

Essential oils used in the poultry industry: Would it be an effective green alternative against *Salmonella* spp. dissemination and antimicrobial resistance?

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ABSTRACT

Due to the increasing emergence of antimicrobial-resistant strains of *Salmonella* spp. isolated from poultry, and the global trend towards reducing antimicrobial use in food-producing animals, alternatives to these drugs are being sought. Among these alternatives, the antimicrobial action of essential oils (EOs) stands out. These compounds are plant-derived compounds with antimicrobial effectiveness against Gram-positive and Gram-negative bacteria, including strains of *Salmonella*. EOs are employed in the food industry due to their aromatic and antimicrobial properties and their role as natural preservatives as well. However, studies have been approaching their weight gain and performance instead of focusing on the applicability of these compounds in poultry challenged with avian salmonellae or other bacterial diseases. Therefore, the present study aimed to review the pros and cons of using EOs against *Salmonella* spp. in poultry. Although there are reports of the antimicrobial effectiveness of EOs against *Salmonella* spp. both *in vitro* and *in vivo*, their use for this purpose has not been deeply studied. Until then, optimal concentrations for controlling *Salmonella* shedding or toxic concentrations for poultry have not been established. On the other hand, it is known that these products can exhibit synergistic effects with other antimicrobials. Thus, investigations related to the pharmacokinetics, mechanisms of action, and adverse effects of EOs in the poultry's body are important to elucidate the treatment with these alternative antimicrobials, as well as to understand their interactions with both pathogenic bacteria and the bacteria that naturally compose the poultry's microbiota.

1. Introduction

Poultry production is crucial in meeting the nutritional demands for low-cost proteins in various countries (El-Hack et al., 2022). However, the increase in production has led to crowding poultry houses, which facilitates the spread of pathogens, such as those from *Salmonella enterica* (*S. enterica*) specie, which are important from One Health's perspective (Saraiva et al., 2022). Antimicrobials with growth promoters are commonly used in poultry production to control these pathogens and optimize poultry productivity (Zhang et al., 2021a, 2021b). Nonetheless, the emergence of multi-drug-resistant bacteria and the indiscriminate use of these drugs in animal production have prompted the search for new products with similar antimicrobial effects (El-Dayem et al., 2024; Galgano et al., 2023).

Faced with a rising rate of antimicrobial resistance, alternative strategies such as probiotics, prebiotics, symbiotics, organic acids, peptides, and essential oils (EOs) are proposed as alternatives to antimicrobials in broilers (Meenu et al., 2023; Stingelin et al., 2023; Roque-Borda et al., 2022). Among these, EOs are phytochemicals extracted from different plant types and are used as herbal additives in the poultry industry due to their antioxidant, and antimicrobial especially against Gram-negative bacteria, antiviral, and antiparasitic effects (El-Hack et al., 2022; Penteado et al., 2021). These compounds are secondary metabolites extracted from different plant parts, such as flowers, leaves, stems, and roots, composed of various bioactive principles that confer their antimicrobial properties (Meenu et al., 2023).

The mechanism of action of EOs varies according to their active principles (Sousa et al., 2023). Although not fully understood, it is

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known that EOs act on the bacterial cell wall and protein denaturation, in addition to making the membrane permeable to cations such as H^+ and K^+ (Liu et al., 2018; Oliveira et al., 2023). The efficacy of the antimicrobial activity of EOs depends on their composition, and the pathogen in which they act on them as well (Nuñez et al., 2024).

Although there is scientific evidence supporting the antimicrobial activity of these oils (Meenu et al., 2023), research on their use for controlling *Salmonella* spp. infections in poultry is still limited. Most studies using EOs in poultry production focus on promoting zootechnical indices, such as weight gain and maintaining intestinal health in broilers (Huang et al., 2024; Nameghi et al., 2019). Furthermore, there is no standardized dose for the effective or toxic use of EOs for poultry phytotherapy (Hu et al., 2023; Stingelin et al., 2023).

In the poultry's body, EOs are rapidly excreted through the renal pathway after ingestion (Szczeapanik et al., 2020). Therefore, their accumulation in the organism is unlikely (Díaz-Sánchez et al., 2015). Although there are few reports of toxicity in poultry, there are reports of embryotoxic effects stimulated by exposure to cinnamaldehyde (Abramovici and Rachmuth-Raizman, 1983).

Based on these observations, we present a deep overview, bringing the pros and cons of the applicability of essential oils as a natural alternative to antimicrobials in poultry production, their potential effects against *Salmonella* genus, and their role in combating antimicrobial resistance.

2. Antimicrobial use in poultry production and antimicrobial resistance

The potential of antimicrobials as growth promoters in poultry farming was first related by Moore et al. in 1946. Since then, in addition to being used to improve zootechnical indicators, these agents have been misused for both prevention of diseases and metaphylaxis purposes (Baptiste and Pokludová, 2020). In 2010, it was estimated that 63,000 tons of antimicrobials were used in animal production. The trend suggests antimicrobial consumption could exceed 100,000 tons by 2030 (Van Boeckel et al., 2015).

The excessive use of antimicrobials in poultry farming has contributed to the selection of resistant strains, a natural evolutionary process accelerated by human intervention (Saraiva et al., 2022). This phenomenon can be attributed to prolonged exposure to these compounds, which may lead to the emergence of resistance genes in both pathogenic microorganisms and commensal microbiota (Xu et al., 2022). One of the factors that correlate poultry farming with the emergence of resistant bacteria is the use of these compounds in sub-therapeutic doses as growth promoters (Saraiva et al., 2018), among the most widely used classes are tetracyclines, penicillins, and aminoglycosides, also used to treat human infections (Rahman et al., 2022). Therefore, these antimicrobials used inappropriately eliminate susceptible microorganisms and select resistant ones, making them predominant in the environment and able to transfer their resistance genes to other bacteria, both within the same species and across distinct species (Saraiva et al., 2022).

The transfer of resistance genes can occur by vertical gene transfer (VGT) or horizontal gene transfer (HGT), through the events of transformation, transduction, and conjugation (Thomas and Nielsen, 2005). Because of these mechanisms, resistant or multi-resistant bacteria are found circulating in domestic and non-domestic species, such as seagulls, turtles, and monkeys (Laborda et al., 2022). Thus, the decline in the effectiveness of antimicrobials has led to the "off-label" use of these drugs in animal production (Caneschi et al., 2023).

Through the consumption of chicken meat and eggs, harboring resistance genes bacteria can be spread to humans (Saraiva et al., 2022), including pathogens from the *Campylobacter*, *Listeria*, and *Salmonella* genera (Chlebicz and Slizewska, 2018). Especially regarding *Salmonella* spp., reports of resistant strains of poultry origin have been described in the literature (Souza et al., 2020; Moreira et al., 2022; Benevides et al., 2024), including human outbreaks associated with the

consumption of poultry meat (CDC, 2018; ECDPC & EFSA, 2022; Gieraltowski et al., 2016).

Faced with the prospect of the emergence of resistance occurring much faster than the development of new antimicrobial molecules (Evangelista et al., 2021) and the difficulty of treating human infections caused by resistant bacteria, the current legislation restricts and rationalizes the use of these drugs in farm animals (Galgano et al., 2023; Nuñez et al., 2024; Oliveira et al., 2023). An example of this practice was the restriction on the use of antimicrobials as growth promoters in the European Union (EU, 2019), a policy that had an economic impact on the EU's main trading partners (Maron et al., 2023). Given the above, there is a clear need to look for alternatives to these drugs (Rahman et al., 2022).

Essential oils, organic acids, competitive exclusion and peptides are proposed alternatives for replacing and reducing the use of conventional antimicrobials in poultry production (Hu et al., 2023; Kolosowski et al., 2024; Roque-Borda et al., 2022). As natural alternatives, EOs exhibit antimicrobial effects against significant pathogens (Meenu et al., 2023). Abers et al. (2021) reported that the antibacterial substances derived from lemon, tea tree, and cinnamon EOs demonstrated lower efficacy in eliminating antibiotic-sensitive *S. aureus* in comparison to Methicillin-resistant *S. aureus* (MRSA). This finding suggests that the resistance mechanism of MRSA may confer susceptibility to the antibacterial activity of these EOs. However, the components of frankincense's EO were effective against antimicrobial-sensitive *Pseudomonas aeruginosa* strains and ineffective against resistant strains (Abers et al., 2021).

3. Essential oils

Essential oils are derived from various plants and contain multiple chemical substances, such as terpenes, sesquiterpenes, and aromatic components (Hoffmann, 2020; Meenu et al., 2023). It's estimated that each oil contains between 20 and 60 chemical components, but some EOs may contain more than 300 (Sousa et al., 2023). However, typically two or three chemical substances are more frequently concentrated than the others in EOs (Aziz et al., 2018), for instance: basil EO has p-allyl-anisole as a major component, concentrated at approximately 70 %, and linalool at 18 % (Nuñez et al., 2024); thyme EO, contains thymol concentrated at approximately 47 %, p-cymene at 20 %, and γ -terpinene at 9 % (Galgano et al., 2023). In general, the evaluation of the constituents of different EOs is performed using Gas Chromatography-Mass Spectrometry (GCMS) (Aziz et al., 2018).

In different countries and cultures, EOs have been employed due to their therapeutic and aromatic effects, particularly within traditional medicine practices (Hoffmann, 2020). In plants' physiology, EOs play a role in antimicrobial protection and attracting or repelling insects due to their aromatic properties (Bakkali et al., 2008). Due to this antimicrobial capability, these compounds are considered potentially effective against pathogens (Sousa et al., 2023) and have passed to be tested against bacteria from food-production animals.

Historically, the extraction of these products was first performed by Arabs during the Middle Ages using the hydrodistillation method (Bakkali et al., 2008). The extraction process determines the quality of the product and composition of EOs; improper extraction can interfere with the chemical and pharmacological activity of the respective components (Aziz et al., 2018). Currently, hydrodistillation remains the most widely used method for industrial EO extraction (Ferrentino et al., 2020), though alternative methods such as steam distillation, hydro-diffusion, solvent extraction, and microwave-assisted extraction are also employed (Aziz et al., 2018).

To optimize the efficacy of EOs and preserve their bioactive components, the pharmaceutical and food industries have adopted microencapsulation or nanoencapsulation technologies (Movahedi et al., 2024; Linh et al., 2022). The primary goal of this technique is to protect these bioactives from oxidative reactions, degradation, and

volatilization (Jia et al., 2016). Encapsulated products allow for broader release in the gastrointestinal tract due to the matrices used in encapsulation (Mo et al., 2022; Movahedi et al., 2024). In previous tests involving microencapsulated EOs for poultry, commonly used matrices included maltodextrin, lipid matrices of palm oil, hydrogenated vegetable fat, and liposomes (Moharreri et al., 2022; Meligy et al., 2023; Stingelin et al., 2023; Wang et al., 2019; Zhang et al., 2019), as well as encapsulation via the spray-drying method (Mo et al., 2022; Moharreri et al., 2022).

There are various methods for microencapsulating EOs, each with their own advantages and disadvantages depending on the desired objectives. Coacervation, spray drying, and interfacial polymerization are some of the techniques employed (Sundar & Parikh, 2023). However, methods involving thermal processes present contribute to the volatilization of the volatile constituents present in EOs (Cimino et al., 2021).

4. Antimicrobial and synergic effects of essential oils

The antibacterial effect varies with each EO and each bactericidal substance they contain. These substances are volatile constituents naturally responsible for the antimicrobial defense of plants (Abers et al., 2021), as discussed above. The use of certain EOs and their components is classified as Generally Recognized as Safe (GRAS) by the U.S. Food and Drug Administration (FDA) (Oliveira Filho et al., 2023; Silva et al., 2012).

The visualization of bacterial morphology allows for understanding the probable mechanisms of action of EOs (Sousa et al., 2023). For instance, after exposure to *Citrus medica* L. EO, *Escherichia coli* and *Staphylococcus aureus* showed holes on the cell surface and a shriveled appearance (Li et al., 2019). In contrast, exposure to mustard essential oil resulted in holes in the cell wall and deformity of the structure in *E. coli* (Turgis et al., 2009). In addition to alterations in the plasma

membrane and cell wall (Li et al., 2019), other observed effects include a decrease in bacterial ATP production (Turgis et al., 2009), changes in bacterial protein synthesis (Burt et al., 2007) and gene modulation (Frazzon et al., 2018), as illustrated in Fig. 1. Even though the bactericidal effect of EOs is damage to the bacterial cell wall, the type of damage to the cell wall can vary in each bacterium species (Turgis et al., 2009). Despite these findings, the exact mechanisms further research is needed to fully elucidate the specific effects, and their specific mechanisms, of EOs on bacterial cell structures.

One of these substances of EOs with bactericidal properties is carvacrol, a compound found in EOs obtained from various aromatic plants such as oregano (*Origanum vulgare*) and marjoram (*Origanum majorana*) (Bona et al., 2012; Silva et al., 2012). During *in vitro* experiments, carvacrol has been shown to be an effective antibacterial agent against *E. coli* and *S. enterica* serovar Typhimurium (*S. Typhimurium*) (Severino et al., 2015). Still regarding *E. coli*, it was observed that carvacrol interfered with flagellin synthesis, rendering the bacteria aflagellate, which compromises bacterial motility and diminishes its pathogenicity (Burt et al., 2007).

Thymol is a substance present in the essential oil of some plants such as thyme (Meenu et al., 2023). A phenolic compound within the monoterpene group, it is extensively utilized in the food industry for its efficacy as a natural preservative (Barbosa et al., 2020). Its antibacterial effect on *Listeria monocytogenes* and some serovars of *S. enterica* (Enteritidis, Typhimurium, Derby, Heidelberg, Livingstone, Paratyphi, and others), have been observed both *in vitro* and *in vivo* (Bona et al., 2012; Meenu et al., 2023). Besides its antibacterial effect, thymol is also used for its antioxidant properties (Barbosa et al., 2020).

Another EO compound, eugenol is included in the group of phenylpropenes, along with cinnamaldehyde, and is extracted respectively from cloves (Oliveira et al., 2020) and cinnamon (Zhang et al., 2022). Both exhibit broad antibacterial activity, being more effective against

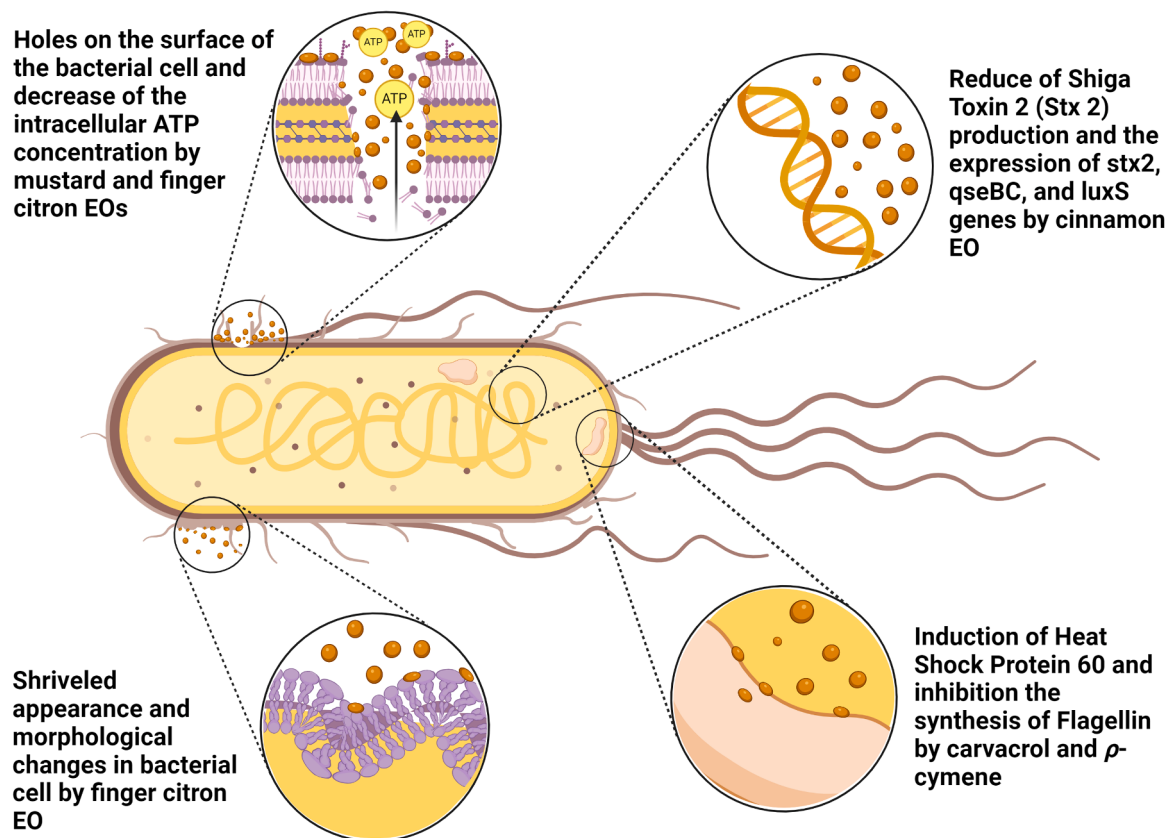


Fig. 1. Main alterations reported in bacterial cells due to contact with essential oils and their chemical components, according to Burt et al. (2007); Frazzon et al. (2018); Li et al. (2019); Turgis et al. (2009). This figure was created using BioRender.com.

Gram-negative bacteria (Meenu et al., 2023). The bactericidal mechanism of cinnamaldehyde against *E. coli* involves disruption of the cell wall and induction of oxidative stress, leading to bacterial cell death (Visvalingam et al., 2017). In addition, eugenol and cinnamaldehyde also possess antifungal activity and are commercially used as natural antimicrobial agents (Oliveira Filho et al., 2023).

EOs have the potential to exhibit synergistic effects with different products, such as antimicrobials, organic acids, or other EOs (Hu et al., 2023; Rapper et al., 2023). Combined with another antimicrobial agent, they can synergize against multidrug-resistant bacteria and significantly reduce their Minimum Inhibitory Concentration (MIC) (Meenu et al., 2023).

The synergism of some EOs has been highlighted in the effect on bacteria. Cinnamaldehyde, when combined with antimicrobials, increased the susceptibility of erythromycin-resistant *E. coli* and maximized the effect of tetracycline and novobiocin (Visvalingam et al., 2017). Organic acids combined with EOs acted synergistically against *Salmonella* Enteritidis in the study by Hu et al. (2023). The combination of tea tree with oregano EOs enhanced the antimicrobial effect of both oils against respiratory pathogens (Rapper et al., 2023). Thus, combining different EOs or other products can further enhance the antimicrobial effect of EOs (Meenu et al., 2023). Alternatively, EOs can be synergistically combined with probiotic bacteria to enhance antimicrobial activity against *Salmonella enterica* strains (Bartkiene et al., 2020).

5. Efficacy of essential oils against *Salmonella* spp. and other pathogens in poultry

Due to their safety for consumption on feed and antimicrobial activity against important poultry pathogens (El-Shall et al., 2020; Mariotti et al., 2022; Puvaca et al., 2020; Shayeganmehr et al., 2018; Zhang et al., 2022), EOs are likely candidates to replace antibiotic therapies in poultry (Puvaca et al., 2020) and preserve the antimicrobials efficiency. However, the main challenge for using EOs to replace antimicrobials in

poultry is that the concentration of active principles, when reaching the intestine, is lower than the MIC set for them (Horky et al., 2019). Although their antimicrobial action is proven, little is known about the non-toxic recommended concentrations for different oils and their mechanisms of action.

Although there are several studies on the *in vitro* antimicrobial efficacy of different essential oils against different serovars of *Salmonella* spp. (Barbosa et al., 2020; Boy et al., 2023; Porter et al., 2020; Robinson et al., 2022; Somrani et al., 2022), few have investigated the potential application of EOs in the treatment of avian salmonellosis, as we disposable de major in Table 1. Overall, cinnamon, clove, and oregano EOs are the most effective antimicrobial agents both *in vitro* and *in vivo* against salmonellosis and other avian pathogens (Barbosa et al., 2020; Ebani et al., 2018; Ebani et al., 2019; Meligy et al., 2023; Stingelin et al., 2023).

In a recent study, essential oils were combined with organic acids in the diet of broiler chickens experimentally infected with *S. Heidelberg*, *S. Minnesota*, and *S. Typhimurium* (Stingelin et al., 2023). A reduction in bacterial load was observed on the 7th and 14th days post-infection in the treated groups (Stingelin et al., 2023). A similar experiment has been carried out using a combination of organic acids with EOs, which not only reduced *S. Enteritidis*, but also facilitated enhanced weight gain among the treated broilers (Hu et al., 2023). However, not all studies were able to achieve similar results, highlighting the divergence between research. For instance, the same antimicrobial effect has not been observed against infections by *S. Enteritidis* and *C. jejuni* in experimentally infected chickens (Girard et al., 2024).

The cecum is recognized as the primary locus of bacterial proliferation during paratyphoid infections in poultry (Barrow et al., 2015). Therefore, in studies aiming to control *Salmonella* paratyphoid infections with EOs, it is crucial to ensure that the active principles of the oils reach the desired concentrations at this location. In a recent study, Stingelin et al. (2023) indicated that microencapsulation of the oils allowed for a reduction of *S. Minnesota*, *S. Heidelberg*, and *S. Typhimurium* in the cecum of broilers. Encapsulation of the oils facilitates better release of

Table 1
Antimicrobial effects of essential oils against *Salmonella* spp. *in vitro* and *in vivo*.

Essential oil	Actives substances identified	Serovar	Concentration	Used methods	Authors
Cinnamon and clove	Cinnamaldehyde, eugenol and others	<i>S. Enteritidis</i> and <i>S. Typhimurium</i>	1.26–0.63 and 2.63–0.16 mg/mL	MIC and DDST	Ebani et al. (2019)
Savory	Thymol, terpinene, p-cymene, and others	<i>S. Enteritidis</i>	0.31–0.62 µL/mL	MIC	Seyedtaghiya et al. (2021)
EOs components and organic acids	Carvacrol, thymol, cinnamaldehyde, and others	<i>S. Enteritidis</i>	800 mg/kg	Experimental infection in broilers chicks	Zhang et al. (2019)
Laurel and zahter	Carvacrol, thymol, terpinene, 1,8-cineole, terpinyl, and others	<i>S. Typhimurium</i>	2 mg/mL and 0.2 µL/mL	MIC	Yilmaz et al. (2024)
Oregano, rosemary, and EOs components	Carvacrol, 1,8-cineole	<i>S. Enteritidis</i>	1.25 µL/mL and 20 µL/mL	MIC	Cariri et al. (2019)
Thyme, savory, peppermint, black pepper	Thymol, carvacrol, p-cymene, terpinene, and others	<i>S. Enteritidis</i>	1.87 mg/mL	MIC	Moharreri et al. (2022)
Clove	Eugenol and others	<i>S. Enteritidis</i>	0.32–0.64 mg/mL	MIC and DDST	Somrani et al. (2022)
Oil mix	Carvacrol, limonene, menthol, cineol, eugenol, and others	<i>S. Typhimurium</i>	0.10–1 % v/v	BMD and MIC	Mariotti et al. (2022)
Oil mix	Eucalyptol, pinene, limonene, thymol, and others	<i>S. Infantis</i> and <i>S. Typhimurium</i>	40 µL/mL	BMD and MIC	Di Vito et al. (2020)
Flowers of Dehesa of Extremadura	Phenolic compounds	<i>S. Choleraesuis</i>	10–100 µL/mL	DDST	Boy et al. (2023)
Oregano	Thymol, carvacrol, and others	<i>S. Enteritidis</i>	130 µg/mL	MIC	Barbosa et al. (2020)
Oil mix and organic acids	Thymol, carvacrol, cinnamaldehyde, butyric acid, and others	<i>S. Enteritidis</i>	500–800 mg/kg	Experimental infection in broilers chicks	Hu et al. (2023)
Oil mix and organic acids	Thymol, carvacrol, and sorbic acid	<i>S. Heidelberg</i> , <i>S. Minnesota</i> and <i>S. Typhimurium</i>	1–2 g/kg	Experimental infection in broilers chicks	Stingelin et al. (2023)
<i>Lippia origanoides</i>	Carvacrol, thymol, diethylphenol, and others	<i>Eimeria</i> and <i>S. Typhimurium</i>	37 ppm	Experimental infection in broilers chicks	Coles et al. (2021)

Legend: Minimum Inhibitory Concentration (MIC); Minimum Bactericidal Concentration (MBC); Disk Diffusion Susceptibility Test (DDST); Broth Microdilution Susceptibility Testing (BMD); Parts Per Million (PPM).

these compounds throughout the gastrointestinal tract and increases their bioavailability, ensuring desired concentrations in the organism (Mo et al., 2022). However, the antimicrobial efficacy of the oils depends not only on the encapsulation of the product but also on the concentration of the active principle and the microorganism (Hu et al., 2023).

In a manner analogous to microencapsulation, nanoencapsulation enhances the bioavailability of phytobiotics through small capsules (Movahedi et al., 2024). Generally, capsules are made from biological materials that will be degraded at specific locations within the organism, such as the duodenum (Linh et al., 2022). Encapsulation techniques can also amplify the antimicrobial effect of EOs. In another study, lavender EO demonstrated three times its antimicrobial potency against *S. aureus* (Yuan et al., 2019).

Recommended dosages of EOs to control *Salmonella* spp. or other avian bacterial pathogens have not been established, and there is variation in suggested dosages across different studies (Bona et al., 2012; Hu et al., 2023; Stingelin et al., 2023). Products based on a mix of EOs and organic acids shown efficacy in reducing *S. Enteritidis* in the cecum using concentrations of 500 mg/kg of feed and 800 mg/kg (Hu et al., 2023), while the concentrations for reducing the excretion of *Salmonella* serovars were found to be 1 g/kg and 2 g/kg of feed (Stingelin et al., 2023). On the other hand, Bona et al. (2012), reported a compound with a minimum guarantee of 4.5 mg/kg of carvacrol, 1 g/kg of cinnamaldehyde, 2 g/kg of cineol, and 20 g of capsaicin per kg of feed was used against *S. Enteritidis* and other bacteria.

Despite the majority of EOs having no established reliable concentrations, the European Food Safety Authority (EFSA) published, in 2019, a report containing the maximum safe concentrations of oregano essential oil for production animals: 22 mg/kg of feed for broilers, 33 mg/kg for laying hens, and 30 mg/kg for turkeys (EFSA et al., 2019). However, considering the variety of EOs and their different active principles, further research is needed to investigate the effective and tolerable doses of these substances in the poultry industry. Although some oils demonstrate *in vitro* antimicrobial activity, not all are recommended for *in vivo* tests due to their minimum inhibitory concentration being higher than acceptable levels in animal production, cost-effectiveness, and palatability (Di Vito et al., 2020).

Even in the absence of a report establishing adequate concentrations of other EOs, some studies documented antimicrobial effectiveness against avian pathogens at non-toxic doses for poultry. For instance, the administration of 100 mg of tea tree EO per kg of feed resulted in a significant reduction in *Mycoplasma synoviae* infection in laying hens (Puvaca et al., 2020). Additionally, a concentration of 0.5 mL of carvacrol and thymol per liter of water reduced *C. jejuni* excretion and increased weight gain in experimentally infected broilers (El-Dayem et al., 2024). The inclusion of 37 ppm of *Lippia origanoides* EO in the diet of experimentally infected broilers resulted in reduced excretion of *S. Typhimurium*, *Eimeria maxima*, and a decrease in the occurrence of necrotic enteritis by *C. perfringens* (Coles et al., 2021).

In addition to their bactericidal effect, EOs possess virucidal properties (El-Shall et al., 2020). Against Avian Infectious Bronchitis Virus (IBV), using 1 and 2 mL of a mixture of EOs and cinnamaldehyde per liter of water resulted in a significant reduction in viral load and increased cure rate in broilers (Zhang et al., 2022). The concentration of 0.5 mL of the mixture of oils (50 g of oregano EO, 10 g of carvacrol, 33 g of thyme, 50 g of eucalyptus EO, 5 g of thymol, and 10 g of eucalyptol) added to 1 L of water reduced Newcastle Disease Virus excretion and decreased poultry mortality (El-Shall et al., 2020).

Another use of EOs in the poultry chain is in the meat industry, as antimicrobials and natural preservatives, aiming to extend the shelf life of products (Andrade et al., 2023). Alternatively, some combinations of EOs have been used to reduce *Salmonella* spp. in poultry meat products. Studies by Porter et al. (2020) and Robinson et al. (2022) evaluated the addition of different EOs to chicken meat to reduce contamination by *Salmonella* spp. Using a formulation made from white mustard essential

oil and carvacrol, a reduction of 0.7 log CFU/g of *S. Enteritidis*, *S. Typhimurium*, *S. Heidelberg*, *S. Kentucky*, and *S. Montevideo* was observed in chicken meat (Porter et al., 2020). In the combination of garlic and ginger oils, a reduction of 2 log CFU/g of *S. Infantis*, *S. Enteritidis*, and *S. Typhimurium* was also observed in chicken meat (Robinson et al., 2022).

A limitation observed in experimental studies involving essential oil treatments is the lack of description regarding the sanitary condition of the environment where the broilers were housed. After all, sanitary conditions are closely linked to the success of antimicrobial treatment (Zhai et al., 2018).

The antioxidant properties of EOs provide benefits when incorporated into meat products, particularly poultry, which is prone to oxidative deterioration due to high concentrations of polyunsaturated fatty acids (Zhai et al., 2018). It was observed that the inclusion of lemongrass and pedestrian EOs in the diet of broilers improved the oxidative stability of the thigh-drumstick meat during storage (Azevedo et al., 2021).

Although these products can be added to foods, the use of EOs in foods can impart desirable or undesirable odors and flavors depending on the type and concentration of oil used (Zhai et al., 2018). As a disadvantage, the alteration in flavor and odor can affect the product's acceptability by the consumer (Ferrentino et al., 2020), even though some reports are conflicting. For instance, although the concentrations capable of eliminating or reducing foodborne bacteria can directly affect sensory acceptability by humans (Meira et al., 2017), the use of essential oils associated with the diet of laying hens did not interfere with the nutritional and sensory quality of the eggs (Puvaca et al., 2020).

Even though there are reports of the therapeutic efficacy of EOs against Gram-negative and Gram-positive bacteria (Fig. 2), both the dysbiotic and toxic effects of these treatments in poultry are not frequently reported in some studies. Recent report showed a reduction in the population of *Bifidobacterium* spp. and *Lactobacillus* spp., alongside an increase in *Campylobacter* spp., was observed in broilers experimentally infected with *S. Enteritidis* and treated with a mixture of garlic, lemon, thyme, and eucalyptus EOs (Laptev et al., 2021). Conversely, in the study by Girard et al. (2024), the authors demonstrated that treatment with EOs in broilers experimentally infected simultaneously with *S. Enteritidis* and *C. jejuni* improved the cecal and ileal microbiota composition of the broilers. It is believed that the beneficial effect of EOs on the cecal and ileal microbiota can be attributed to the promotion of competitive exclusion (Girard et al., 2024).

6. Essential oils interactions on poultry's organism

Although the chemical composition of some oils is well known, the pharmacokinetics of these components and their action remain unclear, and when there is information, these data are based on studies conducted in mammals (Čabarkapa et al., 2020). The literature states that pharmacokinetics is related to absorption in the animal's tissues and is an important topic regarding the possibility of intoxicating the poultry and altering the sensory aspects of the meat intended for human consumption (Ocel'ová et al., 2016). Sensory changes are directly related to the potential to prevent lipid oxidation that some compounds in EOs exhibit, resulting in alterations to the food's chemical properties (Kumar et al., 2015; Ruiz-Hernández et al., 2023).

In poultry, EOs are characterized by rapid metabolism and elimination (Zhai et al., 2018). After absorption, the terpenoids that makeup EOs are eliminated by the kidneys in the form of glucuronides or exhaled as CO₂ (Szczepanik et al., 2020). The predominant method of administration is oral, typically delivered through drinking water or feed (Puvaca et al., 2020; Zhang et al., 2022). Another alternative route for the administration of EOs is through the inhalation of chemical components by adding the oil to poultry litter (Sniegocki et al., 2022), however less common. In the gastrointestinal tract, these components interact with food and are quickly digested in the small intestine along

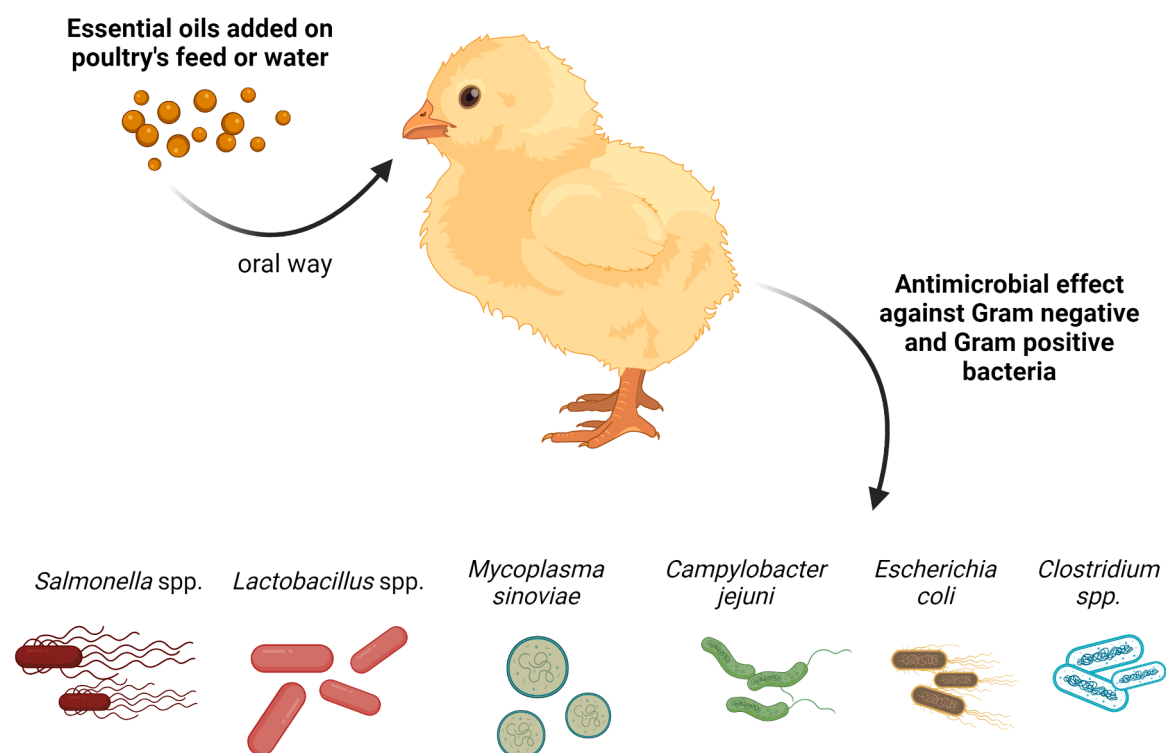


Fig. 2. Antimicrobial effect of EOs added on poultry's feed or water against Gram-positive and Gram-negative bacteria. This figure was created using BioRender.com.

with other dietary lipids and are metabolized in the liver after a first-pass effect (Horky et al., 2019), as illustrated in Fig. 3. During absorption, EOs can increase the production of digestive secretions and alter the microbial composition of the intestine, favoring symbiotic species and controlling pathogens (Attia et al., 2019; Gadde et al., 2017;

Puvaca et al., 2020). The quantification of the chemical components of EOs absorbed by the poultry can be performed using liquid chromatography-mass spectrometry from samples of liver, muscle, plasma, and lung tissue (Sniegocki et al., 2022).

Moreover, studies indicate that the use of EOs can improve both the

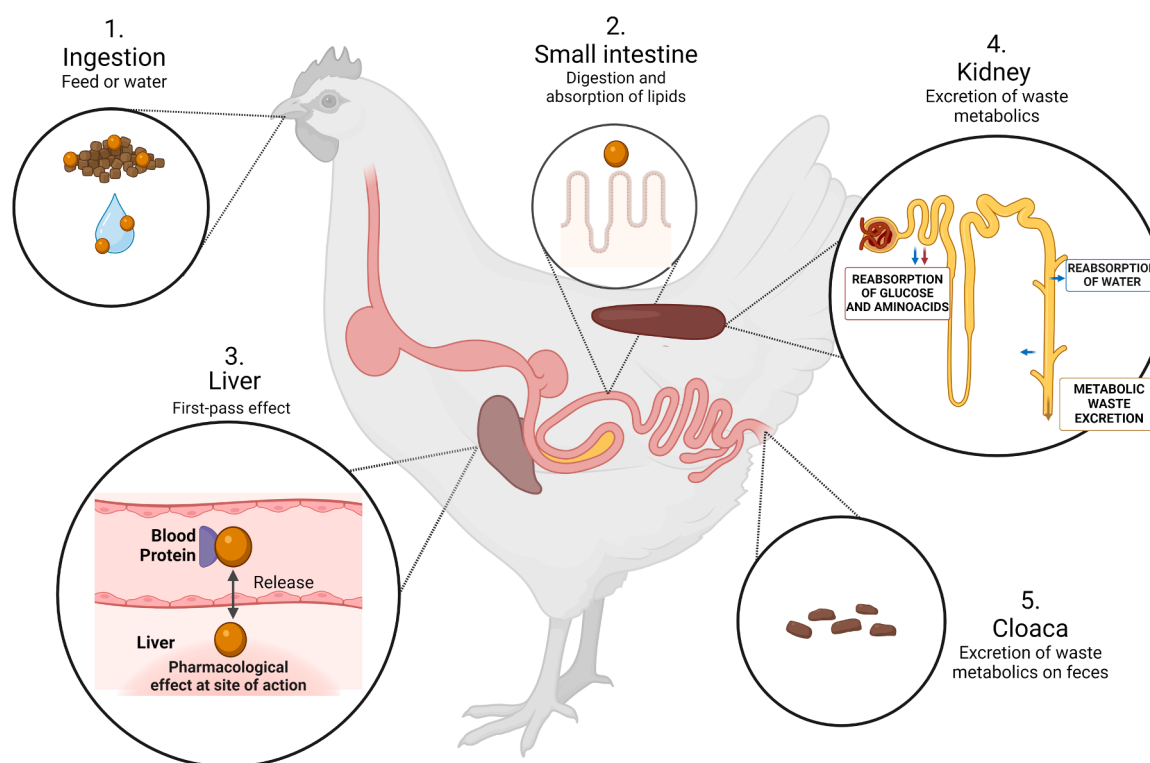


Fig. 3. Probable pharmacokinetic pathways of essential oils in the chicken organism, according to Attia et al. (2019); Gadde et al. (2017); Horky et al. (2019); Puvaca et al. (2020). This figure was created using BioRender.com.

cellular and humoral immune response (Nameghi et al., 2019). For example, an increase in serum titers of anti-Avian Infectious Bronchitis (IB) antibodies was observed in broilers treated with EOs. Similarly, the production of antibodies in vaccination programs can be enhanced in broilers treated with EO. However, in other studies, such as that by El-Shall et al. (2020), treatment with EOs during immunization against Avian Influenza (AI) and IB did not affect the poultry's immune response.

Although most research is focused on investigating the antimicrobial properties of EOs against different pathogens, issues such as toxicity and pharmacological interactions warrant further investigation (Sousa et al., 2023). Despite their rapid elimination from the body, reports have documented toxicity associated with the consumption of high concentrations of EOs, including hepatotoxicity, nephrotoxicity, and vascular changes (Oliveira et al., 2023). Additionally, the literature infrequently reports other adverse effects, such as respiratory disorders, dermal irritation, potential carcinogenicity, and reproductive toxicity (Horky et al., 2019). For instance, Beier et al. (2014) identified hepatomegaly and elevated serum aspartate aminotransferase in poultry treated with 5 % linalool, a compound found in basil oil. Adverse effects in humans, such as oral irritation and allergy, have also been linked to the phenols in clove cinnamon EOs (Triviño et al., 2019). In mice showed no mortality at high doses (200 mg/kg body weight) of oregano essential oil, with the primary effect being fecal rejection due to the high concentration of the oil (EFSA et al., 2019).

However, when compared to antimicrobials, high doses of phyto-biotics present a lower risk of poisoning at high doses (Horky et al., 2019). In addition, according to Linh et al. (2022), microencapsulation may increase the bioavailability and improve the physicochemical stability of active ingredients. In this way, the microencapsulation technique can be a solution to overcome some of the limitations of EO, such as the risk of intoxication, low water solubility, high volatility, and degradation by light (Mo et al., 2022).

7. Other uses of essential oils in the poultry industry

In addition to their use in pathogens control in poultry, EOs may perform other benefits in poultry management due to their antimicrobial properties (Galgano et al., 2023). EOs possess the capability to reduce bacterial contamination at critical points within the poultry environment, such as air, equipment, bedding, facilities, poultry, and eggs (Xu et al., 2022). These products can be used as sanitizers in poultry houses and equipment, ensuring they do not pose health risks to poultry (Oliveira et al., 2024). In the study by Ponomarenko et al. (2021), a 99 % reduction in bacterial load in the air of a poultry house was observed due to the daily use of thyme, fir, and eucalyptus EOs aerosol, without causing toxicity to the animals.

One of the uses of EOs in the poultry industry is to reduce bacterial load on the chicken warehouses. Therefore, studies investigated methods to control the bacteria both in the poultry and the poultry litter to interrupt the epidemiological cycle of paratyphoid salmonellosis (Galgano et al., 2023; Hu et al., 2023; Penha Filho et al., 2010). Experimental data showed that the application of thyme essential oil (EO) was effective in reducing bacterial contaminants within poultry litter, such as *E. coli*, *S. Derby*, and *Mammaliococcus lentus* (Galgano et al., 2023). In addition to its antibacterial effect, the addition of oregano EO and carvacrol to poultry litter can also eliminate *Alphitobius diaperinus* larvae within 24 h (Szczezanik et al., 2020).

A strategic point for optimizing production and hatchability is the sanitization of fertile eggshells (Tebrün et al., 2020). Recently, some studies have proposed EOs as a natural alternative to the sanitizers already used for this purpose (Mustafa et al., 2023; Oliveira et al., 2023). However, some components of EOs, such as thymol, can cause embryonic lesions depending on their application (Oliveira et al., 2024). In the study by Mustafa et al. (2023), lavender EO was used to sanitize fertile eggs, reducing aerobic bacteria and *Salmonella* spp. compared to other

sanitizers.

In another study, the physical quality of chicks from eggs sanitized with clove EO was compared to those sanitized with paraformaldehyde, showing that phytogenic sanitization was superior in quality and did not cause toxicity in poultry (Oliveira et al., 2020). However, some embryonic alterations, such as yolk sac retention, reduced hatchability, and embryonic death, were observed in embryos from fertile eggs sanitized with EOs (Tebrün et al., 2020). Other studies reported that carvacrol and citral, when injected into eggs, caused embryonic malformations and delayed embryonic development (Ali et al., 2019; Zhang et al., 2021a, 2021b). Therefore, the administration method and chemical composition can interfere with embryonic quality (Oliveira et al., 2023).

Phytogenic additives are also proposed as alternatives to conventional growth promoters for poultry. These are divided into four groups: herbs, botanicals, EOs, and oleoresins (El-Hack et al., 2022). The effectiveness of EO-based growth promoters is like that of antimicrobials (Islam et al., 2024). Alternative products to antibiotics are a global trend to reduce antimicrobial resistance while maintaining poultry productivity (Huang et al., 2024). Moreover, some phytogenic additives are still economically unviable for large-scale use.

Intestinal health is a critical point in broiler chicken nutrition (Huang et al., 2024). Various studies have investigated the benefits of thymol, carvacrol, and EOs on intestinal health and their impact on the avian microbiota (Islam et al., 2024; Huang et al., 2024; Nameghi et al., 2019). It is believed that phytogenic additives can enhance digestibility, nutrient absorption, and the reduction of pathogenic microorganisms (Ocel'ová et al., 2016). Liu et al. (2018) reported the effect of carvacrol dosages on the nutritional performance and intestinal health of poultry was analyzed. Groups treated with 300–400 µL of carvacrol showed increased sucrase, lactase, and other enzyme activity, as well as an increase in goblet cell content and a reduction in *Salmonella* spp. and *E. coli* (Liu et al., 2018).

In addition to improving performance, thymol and carvacrol reduced the intestinal lesion score in broiler chickens (Islam et al., 2024). The reduction of intestinal lesions by EO treatment can be attributed to their effectiveness in controlling *C. perfringens* and *Eimeria maxima* (Coles et al., 2021; Gharaibeh et al., 2021; Pham et al., 2022). The control of *C. perfringens*, *E. maxima*, and other enteric pathogens and the selection of beneficial bacteria contribute to maintaining the intestinal health of poultry and, consequently, to the reduction of enteric lesions (Nameghi et al., 2019). In addition to preventing intestinal colonization by pathogenic bacteria, EOs can be combined with certain probiotics, such as *Saccharomyces cerevisiae* (Ebani et al., 2019). However, some oils, such as thyme oil, may inhibit probiotic bacterial strains, including those from the *Lactobacillus* group. Despite this inhibition of beneficial bacteria, their metabolites can exhibit antibacterial activity against strains of *Salmonella enterica* (Bartkiene et al., 2020).

8. Final considerations and future perspectives

As natural substances with antimicrobial efficacy against various serovars of *Salmonella enterica*, EOs are proposed as alternatives to conventional antimicrobials. However, to ensure the safe use of these products *in vivo*, more research is needed to investigate effective and toxic concentration thresholds in broilers and laying hens. Some observed limitations include the scarcity of literature on the *in vivo* control of *Salmonella* spp. using EOs and the cost-benefit relationship of these products for this purpose. Therefore, future research should focus on investigating the interactions between phytobiotics, poultry, and the target bacteria to enhance antimicrobial efficacy. Zootechnical information, such as daily intake and its impact on poultry development, are critical considerations for future research involving *in vivo* experiments with phytobiotics. In the long term, regulation by official agencies regarding the use of these products as antimicrobials in poultry farms will be essential tools for rationalizing and properly managing their application.

CRediT authorship contribution statement

Kolososki Isis Mari Miyashiro: Writing – original draft, Methodology, Investigation, Conceptualization. **Benevides Valdinete P.:** Writing – review & editing, Writing – original draft, Validation, Investigation. **Rodrigues Heitor Leocádio de Souza:** Writing – original draft, Methodology, Formal analysis, Data curation, Conceptualization. **Saraiva Mauro M. S.:** Writing – review & editing, Writing – original draft, Validation, Supervision, Project administration, Methodology, Investigation, Data curation. **Berchieri Junior Angelo:** Writing – review & editing, Writing – original draft, Supervision, Project administration.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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Data availability

Data will be made available on request.

References

- Abers, M., Schroeder, S., Goelz, L., Sulser, A., Rose, T.S., Puchalski, K., Langland, J., 2021. Antimicrobial activity of volatile substances from essential oils. *BMC Complement Med. Ther.* 21 (124). <https://doi.org/10.1186/s12906-021-03285-3>.
- Abramovici, A., Rachmuth-Raizman, P., 1983. Molecular structure-teratogenicity relationships of some fragrance additives. *Toxicology* 29 (1-2), 143–156. [https://doi.org/10.1016/0300-483X\(83\)90046-X](https://doi.org/10.1016/0300-483X(83)90046-X).
- Ali, R.A., Elzahrar, D., Mostafa, F., Aboulqasem, H.E., 2019. The effect of co-treatment with retinoic acid on rescuing citral induced morphological anomalies during chick embryo development. *Assiut Univ. J. Zool.* 1 (1), 42–63.
- Andrade, B.F., Ferreira, V.R.F., Cardoso, G.P., et al., 2023. Allspice (*Pimenta dioica* Lindl.) leaves essential oils as a potential antioxidant and antimicrobial source for use in mechanically deboned poultry meat. *Braz. J. Food Technol.* 26, 2022125. <https://doi.org/10.1590/1981-6723.12522>.
- Attia, Y., Al-Harathi, M., El-Kelawy, M., 2019. Utilization of essential oils as a natural growth promoter for broiler chickens. *Ital. Journ. Anim. Sci.* 18 (1), 1005–1012. <https://doi.org/10.1080/1828051X.2019.1607574>.
- Azevedo, I.L., Nogueira, W.C.L., Almeida, A.C., Guedes, L.L.M., Vieira, C.R., Santos, S.H.S., Carvalho, C.M.C., Fonseca, F.S.A., Souza, R.M., Souza, C.N., 2021. Antioxidant activity and chemical composition of meat from broilers fed diets containing different essential oils. *Vet. World* 14 (6), 1638–1643. <https://doi.org/10.14202/vetworld.2021.1638-1643>.
- Aziz, Z.A.A., Ahmad, A., Setapar, S.H.M., Karakucuk, A., Azim, M.M., Lokhat, D., et al., 2018. Essential oils: extraction techniques, pharmaceutical and therapeutic potential – a review. *Curr. Drug Metab.* 19 (13), 1100–1110. <https://doi.org/10.2174/1389200219666180723144850>.
- Bakkali, F., Averbeck, S., Averbeck, D., Idaomar, M., 2008. Biological effects of essential oils – a review. *Food Chem. Tox* 46 (2), 446–475. <https://doi.org/10.1016/j.fct.2007.09.106>.
- Baptiste, K.E., Pokludová, L., 2020. Mass medications: prophylaxis and metaphylaxis, cascade and off-label use, treatment guidelines and antimicrobial stewardship. Springer. https://doi.org/10.1007/978-3-030-46721-0_7.
- Barbosa, L.N., Alves, F.C.B., Andrade, B.F.M.T., Albano, M., Rall, V.L.M., Fernandes, A.A.H., Buzalaf, M.A.R., Leite, A.L., Pontes, L.G., Santos, L.D., Fernandes Junior, A., 2020. Proteomic analysis and antibacterial resistance mechanisms of *Salmonella* Enteritidis submitted to the inhibitory effect of *Origanum vulgare* essential oil, thymol and carvacrol. *J. Proteom.* 214 (1). <https://doi.org/10.1016/j.jprot.2019.103625>.
- Barrow, P.A., Berchieri Junior, A., Freitas Neto, O.C., Lovell, M., 2015. The contribution of aerobic and anaerobic respiration to intestinal colonization and virulence for *Salmonella* Typhimurium in the chicken. *Avian Pathol.* 44 (5), 401–407. <https://doi.org/10.1080/03079457.2015.1062841>.
- Bartkiene, E., Ruzauskas, M., Bartkevics, V., Pugajeva, I., et al., 2020. Study of the antibiotic residues in poultry meat in some of the EU countries and selection of the best composition of lactic acid bacteria and essential oils against *Salmonella enterica*. *Poult. Sci.* 99 (8), 4065–4076. <https://doi.org/10.1016/j.psj.2020.05.002>.
- Beier, R.C., Byrd, J.A., Kubena, L.F., Hume, M.E., McReynolds, J.L., Anderson, R.C., Nisbet, D.J., 2014. Evaluation of linalool, a natural antimicrobial and insecticidal essential oil from basil: effects on poultry. *Poult. Sci.* 93 (2), 267–272. <https://doi.org/10.3382/ps.2013-03254>.
- Benevides, V.P., Saraiva, M.M.S., Nascimento, C.F., Delgado-Suaréz, E.J., Oliveira, C.J.B., Silva, S.R., Miranda, V.F.O., Christensen, H., Olsen, J.E., Berchieri Junior, A., 2024. Genomic features and phylogenetic analysis of antimicrobial-resistant *Salmonella* Mbandaka ST413 strains. *Microorganisms* 12 (2), 312. <https://doi.org/10.3390/microorganisms12020312>.
- Bona, T.D.M.M., Pickler, L., Miglino, L.B., et al., 2012. Óleo essencial de orégano, alecrim, canela e extrato de pimenta no controle de *Salmonella*, *Eimeria* e *Clostridium* em frangos de corte. *Pesq. Vet. Bras.* 32 (5). <https://doi.org/10.1590/S0100-736X2012000500009>.
- Boy, F.R., Benito, M.J., Córdoba, M.G., et al., 2023. Antimicrobial properties of essential oils obtained from autochthonous aromatics plants. *Int. J. Environ. Res. Public Health* 20 (3), 1657. <https://doi.org/10.3390/ijerph20031657>.
- Burt, S.A., Zee, R.V.D., Koets, A.P., Graaff, A.M., et al., 2007. Carvacrol induces heat shock protein 60 and inhibits synthesis of flagellin in *Escherichia coli* O157:H7. *Appl. Environ. Microb.* 73 (14), 4484–4490. <https://doi.org/10.1128/AEM.00340-07>.
- Çabarkapa, I., Puvaca, N., Popovic, S., Colovic, D., Kostadinovic, L., Tatham, E.K., Levic, J., 2020. Aromatic plants and their extracts pharmacokinetics and *in vitro/in vivo* mechanisms of action. *Feed Addit.* 75–88. <https://doi.org/10.1016/B978-0-12-814700-9.00005-4>.
- Caneschi, A., Bardhi, A., Barbarossa, A., Zaghini, A., 2023. The use of antibiotics and antimicrobial resistance in veterinary medicine, a complex phenomenon: a narrative review. *Antibiotics* 12 (3), 487. <https://doi.org/10.3390/antibiotics12030487>.
- Cariri, M.L., Melo, A.N.F., Mizzi, L., et al., 2019. Quantitative assessment of tolerance response to stress after exposure to oregano and rosemary essential oils, carvacrol and 1,8-cineole in *Salmonella* Enteritidis 86 and its isogenic deletion mutants *Adps*, *ArpoS* and *ΔompR*. *Food Res. Int.* 122 (1), 679–687. <https://doi.org/10.1016/j.foodres.2019.01.046>.
- Centers for Disease Control and Prevention (CDC). 2018. *Salmonella* Typhimurium linked to chicken salad (final update). Available in: (<https://archive.cdc.gov/www.cdc.gov/salmonella/typhimurium-02-18/index.html>).
- Chlebicz, A., Slizewska, K., 2018. Campylobacteriosis, salmonellosis, yersiniosis, and listeriosis as zoonotic foodborne diseases: a review. *Int. J. Environ. Res. Public Health* 15 (5), 863. <https://doi.org/10.3390/ijerph15050863>.
- Ciminio, C., Maurel, O.M., Musumeci, T., et al., 2021. Essential oils: pharmaceutical applications and encapsulation strategies into lipid-based delivery systems. *Pharmaceutics* 13, 327. <https://doi.org/10.3390/pharmaceutics13030327>.
- Coles, M.E., Forga, A.J., Señas-Cuesta, R., et al., 2021. Assessment of *Lippia origanoides* essential oils in a *Salmonella* Typhimurium, *Escherichia coli*, and *Clostridium perfringens* challenge model to induce necrotic enteritis in broiler chickens. *Animals* 11 (4), 1111. <https://doi.org/10.3390/ani11041111>.
- Di Vito, M., Cacaci, M., Barbanti, L., et al., 2020. *Origanum vulgare* essential oil vs. commercial mixture of essential oils: *In vitro* effectiveness on *Salmonella* spp. from poultry and swine intensive livestock. *Antibiotics* 9 (11), 763. <https://doi.org/10.3390/antibiotics9110763>.
- Díaz-Sánchez, S., D'Souza, D., Biswas, D., Hanning, I., 2015. Botanical alternatives to antibiotics for use in organic poultry production. *Poult. Sci.* 94 (6), 1419–1430. <https://doi.org/10.3382/ps/pev014>.
- Ebani, V.V., Najar, B., Bertelloni, F., Pistelli, L., Mancianti, F., Nardoni, S., 2018. Chemical composition and *in vitro* antimicrobial efficacy of sixteen essential oils against *Escherichia coli* and *Aspergillus fumigatus* isolated from poultry. *Vet. Sci.* 5 (3), 62. <https://doi.org/10.3390/vetsci5030062>.
- Ebani, V.V., Nardoni, S., Bertelloni, F., Tosi, G., Massi, P., Pistelli, L., Mancianti, F., 2019. *In vitro* antimicrobial activity of essential oils against *Salmonella enterica* serotypes Enteritidis and Typhimurium strains isolated from poultry. *Molecules* 24 (5). <https://doi.org/10.3390/molecules24050900>.
- EFSA, Bampidis, V., Azimonti, G., Bastos, M.L., et al., 2019. Safety and efficacy of an essential oil from *Origanum vulgare* ssp. *hirtum* (Link) Letsw. for all animal species. *EFSA J.* <https://doi.org/10.2903/j.efsa.2019.5909>.
- El-Dayem, G.A.A., Shalaby, M., Elkenawy, M.E., 2024. Effects of some essential oils on growth performance and *Campylobacter jejuni* in broilers. *J. Adv. Vet. Res.* 14 (3), 384–389. (<https://advetresearch.com/index.php/AVR/article/view/1589>). Available in).
- El-Hack, M.E.A., El-Saadony, M.T., Salem, H.M., et al., 2022. Alternatives to antibiotics for organic poultry production: types, modes of action and impacts on bird's health and production. *Poult. Sci.* 101. <https://doi.org/10.1016/j.psj.2022.101696>.
- El-Shall, N.A., Shewita, R.S., El-Hack, M.E.A., et al., 2020. effect of essential oils on the immune response to some viral vaccines in broiler chickens, with special reference to Newcastle disease virus. *Poult. Sci.* 99 (6), 2944–2954. <https://doi.org/10.1016/j.psj.2020.03.008>.
- European Centre for Disease Prevention and Control (ECDC) and European Food Safety Authority (EFSA). 2022. Multi-country outbreak of *Salmonella* Mbandaka ST413, possibly linked to consumption of chicken meat in the EU/EEA, Israel and the UK. Available in: (<https://efsa.onlinelibrary.wiley.com/doi/abs/10.2903/sp.efsa.2022.EN-7707>).
- European Union (EU). Regulation (EU) on the manufacture, placing on the market and use of medicated feed, amending Regulation (EC) No 1831/2003 of the European Parliament and of the Council and repealing Council Directive 90/167/EEC. Official Journal of the European Union. 2019. Available in: (<https://eur-lex.europa.eu/legal-content/EN/TXT/PDF/?uri=CELEX:32019R0004&from=EN>).
- Evangelista, A.G., Corrêa, J.A.F., Pinto, A.C.S.M., Luciano, F.B., 2021. The impact of essential oils on antibiotic use in animal production regarding antimicrobial

- resistance – a review. *Crit. Rev. Food Sci. Nutr.* 62 (1), 5267–5283. <https://doi.org/10.1080/10408398.2021.1883548>.
- Ferrentino, G., Morozova, K., Horn, C., Scampicchio, M., 2020. Extraction of essential oils from medicinal plants and their utilization as food antioxidants. *Curr. Pharm. Des.* 26 (1), 519–541. <https://doi.org/10.2174/1381612826666200121092018>.
- Frazzon, A.P.G., Saldanha, M., Lauer Junior, C., Frazzon, J., 2018. Modulation of gene expression by essential oils in bacteria. *Adv. Biot. Microb.* 8 (2), 555731. <https://doi.org/10.19080/ABM.2018.08.555731>.
- Gadde, U., Kim, W.H., Oh, S.T., et al., 2017. Alternatives to antibiotics for maximizing growth performance and feed efficiency in poultry: a review. *Anim. Health Res. Rev.* 18 (1), 26–45. <https://doi.org/10.1017/s1466252316000207>.
- Galgano, M., Pellegrini, F., Fracchiolla, G., et al., 2023. Pilot study on the action of *Thymus vulgaris* essential oil in treating the most common bacterial contaminants and *Salmonella enterica* subsp. *enterica* serovar Derby in poultry litter. *Antibiotics* 12 (3), 436. <https://doi.org/10.3390/antibiotics12030436>.
- Gharabeh, M.H., Khalifeh, M.S., Nawasreh, A.N., et al., 2021. Assessment of immune response and efficacy of essential oils applied on controlling necrotic enteritis induced by *Clostridium perfringens* in broiler chickens. *Molecules* 26 (15), 4527. <https://doi.org/10.3390/molecules26154527>.
- Gieraltowski, L., Peralta, J.H.V., Green, A., Schwensohn, C., Rosen, H., et al., 2016. National outbreak of multidrug resistant *Salmonella* Heidelberg infections linked to a single poultry company. *PLOS One*. <https://doi.org/10.1371/journal.pone.0162369>.
- Girard, C., Chabrilat, T., Kerros, S., Fravallo, P., Thibodeau, A., 2024. Essential oils mix effect on chicks ileal and caecal microbiota modulation: a metagenomics sequencing approach. *Front. Vet. Sci.* 11 (4). <https://doi.org/10.3389/fvets.2024.1350151>.
- Hoffmann, K.H., 2020. Essential oils. *Z. Naturforsch.* 75 (1), 7–8. <https://doi.org/10.1515/znc-2020-0124>.
- Horky, P., Skalickova, S., Smerkova, K., Skladanka, J., 2019. Essential oils as a feed additives: pharmacokinetics and potential toxicity in monogastric animals. *Animals* 9 (6), 352. <https://doi.org/10.3390/ani9060352>.
- Hu, Z., Liu, L., Guo, F., et al., 2023. Dietary supplemental coated essential oils and organic acids mixture improves growth performance and gut health along with reduces *Salmonella* load of broiler chickens infected with *Salmonella* Enteritidis. *J. Anim. Sci. Biotechnol.* 14 (95). <https://doi.org/10.1186/s40104-023-00889-2>.
- Huang, J., Guo, F., Abbas, W., 2024. Effects of microencapsulated essential oils and organic acids preparations on growth performance, slaughter characteristics, nutrient digestibility, and intestinal microenvironment of broiler chickens. *Poult. Sci.* 100 (5), 103655. <https://doi.org/10.1016/j.psj.2024.103655>.
- Islam, Z., Sultan, A., Khan, S., et al., 2024. Effects of an organic acids blend and coated essential oils on broiler growth performance, blood biochemical profile, gut health, and nutrient digestibility. *Ital. J. Anim. Sci.* 23 (1). <https://doi.org/10.1080/1828051X.2023.2297562>.
- Jia, Z., Dumont, M.J., Orsat, V., 2016. Encapsulation of phenolic compounds present in plants using protein matrices. *Food Biosci.* 15 (1), 87–104. <https://doi.org/10.1016/j.fbio.2016.05.007>.
- Kolososki, I.M.M., Saraiva, M.M.S., Nascimento, C.F., Campos, I.C., Lima, T.S., et al., 2024. Competitive exclusion products as an antimicrobial alternative to control *Salmonella* Heidelberg in broilers. *L. Appl. Microb.* 77 (8). <https://doi.org/10.1093/lambio/ovae071>.
- Kumar, Y., Yadav, D.N., Ahmad, T., Narsaiah, K., 2015. Recent trends in the use of natural antioxidants for meat and meat products. *Comp. Rev. F. Sci. Saf.* 14 (1). <https://doi.org/10.1111/1541-4337.12156>.
- Laborda, P., Garcia, F.S., Ochoa-Sánchez, L., et al., 2022. Wildlife and antibiotic resistance. *Front. Vet. Sci.* 12. <https://doi.org/10.3389/fvets.2022.873989>.
- Laptev, G.Y., Yildirim, E.A., Ilina, L.A., Filippova, V.A., Kochish, I.I., Gorfunkel, E.P., et al., 2021. Effects of essential oils-based supplement and *Salmonella* infection on gene expression, blood parameters, cecal microbiome, and egg production in laying hens. *Animals* 11 (2), 360. <https://doi.org/10.3390/ani11020360>.
- Li, Z.H., Cai, M., Liu, Y.S., Sun, P.L., Luo, S.L., 2019. Antibacterial activity and mechanisms of essential oil from *Citrus medica* L. var. *sarcodactylis*. *Molecules* 24 (8), 1577. <https://doi.org/10.3390/molecules24081577>.
- Linh, N.T., Qui, N.H., Triatmojo, A., 2022. The effect of nano-encapsulated herbal essential oils on poultry's health. *Arch. Razi Inst.* 77 (6), 2013–2021. <https://doi.org/10.22092/ari.2022.358842.2318>.
- Liu, S., Song, M., Yun, W., Lee, J., Lee, C., Kwak, W., Han, N., Kim, H., Cho, J., 2018. Effects of oral administration of different dosages of carvacrol essential oils on intestinal barrier function in broilers. *J. Anim. Physiol. Anim. Nutr.* 102 (5), 1257–1265. <https://doi.org/10.1111/jpn.12944>.
- Mariotti, M., Lombardini, G., Rizzo, S., et al., 2022. Potential applications of essential oils for environmental sanitation and antimicrobial treatment of intensive livestock infections. *Microorganisms* 10 (4), 822. <https://doi.org/10.3390/microorganisms10040822>.
- Maron, D.F., Smith, T.J.S., Nachman, K.E., 2023. Restrictions on antimicrobial use in food animal production: an international regulatory and economic survey. *Glob. Health* 9, 48. <https://doi.org/10.1186/1744-8603-9-48>.
- Meenu, M., Padhan, B., Patel, M., et al., 2023. Antibacterial activity of essential oils from different parts of plants against *Salmonella* and *Listeria* spp. *Food Chem.* 404. <https://doi.org/10.1016/j.foodchem.2022.134723>.
- Meira, N.V.B., Holley, R.A., Bordin, K., Macedo, R.E.F., Luciano, F.B., 2017. Combination of essential oil compounds and phenolic acids against *Escherichia coli* O157:H7 in vitro and in dry-fermented sausage production. *Int. J. Food Microbiol.* 2 (260), 59–64. <https://doi.org/10.1016/j.ijfoodmicro.2017.08.010>.
- Meligy, A.M.A., El-Hamid, M.I.A., Yones, A.E., et al., 2023. Liposomal encapsulated oregano, cinnamon, and clove oils enhanced the performance, bacterial metabolites antioxidant potential, and intestinal microbiota of broiler chickens. *Poult. Sci.* 102 (6). <https://doi.org/10.1016/j.psj.2023.102683>.
- Mo, K., Li, J., Liu, F., et al., 2022. Superiority of microencapsulated essential oils compared with common essential oils and antibiotics: effects on the intestinal health and gut microbiota of weaning piglet. *Front. Nutr.* 8 (12). <https://doi.org/10.3389/fnut.2021.808106>.
- Moharri, M., Vakili, R., Oskoueian, E., Rajabzadeh, G.H., 2022. Evaluation of microencapsulated essential oils in broilers challenged with *Salmonella* Enteritidis: a focus on the Body's antioxidant status, gut microbiology, and morphology. *Arch. Raz. Inst.* 77 (2), 629–639. <https://doi.org/10.1001/1.03653439.2022.77.2.16.7>.
- Moreira, J.P.F.F., Monte, D.F.M., Lima, C.A., Oliveira, C.J.B., Martins, N.R.S., Berchieri Junior, A., Freitas Neto, O.C., 2022. Molecular genotyping reveals inter-regional relatedness among antimicrobial resistant *Salmonella* Minnesota strains isolated from poultry farm and humans, Brazil. *Braz. J. Microbiol.* 53 (1), 503–508. <https://doi.org/10.1007/s42770-021-00666-1>.
- Movahedi, F., Nirmal, N., Wang, P., et al., 2024. Recent advances in essential oils and their nanoformulations for poultry feed. *J. Anim. Sci. Biotechnol.* 15, 110. <https://doi.org/10.1186/s40104-024-01067-8>.
- Mustafa, A.A., Mirza, R.A., Aziz, H.I., 2023. Lavender essential oil in sanitization on fertile egg. *Passer J. Basic Appl. Sci.* 5 (2), 377–381. <https://doi.org/10.24271/psr.2023.403768.1340>.
- Nameghi, A.H., Edalatian, O., Bakhshalinejad, R., 2019. Effects of a blend of thyme, peppermint and eucalyptus essential oils on growth performance, serum lipid and hepatic enzyme indices, immune response and ileal morphology and microflora in broilers. *J. Anim. Physiol. Anim. Nutr.* 103 (5), 1388–1398. <https://doi.org/10.1111/jpn.13122>.
- Núñez, K.V.M., Abreu, A.C.S., Almeida, J.M., et al., 2024. Antimicrobial activity of selected essential oils against *Staphylococcus aureus* from bovine mastitis. *Dairy* 5 (1), 54–65. <https://doi.org/10.3390/dairy5010005>.
- Ocel'ová, V., Chizzola, R., Pisarciková, J., Novák, J., Ivanisina, O., Faix, S., 2016. Plachá, I. Effect of thyme essential oil supplementation on thymol content in blood plasma, liver, kidney and muscle in broilers chickens. *Nat. Prod. Com.* 11 (10), 1545–1550.
- Oliveira Filho, J.G.O., Duarte, L.G.R., Silva, Y.B.B., et al., 2023. Novel approach for improving papaya fruit storage with carnauba wax nanoemulsion in combination with *Syzygium aromaticum* and *Mentha spicata* Essential Oils. *Coatings*. <https://doi.org/10.3390/coatings13050847>.
- Oliveira, G.S., McManus, C., Vale, I.R.R., et al., 2024. Obtaining microbiologically safe hatching eggs from hatcheries: using essential oils integrated sanitization strategies in hatching eggs, poultry houses and poultry. *Pathogens* 13 (3), 260. <https://doi.org/10.3390/pathogens13030260>.
- Oliveira, G.S., Nascimento, S.T., Santos, V.M., et al., 2020. Clove essential oil in the sanitation of fertile eggs. *Poult. Sci.* 99 (11), 5509–5516. <https://doi.org/10.1016/j.psj.2020.07.014>.
- Oliveira, G.S., McManus, C., Araújo, M.V., Sousa, D.E.R., Macêdo, I.L., Castro, M.B., Santos, V.M., 2023. Sanitizing hatching eggs with essential oils: avian and microbiological safety. *Microorg* 11 (8), 1890. <https://doi.org/10.3390/microorganisms11081890>.
- Penha Filho, R.A.C., Paiva, J.B., Silva, M.D., Almeida, A.M., Berchieri Jr, 2010. A. Control of *Salmonella* Enteritidis and *Salmonella* Gallinarum in birds by using live vaccine candidate containing attenuated *Salmonella* Gallinarum mutant strain. *Vaccine* 28 (16), 2853–2859. <https://doi.org/10.1016/j.vaccine.2010.01.058>.
- Penteado, A.L., Eschionato, R.A., Souza, D.R.C., et al., 2021. Avaliação in vitro de atividade antimicrobiana de óleos essenciais contra *Salmonella* Typhimurium e *Staphylococcus aureus*. *Rev. Hig. Aliment.* 35 (293). <https://doi.org/10.37585/HA2021.02avaliacao>.
- Pham, V.H., Abbas, W., Huang, J., et al., 2022. Effect of blending encapsulated essential oils and organic acids as an antibiotic growth promoter alternative on growth performance and intestinal health in broilers with necrotic enteritis. *Poult. Sci.* 101 (1), 101563. <https://doi.org/10.1016/j.psj.2021.101563>.
- Ponomarenko, G.V., Kovalenko, V.L., Balatskiy, Y.O., et al., 2021. Bactericidal efficiency of preparation based on essential oils used in aerosol disinfection in the presence of poultry. *Regul. Mech. Biosyst.* 12 (4), 635–641. <https://doi.org/10.15421/022187>.
- Porter, J.A., Morey, A., Manu, E.A., 2020. Antimicrobial efficacy of white mustard essential oil and carvacrol against *Salmonella* in refrigerated ground chicken. *Poult. Sci.* 99 (10), 5091–5095. <https://doi.org/10.1016/j.psj.2020.06.027>.
- Puvaca, N., Lika, E., Tufarelli, V., et al., 2020. Influence of different tetracycline antimicrobial therapy of *Mycoplasma* (*Mycoplasma synoviae*) in laying hens compared to Tea Tree essential oil on table egg quality and antibiotics residues. *Foods* 9 (5), 612. <https://doi.org/10.3390/foods9050612>.
- Rahman, M.R.T., Fliss, I., Biron, E., 2022. Insights in the development and uses of alternatives to antibiotic growth promoters in poultry and swine production. *Antibiotics* 11 (6), 766. <https://doi.org/10.3390/antibiotics11060766>.
- Rapper, S.L., Viljoen, A., Vuuren, S.V., 2023. Optimizing the antimicrobial synergism of *Melaleuca alternifolia* (Tea Tree) essential oil combinations for application against respiratory related pathogens. *Planta Med* 89 (4), 454–463. <https://doi.org/10.1055/a-1947-5680>.
- Robinson, K., Assumpcao, A.L.F.V., Arsi, K., Donoghue, A., Jesudhasan, P.R.R., 2022. Ability of garlic and ginger oil to reduce *Salmonella* in post-harvest poultry. *Animals* 12 (21), 2974. <https://doi.org/10.3390/ani12212974>.
- Roque-Borda, C.A., Saraiva, M.M.S., Monte, D.F.M., Rodrigues-Alves, L.B., Almeida, A.M., et al., 2022. HPMCAS-Coated alginate microparticles loaded with Ctx(Ile²¹)-Ha a promising antimicrobial agent against *Salmonella* Enteritidis in a chicken infection model. *ACS Infect. Dis.* 8 (1), 472–481. <https://doi.org/10.1021/acscinfecdis.1c00264>.
- Ruiz-Hernández, K., Sosa-Morales, M.E., Cerón-García, A., Gómez-Salazar, J.A., 2023. Physical, Chemical and sensory changes in meat and meat products induced by the

- addition of essential oils: a concise review. *Food Rev. Int.* 39 (4), 2027–2056. <https://doi.org/10.1080/87559129.2021.1939369>.
- Saraiva, M.M.S., Lim, K., Monte, D.F.M., et al., 2022. Antimicrobial resistance in the globalized food chain: a *One Health* perspective applied to the poultry industry. *Braz. J. Microbiol.* 53 (1), 465–486. <https://doi.org/10.1007/s42770-021-00635-8>.
- Saraiva, M.M.S., Moreira Filho, A.L.B., Freitas Neto, O.C., Silva, N.M.V., Givisiez, P.E.N., Gebreyes, W.A., Oliveira, C.J.B., 2018. Off-label use of ceftiofur in one-day chicks triggers a short-term increase of ESBL-producing *E. coli* in the gut. *PLOS ONE*. <https://doi.org/10.1371/journal.pone.0203158>.
- Severino, R., Ferrari, G., Vu, K.D., et al., 2015. Antimicrobial effects of modified chitosan based coating containing nanoemulsion of essential oils, modified atmosphere packaging and gamma irradiation against *Escherichia coli* O157:H7 and *Salmonella* Typhimurium on green beans. *Food Control* 215–222. <https://doi.org/10.1016/j.foodcont.2014.08.029>.
- Seyedtaghiya, M.H., Fasaie, B.N., Peighambari, S.M., 2021. Antimicrobial and antibiofilm effects of *Satureja hortensis* essential oil against *Escherichia coli* and *Salmonella* isolated from poultry. *Iran. J. Microbiol* 13 (1), 74–80. <https://doi.org/10.18502/ijm.v13i1.5495>.
- Shayegannmehr, A., Marandi, M.V., Karimi, V., Barin, A., Ghalyanchilangeroudi, A., 2018. *Zataria multiflora* essential oil reduces replication rate of avian influenza virus (H9N2 subtype) in challenged broiler chicks. *Immun. Heal. Dis.* 59 (4), 389–395. <https://doi.org/10.1080/00071668.2018.1478064>.
- Silva, F.V., Guimarães, A.G., Silva, E.R.S., et al., 2012. Anti-inflammatory and anti-ulcer activities of carvacrol, a monoterpene present in the essential oil of oregano. *J. Med. Food* 15 (11). <https://doi.org/10.1089/jmf.2012.0102>.
- Sniegocki, T., Raszkowska-Kaczor, A., Bajer, K., Sell, B., Kozdrun, W., Giergiel, M., Posnyak, A., 2022. A preliminary study of the poultry body weight effect of carvacrol in litter and of carvacrol residue in organ tissue of exposed chickens. *J. Vet. Res.* 66 (4), 613–617. <https://doi.org/10.2478/jvetres-2022-0054>.
- Somrani, M., Debbabi, H., Palop, A., 2022. Antibacterial and antibiofilm activity of essential oil of clove against *Listeria monocytogenes* and *Salmonella* Enteritidis. *Food Sci. Technol. Int.* 28 (4), 331–339. <https://doi.org/10.1177/10820132211013273>.
- Sousa, D.P., Damasceno, R.O.S., Amorati, R., Elshabrawy, H.Á., Castro, R.D., et al., 2023. Essential oils: chemistry and pharmacological activities. *Biom* 13 (7), 1144. <https://doi.org/10.3390/biom13071144>.
- Souza, A.I.S., Saraiva, M.M.S., Casas, M.R.T., Oliveira, G.M., Cardozo, M.V., Benevides, V.P., Barbosa, F.O., Freitas Neto, O.C., Almeida, A.M., Berchieri Junior, A., 2020. High occurrence of β -lactamase-producing *Salmonella* Heidelberg from poultry origin. *PLOS ONE*. <https://doi.org/10.1371/journal.pone.0230676>.
- Stingelin, G.M., Scherer, R.S., Machado, A.C., et al., 2023. The use of thymol, carvacrol and sorbic acid in microencapsules to control *Salmonella* Heidelberg, *S. Minnesota* and *S. Typhimurium* in broilers. *Front. Vet. Sci.* <https://doi.org/10.3389/fvets.2022.1046395>.
- Sundar, S.K., Parikh, J.K., 2023. Advances and trends in encapsulation of essential oils. *Int. J. Pharm.* 635 (25). <https://doi.org/10.1016/j.ijpharm.2023.122668>.
- Szczepanik, M., Raszkowska-Kaczor, A., Olkiewicz, D., Bajer, D., Bajer, K., 2020. Use of starch granules enriched with carvacrol for the lesser mealworm, *Alphitobius diaperinus* control in chicken house: effects on insects and poultry. *J. Poult. Sci.* 57 (2), 168–174. <https://doi.org/10.2141/jpsa.0190068>.
- Tebrün, W., Motola, G., Hafez, M.H., Bachmeier, J., Schmidt, V., Renfert, K., Reichelt, C., Bruggemann-Schwarze, S., Pees, M., 2020. Preliminary study: health and performance assessment in broiler chicks following application of six different hatching egg disinfection protocols. *Plos One*. <https://doi.org/10.1371/journal.pone.0232825>.
- Thomas, C.M., Nielsen, K.M., 2005. Mechanisms of, and barriers to, horizontal gene transfer between bacteria. *Nat. Rev. Microbiol.* 3 (1), 711–721. <https://doi.org/10.1038/nrmicro1234>.
- Triviño, F.J.N., Cuenca-Barrales, C., Ruiz-Villaverde, R., 2019. Eugenol allergy mimicking recurrent aphthous stomatitis and burning mouth syndrome. *Contact Dermat.* 81 (6), 462–463. <https://doi.org/10.1111/cod.13365>.
- Turgis, M., Han, J., Caillet, S., Lacroix, M., 2009. Antimicrobial activity of mustard essential oil against *Escherichia coli* O157:H7 and *Salmonella typhi*. *Food Control* 20 (12), 1073–1079. <https://doi.org/10.1016/j.foodcont.2009.02.001>.
- Van Boeckel, T.P., Brower, C., Gilberto, C., et al., 2015. Global trends in antimicrobial use in food animals. *PNAS* 112 (18), 5649–5654. <https://doi.org/10.1073/pnas.1503141112>.
- Visvalingam, J., Palaniappan, K., Holley, R.A., 2017. In vitro enhancement of antibiotic susceptibility of drug-resistant *Escherichia coli* by cinnamaldehyde. *Food Control* 79 (1), 288–291. <https://doi.org/10.1016/j.foodcont.2017.04.011>.
- Wang, H., Liang, S., Li, X., Yang, X., et al., 2019. Effects of encapsulated essential oils and organic acids on laying performance, egg quality, intestinal morphology, barrier function, and microflora count of hens during the early laying period. *Poult. Sci.* 98 (12), 6751–6760. <https://doi.org/10.3382/ps/pez391>.
- Xu, X., Zhou, W., Xie, C., et al., 2022. Airborne bacterial communities in the poultry farm and their relevance with environmental factors and antibiotics resistance genes. *Sci. Total Environ.* 846, 157420. <https://doi.org/10.1016/j.scitotenv.2022.157420>.
- Yilmaz, E.A., Yalçın, H., Polat, Z., 2024. Antimicrobial effects of laurel extract, laurel essential oil, zahter extract, and zahter essential oil on chicken wings contaminated with *Salmonella* Typhimurium. *Vet. Med. Sci.* 10, e1445. <https://doi.org/10.1002/vms3.1445>.
- Yuan, C., Wang, Y., Liu, Y., Cui, B., 2019. Physicochemical characterization and antibacterial activity assessment of lavender essential oil encapsulated in hydroxypropyl-beta-cyclodextrin. *Ind. Crops Prod.* 130 (1), 104–110. <https://doi.org/10.1016/j.indcrop.2018.12.067>.
- Zhai, H., Liu, H., Wang, S., et al., 2018. Potential of essential oils for poultry and pigs. *Anim. Nutr.* 4 (2), 179–186. <https://doi.org/10.1016/j.aninu.2018.01.005>.
- Zhang, L.Y., Peng, Q.Y., Liu, Y.R., et al., 2021. Effects of oregano essential oil as an antibiotic growth promoter alternative on growth performance, antioxidant status, and intestinal health of broilers. *Poult. Sci.* 100. <https://doi.org/10.1016/j.psj.2021.101163>.
- Zhang, S., Shen, Y.R., Wu, S., Xiao, Y.Q., et al., 2019. The dietary combination of essential oils and organic acids reduces *Salmonella* Enteritidis in challenged chicks. *Poult. Sci.* 98 (12), 6349–6355. <https://doi.org/10.3382/ps/pez457>.
- Zhang, X., Peng, Y., Wu, C., 2021. Chicken embryonic toxicity and potential in vitro estrogenic and mutagenic activity of carvacrol and thymol in low dose/concentration. *Food Chem. Toxicol.* 150 (1), 112038. <https://doi.org/10.1016/j.fct.2021.112038>.
- Zhang, Y., Li, X.Y., Zhang, B.S., et al., 2022. In vivo antiviral effect of plant essential oils against avian infectious bronchitis virus. *BMC Vet. Res.* 18, 90. <https://doi.org/10.1186/s12917-022-03183-x>.