

# RESSALVA

Atendendo solicitação do(a) autor(a), o texto completo desta tese será disponibilizado somente a partir de 09/04/2026.

UNIVERSIDADE ESTADUAL PAULISTA  
FACULDADE DE MEDICINA VETERINÁRIA E ZOOTECNIA

**ASSINATURAS PROTEICAS DAS GLÂNDULAS ANEXAS E  
TESTÍCULOS DE RATOS OBESOS E DIABÉTICOS**

MARCOS GOMIDES CARVALHO

BOTUCATU, SÃO PAULO  
MARÇO – 2024

UNIVERSIDADE ESTADUAL PAULISTA  
FACULDADE DE MEDICINA VETERINÁRIA E ZOOTECNIA

**ASSINATURAS PROTEICAS DAS GLÂNDULAS ANEXAS E  
TESTÍCULOS DE RATOS OBESOS E DIABÉTICOS**

Tese apresentada junto ao Programa de Pós-  
graduação em Biotecnologia Animal para  
obtenção do título de DOUTOR.  
Marcos Gomides Carvalho

Orientadora: Profa. Dra. Fabiana Ferreira de  
Souza  
Coorientador: Prof. Dr. João Luiz Amaro

BOTUCATU, SÃO PAULO  
MARÇO – 2024

FICHA CATALOGRÁFICA ELABORADA PELA SEÇÃO TÊC. AQUIS. TRATAMENTO DA INFORM.  
DIVISÃO TÉCNICA DE BIBLIOTECA E DOCUMENTAÇÃO - CAMPUS DE BOTUCATU - UNESP  
BIBLIOTECÁRIA RESPONSÁVEL: MARIA CAROLINA A. CRUZ E SANTOS-CRB 8/10188

Carvalho, Marcos Gomides.

Assinaturas proteicas das glândulas anexas e testículos  
de ratos obesos e diabéticos / Marcos Gomides Carvalho. -  
Botucatu, 2024

Tese (doutorado) - Universidade Estadual Paulista "Júlio  
de Mesquita Filho", Faculdade de Medicina Veterinária e  
Zootecnia

Orientador: Fabiana Ferreira de Sousa

Coorientador: João Luis Amaro

Capes: 50504002

1. Pesquisa médica translacional - Medicina veterinária.  
2. Fertilidade. 3. Próstata. 4. Síndrome metabólica.  
5. Vesícula seminal.

Palavras-chave: Estudo translacional; Fertilidade;  
Próstata; Síndrome metabólica; Vesícula seminal.

Nome do autor: Marcos Gomides Carvalho

Título: ASSINATURAS PROTEICAS DAS GLÂNDULAS ANEXAS E TESTÍCULOS  
DE RATOS OBESOS E DIABÉTICOS

**BANCA EXAMINADORA**

Profa. Dra. Fabiana Ferreira de Souza

Presidente e Orientadora

Departamento de Cirurgia Veterinária e Reprodução Animal, FMVZ - UNESP -  
Botucatu/SP

Profa. Dra. Camila Renata Correa Camacho

Membro

Departamento de Clínica Médica, FMB - UNESP - Botucatu/SP

Profa. Dra. Kelry Mayara Da Silva

Membro

UNIFATECIE, Paranavaí/PR

Profa. Dra. Thaís Regiani Cataldi

Membro

Departamento de Genética, ESALQ - USP - Piracicaba/SP

Profa. Dra Jaqueline Candido de Carvalho

Membro

UNISA, São Paulo/SP

Data da defesa: 28 de março 2024.

## AGRADECIMENTOS

Agradeço em primeiro lugar a minha Família, em especial os meus pais Ginaldo e Maria Terezinha, e os meus irmãos Ricardina, Mariana e Ginaldo Júnior e meus sobrinhos, João Paulo e Pedro Lucas, pelo amor e apoio incondicional durante todos esses anos de jornada acadêmica. Pela compreensão e apoio durante tanta ausência. São minhas inspirações e de onde sempre obtenho forças para seguir. E a Deus e Nossa Senhora por toda a proteção e interseção.

A todos os meus amigos de Goiandira, Uberaba, Botucatu e Colômbia, por compartilharmos momentos, por serem minha rede de apoio e a família que escolhi. Sem vocês definitivamente, eu não seria tão feliz e realizado.

A minha orientadora Fabiana Ferreira de Souza, por todo esse tempo de parceria, pela forma que conduziu meus ensinamentos e aprendizado, e pela amizade e admiração construídas. Estendo a todos os professores que passaram pelo meu caminho durante toda a minha formação, aos quais eu tenho total admiração e respeito.

A CAPES e ao CNPq pela concessão de bolsas no Brasil e no Canadá.

## RESUMO

CARVALHO, M.G. ASSINATURAS PROTEICAS DAS GLÂNDULAS ANEXAS E TESTÍCULOS DE RATOS OBESOS E DIABÉTICOS. Botucatu – SP 2024. 424p. Defesa (Doutorado). Faculdade de Medicina Veterinária e Zootecnia, Campus Botucatu, Universidade Estadual Paulista.

A incidência crescente da obesidade e do diabetes representa um desafio significativo para a saúde pública mundial. Na faixa etária dos jovens em idade reprodutiva, a prevalência dessas condições é alarmante, atingindo 19,8% para a obesidade e 8,5% para o diabetes. Além disso, essas doenças estão ligadas à disfunção hormonal e erétil, comprometendo a morfologia e a qualidade dos espermatozoides. Os processos inflamatórios e o estresse oxidativo resultantes de irregularidades na sinalização da glicose e insulina são fatores determinantes para a atrofia e deterioração funcional dos testículos e glândulas associadas, contribuindo significativamente para a infertilidade masculina. Nesse sentido, pretende-se compreender os efeitos da síndrome metabólica na função testicular, próstata e vesícula seminal, utilizando a proteômica para identificar biomarcadores para diagnóstico ou prognóstico da infertilidade masculina. Foram utilizados 30 ratos Wistar com aproximadamente 50 dias de idade, separados em 3 grupos: controle (n = 10), diabéticos (n = 10) e obesos (n = 10). O grupo obeso recebeu dieta de cafeteria e água *ad libitum* com 5% de sacarose, por 38 semanas, simultaneamente os grupos controle e diabéticos receberam ração comercial balanceada e água *ad libitum*. Os animais foram pesados uma vez por semana, para avaliação da evolução do peso para cálculo do índice de *Lee*. O diabetes foi induzido com aplicação de 35 mg/kg de estreptozotocina, dose única, e foram considerados diabéticos animais com perda de peso corporal e níveis de glicemia maiores ou iguais a 200 mg/dL. Os testículos, próstata e vesícula seminal, foram dissecados e preparados para proteômica a qual utilizou a abordagem *shotgun* (CEUA, n 0212/2022). Os dados foram submetidos a análise multivariada (*principal component analysis*) e a univariada (ANOVA e teste de Fisher LSD). Curvas ROC foram construídas para grupos tratados (diabéticos ou obesos) *versus* controle, considerando AUC > 0,80 e P < 0,05. Na análise de componentes principais (PCA) realizada na próstata e no fluido da vesícula seminal, observou-se uma clara separação apenas no grupo de diabéticos, destacando a identificação de 23 e 8 proteínas, respectivamente, com abundância diferencial. Já no testículo, a PCA revelou uma distinção entre os grupos controle, diabético e obeso, conforme evidenciado pelo *heatmap*. Nesse contexto, foram identificadas 80 proteínas com abundância diferencial, distribuídas em diferentes localizações celulares, como citoplasma, acrossomo da célula espermática, mitocôndria e núcleo. A maioria dessas proteínas são produzidas nos testículos humanos. Além disso, na análise das interações proteína-proteína, foram identificadas vias relevantes, como Wnt e spliceossomo, juntamente com alterações nas atividades enzimáticas das vias de glicólise/gliconeogênese. Este estudo revela que as proteínas identificadas estão intimamente ligadas à atividade androgênica, motilidade espermática, integridade celular, proteção contra estresse oxidativo e fertilização. Essas descobertas oferecem *insights* valiosos sobre possíveis biomarcadores associados às glândulas sexuais acessórias e células testiculares em contextos de diabetes e obesidade. Notavelmente, os efeitos foram mais pronunciados entre os indivíduos diabéticos, enquanto nos obesos foram observados mecanismos compensatórios. Essas conclusões destacam a importância crítica de entender as complexas interações entre condições metabólicas e saúde reprodutiva masculina.

**Palavras chaves:** síndrome metabólica; próstata; vesícula seminal, fertilidade, estudo translacional

## ABSTRACT

**CARVALHO, M.G. PROTEIN SIGNATURES OF THE ANNEX GLANDS AND TESTICLES OF OBESE AND DIABETIC RATS.** Botucatu – SP 2024. 424p. Defesa (Doutorado). Faculdade de Medicina Veterinária e Zootecnia, Campus Botucatu, Universidade Estadual Paulista.

The increasing incidence of obesity and diabetes represents a significant challenge for global public health. In the age group of young individuals in reproductive age, the prevalence of these conditions is alarming, reaching 19.8% for obesity and 8.5% for diabetes. Furthermore, these diseases are linked to hormonal and erectile dysfunction, compromising the morphology and quality of spermatozoa. The inflammatory processes and oxidative stress resulting from irregularities in glucose and insulin signaling are determining factors for the atrophy and functional deterioration of testicles and associated glands, significantly contributing to male infertility. In this regard, there is an intention to understand the effects of metabolic syndrome on testicular, prostatic, and seminal vesicle function, using proteomics to identify biomarkers for the diagnosis or prognosis of male infertility. Thirty Wistar rats approximately 50 days old were used, divided into 3 groups: control (n = 10), diabetic (n = 10), and obese (n = 10). The obese group received a cafeteria diet and ad libitum water with 5% sucrose for 38 weeks, while the control and diabetic groups received balanced commercial feed and ad libitum water simultaneously. The animals were weighed once a week to evaluate weight evolution for Lee index calculation. Diabetes was induced by administering a single dose of 35 mg/kg streptozotocin, and animals with body weight loss and blood glucose levels equal to or greater than 200 mg/dL were considered diabetic. The testicles, prostate, and seminal vesicle were dissected and prepared for proteomics using shotgun approach (CEUA, n 0212/2022). The data underwent multivariate analysis (principal component analysis) and univariate analysis (ANOVA and Fisher LSD test). ROC curves were constructed for treated groups (diabetic or obese) versus control, considering AUC > 0.80 and P < 0.05. In the principal component analysis (PCA) performed on the prostate and seminal vesicle fluid, a clear separation was observed only in the diabetic group, highlighting the identification of 23 and 8 proteins, respectively, with differential abundance. In the testicle, PCA revealed a distinction between control, diabetic, and obese groups, as evidenced by the heatmap. In this context, 80 proteins with differential abundance were identified, distributed in different cellular locations such as cytoplasm, sperm cell acrosome, mitochondria, and nucleus. Most of these proteins are produced in human testicles. Additionally, in the analysis of protein-protein interactions, relevant pathways were identified, such as Wnt and spliceosome, along with alterations in enzymatic activities of glycolysis/gluconeogenesis pathways. This study reveals that the identified proteins are closely linked to androgenic activity, sperm motility, cellular integrity, protection against oxidative stress, and fertilization. These findings offer valuable insights into potential biomarkers associated with accessory glands and testicular cells in the context of diabetes and obesity. Notably, the effects were more pronounced among diabetic individuals, while compensatory mechanisms were observed in obese individuals. These conclusions highlight the critical importance of understanding the complex interactions between metabolic conditions and male reproductive health.

**Keywords:** metabolic syndrome; prostate; seminal vesicle, fertility, translational study

## SUMÁRIO

|                                                                                             |           |
|---------------------------------------------------------------------------------------------|-----------|
| <b>CAPÍTULO 1 .....</b>                                                                     | <b>2</b>  |
| 1 INTRODUÇÃO E JUSTIFICATIVA.....                                                           | 2         |
| 2 REVISÃO DE LITERATURA .....                                                               | 3         |
| 2.1 A relação da obesidade, diabetes e síndrome metabólica com a fertilidade masculina..... | 3         |
| 2.2 Proteômica .....                                                                        | 8         |
| 2.2.1 Testículo .....                                                                       | 9         |
| 2.2.2 Vesícula seminal e Próstata .....                                                     | 13        |
| 3 HIPÓTESE .....                                                                            | 18        |
| 4 OBJETIVO GERAL.....                                                                       | 18        |
| 5 OBJETIVOS ESPECÍFICOS .....                                                               | 18        |
| 6 REFERÊNCIAS.....                                                                          | 19        |
| <b>CAPÍTULO 2.....</b>                                                                      | <b>39</b> |
| ARTIGO CIENTÍFICO .....                                                                     | 40        |
| INTRODUÇÃO.....                                                                             | 42        |
| SEÇÃO EXPERIMENTAL .....                                                                    | 43        |
| Preceitos éticos .....                                                                      | 43        |
| Design experimental.....                                                                    | 43        |
| Proteômica .....                                                                            | 43        |
| Análise estatística .....                                                                   | 45        |
| RESULTADOS .....                                                                            | 47        |
| Análise comparativa dos 3 grupos .....                                                      | 47        |
| DISCUSSÃO.....                                                                              | 54        |
| Tecido prostático.....                                                                      | 54        |
| Fluido da vesícula seminal .....                                                            | 59        |
| CONCLUSÃO .....                                                                             | 62        |

|                                           |            |
|-------------------------------------------|------------|
| <b>REFERÊNCIAS .....</b>                  | <b>63</b>  |
| <b>CAPÍTULO 3 .....</b>                   | <b>359</b> |
| <b>ARTIGO CIENTÍFICO .....</b>            | <b>360</b> |
| <b>INTRODUÇÃO .....</b>                   | <b>363</b> |
| <b>SEÇÃO EXPERIMENTAL .....</b>           | <b>364</b> |
| <b>Reagentes.....</b>                     | <b>364</b> |
| <b>Design experimental.....</b>           | <b>364</b> |
| <b>Proteômica .....</b>                   | <b>364</b> |
| <b>Digestão triptica.....</b>             | <b>365</b> |
| <b>LC–MS/MS Análise.....</b>              | <b>365</b> |
| <b>Análise estatística .....</b>          | <b>367</b> |
| <b>RESULTADOS .....</b>                   | <b>367</b> |
| <b>Proteômica .....</b>                   | <b>367</b> |
| <b>DISCUSSÃO .....</b>                    | <b>375</b> |
| <b>Citoplasma .....</b>                   | <b>375</b> |
| <b>Célula espermática .....</b>           | <b>379</b> |
| <b>Mitocôndria.....</b>                   | <b>381</b> |
| <b>Núcleo .....</b>                       | <b>383</b> |
| <b>Via glicólise/gliconeogênese .....</b> | <b>388</b> |
| <b>CONCLUSÃO .....</b>                    | <b>388</b> |
| <b>REFERÊNCIA .....</b>                   | <b>389</b> |

# CAPÍTULO 1

## 1 INTRODUÇÃO E JUSTIFICATIVA

O peso médio da população tem se modificado rapidamente ao longo dos anos e, durante gerações, a curva de distribuição tem crescido de forma exponencial, e o peso humano continua se elevando, afetando homens, muitas vezes jovens e na fase reprodutiva. Em 2019, a obesidade já ocupava o 5º lugar no ranking mundial de doenças que levam ao óbito (Ritchie and Roser, 2020). A evolução global (%), em relação ao aumento da prevalência da obesidade em adultos (> 18 anos), com excesso de peso (IMC > 30), é de 19,8% sendo 16,2% em homens. No Brasil a prevalência é de 22,1% na população e 18,5% em homens. A estimativa é, que até 2035, a prevalência no Brasil seja 41% na população adulta (Lobstein et al., 2023; Wisevoter, 2023a).

Segundo a Sociedade Brasileira de Endocrinologia e Metabologia (SBEM) (Tschiedel, 2014), a obesidade é caracterizada pelo acúmulo excessivo de gordura corporal no indivíduo, causada pela predisposição genética, maus hábitos alimentares ou disfunção endócrina. Em homens, o ideal é que o tecido adiposo corresponda a 20% do peso corporal (Marques and Langouche, 2013). O diagnóstico da obesidade é realizado pelo índice de massa corporal (IMC), parâmetro utilizado pela OMS (WHO, 2018). A obesidade tem etiologia complexa e é associada a diversas comorbidades, elevando o risco de doenças crônicas cardíacas, derrame e diabetes *mellitus* (Pedersen, 2013; Wisevoter, 2023a).

O termo diabetes *mellitus* descreve o distúrbio metabólico com diversas etiologias, alterando o metabolismo de proteínas, gorduras, carboidratos, devido a danos na ação e/ou secreção da insulina (Alberti and Zimmet, 1999; Roglic et al., 2005). Assim como a obesidade, a doença apresenta elevada incidência e abrangência mundial, acometendo também jovens na fase reprodutiva, cuja prevalência é de 9,3% em pessoas com idades acima de 18 anos, de 2000 a 2019 a taxa de mortalidade aumentou 3%, sendo a causa direta de 1,5 milhões de mortes ao ano (WHO, 2023a). No Brasil a prevalência é de 8,8% (Wisevoter, 2023b). Portanto, considerando que essas comorbidades sistêmicas possam coexistir, a avaliação, compreensão e o tratamento voltado para reprodução, de ambas as doenças, assumem uma importância crucial.

Segundo a Organização Mundial da Saúde (WHO, 2023b) estima-se que 17,5% (1 a cada 6 indivíduos) das pessoas adultas ao redor do mundo apresentam infertilidade, sendo a síndrome metabólica, um dos fatores masculinos (Heryanto et al., 2022; Y. Zhang et al., 2023). A fertilidade é definida como a capacidade de gerar descendentes normais, condição essencial para o progresso genético, podendo ser alterada por condições que

afetam a capacidade de cópula e/ou habilidade dos espermatozoides fecundarem o oócito (Bicudo et al., 2007). Em humanos, atualmente estima-se que a infertilidade atinja 8% a 15% dos casais em idade reprodutiva (Cui, 2010). São considerados inférteis casais que não usam métodos contraceptivos durante 12 meses, e não conseguem engravidar, (WHO, 2023b) aproximadamente 35% dos casos, a infertilidade é causada por fatores masculinos ou femininos, 20% fatores masculinos e 10% fatores desconhecidos (Matos, 2019). Assim poderíamos dizer, que 55% são fatores masculinos.

As implicações genéticas na ocorrência de obesidade são raras, sendo originada por diversos fatores, como alterações ambientais e comportamentais (Malik, Hu, 2012; Nascimento et al., 2008). Por esse motivo modelos experimentais poligênicos de obesidade induzida por dietas, são preferencialmente utilizados, mimetizando a fisiopatologia da doença, levando a melhor compreensão da síndrome metabólica, e consequente diabetes (Brown, Panchal, 2011; Nilsson et al., 2012).

Em vista da prevalência da obesidade e das comorbidades associadas, como o diabetes, ambas atuando na síndrome metabólica e consequentemente sobre a fertilidade masculina, este estudo teve como objetivo identificar os efeitos da doença sobre a capacidade reprodutiva de ratos obesos e diabéticos. Embora sejam de abrangência mundial ainda permanece inconclusivo, a influência dessas doenças e suas associações no sistema reprodutivo. Desta forma, este estudo visa identificar suas consequências no eixo reprodutivo, utilizando a abordagem proteômica do testículo, próstata e vesícula seminal, em busca de possíveis biomarcadores da infertilidade masculina.

A associação entre as comorbidades e as ações deletérias na função reprodutiva masculina é multifatorial, isso dificulta o entendimento das vias pelas quais ocorrem as principais alterações moleculares, levando a uma diversificada tradução de proteínas no tecido alvo. Ademais, os estudos translacionais utilizam diferentes metodologias e modelos experimentais, para indução da obesidade e diabetes, somado a isso, existem poucos estudos que avaliam ambas as doenças no mesmo delineamento experimental e que usam proteômica do sistema reprodutivo de obesos e diabético.

## 6 REFERÊNCIAS

Aalberts M, Stout TAE, Stoorvogel W. Prostatosomes: extracellular vesicles from the prostate. *REPRODUCTION* 2014;147:R1–14. <https://doi.org/10.1530/REP-13-0358>.

Aaron-Brooks LM, Sasaki T, Vickman RE, Wei L, Franco OE, Ji Y, et al. Hyperglycemia and T Cell infiltration are associated with stromal and epithelial prostatic hyperplasia in the nonobese diabetic mouse. *Prostate* 2019;79:980–93. <https://doi.org/10.1002/pros.23809>.

Abbas W, Kumar A, Herbein G. The eEF1A Proteins: At the Crossroads of Oncogenesis, Apoptosis, and Viral Infections. *Front Oncol* 2015;5. <https://doi.org/10.3389/fonc.2015.00075>.

Ajayi AF, Lateef OM, Adebayo OI, Adebayo ET. Hsps70 and 90 protect the heart of hyperthyroid rats via nitric oxide production and VEGF inhibition of apoptosis. *Endocrine and Metabolic Science* 2021;4:100097. <https://doi.org/10.1016/j.endmts.2021.100097>.

Alberti KGMM, Zimmet PZ. Part 1: Diagnosis and Classification of Diabetes Mellitus. Definition, Diagnosis and Classification of Diabetes Mellitus and its Complications, 1999, p. 49. <https://doi.org/WHO/NCD/NCS/99.2>.

Almanza A, Carlesso A, Chintia C, Creedican S, Doultisinos D, Leuzzi B, et al. Endoplasmic reticulum stress signalling – from basic mechanisms to clinical applications. *FEBS J* 2019;286:241–78. <https://doi.org/10.1111/febs.14608>.

Alur M, Nguyen MM, Eggener SE, Jiang F, Dadras SS, Stern J, et al. Suppressive Roles of Calreticulin in Prostate Cancer Growth and Metastasis. *Am J Pathol* 2009;175:882–90. <https://doi.org/10.2353/ajpath.2009.080417>.

Alvarez-Rodríguez M, Alvarez M, Borrigan S, Martinez-Pastor F, Holt WV, Fazeli A, et al. The addition of heat shock protein HSPA8 to cryoprotective media improves the survival of brown bear (*Ursus arctos*) spermatozoa during chilling and after cryopreservation. *Theriogenology* 2013;79:541–50. <https://doi.org/10.1016/j.theriogenology.2012.11.006>.

An T, Zhang J, Ma Y, Lian J, Wu Y-X, Lv B-H, et al. Relationships of Non-coding RNA with diabetes and depression. *Sci Rep* 2019;9:10707. <https://doi.org/10.1038/s41598-019-47077-9>.

Anamthathmakula P, Erickson JA, Winuthayanon W. Blocking serine protease activity prevents semenogelin degradation leading to hyperviscous semen in humans. *Biol Reprod* 2022;106:879–87. <https://doi.org/10.1093/biolre/ioac023>.

Araki N, Kawano N, Kang W, Miyado K, Yoshida K, Yoshida M. Seminal vesicle proteins SVS3 and SVS4 facilitate SVS2 effect on sperm capacitation. *Reproduction* 2016;152:313–21. <https://doi.org/10.1530/REP-15-0551>.

Arispe N, Doh M, Simakova O, Kurganov B, Maio A De. Hsc70 and Hsp70 interact with phosphatidylserine on the surface of PC12 cells resulting in a decrease of viability. *The FASEB Journal* 2004;18:1636–45. <https://doi.org/10.1096/fj.04-2088com>.

Aumüller G, Riva A. Morphology and functions of the human seminal vesicle. *Andrologia* 2009;24:183–96. <https://doi.org/10.1111/j.1439-0272.1992.tb02636.x>.

Aziz NM, Kamel MY, Mohamed MS, Ahmed SM. Antioxidant, anti-inflammatory, and anti-apoptotic effects of zinc supplementation in testes of rats with experimentally induced diabetes. *Applied Physiology, Nutrition, and Metabolism* 2018;43:1010–8. <https://doi.org/10.1139/apnm-2018-0070>.

Baker SS, Cardullo RA, Thaler CD. Sonication of Mouse Sperm Membranes Reveals Distinct Protein Domains1. *Biol Reprod* 2002;66:57–64. <https://doi.org/10.1095/biolreprod66.1.57>.

Besiroglu H, Ozbek E, Dursun M, Otunctemur A. Visceral adiposity index is associated with benign prostatic enlargement in non-diabetic patients: a cross-sectional study. *The Aging Male* 2018;21:40–7. <https://doi.org/10.1080/13685538.2017.1365833>.

Bezerra MJB, Arruda-Alencar JM, Martins JAM, Viana AGA, Viana Neto AM, Rêgo JPA, et al. Major seminal plasma proteome of rabbits and associations with sperm quality. *Theriogenology* 2019;128:156–66. <https://doi.org/10.1016/j.theriogenology.2019.01.013>.

Bicudo SD, Siqueira JB, Meira C. Patologias do sistema reprodutor de touros. *Biológico* 2007;69:43–8.

Blando J, Moore T, Hursting S, Jiang G, Saha A, Beltran L, et al. Dietary Energy Balance Modulates Prostate Cancer Progression in Hi-Myc Mice. *Cancer Prevention Research* 2011;4:2002–14. <https://doi.org/10.1158/1940-6207.CAPR-11-0182>.

Blochberger TC, Vergnes JP, Hempel J, Hassell JR. cDNA to chick lumican (corneal keratan sulfate proteoglycan) reveals homology to the small interstitial proteoglycan gene family and expression in muscle and intestine. *J Biol Chem* 1992;267:347–52.

Boden G, Duan X, Homko C, Molina EJ, Song W, Perez O, et al. Increase in Endoplasmic Reticulum Stress–Related Proteins and Genes in Adipose Tissue of Obese,

Insulin-Resistant Individuals. *Diabetes* 2008;57:2438–44. <https://doi.org/10.2337/db08-0604>.

Bouker A, Halouani L, Kharouf M, Latrous H, Makni M, Marrakchi O, et al. Step-by-step loupes-mTESE in non-obstructive azoospermic men, a retrospective study. *Basic Clin Androl* 2019;29:11. <https://doi.org/10.1186/s12610-019-0091-9>.

Campbell KR, Chaudhary R, Montano M, Iozzo R V., Bushman WA, Campagnola PJ. Second-harmonic generation microscopy analysis reveals proteoglycan decorin is necessary for proper collagen organization in prostate. *J Biomed Opt* 2019;24:1. <https://doi.org/10.1117/1.JBO.24.6.066501>.

Carvalho MDGDC, Carvalho JF, Merrick WC. Biological characterization of various forms of elongation factor 1 from rabbit reticulocytes. *Arch Biochem Biophys* 1984;234:603–11. [https://doi.org/10.1016/0003-9861\(84\)90310-2](https://doi.org/10.1016/0003-9861(84)90310-2).

Carvalho MG, Silva KM, Aristizabal VHV, Ortiz PEO, Paranzini CS, Melchert A, et al. Effects of Obesity and Diabetes on Sperm Cell Proteomics in Rats. *J Proteome Res* 2021;20:2628–42. <https://doi.org/10.1021/acs.jproteome.0c01044>.

Cheng L, Sun P, Xie X, Sun D, Zhou Q, Yang S, et al. Hepatitis B virus surface protein induces oxidative stress by increasing peroxides and inhibiting antioxidant defences in human spermatozoa. *Reprod Fertil Dev* 2020;32:1180. <https://doi.org/10.1071/RD20130>.

Chiu PCN, Chung M-K, Tsang H-Y, Koistinen R, Koistinen H, Seppala M, et al. Glycodelin-S in Human Seminal Plasma Reduces Cholesterol Efflux and Inhibits Capacitation of Spermatozoa. *Journal of Biological Chemistry* 2005;280:25580–9. <https://doi.org/10.1074/jbc.M504103200>.

Clavert A, Cranz C, Bollack C. Functions of the seminal vesicle: Funktionen der Bläschendrüse. *Andrologia* 1990;22:185–92. <https://doi.org/10.1111/j.1439-0272.1990.tb02084.x>.

Cui W. Mother of nothing: the agony of infertility. *Bull World Health Organ* 2010;88:881–2. <https://doi.org/10.2471/BLT.10.011210>.

Cultrara CN, Kozuch SD, Ramasundaram P, Heller CJ, Shah S, Beck AE, et al. GRP78 modulates cell adhesion markers in prostate Cancer and multiple myeloma cell lines. *BMC Cancer* 2018;18:1263. <https://doi.org/10.1186/s12885-018-5178-8>.

Curci L, Brukman NG, Weigel Muñoz M, Rojo D, Carvajal G, Sulzyk V, et al. Functional redundancy and compensation: Deletion of multiple murine *Crisp* genes

reveals their essential role for male fertility. *The FASEB Journal* 2020;34:15718–33. <https://doi.org/10.1096/fj.202001406R>.

Dickerman BA, Ahearn TU, Giovannucci E, Stampfer MJ, Nguyen PL, Mucci LA, et al. Weight change, obesity and risk of prostate cancer progression among men with clinically localized prostate cancer. *Int J Cancer* 2017;141:933–44. <https://doi.org/10.1002/ijc.30803>.

Ebert B, Kisiela M, Malátková P, El-Hawari Y, Maser E. Regulation of Human Carbonyl Reductase 3 (CBR3; SDR21C2) Expression by Nrf2 in Cultured Cancer Cells. *Biochemistry* 2010;49:8499–511. <https://doi.org/10.1021/bi100814d>.

Elango K, Karuthadurai T, Kumaresan A, Sinha MK, Ebenezer Samuel King JP, Nag P, et al. High-throughput proteomic characterization of seminal plasma from bulls with contrasting semen quality. *3 Biotech* 2023a;13:60. <https://doi.org/10.1007/s13205-023-03474-6>.

Elango K, Karuthadurai T, Kumaresan A, Sinha MK, Ebenezer Samuel King JP, Nag P, et al. High-throughput proteomic characterization of seminal plasma from bulls with contrasting semen quality. *3 Biotech* 2023b;13:60. <https://doi.org/10.1007/s13205-023-03474-6>.

ElBanna AH, Osman AS, Hossny A, ElBanna H, Abo El-Ela FI. Dose-dependent effects of taurine against testicular damage in a streptozotocin-induced type 1 diabetes mellitus rat model. *Int J Immunopathol Pharmacol* 2023;37:039463202311727. <https://doi.org/10.1177/03946320231172745>.

Elliott RMA, Lloyd RE, Fazeli A, Sostaric E, Georgiou AS, Satake N, et al. Effects of HSPA8, an evolutionarily conserved oviductal protein, on boar and bull spermatozoa. *REPRODUCTION* 2009;137:191–203. <https://doi.org/10.1530/REP-08-0298>.

Erata GÖ, Koçak Toker N, Durlanık Ö, Kadioğlu A, Aktan G, Aykaç Toker G. The role of heat shock protein 70 (Hsp 70) in male infertility: is it a line of defense against sperm DNA fragmentation? *Fertil Steril* 2008;90:322–7. <https://doi.org/10.1016/j.fertnstert.2007.06.021>.

Fan L, Goh B-C, Wong C-I, Sukri N, Lim S-E, Tan S-H, et al. Genotype of human carbonyl reductase CBR3 correlates with doxorubicin disposition and toxicity. *Pharmacogenet Genomics* 2008;18:623–31. <https://doi.org/10.1097/FPC.0b013e328301a869>.

Fan Y, Guo Y, Hamblin M, Chang L, Zhang J, Chen YE. Inhibition of Gluconeogenic Genes by Calcium-regulated Heat-stable Protein 1 via Repression of

Peroxisome Proliferator-activated Receptor  $\alpha$ . *Journal of Biological Chemistry* 2011;286:40584–94. <https://doi.org/10.1074/jbc.M111.232918>.

Ferdous Z, Wei VM, Iozzo R, Höök M, Grande-Allen KJ. Decorin-transforming Growth Factor- $\beta$  Interaction Regulates Matrix Organization and Mechanical Characteristics of Three-dimensional Collagen Matrices. *Journal of Biological Chemistry* 2007;282:35887–98. <https://doi.org/10.1074/jbc.M705180200>.

Feske S, Wulff H, Skolnik EY. Ion Channels in Innate and Adaptive Immunity. *Annu Rev Immunol* 2015;33:291–353. <https://doi.org/10.1146/annurev-immunol-032414-112212>.

Ficarro S, Chertihin O, Westbrook VA, White F, Jayes F, Kalab P, et al. Phosphoproteome Analysis of Capacitated Human Sperm. *Journal of Biological Chemistry* 2003;278:11579–89. <https://doi.org/10.1074/jbc.M202325200>.

Fontana C. Tipos de diabetes 2023. <https://www.euroclinux.net/pt/diabetes/tipos-de-diabetes> (accessed August 22, 2023).

Fujihara Y, Noda T, Kobayashi K, Oji A, Kobayashi S, Matsumura T, et al. Identification of multiple male reproductive tract-specific proteins that regulate sperm migration through the oviduct in mice. *Proceedings of the National Academy of Sciences* 2019;116:18498–506. <https://doi.org/10.1073/pnas.1908736116>.

Furmanik M, van Gorp R, Whitehead M, Ahmad S, Bordoloi J, Kapustin A, et al. Endoplasmic Reticulum Stress Mediates Vascular Smooth Muscle Cell Calcification via Increased Release of Grp78 (Glucose-Regulated Protein, 78 kDa)-Loaded Extracellular Vesicles. *Arterioscler Thromb Vasc Biol* 2021;41:898–914. <https://doi.org/10.1161/ATVBAHA.120.315506>.

Garciaparamio P, Carmena MJ, Gutierrezocana MT, Recio MN, Prieto JC. Alteration of Protein Kinase C Activity in Diabetic Rat Prostate. *Biochem Biophys Res Commun* 1993;195:166–72. <https://doi.org/10.1006/bbrc.1993.2025>.

Ge T, Yuan L, Xu L, Yang F, Xu W, Niu C, et al. Coiled-coil domain containing 159 (CCDC159) is required for spermatid head and tail assembly in mice. *Biol Reprod* 2024. <https://doi.org/10.1093/biolre/ioae012>.

van Gestel RA, Brewis IA, Ashton PR, Brouwers JF, Gadella BM. Multiple proteins present in purified porcine sperm apical plasma membranes interact with the zona pellucida of the oocyte. *MHR: Basic Science of Reproductive Medicine* 2007;13:445–54. <https://doi.org/10.1093/molehr/gam030>.

Ghaderi S, Ahmadian S, Soheili Z, Ahmadi H, Samiei S, Kheitan S, et al. AAV delivery of GRP78/BiP promotes adaptation of human RPE cell to ER stress. *J Cell Biochem* 2018;119:1355–67. <https://doi.org/10.1002/jcb.26296>.

Gomes FP, Park R, Viana AG, Fernandez-Costa C, Topper E, Kaya A, et al. Protein signatures of seminal plasma from bulls with contrasting frozen-thawed sperm viability. *Sci Rep* 2020;10:14661. <https://doi.org/10.1038/s41598-020-71015-9>.

Greenberg NM, DeMayo FJ, Sheppard PC, Barrios R, Lebovitz R, Finegold M, et al. The rat probasin gene promoter directs hormonally and developmentally regulated expression of a heterologous gene specifically to the prostate in transgenic mice. *Molecular Endocrinology* 1994;8:230–9. <https://doi.org/10.1210/mend.8.2.8170479>.

Groblewski GE, Yoshida M, Bragado MJ, Ernst SA, Leykam J, Williams JA. Purification and Characterization of a Novel Physiological Substrate for Calcineurin in Mammalian Cells. *Journal of Biological Chemistry* 1998;273:22738–44. <https://doi.org/10.1074/jbc.273.35.22738>.

Gruber CW, Čemažar M, Heras B, Martin JL, Craik DJ. Protein disulfide isomerase: the structure of oxidative folding. *Trends Biochem Sci* 2006;31:455–64. <https://doi.org/10.1016/j.tibs.2006.06.001>.

Heryanto A, I Gusti Ngurah Pramesemara, Yukhi Kurniawan, I Gusti Ayu Widiyanti. Relationship Between Body Mass Index and Male Infertility at Kasih Ibu General Hospital Denpasar. *Indonesian Andrology and Biomedical Journal* 2022;3:1–6. <https://doi.org/10.20473/iabj.v3i1.35876>.

Heutinck KM, ten Berge IJM, Hack CE, Hamann J, Rowshani AT. Serine proteases of the human immune system in health and disease. *Mol Immunol* 2010;47:1943–55. <https://doi.org/10.1016/j.molimm.2010.04.020>.

Hiller-Sturmhöfel S, Bartke A. The endocrine system: an overview. *Alcohol Health Res World* 1998;22:153–64.

Holland JW, Meehan KL, Redmond SL, Dawkins HJS. Purification of the keratan sulfate proteoglycan expressed in prostatic secretory cells and its identification as lumican. *Prostate* 2004;59:252–9. <https://doi.org/10.1002/pros.20002>.

Holt M. Cell motility: proline-rich proteins promote protrusions. *Trends Cell Biol* 2001;11:38–46. [https://doi.org/10.1016/S0962-8924\(00\)01876-6](https://doi.org/10.1016/S0962-8924(00)01876-6).

Hou H, Wang F, Zhang W, Wang D, Li X, Bartlam M, et al. Structure-Functional Analyses of CRHSP-24 Plasticity and Dynamics in Oxidative Stress Response. *Journal of Biological Chemistry* 2011;286:9623–35. <https://doi.org/10.1074/jbc.M110.177436>.

Hu Y, Sun H, Owens RT, Wu J, Chen YQ, Berquin IM, et al. Decorin Suppresses Prostate Tumor Growth through Inhibition of Epidermal Growth Factor and Androgen Receptor Pathways. *Neoplasia* 2009;11:1042–53. <https://doi.org/10.1593/neo.09760>.

Huang SY, Kuo YH, Lee WC, Tsou HL, Lee YP, Chang HL, et al. Substantial decrease of heat-shock protein 90 precedes the decline of sperm motility during cooling of boar spermatozoa. *Theriogenology* 1999;51:1007–16. [https://doi.org/10.1016/S0093-691X\(99\)00046-1](https://doi.org/10.1016/S0093-691X(99)00046-1).

Huang SY, Kuo YH, Lee YP, Tsou HL, Lin EC, Ju CC, et al. Association of heat shock protein 70 with semen quality in boars. *Anim Reprod Sci* 2000;63:231–40. [https://doi.org/10.1016/S0378-4320\(00\)00175-5](https://doi.org/10.1016/S0378-4320(00)00175-5).

Inoue N, Ikawa M, Nakanishi T, Matsumoto M, Nomura M, Seya T, et al. Disruption of Mouse CD46 Causes an Accelerated Spontaneous Acrosome Reaction in Sperm. *Mol Cell Biol* 2003;23:2614–22. <https://doi.org/10.1128/MCB.23.7.2614-2622.2003>.

Iso-Touru T, Wurmser C, Venhoranta H, Hiltbold M, Savolainen T, Sironen A, et al. A splice donor variant in CCDC189 is associated with asthenospermia in Nordic Red dairy cattle. *BMC Genomics* 2019;20:286. <https://doi.org/10.1186/s12864-019-5628-y>.

Jackson FL, Hutson JC. Altered Responses to Androgen in Diabetic Male Rats. *Diabetes* 1984;33:819–24. <https://doi.org/10.2337/diab.33.9.819>.

Jakobs C, Bartok E, Kubarenko A, Bauernfeind F, Hornung V. Immunoblotting for Active Caspase-1. *Methods in Molecular Biology*, vol. 1040. Clifton, N.J., New York : 2013, p. 103–15. [https://doi.org/10.1007/978-1-62703-523-1\\_9](https://doi.org/10.1007/978-1-62703-523-1_9).

Jarvis S, Gethings LA, Samanta L, Pedroni SMA, Withers DJ, Gray N, et al. High fat diet causes distinct aberrations in the testicular proteome. *Int J Obes* 2020;44:1958–69. <https://doi.org/10.1038/s41366-020-0595-6>.

Jesik CJ, Holland JM, Lee C. An anatomic and histologic study of the rat prostate. *Prostate* 1982;3:81–97. <https://doi.org/10.1002/pros.2990030111>.

Kherraf Z, Christou-Kent M, Karaouzene T, Amiri-Yekta A, Martinez G, Vargas AS, et al. SPINK2 deficiency causes infertility by inducing sperm defects in heterozygotes and azoospermia in homozygotes. *EMBO Mol Med* 2017;9:1132–49. <https://doi.org/10.15252/emmm.201607461>.

Kim J-H, Park S-J, Kim T-S, Park H-J, Park J, Kim BK, et al. Testicular hyperthermia induces Unfolded Protein Response signaling activation in spermatocyte.

Biochem Biophys Res Commun 2013;434:861–6.  
<https://doi.org/10.1016/j.bbrc.2013.04.032>.

KOCHAKIAN CD. Hypotaurine: Regulation of Production in Seminal Vesicles and Prostate of Guinea-pig by Testosterone. *Nature* 1973;241:202–3.  
<https://doi.org/10.1038/241202a0>.

Koninger J. The ECM proteoglycan decorin links desmoplasia and inflammation in chronic pancreatitis. *J Clin Pathol* 2006;59:21–7.  
<https://doi.org/10.1136/jcp.2004.023135>.

Kornberg RD. Structure of Chromatin. *Annu Rev Biochem* 1977;46:931–54.  
<https://doi.org/10.1146/annurev.bi.46.070177.004435>.

Lallet-Daher H, Roudbaraki M, Bavencoffe A, Mariot P, Gackière F, Bidaux G, et al. Intermediate-conductance  $\text{Ca}^{2+}$ -activated  $\text{K}^{+}$  channels (IKCa1) regulate human prostate cancer cell proliferation through a close control of calcium entry. *Oncogene* 2009;28:1792–806. <https://doi.org/10.1038/onc.2009.25>.

Lea OA, Petrusz P, French FS. Prostatein. A major secretory protein of the rat ventral prostate. *J Biol Chem* 1979;254:6196–202.

Leisegang K, Bouic PJD, Henkel RR. Metabolic syndrome is associated with increased seminal inflammatory cytokines and reproductive dysfunction in a case-controlled male cohort. *American Journal of Reproductive Immunology* 2016;76:155–63.  
<https://doi.org/10.1111/aji.12529>.

Lin M-H, Lee RK-K, Hwu Y-M, Lu C-H, Chu S-L, Chen Y-J, et al. SPINKL, a Kazal-type serine protease inhibitor-like protein purified from mouse seminal vesicle fluid, is able to inhibit sperm capacitation. *REPRODUCTION* 2008a;136:559–71.  
<https://doi.org/10.1530/REP-07-0375>.

Lin M-H, Lee RK-K, Hwu Y-M, Lu C-H, Chu S-L, Chen Y-J, et al. SPINKL, a Kazal-type serine protease inhibitor-like protein purified from mouse seminal vesicle fluid, is able to inhibit sperm capacitation. *REPRODUCTION* 2008b;136:559–71.  
<https://doi.org/10.1530/REP-07-0375>.

Lin S-R, Li Y-C, Luo M-L, Guo H, Wang T-T, Chen J-B, et al. Identification and characteristics of the testes-specific gene, *Ccdc38*, in mice. *Mol Med Rep* 2016;14:1290–6. <https://doi.org/10.3892/mmr.2016.5360>.

Liu J, Yin G, Hu K, Huang H, Xu F, Yang Y, et al. Parental uveitis causes elevated hair loss in offspring of C57BL/6J mice. *Exp Eye Res* 2022;219:109056.  
<https://doi.org/10.1016/j.exer.2022.109056>.

Lloyd RE, Fazeli A, Watson PF, Holt W V. The oviducal protein, heat-shock 70-kDa protein 8, improves the long-term survival of ram spermatozoa during storage at 17°C in a commercial extender. *Reprod Fertil Dev* 2012;24:543. <https://doi.org/10.1071/RD11173>.

Lobstein T, Jackson-Leach R, Powis J, Brinsden H, Gray M. Country scorecards. *World Obesity Atlas 2023*, vol. 5, 2023, p. 60.

Long L, Qiu H, Cai B, Chen N, Lu X, Zheng S, et al. Hyperglycemia induced testicular damage in type 2 diabetes mellitus rats exhibiting microcirculation impairments associated with vascular endothelial growth factor decreased via PI3K/Akt pathway. *Oncotarget* 2018;9:5321–36. <https://doi.org/10.18632/oncotarget.23915>.

Lu A, Pallero MA, Owusu BY, Borovjagin A V., Lei W, Sanders PW, et al. Calreticulin is important for the development of renal fibrosis and dysfunction in diabetic nephropathy. *Matrix Biol Plus* 2020;8:100034. <https://doi.org/10.1016/j.mbplus.2020.100034>.

Lu C-H, Lee RK-K, Hwu Y-M, Chu S-L, Chen Y-J, Chang W-C, et al. SERPINE2, a Serine Protease Inhibitor Extensively Expressed in Adult Male Mouse Reproductive Tissues, May Serve as a Murine Sperm Decapacitation Factor1. *Biol Reprod* 2011;84:514–25. <https://doi.org/10.1095/biolreprod.110.085100>.

Luo J, Dunn T, Ewing C, Sauvageot J, Chen Y, Trent J, et al. Gene expression signature of benign prostatic hyperplasia revealed by cDNA microarray analysis. *Prostate* 2002;51:189–200. <https://doi.org/10.1002/pros.10087>.

Luo S, Wehr NB, Levine RL. Quantitation of protein on gels and blots by infrared fluorescence of Coomassie blue and Fast Green. *Anal Biochem* 2006;350:233–8. <https://doi.org/10.1016/j.ab.2005.10.048>.

Lv Z-M, Ling M-Y, Chen C. Comparative proteomics reveals protective effect of resveratrol on a high-fat diet-induced damage to mice testis. *Syst Biol Reprod Med* 2020;66:37–49. <https://doi.org/10.1080/19396368.2019.1701138>.

Ma L, Yu H, Ni Z, Hu S, Ma W, Chu C, et al. Spink13, an Epididymis-specific Gene of the Kazal-type Serine Protease Inhibitor (SPINK) Family, Is Essential for the Acrosomal Integrity and Male Fertility. *Journal of Biological Chemistry* 2013;288:10154–65. <https://doi.org/10.1074/jbc.M112.445866>.

Mahajan K, Malla P, Lawrence HR, Chen Z, Kumar-Sinha C, Malik R, et al. ACK1/TNK2 Regulates Histone H4 Tyr88-phosphorylation and AR Gene Expression in

Castration-Resistant Prostate Cancer. *Cancer Cell* 2017;31:790-803.e8. <https://doi.org/10.1016/j.ccell.2017.05.003>.

Manandhar G, Miranda-Vizuet A, Pedrajas JR, Krause WJ, Zimmerman S, Sutovsky M, et al. Peroxiredoxin 2 and Peroxidase Enzymatic Activity of Mammalian Spermatozoa1. *Biol Reprod* 2009;80:1168–77. <https://doi.org/10.1095/biolreprod.108.071738>.

Marques MB, Langouche L. Endocrine, metabolic, and morphologic alterations of adipose tissue during critical illness. *Crit Care Med* 2013;41:317–25. <https://doi.org/DOI:10.1097/CCM.0b013e318265f21c>.

de Mateo S, Castillo J, Estanyol JM, Ballescà JL, Oliva R. Proteomic characterization of the human sperm nucleus. *Proteomics* 2011;11:2714–26. <https://doi.org/10.1002/pmic.201000799>.

Matos F. Infertilidade: como enfrentar o diagnóstico e buscar o tratamento adequado 2019. <https://sbra.com.br/noticias/infertilidade-como-enfrentar-o-diagnostico-e-buscar-o-tratamento-adequado/> (accessed August 16, 2023).

Mcgee RS, Herr JC. Human Seminal Vesicle-Specific Antigen is a Substrate for Prostate-Specific Antigen (or P-30)1. *Biol Reprod* 1988;39:499–510. <https://doi.org/10.1095/biolreprod39.2.499>.

Mi K, Chen Y, Lizhong Z, Shuanying Y. RNA-binding Protein ZC3H18 Enhances Cell Proliferation and Metastasis in Lung Adenocarcinoma. *Mol Biol Rep* 2023;1:1–19. <https://doi.org/https://doi.org/10.21203/rs.3.rs-2783463/v1>.

Miura T, Nishinaka T, Terada T. Different functions between human monomeric carbonyl reductase 3 and carbonyl reductase 1. *Mol Cell Biochem* 2008;315:113–21. <https://doi.org/10.1007/s11010-008-9794-5>.

Mogielnicka-Brzozowska M, Fraser L, Dziekońska A, Gackowska K, Sobiewska M, Kuzborska A, et al. Identification of proteoforms of albumin and kallikrein in stallion seminal plasma and their correlations with sperm motility. *Pol J Vet Sci* 2019;227–227. <https://doi.org/10.24425/pjvs.2019.127090>.

Moradi M, Karimi J, Khodadadi I, Amiri I, Karami M, Saidijam M, et al. Evaluation of the p53 and Thioredoxin reductase in sperm from asthenozoospermic males in comparison to normozoospermic males. *Free Radic Biol Med* 2018;116:123–8. <https://doi.org/10.1016/j.freeradbiomed.2017.12.038>.

Motrich RD, Maccioni M, Ponce AA, Gatti GA, Oberti JPM, Rivero VE. Pathogenic Consequences in Semen Quality of an Autoimmune Response against the

Prostate Gland: From Animal Models to Human Disease. *The Journal of Immunology* 2006;177:957–67. <https://doi.org/10.4049/jimmunol.177.2.957>.

Murer V, Spetz JF, Hengst U, Altrogge LM, de Agostini A, Monard D. Male fertility defects in mice lacking the serine protease inhibitor protease nexin-1. *Proceedings of the National Academy of Sciences* 2001;98:3029–33. <https://doi.org/10.1073/pnas.051630698>.

Murtola TJ, Vihervuori VJ, Lahtela J, Talala K, Taari K, Tammela TL, et al. Fasting blood glucose, glycaemic control and prostate cancer risk in the Finnish Randomized Study of Screening for Prostate Cancer. *Br J Cancer* 2018;118:1248–54. <https://doi.org/10.1038/s41416-018-0055-4>.

Neuhoff V, Stamm R, Eibl H. Clear background and highly sensitive protein staining with Coomassie Blue dyes in polyacrylamide gels: A systematic analysis. *Electrophoresis* 1985;6:427–48. <https://doi.org/10.1002/elps.1150060905>.

Noda T, Fujihara Y, Matsumura T, Oura S, Kobayashi S, Ikawa M. Seminal vesicle secretory protein 7, PATE4, is not required for sperm function but for copulatory plug formation to ensure fecundity†. *Biol Reprod* 2019;100:1035–45. <https://doi.org/10.1093/biolre/iyoy247>.

NUMEB. Insulina e Glucagon como modelos de estrutura protéica. Núcleo de Produção de Material Educacional e Divulgação Científica Em Biologia - FURG 2023. [https://numeb.furg.br/index.php?option=com\\_content&view=article&id=41&Itemid](https://numeb.furg.br/index.php?option=com_content&view=article&id=41&Itemid) (accessed August 22, 2023).

O’Flaherty C, Rico de Souza A. Hydrogen Peroxide Modifies Human Sperm Peroxiredoxins in a Dose-Dependent Manner. *Biol Reprod* 2011;84:238–47. <https://doi.org/10.1095/biolreprod.110.085712>.

Ohmuraya M, Sugano A, Hirota M, Takaoka Y, Yamamura K. Role of Intrapancreatic SPINK1/Spink3 Expression in the Development of Pancreatitis. *Front Physiol* 2012;3. <https://doi.org/10.3389/fphys.2012.00126>.

Ou C -M., Tang J -B., Huang M -S., Sudhakar Gandhi PS, Geetha S, Li S -H., et al. The mode of reproductive-derived Spink (serine protease inhibitor Kazal-type) action in the modulation of mammalian sperm activity. *Int J Androl* 2012;35:52–62. <https://doi.org/10.1111/j.1365-2605.2011.01159.x>.

Page MJ, Di Cera E. Serine peptidases: Classification, structure and function. *Cellular and Molecular Life Sciences* 2008;65:1220–36. <https://doi.org/10.1007/s00018-008-7565-9>.

Park Y-J, Pang M-G. Mitochondrial Functionality in Male Fertility: From Spermatogenesis to Fertilization. *Antioxidants* 2021;10:98. <https://doi.org/10.3390/antiox10010098>.

Parker M, Needham M, White R. Prostatic steroid binding protein: gene duplication and steroid binding. *Nature* 1982a;298:92–4. <https://doi.org/10.1038/298092a0>.

Parker M, Needham M, White R, Hurst H, Page M. Prostatic steroid binding protein: organisation of C1 and C2 genes. *Nucleic Acids Res* 1982b;10:5121–32. <https://doi.org/10.1093/nar/10.17.5121>.

Parmar KH, Singh V, Savsani HH, Kavani FS, Rajoriya JS, Lone SA. Seasonal variation in heat shock proteins (hsp70 and hsp90) and their association with frozen semen quality and fertility in buffaloes. *Cryo Letters* 2021;42:261–6.

Parte PP, Rao P, Redij S, Lobo V, D'Souza SJ, Gajbhiye R, et al. Sperm phosphoproteome profiling by ultra performance liquid chromatography followed by data independent analysis (LC–MSE) reveals altered proteomic signatures in asthenozoospermia. *J Proteomics* 2012;75:5861–71. <https://doi.org/10.1016/j.jprot.2012.07.003>.

Peach M, Marsh N, MacPhee DJ. Protein Solubilization: Attend to the Choice of Lysis Buffer. *Methods in Molecular Biology*, vol. 869. Humana Press, Totowa: 2012, p. 37–47. [https://doi.org/10.1007/978-1-61779-821-4\\_4](https://doi.org/10.1007/978-1-61779-821-4_4).

Pedersen SD. Metabolic complications of obesity. *Best Pract Res Clin Endocrinol Metab* 2013;27:179–93. <https://doi.org/10.1016/j.beem.2013.02.004>.

Pei L, Yang G, Jiang J, Jiang R, Deng Q, Chen B, et al. Expression of Aquaporins in Prostate and Seminal Vesicles of Diabetic Rats. *J Sex Med* 2013;10:2975–85. <https://doi.org/10.1111/jsm.12276>.

Peng Q, Yang H, Xue S, Shi L, Yu Q, Kuang Y. Secretome profile of mouse oocytes after activation using mass spectrum. *J Assist Reprod Genet* 2012a;29:765–71. <https://doi.org/10.1007/s10815-012-9789-8>.

Peng Q, Yang H, Xue S, Shi L, Yu Q, Kuang Y. Secretome profile of mouse oocytes after activation using mass spectrum. *J Assist Reprod Genet* 2012b;29:765–71. <https://doi.org/10.1007/s10815-012-9789-8>.

Pérez-Patiño C, Parrilla I, Barranco I, Vergara-Barberán M, Simó-Alfonso EF, Herrero-Martínez JM, et al. New In-Depth Analytical Approach of the Porcine Seminal

Plasma Proteome Reveals Potential Fertility Biomarkers. *J Proteome Res* 2018;17:1065–76. <https://doi.org/10.1021/acs.jproteome.7b00728>.

Pfeiffer JR, McAvoy BL, Fecteau RE, Deleault KM, Brooks SA. CARHSP1 Is Required for Effective Tumor Necrosis Factor Alpha mRNA Stabilization and Localizes to Processing Bodies and Exosomes. *Mol Cell Biol* 2011;31:277–86. <https://doi.org/10.1128/MCB.00775-10>.

Pilatz A, Lochnit G, Karnati S, Paradowska-Dogan A, Lang T, Schultheiss D, et al. Acute epididymitis induces alterations in sperm protein composition. *Fertil Steril* 2014;101:1609-1617.e5. <https://doi.org/10.1016/j.fertnstert.2014.03.011>.

Powell MD, Read KA, Sreekumar BK, Oestreich KJ. Ikaros Zinc Finger Transcription Factors: Regulators of Cytokine Signaling Pathways and CD4+ T Helper Cell Differentiation. *Front Immunol* 2019;10. <https://doi.org/10.3389/fimmu.2019.01299>.

Rakhra Gurseen, Rakhra Gurmeen. Zinc finger proteins: insights into the transcriptional and post transcriptional regulation of immune response. *Mol Biol Rep* 2021;48:5735–43. <https://doi.org/10.1007/s11033-021-06556-x>.

Rego JPA, Crisp JM, Moura AA, Nouwens AS, Li Y, Venus B, et al. Seminal plasma proteome of electroejaculated *Bos indicus* bulls. *Anim Reprod Sci* 2014;148:1–17. <https://doi.org/10.1016/j.anireprosci.2014.04.016>.

Riley LK, Rupert J. Evaluation of Patients with Leukocytosis. *Am Fam Physician* 2015;92:1004–11.

Ritchie H, Roser M. Obesity. *Our World in Data* 2020. <https://ourworldindata.org/obesity> (accessed March 21, 2021).

Robinson MB, Taylor AR, Gifondorwa DJ, Tytell M, Milligan CE. Exogenous Hsc70, but not thermal preconditioning, confers protection to motoneurons subjected to oxidative stress. *Dev Neurobiol* 2008;68:1–17. <https://doi.org/10.1002/dneu.20550>.

Roglic G, Unwin N, Bennett P, Mathers C, Tuomilehto J, Nag S, et al. The Burden of Mortality Attributable to Diabetes. *Diabetes Care* 2005;28:6.

Rusco G, Słowińska M, Di Iorio M, Cerolini S, Maffione AB, Ciereszko A, et al. Proteomic analysis of rabbit fresh and cryopreserved semen provides an important insight into molecular mechanisms of cryoinjuries to spermatozoa. *Theriogenology* 2022;191:77–95. <https://doi.org/10.1016/j.theriogenology.2022.07.018>.

Samanta L, Parida R, Dias TR, Agarwal A. The enigmatic seminal plasma: a proteomics insight from ejaculation to fertilization. *Reproductive Biology and Endocrinology* 2018;16:41. <https://doi.org/10.1186/s12958-018-0358-6>.

Sampanang A, Arun S, Sukhorum W, Burawat J, Nualkaew S, Maneenin C, et al. Antioxidant and Hypoglycemic Effects of *Momordica cochinchinensis* Spreng: (Gac) Aril Extract on Reproductive Damages in Streptozotocin (STZ)-Induced Hyperglycemia Mice. *International Journal of Morphology* 2017;35:667–75. <https://doi.org/10.4067/S0717-95022017000200046>.

Schon SB, Luense LJ, Wang X, Bartolomei MS, Coutifaris C, Garcia BA, et al. Histone modification signatures in human sperm distinguish clinical abnormalities. *J Assist Reprod Genet* 2019;36:267–75. <https://doi.org/10.1007/s10815-018-1354-7>.

Seisen T, Rouprêt M, Faix A, Droupy S. La prostate : une glande au carrefour urogénital. *Progrès En Urologie* 2012;22:S2–6. [https://doi.org/10.1016/S1166-7087\(12\)70028-3](https://doi.org/10.1016/S1166-7087(12)70028-3).

Sha Y, Xu Y, Wei X, Liu W, Mei L, Lin S, et al. CCDC9 is identified as a novel candidate gene of severe asthenozoospermia. *Syst Biol Reprod Med* 2019;65:465–73. <https://doi.org/10.1080/19396368.2019.1655812>.

Sharma R, Agarwal A, Mohanty G, Du Plessis SS, Gopalan B, Willard B, et al. Proteomic analysis of seminal fluid from men exhibiting oxidative stress. *Reproductive Biology and Endocrinology* 2013;11:85. <https://doi.org/10.1186/1477-7827-11-85>.

Sherman BT, Hao M, Qiu J, Jiao X, Baseler MW, Lane HC, et al. DAVID: a web server for functional enrichment analysis and functional annotation of gene lists (2021 update). *Nucleic Acids Res* 2022;50:W216–21. <https://doi.org/10.1093/nar/gkac194>.

Shevchenko A, Wilm M, Vorm O, Mann M. Mass Spectrometric Sequencing of Proteins from Silver-Stained Polyacrylamide Gels. *Anal Chem* 1996;68:850–8. <https://doi.org/10.1021/ac950914h>.

Shi CY, Kingston ER, Kleaveland B, Lin DH, Stubna MW, Bartel DP. The ZSWIM8 ubiquitin ligase mediates target-directed microRNA degradation. *Science* (1979) 2020;370. <https://doi.org/10.1126/science.abc9359>.

Silva HVR, Rodriguez-Villamil P, Magalhães FF de, Nunes TGP, Freitas LA de, Ribeiro LR, et al. Seminal plasma and sperm proteome of ring-tailed coatis (*Nasua nasua*, Linnaeus, 1766). *Theriogenology* 2018;111:34–42. <https://doi.org/10.1016/j.theriogenology.2017.12.036>.

Singh R, Sengar G, Singh U, Deb R, Junghare V, Hazra S, et al. Functional proteomic analysis of crossbred (Holstein Friesian × Sahiwal) bull spermatozoa. *Reproduction in Domestic Animals* 2018;53:588–608. <https://doi.org/10.1111/rda.13146>.

Skerrett-Byrne DA, Anderson AL, Hulse L, Wass C, Dun MD, Bromfield EG, et al. Proteomic analysis of koala ( *phascolarctos cinereus* ) spermatozoa and prostatic bodies. *Proteomics* 2021;21. <https://doi.org/10.1002/pmic.202100067>.

Smyth SP, Nixon B, Anderson AL, Murray HC, Martin JH, MacDougall LA, et al. Elucidation of the protein composition of mouse seminal vesicle fluid. *Proteomics* 2022;22. <https://doi.org/10.1002/pmic.202100227>.

Souza F, Chirinéa V, Martins M, Lopes M. Osteopontin in Seminal Plasma and Sperm Membrane of Dogs. *Reproduction in Domestic Animals* 2009;44:283–6. <https://doi.org/10.1111/j.1439-0531.2009.01447.x>.

Spinaci M, Volpe S, Bernardini C, De Ambrogi M, Tamanini C, Seren E, et al. Immunolocalization of heat shock protein 70 (Hsp 70) in boar spermatozoa and its role during fertilization. *Mol Reprod Dev* 2005;72:534–41. <https://doi.org/10.1002/mrd.20367>.

Springer M, Moco S. Resveratrol and Its Human Metabolites—Effects on Metabolic Health and Obesity. *Nutrients* 2019;11:143. <https://doi.org/10.3390/nu11010143>.

Tan Z, Gao L, Wang Yan, Yin H, Xi Y, Wu X, et al. PRSS contributes to cetuximab resistance in colorectal cancer. *Sci Adv* 2020;6. <https://doi.org/10.1126/sciadv.aax5576>.

The Human Protein Atlas. Coiled-Coil Domain Containing 175 n.d. The Human Protein Atlas (accessed February 6, 2024).

The UniProt Consortium. UniProt: A worldwide hub of protein knowledge. *Nucleic Acids Res* 2019;47:D506–5012. <https://doi.org/10.1093/nar/gky1049>.

Tokuhiro K, Ikawa M, Benham AM, Okabe M. Protein disulfide isomerase homolog PDILT is required for quality control of sperm membrane protein ADAM3 and male fertility. *Proceedings of the National Academy of Sciences* 2012;109:3850–5. <https://doi.org/10.1073/pnas.1117963109>.

Truebestein L, Leonard TA. Coiled-coils: The long and short of it. *BioEssays* 2016;38:903–16. <https://doi.org/10.1002/bies.201600062>.

Tschiedel B. A História do Diabetes. Sociedade Brasileira de Endocrinologia e Metabologia 2014.

Tsounapi P, Honda M, Dimitriadis F, Kawamoto B, Hikita K, Muraoka K, et al. Impact of antioxidants on seminal vesicles function and fertilizing potential in diabetic rats. *Asian J Androl* 2017;19:639. <https://doi.org/10.4103/1008-682X.186871>.

Turunen T, Magris M, Malinen M, Kekäläinen J. Seminal-Plasma-Mediated Effects on Sperm Performance in Humans. *Cells* 2022;11:2147. <https://doi.org/10.3390/cells11142147>.

Ul Hassan I, Mamoon Rehman H, Liu Z, Zhou L, Kaleem Samma M, Wang C, et al. Genome-wide identification and spatiotemporal expression profiling of zinc finger SWIM domain-containing protein family genes. *Zool Res* 2023;44:663–74. <https://doi.org/10.24272/j.issn.2095-8137.2022.418>.

Umar S, Ahmad M, Ahmad I, Zubair M, Umar Z, Qureshi AS, et al. Correlation of biochemical constituents of seminal plasma with semen quality in Teddy goat (*Capra hircus*) bucks. *Andrologia* 2018;50:e12940. <https://doi.org/10.1111/and.12940>.

Vaughan PS, van der Meijden CMJ, Aziz F, Harada H, Taniguchi T, van Wijnen AJ, et al. Cell Cycle Regulation of Histone H4 Gene Transcription Requires the Oncogenic Factor IRF-2. *Journal of Biological Chemistry* 1998;273:194–9. <https://doi.org/10.1074/jbc.273.1.194>.

Verze P, Cai T, Lorenzetti S. The role of the prostate in male fertility, health and disease. *Nat Rev Urol* 2016;13:379–86. <https://doi.org/10.1038/nrurol.2016.89>.

Wang L, Chen W, Zhao C, Huo R, Guo X-J, Lin M, et al. The role of ezrin-associated protein network in human sperm capacitation. *Asian J Androl* 2010;12:667–76. <https://doi.org/10.1038/aja.2010.79>.

Wang M, Liu X, Chang G, Chen Yidong, An G, Yan L, et al. Single-Cell RNA Sequencing Analysis Reveals Sequential Cell Fate Transition during Human Spermatogenesis. *Cell Stem Cell* 2018;23:599-614.e4. <https://doi.org/10.1016/j.stem.2018.08.007>.

Wang P, Huo HL, Wang SY, Miao YW, Zhang YY, Zhang QL, et al. Cloning, sequence characterization, and expression patterns of members of the porcine TSSK family. *Genetics and Molecular Research* 2015;14:14908–19. <https://doi.org/10.4238/2015.October.18.56>.

Wang Y-B, Yang L, Deng Y-Q, Yan S-Y, Luo L-S, Chen P, et al. Causal relationship between obesity, lifestyle factors and risk of benign prostatic hyperplasia: a univariable and multivariable Mendelian randomization study. *J Transl Med* 2022;20:495. <https://doi.org/10.1186/s12967-022-03722-y>.

Wang Z, Tufts R, Haleem R, Cai X. Genes regulated by androgen in the rat ventral prostate. *Proceedings of the National Academy of Sciences* 1997;94:12999–3004. <https://doi.org/10.1073/pnas.94.24.12999>.

Weaver EM, Zamora FJ, Puplampu-Dove YA, Kiessu E, Hearne JL, Martin-Caraballo M. Regulation of T-type calcium channel expression by sodium butyrate in prostate cancer cells. *Eur J Pharmacol* 2015;749:20–31. <https://doi.org/10.1016/j.ejphar.2014.12.021>.

Wells AD, Malkovsky M. Heat shock proteins, tumor immunogenicity and antigen presentation: an integrated view. *Immunol Today* 2000;21:129–32. [https://doi.org/10.1016/S0167-5699\(99\)01558-3](https://doi.org/10.1016/S0167-5699(99)01558-3).

Welter H, Herrmann C, Fröhlich T, Flenkenthaler F, Eubler K, Schorle H, et al. Filamin A Orchestrates Cytoskeletal Structure, Cell Migration and Stem Cell Characteristics in Human Seminoma TCam-2 Cells. *Cells* 2020;9:2563. <https://doi.org/10.3390/cells9122563>.

WHO. Diabetes 2023a. <https://www.who.int/news-room/fact-sheets/detail/diabetes> (accessed August 15, 2023).

WHO. 1 in 6 people globally affected by infertility: WHO 2023b. <https://www.who.int/news/item/04-04-2023-1-in-6-people-globally-affected-by-infertility#:~:text=Large%20numbers%20of%20people%20are,care%20for%20those%20in%20need>. (accessed August 15, 2023).

WHO. Sexual and reproductive health. World Health Organization 2018. <https://www.who.int/reproductivehealth/%0Apublications/guidelines-treatment-infertility/en/> (accessed June 12, 2021).

Wijeyesakere SJ, Bedi SK, Huynh D, Raghavan M. The C-Terminal Acidic Region of Calreticulin Mediates Phosphatidylserine Binding and Apoptotic Cell Phagocytosis. *The Journal of Immunology* 2016;196:3896–909. <https://doi.org/10.4049/jimmunol.1502122>.

Williams DB. Beyond lectins: the calnexin/calreticulin chaperone system of the endoplasmic reticulum. *J Cell Sci* 2006;119:615–23. <https://doi.org/10.1242/jcs.02856>.

Winderickx J, Hemschoote K, De Clercq N, Van Dijck P, Peeters B, Rombauts W, et al. Tissue-Specific Expression and Androgen Regulation of Different Genes Encoding Rat Prostatic 22-Kilodalton Glycoproteins Homologous to Human and Rat Cystatin. *Molecular Endocrinology* 1990;4:657–67. <https://doi.org/10.1210/mend-4-4-657>.

Wisevoter. Most Obese Countries 2023a. <https://wisevoter.com/country-rankings/most-obese-countries/#map> (accessed August 15, 2023).

Wisevoter. Diabetes Rates by Country 2023b. <https://wisevoter.com/country-rankings/diabetes-rates-by-country/> (accessed August 15, 2023).

Wu C-T, Altuwaijri S, Ricke WA, Huang S-P, Yeh S, Zhang C, et al. Increased prostate cell proliferation and loss of cell differentiation in mice lacking prostate epithelial androgen receptor. *Proceedings of the National Academy of Sciences* 2007;104:12679–84. <https://doi.org/10.1073/pnas.0704940104>.

Wu H, Liu Y, Li Y, Li K, Xu C, Gao Y, et al. DNALI1 deficiency causes male infertility with severe asthenozoospermia in humans and mice by disrupting the assembly of the flagellar inner dynein arms and fibrous sheath. *Cell Death Dis* 2023;14:127. <https://doi.org/10.1038/s41419-023-05653-y>.

Xia J, Wishart DS. Using MetaboAnalyst 3.0 for Comprehensive Metabolomics Data Analysis. *Curr Protoc Bioinformatics* 2016;55. <https://doi.org/10.1002/cpbi.11>.

Xiu-Yan Y, Gu YJ, An T, Liu JX, Pan YY, Mo FF, et al. Proteomics Analysis of Testis of Rats Fed a High-Fat Diet. *Cellular Physiology and Biochemistry* 2018;47:378–89. <https://doi.org/10.1159/000489918>.

Xu X, Wang Yaohui, Chen J, Du X, Yao L, Xu J, et al. Mutation of Serine protease 1 Induces Male Sterility in *Bombyx mori*. *Front Physiol* 2022;13. <https://doi.org/10.3389/fphys.2022.828859>.

Xue B, Wu S, Sharkey C, Tabatabaei S, Wu C-L, Tao Z, et al. Obesity-associated inflammation induces androgenic to estrogenic switch in the prostate gland. *Prostate Cancer Prostatic Dis* 2020;23:465–74. <https://doi.org/10.1038/s41391-020-0208-4>.

Yamaguchi A, Kaneko T, Inai T, Iida H. Molecular Cloning and Subcellular Localization of Tektin2-Binding Protein 1 (Ccdc 172) in Rat Spermatozoa. *Journal of Histochemistry & Cytochemistry* 2014;62:286–97. <https://doi.org/10.1369/0022155413520607>.

Yan Z, Zhou Y, Yang Y, Zeng C, Li P, Tian H, et al. Zinc finger protein 384 enhances colorectal cancer metastasis by upregulating MMP2. *Oncol Rep* 2022;47:49. <https://doi.org/10.3892/or.2022.8260>.

Yannasithinon S, Iamsaard S. Alterations of morphology and phosphorylated protein expression in the seminal vesicles of diabetic mice. *Andrologia* 2019;51. <https://doi.org/10.1111/and.13406>.

Yean Wong RL, Wang Q, Treviño LS, Bosland MC, Chen J, Medvedovic M, et al. Identification of Secretoglobin *Scgb2a1* as a target for developmental reprogramming by BPA in the rat prostate. *Epigenetics* 2015;10:127–34. <https://doi.org/10.1080/15592294.2015.1009768>.

Yeung C. Studies on the origin of redox enzymes in seminal plasma and their relationship with results of in-vitro fertilization. *Mol Hum Reprod* 1998;4:835–9. <https://doi.org/10.1093/molehr/4.9.835>.

Yeung WSB, Lee K-F, Koistinen R, Koistinen H, Seppälä M, Chiu PCN. Effects of glycodepins on functional competence of spermatozoa. *J Reprod Immunol* 2009;83:26–30. <https://doi.org/10.1016/j.jri.2009.04.012>.

Yu Z, Luo H, Fu W, Mattson MP. The Endoplasmic Reticulum Stress-Responsive Protein GRP78 Protects Neurons Against Excitotoxicity and Apoptosis: Suppression of Oxidative Stress and Stabilization of Calcium Homeostasis. *Exp Neurol* 1999;155:302–14. <https://doi.org/10.1006/exnr.1998.7002>.

Zannoni A, Bernardini C, Zaniboni A, Ferlizza E, Ventrella D, Bacci ML, et al. Relative abundance of heat shock proteins and clusterin transcripts in spermatozoa collected from boar routinely utilised in an artificial insemination centre: preliminary results. *Vet Res Commun* 2017;41:233–9. <https://doi.org/10.1007/s11259-017-9689-6>.

Zhang C, Kho Y-S, Wang Z, Chiang YT, Ng GKH, Shaw P-C, et al. Transmembrane and Coiled-Coil Domain Family 1 Is a Novel Protein of the Endoplasmic Reticulum. *PLoS One* 2014;9:e85206. <https://doi.org/10.1371/journal.pone.0085206>.

Zhang L, Qin Y, Gong X, Peng R, Cai C, Zheng Y, et al. A promoter variant in ZNF804A decreasing its expression increases the risk of autism spectrum disorder in the Han Chinese population. *Transl Psychiatry* 2019;9:31. <https://doi.org/10.1038/s41398-019-0369-x>.

Zhang X, Peng J, Wu M, Sun A, Wu X, Zheng J, et al. Broad phosphorylation mediated by testis-specific serine/threonine kinases contributes to spermiogenesis and male fertility. *Nat Commun* 2023;14:2629. <https://doi.org/10.1038/s41467-023-38357-0>.

Zhang Y, Qi J, Zhao J, Li M, Zhang Yulin, Hu H, et al. Effect of Dietetic Obesity on Testicular Transcriptome in Cynomolgus Monkeys. *Genes (Basel)* 2023;14:557. <https://doi.org/10.3390/genes14030557>.

Zhang YL, Zhou ZX, Zhang YD, Parker MG. Expression of androgen receptors and prostatic steroid-binding protein during development of the rat ventral prostate. *Journal of Endocrinology* 1988;117:361-NP. <https://doi.org/10.1677/joe.0.1170361>.

Zhao J, Dong X, Hu X, Long Z, Wang L, Liu Q, et al. Zinc levels in seminal plasma and their correlation with male infertility: A systematic review and meta-analysis. *Sci Rep* 2016;6:22386. <https://doi.org/10.1038/srep22386>.

Zini A, Lamirande E, Gagnon C. Reactive oxygen species in semen of infertile patients: levels of superoxide dismutase- and catalase-like activities in seminal plasma and spermatozoa. *Int J Androl* 1993;16:183–8. <https://doi.org/10.1111/j.1365-2605.1993.tb01177.x>.