L. et al. Structural insights for fatty acid binding in a Lys49-phospholipase A2: crystal structure of myotoxin II from *Bothrops moojeni* complexed with stearic acid. **Biochimie**, v. 87, n. 2, p. 161–167, fev. 2005.
- WATSON, J. D.; CRICK, F. H. Molecular structure of nucleic acids; a structure for deoxyribose nucleic acid. **Nature**, v. 171, n. 4356, p. 737–738, 25 abr. 1953.
- WINN, M. D. et al. Overview of the CCP4 suite and current developments. **Acta Crystallographica Section D Biological Crystallography**, v. 67, n. 4, p. 235–242, 18 mar. 2011.
- WORLD HEALTH ORGANIZATION. **Rabies and envenomings: a neglected public health issue**. Geneva: W.H.O., 2007.
- XIMENES, R. M. et al. Harpalycin 2 inhibits the enzymatic and platelet aggregation activities of PrTX-III, a D49 phospholipase A2 from *Bothrops pirajai* venom. **BMC complementary and alternative medicine**, v. 12, p. 139, 2012.
- XU, Q. et al. Statistical analysis of interface similarity in crystals of homologous proteins. **Journal of Molecular Biology**, v. 381, n. 2, p. 487–507, 29 ago. 2008.

XU, Q.; DUNBRACK, R. L. The protein common interface database (ProtCID)--a comprehensive database of interactions of homologous proteins in multiple crystal forms. **Nucleic Acids Research**, v. 39, n. Database issue, p. D761-770, jan. 2011.

YUNES QUARTINO, P. J.; BARRA, J. L.; FIDELIO, G. D. Cloning and functional expression of secreted phospholipases A(2) from *Bothrops diporus* (Yarará Chica). **Biochemical and Biophysical Research Communications**, v. 427, n. 2, p. 321-325, 19 out. 2012.

ZWART, P.; GROSSE-KUNSTLEVE, R.; ADAMS, P. Xtriage and Fest: automatic assessment of X-ray data and substructure structure factor estimation. **CCP4 newsletter**, v. 43, Winter 2005.