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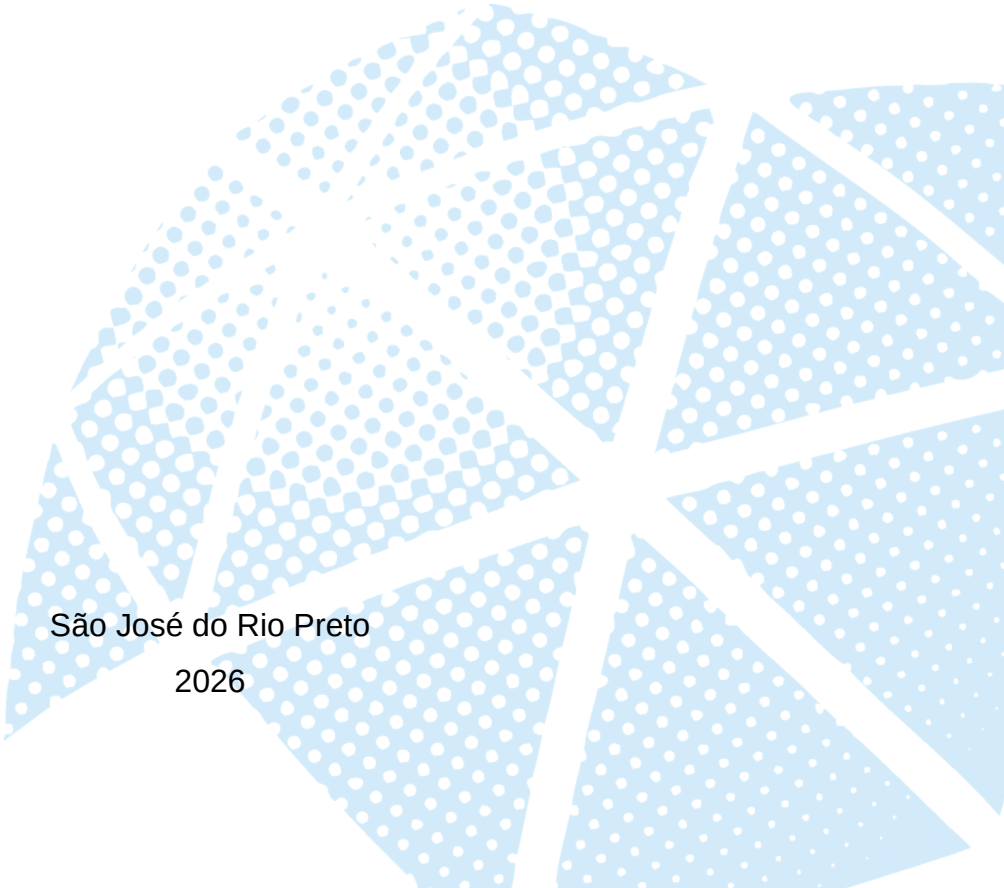
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Instituto de Biociências, Letras e Ciências Exatas - Campus de São José
do Rio Preto

PATRICK RÔMBOLA OZANIQUE

Síntese e avaliação de retrochalconas como agentes antibacterianos e
antivirais

São José do Rio Preto
2026



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**Síntese e avaliação de retrochalconas como agentes antibacterianos e
antivirais**

Tese apresentada à Universidade Estadual Paulista (UNESP), Instituto de Biociências, Letras e Ciências Exatas, São José do Rio Preto, para obtenção do título de Doutor em Química.

Área de Concentração: Química

Orientador: Prof. Dr. Luis Octavio Regasini

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IMPACTO POTENCIAL DESTA PESQUISA

O processo de mutação dos patógenos que ameaçam a saúde pública é contínuo, resultando no frequente surgimento de novas cepas resistentes aos fármacos convencionais. Dessa forma, a busca por novos agentes terapêuticos é incessante, sendo particularmente desejável a descoberta de novas classes de compostos com mecanismos de ação inovadores. No presente trabalho, são propostas sínteses inéditas e viáveis de produtos naturais de alto valor agregado e seus análogos sintéticos, os quais foram avaliados contra patógenos listados pela Organização Mundial da Saúde como prioritários para pesquisa e desenvolvimento de tratamentos alternativos. Entre esses, incluem-se microrganismos frequentemente negligenciados pela indústria farmacêutica, devido à sua ocorrência predominante em regiões subdesenvolvidas. Destaca-se, ainda, a exploração de substâncias bioativas com potencial atividade contra arbovírus endêmicos na região de São José do Rio Preto, como Zika e Chikungunya, além da investigação de seus mecanismos de ação.

POTENTIAL IMPACT OF THIS RESEARCH

The mutation process of pathogens that threaten public health is continuous, leading to the frequent emergence of new strains resistant to conventional drugs. Consequently, the search for new therapeutic agents is relentless, with a particular emphasis on discovering novel classes of compounds with innovative mechanisms of action. In this study, it was proposed unprecedented and feasible syntheses of high-value-added natural products and their synthetic analogs, which were evaluated against pathogens listed by the World Health Organization as priorities for research and the development of alternative treatments. Among these are microorganisms often neglected by the pharmaceutical industry due to their predominant occurrence in underdeveloped regions. Furthermore, this study highlights the exploration of bioactive compounds against endemic arboviruses in the São José do Rio Preto region, such as Zika and Chikungunya, as well as the investigation of their mechanisms of action.

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Banca Examinadora:

Prof. Dr. Luis Octavio Regasini – Universidade Estadual Paulista (UNESP)

Prof. Dr. Otávio Augusto Chaves – Universidade Estadual Paulista (UNESP)

Prof. Dr. Arthur Eugen Kümmerle – Universidade Rural do Rio de Janeiro (UFFRJ)

Prof. Dr. Ricardo Olímpio de Moura – Universidade Estadual da Paraíba (UEPB)

Profa. Dra. Caroline Souza de Freitas – Fundação Oswaldo Cruz (FIOCRUZ Minas)

*Dedico este trabalho à minha filha,
Emily. Minha fonte diária de vida,
amor, motivação e persistência.*

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RESUMO

As doenças infecciosas causadas por bactérias resistentes e arbovírus representam importantes desafios à saúde pública mundial, reforçando a necessidade do desenvolvimento de novos agentes terapêuticos com mecanismos de ação inovadores. Nesse contexto, retrochalconas encontradas em espécies de plantas do gênero *Glycyrrhiza* destacam-se como estruturas promissoras, devido à sua diversidade química e amplo espectro biológico. Assim, esta tese teve como objetivo desenvolver novos agentes antivirais e antibacterianos, utilizando como ponto de partida retrochalconas naturais. Foram utilizadas ferramentas da Química Medicinal, como variação de posição de substituintes, método de Topliss, isosterismo de anel e extensão do sistema π . Foram aplicadas diversas estratégias sintéticas para obtenção de cinco retrochalconas naturais e trinta e dois análogos estruturais inéditos, incluindo reações de proteção/desproteção com grupos protetores metoximetila e tetrahidropiranila, C-acilação, C-prenilação, O-prenilação, O-metilação, iodação regioseletiva, condensação de Claisen–Schmidt, acoplamento de Suzuki–Miyaura e rearranjo de Claisen. Inicialmente, a licochalcona C (LCC) foi sintetizada por uma rota de cinco etapas, com rendimento global de 13%, e avaliada frente a bactérias Gram-positivas, Gram-negativas e micobactérias, incluindo cepas resistentes aos fármacos comumente utilizados. LCC apresentou atividade contra cepas Gram-positivas ($\text{CIM}_{90} = 6.2 - 50 \mu\text{g mL}^{-1}$), *Helicobacter pylori* ($\text{CIM}_{90} = 25 \mu\text{g mL}^{-1}$) e micobactérias ($\text{CIM}_{90} = 31.2 - 125 \mu\text{g mL}^{-1}$), além de efeitos sobre formação de biofilme ($\text{CIMB}_{50} = 6.2 \mu\text{g mL}^{-1}$) e ruptura de membrana de *Staphylococcus aureus*. A avaliação *in vivo* de LCC em *Galleria mellonella* não demonstrou toxicidade sistêmica até $100 \mu\text{g mL}^{-1}$, oito vezes o CIM_{90} contra *S. aureus*. Posteriormente, uma nova rota sintética regioseletiva foi desenvolvida, permitindo a obtenção da LCC com rendimento global de 30%, além da síntese de seus análogos e do produto natural licoagrochalcona C (LACC). Para explorar outros padrões de prenilação, licochalcona E (LCE) e seus análogos foram sintetizados pelo rearranjo anormal de Claisen do produto natural xinjiachalcona A com diversos padrões de anel A, obtendo rendimentos entre 38 e 71% na conversão. As chalcones sintetizadas foram avaliadas contra arbovírus, onde LCC se destacou e apresentou valores

de CE_{50} de $6,2 \pm 0,2 \mu\text{M}$ contra o vírus Zika (ZIKV), $8,1 \pm 0,9 \mu\text{M}$ contra Mayaro e inibiu 48% da replicação do vírus Chikungunya a $10 \mu\text{M}$. As retrochalconas apresentaram efeito inibitório a atividade proteolítica de NS2B-NS3^{pro} de ZIKV, destacando os valores de CI_{50} dos produtos naturais (LCC: $53 \mu\text{M}$; LCE: $46 \mu\text{M}$; LACC: $44 \mu\text{M}$; ECH: $12 \mu\text{M}$). Os análogos de LCC e LCE também demonstraram capacidade perturbar a estabilidade térmica da NS3^{hel} de ZIKV, chegando a reduções de 9°C na temperatura de desnaturação da proteína. Nenhum dos análogos sintetizados demonstrou inibição da atividade de NS5^{RdRp} na concentração testada ($500 \mu\text{M}$). Os estudos de Relação Estrutura–Atividade com os análogos sintetizados evidenciaram a importância da presença do grupo hidroxila no anel A das retrochalconas tanto para a capacidade de inibição da NS2B-NS3^{pro} do ZIKV, quanto para a atividade anti-arboviral fenotípica. Por fim, técnicas computacionais de modelagem molecular auxiliaram no entendimento das possíveis interações entre as retrochalconas mais ativas os resíduos da NS2-NS3^{pro}, corroborando os resultados dos ensaios bioquímicos.

Palavras-chave: química medicinal; chalcona; síntese orgânica; antibacterianos; antivirais.

ABSTRACT

Infectious diseases caused by resistant bacteria and arboviruses represent important challenges to global public health, reinforcing the need for the development of new therapeutic agents with innovative mechanisms of action. In this context, retrochalcones found in plant species of the genus *Glycyrrhiza* stand out as promising structures due to their chemical diversity and broad biological spectrum. This thesis aimed to develop new antiviral and antibacterial agents, using natural retrochalcones as a starting point. Medicinal Chemistry tools were used, such as variation of substituent position, the Topliss method, ring isosterism, and extension of the π system. Several synthetic strategies were applied to obtain five natural retrochalcones and thirty-two unprecedented structural analogs, including protection/deprotection reactions with methoxymethyl and tetrahydropyranyl protecting groups, C-acylation, C-prenylation, O-prenylation, O-methylation, regioselective iodination, Claisen–Schmidt condensation, Suzuki–Miyaura coupling, and Claisen rearrangement. Initially, licochalcone C (LCC) was synthesized through a five-step route, with an overall yield of 13%, and evaluated against Gram-positive bacteria, Gram-negative bacteria, and mycobacteria, including strains resistant to commonly used drugs. LCC showed activity against Gram-positive strains ($\text{MIC}_{90} = 6.2 - 50 \mu\text{g mL}^{-1}$), *Helicobacter pylori* ($\text{MIC}_{90} = 25 \mu\text{g mL}^{-1}$), and mycobacteria ($\text{MIC}_{90} = 31.2 - 125 \mu\text{g mL}^{-1}$), in addition to effects on biofilm formation ($\text{MBIC}_{50} = 6.2 \mu\text{g mL}^{-1}$) and membrane disruption of *Staphylococcus aureus*. The *in vivo* evaluation of LCC in *Galleria mellonella* model did not demonstrate systemic toxicity up to $100 \mu\text{g mL}^{-1}$, eight times the MIC_{90} against *S. aureus*. Subsequently, a new regioselective synthetic route was developed, allowing the obtainment of LCC with an overall yield of 30%, in addition to the synthesis of its analogs and the natural product licoagrochalcone C (LACC). To explore other prenylation patterns, licochalcone E (LCE) and its analogs were synthesized through the abnormal Claisen rearrangement of the natural product xinjiachalcone A with different A-ring patterns, obtaining conversion yields ranging from 38 to 71%. The synthesized chalcones were evaluated against arboviruses, where LCC stood out and showed EC_{50} values of $6.2 \pm 0.2 \mu\text{M}$ against Zika virus (ZIKV), $8.1 \pm 0.9 \mu\text{M}$ against Mayaro virus, and inhibited 48% of Chikungunya virus replication at 10

μM . The retrochalcones showed an inhibitory effect on the proteolytic activity of ZIKV NS2B-NS3^{pro}, with the IC_{50} values of the natural products standing out (LCC: 53 μM ; LCE: 46 μM ; LACC: 44 μM ; ECH: 12 μM). LCC and LCE analogs also demonstrated the ability to disturb the thermal stability of ZIKV NS3^{hel}, reaching reductions up to 9 °C in the protein denaturation temperature. None of the synthesized analogs demonstrated inhibition of NS5^{RdRp} activity at the tested concentration (500 μM). Structure–Activity Relationship studies with the synthesized analogs demonstrated the importance of the presence of the hydroxyl group on the A ring of retrochalcones for both their ability to inhibit ZIKV NS2B-NS3^{pro} and their phenotypic anti-arboviral activity. Finally, computational molecular modeling techniques assisted in understanding the possible interactions between the most active retrochalcones and the residues of NS2B-NS3^{pro}, corroborating the results of the biochemical assays.

Keywords: medicinal chemistry; chalcone; organic synthesis; antibacterial; antiviral.

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LIST OF ABBREVIATIONS

AMPK	AMP-activated protein kinase
AMR	Antimicrobial resistance
ATCC	American Type Culture Collection
Bcl-2	B-cell lymphoma 2
CC₅₀	Half-maximal cytotoxic concentration
CHIKV	Chikungunya virus
COX	Cyclooxygenase
DMSO	Dimethyl sulfoxide
EC₅₀	Half-maximal effective concentration
ECH	Echinatin
eEF2	Eukaryotic elongation factor 2
EtOH	Ethanol
Ger	Geranyl
GI₅₀	Half-maximal growth inhibition
HPLC	High-performance liquid chromatography
IC₅₀	Half-maximal inhibitory concentration
ID₅₀	Half-maximal inhibition dose
LACC	Licoagrochalcone C
LC	Licochalcones
LCA	Licochalcone A
LCC	Licochalcone C
LCE	Licochalcone E
LOX	Lipoxygenase
MAYV	Mayaro virus
MBC	Minimum bactericidal concentration
MBIC	Minimum biofilm inhibitory concentration
MCF-7	Michigan Cancer Foundation-7 breast cancer cell line
Me	Methyl group
MFC	Minimum fungicidal concentration
MIC	Minimum inhibitory concentration
MIC₅₀	Minimum inhibitory concentration for 50%
MIC₉₀	Minimum inhibitory concentration for 90%

MMP	Matrix metalloproteinase
MOM	Methoxymethyl ether
MPO	Myeloperoxidase
MRSA	Methicillin-resistant <i>Staphylococcus aureus</i>
MS	Mass spectrometry
MSSA	Methicillin-susceptible <i>Staphylococcus aureus</i>
NMR	Nuclear magnetic resonance
NO	Nitric oxide
NS2B-NS3^{pro}	NS2B-NS3 protease
NS3^{hel}	NS3 helicase
NS5^{RdRp}	NS5 RNA-dependent RNA polymerase
NSP	Non-structural protein
NTDs	Neglected tropical diseases
OH	Hydroxyl group
OMe	Methoxyl group
PAINS	Pan-assay interference compounds
PDB	Protein Data Bank
Ph	Phenyl
ROS	Reactive oxygen species
SAR	Structure-activity relationship
SI	Selectivity index
THF	Tetrahydrofuran
THP	Tetrahydropyran
TLC	Thin-layer chromatography
WHO	World Health Organization
XCA	Xinjiachalcone A
ZIKV	Zika virus

LIST OF SYMBOLS

α	Alpha
β	Beta
Δ	Delta (variation)
δ	Delta (chemical shift)
π	Pi (electron system)
λ	Lambda (wavelength)
$^{\circ}\text{C}$	Degree Celsius
s	Second
h	Hour
mg	Milligram
μg	Microgram
g	Gram
mL	Milliliter
L	Liter
M	Molar concentration
mmol	Milimole
μmol	Micromole
MHz	Megahertz
<i>J</i>	Coupling constant
<i>o</i>	<i>ortho</i> position
<i>m</i>	<i>meta</i> position
<i>p</i>	<i>para</i> position
<i>E</i>	<i>Entgegen</i> (<i>Trans</i> configuration)
<i>R</i>	<i>Rectus</i> configuration
<i>S</i>	<i>Sinister</i> configuration

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1 INTRODUCTION

1.1 GENERAL INTRODUCTION AND OVERVIEW

A long-term analysis of World Health Organization (WHO) communications highlights the urgent need to address Neglected Tropical Diseases (NTDs), which remain highly prevalent in underdeveloped countries and impoverished regions where access to basic healthcare and sanitation is severely limited [1]. The WHO's Immunization Agenda 2023 represents a significant initiative aimed at raising scientific awareness and promoting global collaboration against these widespread health challenges [2]. However, progress remains slow, failing to match the severity and high mortality rates associated with NTDs [3]. An obvious example of this is *Mycobacterium tuberculosis*, the pathogen responsible for tuberculosis, which, despite being identified in 1882, remains one of the leading causes of death worldwide [4]. Regardless of the emergence and decline of COVID-19, tuberculosis remains the world's deadliest infectious disease, continuing to pose a major global health threat [5].

Among priority pathogens, resistant bacteria, *Mycobacterium* species, and arboviruses stand out as key targets in the research and development of novel drugs and vaccines due to their high capacity to emerge as new epidemics or pandemics [6, 7]. However, overcoming resistance mechanisms and adapting to rapid viral mutations remain significant challenges in drug discovery [8, 9]. In this context, natural compounds and their structural inspiration have emerged as promising strategies, given their chemical diversity, which enables the discovery of broad-spectrum bioactive molecules with diverse mechanisms of action [10].

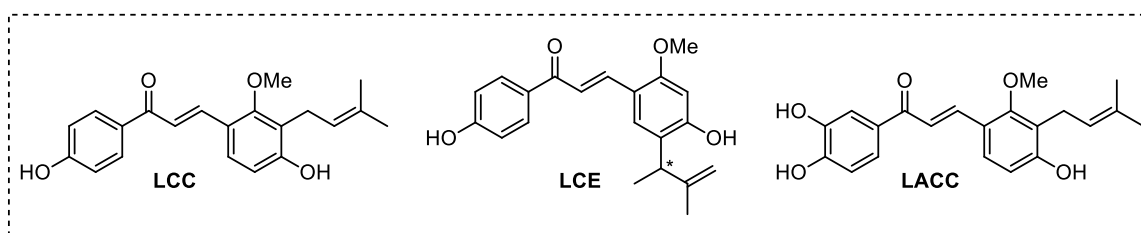
Flavonoids are a rich class of natural products widely distributed in plants across the globe, characterized by diverse structural patterns and a broad spectrum of bioactivities [11]. Among them, retrochalcones stand out as a less-explored subgroup with significant biological potential. These compounds feature an open-chain structure, consisting of two aryl rings (A and B) connected by an α,β -unsaturated ketone bridge, along with a distinct oxidation pattern that contributes to their unique pharmacological properties [12].

Licochalcones derived from *Glycyrrhiza* species have recently gained growing attention in biomedical research, particularly licochalcone A (LCA), which is currently undergoing clinical trials for its anti-inflammatory and antimicrobial

properties [13]. In contrast, other licochalcones remain far less studied, despite preliminary evidence of a wide range of bioactivities. This underexplored potential offers a valuable opportunity for further investigation – both of the natural products and their structural analogs. Motivated by this, our research group has dedicated efforts and resources to evaluating the antibacterial and anti-arboviral properties of licochalcones [14].

This thesis is structured as a collection of interconnected studies focused on novel synthetic strategies and biological evaluation of licochalcone C (LCC), licochalcone E (LCE), and licoagrochalcone C (LACC) (Figure 1). These retrochalcones are found in *Glycyrrhiza* species, presenting limited reports of synthesis to date, typically yielding low efficiency, despite its promising potential against microbial infections and multiple unexplored applications.

Figure 1. Chemical structure of LCC, LCE, and LACC.



Source: Prepared by the author.

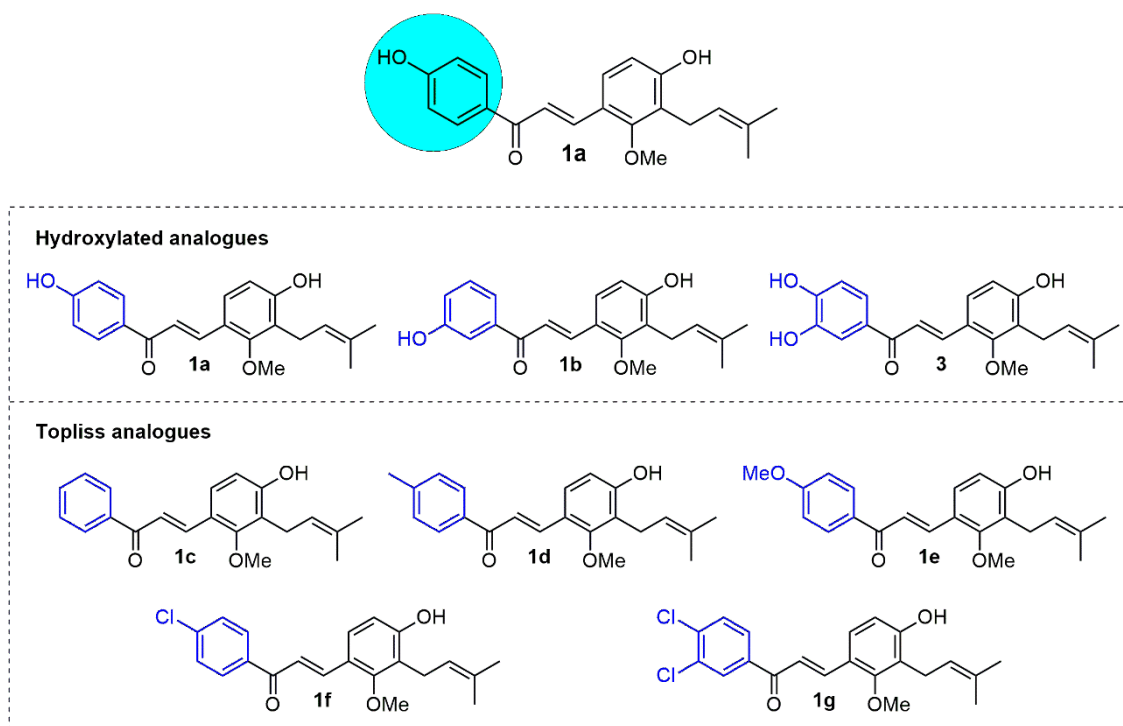
The project was initially conceived with a focus on the antibacterial potential of LCC, motivated by the urgent need for new agents capable of overcoming resistance in clinically relevant bacterial pathogens. The selection of LCC was based on preliminary evidence of antimicrobial activity within prenylated chalcones [15, 16], combined with its relatively unexplored synthetic accessibility and structural features susceptible to modifications.

Early biological results confirmed the antibacterial potential of LCC, particularly against Gram-positive strains. These results provided initial insights into its distinct mechanism of action, as it was able to inhibit the growth of both susceptible and resistant to methicillin strains of *Staphylococcus aureus* with the same potency. The following steps were the evaluation of LCC against biofilm formation, its synergy with reference drugs, investigations towards its mechanism of action, and its systemic toxicity using *Galleria mellonella* larvae model [17].

The synthetic method described for LCC presented some limitations, not being feasible with higher scales, which limited the obtainment and evaluation of the chalcone with ease. Because of this, a new strategy was developed aiming at a mass-scalable and versatile synthesis able to afford structural analogs of LCC.

After optimizing a regioselective synthesis to afford LCC with higher yields and accumulate a considerable quantity of the compound to perform biological trials, structural analogs were synthesized (Figure 2). These analogs were based both on position modifications of the hydroxyl group and the Topliss manual method, to evaluate physicochemical parameters, such as hydrophobicity, and electronic and steric features, performing the modifications on the A ring to preserve the distinct pattern of the retrochalcone. Interestingly, the Topliss modifications resulted in a complete loss of antibacterial activity.

Figure 2. Analog design on LCC series, based on modification on hydroxyl position and the Topliss method.



Source: Prepared by the author.

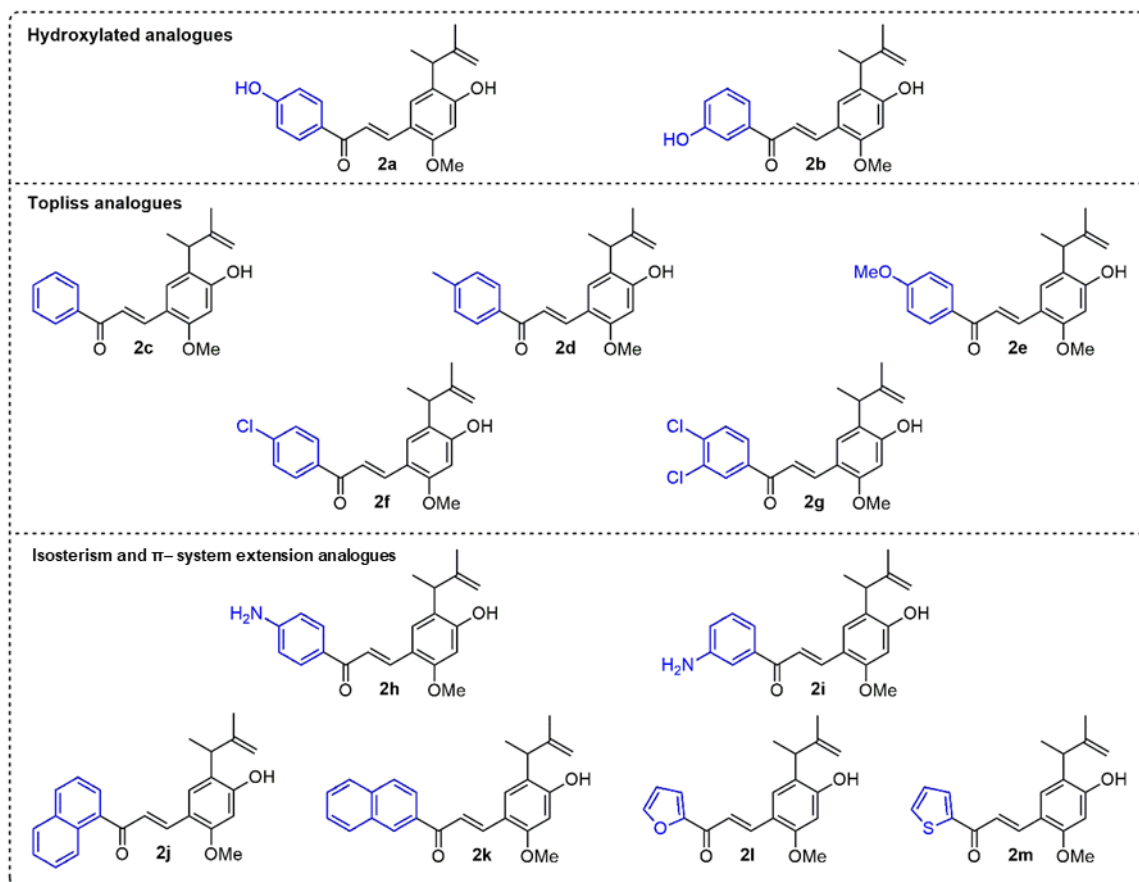
Considering that the selected modification pattern did not exert an improvement in antibacterial effects, the compounds were submitted to other biological screenings. Inspired in the growing interest in arboviral diseases and

the structural features that indicated the potential of licochalcones, initial assays were performed and led to promising results in phenotypic assays of replication inhibition of arboviruses. One more time LCC stood out in comparison to its analogs and demonstrated a broad-spectrum anti-arboviral effect, inhibiting the replication of ZIKV, MAYV, and CHIKV. Taking advantage of the broad antiviral activity of LCC, Section 4 explored further effects on non-structural proteins of ZIKV, and dose-response assays on phenotypic model. However, distinct substitution patterns on its A ring resulted in different selective patterns against those arboviruses.

The promising licochalcone pattern against non-structural proteins of ZIKV, highlighting NS2B-NS3^{pro}, inspired the synthesis of other natural products. As LCA is the most explored retrochalcone reported to date, it was decided to evaluate a different prenylation pattern, and LCE was chosen as a second prototype in this work, containing the only prenyl group with an asymmetric carbon. To this end, other synthetic strategies were explored, and it was successfully obtained through Claisen-rearrangement as a racemate. The concise and versatile synthesis allowed diverse structural modifications, which were planned after an initial screening of LCE and its structural analogs containing Topliss substituents (*p*-Me, *p*-OMe, *p*-Cl, *m,p*-diCl, and the unsubstituted aryl ring).

Preliminary results indicated, again, the importance of the hydroxyl group on the A ring, but also indicated high potential of the analog with unsubstituted ring as a NS2B-NS3^{pro} inhibitor. Because of this, modifications on the position of the OH group were performed, as well as its substitution with an NH₂ group. Additionally, analogs with naphthyl, furyl, and thienyl patterns were also synthesized to identify ring bioisosterism. The structure-activity (SAR) studies on LCE pattern were then established with 12 synthetic analogs (Figure 3).

Figure 3. Analog design on LCE series, based on modification on hydroxyl position, the Topliss method, isosterism (ring and hydroxyl group), and π -system extension.



Source: Prepared by the author.

Collectively, it was identified that the hydroxyl moiety on the A ring from both LCC and LCE poses an important influence in antibacterial and antiviral activities. Because of this, analogs of LCC were synthesized modifying the position and number of hydroxyl groups: *m*-OH and *p,m*-diOH. The dihydroxylated analog presents a catechol pattern on ring A, and is a known natural product, LACC, which demonstrated greater potency than LCC and its *m*-OH analog, both in ZIKV protease inhibition and in replication inhibition on cellular assays, resulting in Section 6.

Additionally, the synthesis of other compounds and other bioassays were performed along the development of the project. They were summarized in Section 7 as “additional results” and will integrate further investigations carried out by the research group in future projects.

- **Section II** presents a comprehensive review of natural and synthetic chalcones and their biological activities, highlighting the broad-spectrum effects of this class of compounds from the perspective of Brazilian research.
- **Section III** presents an evaluation of LCC against critical bacterial pathogens, including drug-resistant strains. The compound demonstrated activity against Gram-positive bacteria, *Mycobacterium tuberculosis*, and *Helicobacter pylori*, and its mechanism of action was elucidated. Additionally, the systemic toxicity of LCC in *Galleria mellonella* model was assessed.
- **Section IV** details the development of a new regioselective synthetic route to LCC and novel analogs, achieving the highest yield reported in the literature. These chalcones were assessed for anti-arboviral activity, with the natural prototype exhibiting remarkable activity against Chikungunya virus (CHIKV), Mayaro virus (MAYV), and Zika virus (ZIKV) and targeting ZIKV NS2B-NS3^{pro} and NS3^{hel}.
- **Section V** explores LCE as an inhibitor of ZIKV NS2B-NS3^{pro}, demonstrating the SAR of the natural product and its structural analogs towards their protease inhibition. The importance of the presence and position of hydroxyl group to enhance the inhibition was highlighted.
- **Section VI** demonstrates the influence of number and position of hydroxyl groups in the licochalcone scaffold. To this end, LACC was highlighted as an effective antiviral agent, presenting enhanced potency in comparison to LCC.
- **Section VII** presents general conclusions towards the synthesis and antiviral and antibacterial effects of licochalcones. Additionally, results that were not organized as publications and will be further explored were summarized.
- **Section VIII** provides the supplementary material, containing raw in silico data, ¹H and ¹³C NMR and MS spectra for synthesized compounds.

1.1.1 General aim

To design, synthesize, and evaluate natural retrochalcones (LCC, LCE, and LACC) and its structural analogs as potential antibacterial and anti-arboviral agents against clinically relevant pathogens.

1.1.2 Specific aims

- To develop efficient and regioselective synthetic routes to obtain LCC, LCE, and LACC.
- To synthesize and structurally characterize novel retrochalcone analogs inspired on natural scaffolds.
- To evaluate the antibacterial activity of the synthesized compounds against Gram-positive, Gram-negative, and *Mycobacterium* strains.
- To investigate the effects of the most active antibacterial retrochalcones on bacterial biofilm formation, membrane disruption, and synergism with conventional drugs.
- To evaluate the *in vivo* systemic toxicity of LCC using *Galleria mellonella* larvae model.
- To evaluate the antiviral activity of retrochalcones against arboviruses, including MAYV, CHIKV, and ZIKV.
- To investigate the inhibitory effect of selected retrochalcones against viral non-structural proteins.
- To explore SAR associated with antibacterial and antiviral activities within licochalcone scaffolds.
- To compile and disseminate the most relevant findings through scientific publications, highlighting the potential of licochalcones as broad-spectrum antimicrobial agents.

In this thesis, a comprehensive bibliographic survey highlighted the relevance of chalcones as broad-spectrum bioactive agents, with special emphasis on the contributions of Brazilian research to the chemistry and pharmacology of this flavonoid class. Inspired on that, thirty-seven compounds were synthesized, including five naturally occurring retrochalcones (LCC, LCE, LACC, XCA and ECH) and thirty-two unprecedented synthetic analogs. These compounds were evaluated for their antibacterial and antiviral activities, as well as in parallel antiparasitic screenings. Collectively, the results demonstrated the potential of retrochalcones from *Glycyrrhiza* species as promising scaffolds for the development of novel antimicrobial agents. It involves synthetic organic chemistry, biological evaluation, and SAR investigation.

Initially, LCC was identified as an important antibacterial prototype, exhibiting activity against clinically relevant Gram-positive bacteria, including MRSA. Further investigations indicated that membrane disruption and inhibition of biofilm formation are important antibacterial effects of LCC. *In vivo* toxicity assays using *Galleria mellonella* larvae model demonstrated a favorable preliminary safety profile.

After that, the development of a new regioselective synthetic route to LCC was achieved and enabled the preparation of multiple structural analogs, including LACC. Biological screening of these derivatives demonstrated the importance of hydroxylated substitution patterns within the LCC scaffold. LACC also demonstrated that increasing the hydroxylation pattern of the aromatic ring enhanced antiviral potency in both enzymatic and cellular assays. The antiviral investigations revealed that LCC displayed broad-spectrum anti-arboviral activity against endemic arboviruses, including ZIKV, MAYV, and CHIKV. The observed effects on viral non-structural proteins demonstrated that LCC may act through distinct antiviral mechanisms

The synthesis of LCE was performed through an innovative approach based on the rearrangement of a natural precursor (XCA), affording a racemic mixture. The evaluation of LCE and its novel analogs further expanded the understanding of SAR within prenylated retrochalcones. The results corroborated the importance of the 4'-hydroxyl group for ZIKV NS2B-NS3^{pro} inhibition.

Collectively, the compilation and dissemination of these results demonstrate that retrochalcones constitute versatile and biologically relevant scaffolds, capable of acting against bacterial and viral pathogens through multiple and distinct mechanisms of action. Furthermore, the synthetic methodologies developed enabled efficient obtainment to structurally diverse analogs, open avenues to future medicinal chemistry optimization studies.

REFERENCES

1. Hasso-Agopsowicz M, Hwang A, Holm-Delgado M-G, et al (2024) Identifying WHO global priority endemic pathogens for vaccine research and development (R&D) using multi-criteria decision analysis (MCDA): an objective of the Immunization Agenda 2030. *EBioMedicine* 110:105424. <https://doi.org/10.1016/j.ebiom.2024.105424>
2. O'Brien KL, Lemango E, Nandy R, Lindstrand A (2024) The immunization Agenda 2030: A vision of global impact, reaching all, grounded in the realities of a changing world. *Vaccine* 42:S1–S4. <https://doi.org/10.1016/j.vaccine.2022.02.073>
3. Weng H-B, Chen H-X, Wang M-W (2018) Innovation in neglected tropical disease drug discovery and development. *Infect Dis Poverty* 7:67. <https://doi.org/10.1186/s40249-018-0444-1>
4. Riccardi N, Canetti D, Martini M, et al (2020) The evolution of a neglected disease: tuberculosis discoveries in the centuries. *J Prev Med Hyg* 61:E9–E12. <https://doi.org/10.15167/2421-4248/jpmh2020.61.1s1.1353>
5. Goletti D, Meintjes G, Andrade BB, et al (2025) Insights from the 2024 WHO Global Tuberculosis Report – More Comprehensive Action, Innovation, and Investments required for achieving WHO End TB goals. *International Journal of Infectious Diseases* 150:107325. <https://doi.org/10.1016/j.ijid.2024.107325>
6. Ukoaka BM, Okesanya OJ, Daniel FM, et al (2024) Updated WHO list of emerging pathogens for a potential future pandemic: Implications for public health and global preparedness. *Infezioni in Medicina* 4:. <https://doi.org/10.53854/liim-3204-5>
7. Jesudason T (2024) WHO publishes updated list of bacterial priority pathogens. *Lancet Microbe* 5:100940. <https://doi.org/10.1016/j.lanmic.2024.07.003>
8. Singh SP, Qureshi A, Hassan W (2021) Mechanisms of action by antimicrobial agents: A review. *McGill Journal of Medicine* 19:. <https://doi.org/10.26443/mjm.v19i1.217>
9. von Delft A, Hall MD, Kwong AD, et al (2023) Accelerating antiviral drug discovery: lessons from COVID-19. *Nat Rev Drug Discov* 22:585–603. <https://doi.org/10.1038/s41573-023-00692-8>

10. Atanasov AG, Zotchev SB, Dirsch VM, et al (2021) Natural products in drug discovery: advances and opportunities. *Nat. Rev. Drug Discov.* 20:200–216
11. Badshah SL, Faisal S, Muhammad A, et al (2021) Antiviral activities of flavonoids. *Biomedicine and Pharmacotherapy* 140
12. Malaník M (2025) Natural retrochalcones: rare compounds with diverse biological activities. *Phytochemistry Reviews*. <https://doi.org/10.1007/s11101-025-10103-y>
13. Li M-T, Xie L, Jiang H-M, et al (2022) Role of Licochalcone A in Potential Pharmacological Therapy: A Review. *Front Pharmacol* 13:. <https://doi.org/10.3389/fphar.2022.878776>
14. Mittal A, Kakkar R (2021) Synthetic methods and biological applications of retrochalcones isolated from the root of *Glycyrrhiza* species: A review. *Results Chem* 3:100216. <https://doi.org/10.1016/j.rechem.2021.100216>
15. Assis LR de, Theodoro R dos S, Costa MBS, et al (2022) Antibacterial Activity of Isobavachalcone (IBC) Is Associated with Membrane Disruption. *Membranes (Basel)* 12:269. <https://doi.org/10.3390/membranes12030269>
16. Tempone AG, Theodoro R dos S, Romanelli MM, et al (2022) A new reduced chalcone-derivative affects the membrane permeability and electric potential of multidrug-resistant *Enterococcus faecalis*. *Chem Biol Interact* 365:110086. <https://doi.org/10.1016/j.cbi.2022.110086>
17. Ozanique PR, Helena AL, Menezes R de P, et al (2024) Synthesis, Antibacterial Effects, and Toxicity of Licochalcone C. *Pharmaceuticals* 17:. <https://doi.org/10.3390/ph17050634>
18. Batovska DI, Todorova IT (2010) Trends in Utilization of the Pharmacological Potential of Chalcones
19. Sahu NK, Balbhadra SS, Choudhary J, Kohli D V (2012) Exploring Pharmacological Significance of Chalcone Scaffold: A Review
20. Santos AM, Carvalho Nascimento Júnior JA, Silva Cezar SV, et al (2024) Exploring the therapeutic potential and chemical properties of trans-chalcone: a comprehensive review. *Future Med Chem* 16:2547–2562. <https://doi.org/10.1080/17568919.2024.2424148>
21. Miranda-Sapla MM, Tomiotto-Pellissier F, Assolini JP, et al (2019) trans-Chalcone modulates *Leishmania amazonensis* infection in vitro by Nrf2 overexpression affecting iron availability. *Eur J Pharmacol* 853:275–288. <https://doi.org/10.1016/j.ejphar.2019.03.049>

22. Siqueira E da S, Concato VM, Tomiotto-Pellissier F, et al (2021) Trans-chalcone induces death by autophagy mediated by p53 up-regulation and β -catenin down-regulation on human hepatocellular carcinoma HuH7.5 cell line. *Phytomedicine* 80:153373. <https://doi.org/10.1016/j.phymed.2020.153373>
23. Piva M, Yaekashi KM, Pereira TGO, et al (2026) Trans-Chalcone alleviates overt pain-like behavior by targeting the activation of nociceptive neuron TRPV1 and TRPA1 channels. *Inflammopharmacology* 34:1213–1232. <https://doi.org/10.1007/s10787-025-02099-w>
24. Mellado M, Sariego-Kluge R, Valdés-Navarro F, et al (2023) Synthesis of fluorescent chalcones, photophysical properties, quantitative structure-activity relationship and their biological application. *Spectrochim Acta A Mol Biomol Spectrosc* 291:. <https://doi.org/10.1016/j.saa.2023.122332>
25. Wangngae S, Pewklang T, Chansaenpak K, et al (2021) A chalcone-based fluorescent responsive probe for selective detection of nitroreductase activity in bacteria. *New Journal of Chemistry* 45:11566–11573. <https://doi.org/10.1039/d1nj01794b>
26. Pelosi AG, Silveira-Alves E, Cocca LHZ, et al (2023) Two-Photon Absorption and Multiphoton Excited Fluorescence of Acetamide-Chalcone Derivatives: The Role of Dimethylamine Group on the Nonlinear Optical and Photophysical Properties. *Molecules* 28:. <https://doi.org/10.3390/molecules28041572>
27. Kaide S, Watanabe H, Ikuni S, et al (2022) Chalcone Analogue as New Candidate for Selective Detection of α -Synuclein Pathology. *ACS Chem Neurosci* 13:16–26. <https://doi.org/10.1021/acscchemneuro.1c00441>
28. Zhuang C, Zhang W, Sheng C, et al (2017) Chalcone: A Privileged Structure in Medicinal Chemistry. *Chem. Rev.* 117:7762–7810
29. Matos MJ, Vazquez-Rodriguez S, Uriarte E, Santana L (2015) Potential pharmacological uses of chalcones: A patent review (from June 2011-2014). *Expert Opin. Ther. Pat.* 25:351–366
30. Salehi B, Quispe C, Chamkhi I, et al (2021) Pharmacological Properties of Chalcones: A Review of Preclinical Including Molecular Mechanisms and Clinical Evidence. *Front. Pharmacol.* 11
31. de Mello MVP, Abrahim-Vieira B de A, Domingos TFS, et al (2018) A comprehensive review of chalcone derivatives as antileishmanial agents. *Eur. J. Med. Chem.* 150:920–929

32. de Oliveira AS, Cenci AR, Gonçalves L, et al (2023) Chalcone Derivatives as Antibacterial Agents: An Updated Overview. *Curr Med Chem* 31:2314–2329. <https://doi.org/10.2174/0929867330666230220140819>
33. Gomes MN, Muratov EN, Pereira M, et al (2017) Chalcone derivatives: Promising starting points for drug design. *Molecules* 22
34. Yang T, Liu D, Li Y, et al (2023) Chemoproteomics reveals Sofalcone inhibits the inflammatory response of Caco-2 cells by covalently targeting HMGB1. *Chemical Communications* 59:8981–8984. <https://doi.org/10.1039/d3cc00577a>
35. Kim W, Lee H, Kim S, et al (2019) Sofalcone, a gastroprotective drug, covalently binds to KEAP1 to activate Nrf2 resulting in anti-colitic activity. *Eur J Pharmacol* 865:. <https://doi.org/10.1016/j.ejphar.2019.172722>
36. Helvacı Ö, Helvacı B (2023) A Story of Serendipities: From Phlorizin to Gliflozins. *Experimental and Clinical Transplantation* 21:105–108. <https://doi.org/10.6002/ect.IAHNCongress.25>
37. O'Hara D V., Lam CSP, McMurray JJV, et al (2024) Applications of SGLT2 inhibitors beyond glycaemic control. *Nat. Rev. Nephrol.* 20:513–529
38. Medina-Alarcón KP, L Singulani J de, Dutra LA, et al (2020) Antifungal Activity of 2'-Hydroxychalcone Loaded in Nanoemulsion Against *Paracoccidioides* Spp. *Future Microbiol* 15:21–33. <https://doi.org/10.2217/fmb-2019-0095>
39. Ribeiro Lima FR, Figueiredo LC de, Oliveira Braga AR, et al (2025) Antimicrobial and anti-biofilm activity of a mucoadhesive hydrogel functionalized with aminochalcone on titanium surfaces and in *Galleria mellonella* model: In vitro and in vivo study. *Microb Pathog* 200:107286. <https://doi.org/10.1016/j.micpath.2025.107286>
40. Acho L, Neto SF, Barros A, et al (2026) Polymeric nanoparticles containing 4-methoxychalcone attenuates hyperglycemia in diabetes-induced mice. *Nanomed J* 13:159–169. <https://doi.org/10.22038/NMJ.2025.89217.2260>
41. Maschio-Lima T, Lemes TH, Marques MDR, et al (2025) Synergistic activity between conventional antifungals and chalcone-derived compound against dermatophyte fungi and *Candida* spp. *International Microbiology* 28:265–275. <https://doi.org/10.1007/s10123-024-00541-7>
42. Bila NM, Costa-Orlandi CB, Vaso CO, et al (2021) 2-Hydroxychalcone as a Potent Compound and Photosensitizer Against Dermatophyte Biofilms. *Front Cell Infect Microbiol* 11:. <https://doi.org/10.3389/fcimb.2021.679470>

43. Rozmer Z, Perjési P (2016) Naturally occurring chalcones and their biological activities. *Phytochemistry Reviews* 15:87–120
44. Machado TDB, Leal ICR, Kuster RM, et al (2005) Brazilian phytopharmaceuticals - Evaluation against hospital bacteria. *Phytotherapy Research* 19:519–525. <https://doi.org/10.1002/ptr.1696>
45. Brandão GC, Kroon EG, Duarte MGR, et al (2010) Antimicrobial, antiviral and cytotoxic activity of extracts and constituents from *Polygonum spectabile* Mart. *Phytomedicine* 17:926–929. <https://doi.org/10.1016/j.phymed.2010.03.004>
46. Siqueira EP, Oliveira DM, Johann S, et al (2011) Bioactivity of the compounds isolated from *Blepharocalyx salicifolius*. *Revista Brasileira de Farmacognosia* 21:645–651. <https://doi.org/10.1590/S0102-695X2011005000111>
47. De Castro CCB, Costa PS, Laktin GT, et al (2015) Cardamonin, a schistosomicidal chalcone from *Piper aduncum* L. (Piperaceae) that inhibits *Schistosoma mansoni* ATP diphosphohydrolase. *Phytomedicine* 22:921–928. <https://doi.org/10.1016/j.phymed.2015.06.009>
48. de Carvalho LSA, Silva LM, de Souza VC, et al (2021) Cardamonin Presents in Vivo Activity against *Schistosoma mansoni* and Inhibits Potato Apyrase. *Chem Biodivers* 18:. <https://doi.org/10.1002/cbdv.202100604>
49. Nesello LAN, Campos A, Wagner T, et al (2016) Chemical Composition and Antinociceptive Potential of *Campomanesia reitziana* Fruits. *J Med Food* 19:518–520. <https://doi.org/10.1089/jmf.2015.0092>
50. Cury BJ, Boeing T, Somensi LB, et al (2022) Dimethyl Cardamonin from Fruits of *Campomanesia reitziana* D. Legrand Promotes Gastroprotection and Gastric Healing Effects in Rodents. *Chem Biodivers* 19:. <https://doi.org/10.1002/cbdv.202200727>
51. Rossette MC, Moraes DC, Sacramento EK, et al (2017) The In Vitro and In Vivo Antiangiogenic Effects of Flavokawain B. *Phytotherapy Research* 31:1607–1613. <https://doi.org/10.1002/ptr.5891>
52. Rodrigues DF, Maniscalco DA, Silva FAJ, et al (2017) Trypanocidal Activity of Flavokawin B, a Component of *Polygonum ferrugineum* Wedd. *Planta Med* 83:239–244. <https://doi.org/10.1055/s-0042-112031>
53. Rezende-Júnior LM, Andrade LM de S, Leal ALAB, et al (2020) Chalcones isolated from *Arrabidaea brachypoda* flowers as inhibitors of nora and mepa

- multidrug efflux pumps of *Staphylococcus aureus*. *Antibiotics* 9:1–12. <https://doi.org/10.3390/antibiotics9060351>
54. Lima EM, Fernando LM, Felix LP, et al (2021) First complete NMR data and theoretical study of an antimicrobial formylated dihydrochalcone from *Psidium guineense* Sw. *Nat Prod Res* 36:419–423. <https://doi.org/10.1080/14786419.2020.1771709>
 55. Vieira NC, Espíndola LS, Santana JM, et al (2008) Trypanocidal activity of a new pterocarpan and other secondary metabolites of plants from Northeastern Brazil flora. *Bioorg Med Chem* 16:1676–1682. <https://doi.org/10.1016/j.bmc.2007.11.027>
 56. Cursino LMC, Lima NM, Murillo R, et al (2016) Isolation of flavonoids from *deguelia duckeana* and their effect on cellular viability, AMPK, eEF2, eIF2 and eIF4E. *Molecules* 21:. <https://doi.org/10.3390/molecules21020192>
 57. Lima NM, Cursino-Hron LM, Lima AM, et al (2018) Antifungal activity of extracts and phenolic compounds from *Deguelia duckeana*. *Revista Brasileira de Farmacognosia* 28:697–702. <https://doi.org/10.1016/j.bjp.2018.08.004>
 58. Souza RL, Gonçalves UO, Badoco FR, et al (2017) Licochalcone A induces morphological and biochemical alterations in *Schistosoma mansoni* adult worms. *Biomedicine and Pharmacotherapy* 96:64–71. <https://doi.org/10.1016/j.biopha.2017.09.128>
 59. da Silva Landim EMBM, Ruiz ALTG, de Carvalho JE, et al (2021) Antiproliferative activity and chemical constituents of *Lonchocarpus cultratus* (Fabaceae). *Nat Prod Res* 35:2056–2059. <https://doi.org/10.1080/14786419.2019.1647427>
 60. Mottin M, Caesar LK, Brodsky D, et al (2022) Chalcones from *Angelica keiskei* (ashitaba) inhibit key Zika virus replication proteins. *Bioorg Chem* 120:. <https://doi.org/10.1016/j.bioorg.2022.105649>
 61. De RJ, Albuquerque M, Kalyne L, et al Chalcones from *Myracrodruon urundeuva* are efficacious in guinea pig ovalbumin-induced allergic conjunctivitis. *Revista Brasileira de Farmacognosia Brazilian Journal of Pharmacognosy* 21:953–962. <https://doi.org/10.1590/S0102>
 62. Nobre-Júnior H V., Oliveira RA, Maia FD, et al (2009) Neuroprotective effects of chalcones from *myracrodruon urundeuva* on 6-hydroxydopamine-induced cytotoxicity in rat mesencephalic cells. *Neurochem Res* 34:1066–1075. <https://doi.org/10.1007/s11064-008-9876-5>

63. Sarria ALF, Silva TL, de Oliveira JM, et al (2018) Dimeric chalcones derivatives from *Myracrodruon urundeuva* act as cathepsin V inhibitors. *Phytochemistry* 154:31–38. <https://doi.org/10.1016/j.phytochem.2018.06.009>
64. Moreira BO, Vilar VLS, de Almeida RNS, et al (2022) New dimer and trimer of chalcone derivatives from anti-inflammatory and antinociceptive extracts of *Schinopsis brasiliensis* roots. *J Ethnopharmacol* 289:. <https://doi.org/10.1016/j.jep.2022.115089>
65. da Silva EP, David JM, David JP, et al (2024) Chalcone dimers from *Astronium graveolens* (Anacardiaceae) and their biological activities. *Phytochem Lett* 63:38–42. <https://doi.org/10.1016/j.phytol.2024.07.013>
66. Messana I, Ferrari F, de Moraes e Souza MA, Gács-Baitz E (1990) (–)-Salzol, an isopimarane diterpene, and a chalcone from *Hyptis salzmanii*. *Phytochemistry* 29:329–332. [https://doi.org/10.1016/0031-9422\(90\)89065-H](https://doi.org/10.1016/0031-9422(90)89065-H)
67. Monteiro V de FF, Mathias L, Vieira IJC, et al (2002) Prenylated Coumarins, Chalcone and New Cinnamic Acid and Dihydrocinnamic Acid Derivatives from *Brosimum gaudichaudii*. *J Braz Chem Soc* 13:281–287. <https://doi.org/10.1590/S0103-50532002000200023>
68. Tomazela DM, Pupo MT, Passador EAP, et al (2000) Pyrano chalcones and a flavone from *Neoraputia magnifica* and their *Trypanosoma cruzi* glycosomal glyceraldehyde-3-phosphate dehydrogenase-inhibitory activities. *Phytochemistry* 55:643–651. [https://doi.org/10.1016/S0031-9422\(00\)00248-X](https://doi.org/10.1016/S0031-9422(00)00248-X)
69. Magalhães AF, Azevedo Tozzi AMG, Noronha Sales BHL, Magalhães EG (1996) Twenty-three flavonoids from *Lonchocarpus subglaucens*. *Phytochemistry* 42:1459–1471. [https://doi.org/10.1016/0031-9422\(96\)00134-3](https://doi.org/10.1016/0031-9422(96)00134-3)
70. Ramalho SD, Bernades A, Demetrius G, et al (2013) Synthetic chalcone derivatives as inhibitors of cathepsins K and B, and their cytotoxic evaluation. *Chem Biodivers* 10:1999–2006. <https://doi.org/10.1002/cbdv.201200344>
71. Borges ID, Faria ECM, Custódio JFM, et al (2022) Insights into chalcone analogues with potential as antioxidant additives in diesel-biodiesel blends. *RSC Adv* 12:34746–34759. <https://doi.org/10.1039/d2ra07300e>
72. Kobelnik M, Ferreira LMB, Regasini LO, et al (2018) Thermal study of chalcones: Thermal decomposition of chalcone and its hydroxylated derivatives. *J Therm Anal Calorim* 132:425–431. <https://doi.org/10.1007/s10973-017-6956-2>

73. Claisen L, Claparède A (1881) Condensationen von Ketonen mit Aldehyden. *Berichte der deutschen chemischen Gesellschaft* 14:2460–2468. <https://doi.org/10.1002/cber.188101402192>
74. Schmidt JG (1881) Ueber die Einwirkung von Aceton auf Furfurol und auf Bittermandelöl bei Gegenwart von Alkalilauge. *Berichte der deutschen chemischen Gesellschaft* 14:1459–1461. <https://doi.org/10.1002/cber.188101401306>
75. Perrin CL, Chang KL (2016) The Complete Mechanism of an Aldol Condensation. *Journal of Organic Chemistry* 81:5631–5635. <https://doi.org/10.1021/acs.joc.6b00959>
76. Torrezan GS, Polaquini CR, Lima MF, Regasini LO (2020) Use of glycerol, waste glycerol from biodiesel production and other protic solvents in bioactive α,β -unsaturated ketones synthesis. *Sustain Chem Pharm* 16:. <https://doi.org/10.1016/j.scp.2020.100250>
77. Jardim GAM, Guimarães TT, Pinto MDCFR, et al (2015) Naphthoquinone-based chalcone hybrids and derivatives: Synthesis and potent activity against cancer cell lines. *Medchemcomm* 6:120–150. <https://doi.org/10.1039/c4md00371c>
78. Almeida LR, Anjos MM, Ribeiro GC, et al (2017) Synthesis, structural characterization and computational study of a novel amino chalcone: a potential nonlinear optical material. *New Journal of Chemistry* 41:1744–1754. <https://doi.org/10.1039/c5nj03214h>
79. Firmino PP, Queiroz JE, Dias LD, et al (2022) Synthesis, Molecular Structure, Thermal and Spectroscopic Analysis of a Novel Bromochalcone Derivative with Larvicidal Activity. *Crystals (Basel)* 12:. <https://doi.org/10.3390/cryst12040440>
80. Ribeiro MA, de-Farias IF, Freire PTC, et al (2023) Synthesis, crystal structure, structural and spectroscopic analysis of (2E)-1-(4-chlorophenyl)-3-(4-methoxyphenyl)prop-2-en-1-one. *J Mol Struct* 1294:. <https://doi.org/10.1016/j.molstruc.2023.136410>
81. da Silva AA, Maia PI da S, Lopes CD, et al (2021) Synthesis, characterization and antichagasic evaluation of thiosemicarbazones prepared from chalcones and dibenzalacetones. *J Mol Struct* 1232:. <https://doi.org/10.1016/j.molstruc.2021.130014>

82. Ternavisk RR, Camargo AJ, Machado FBC, et al (2014) Synthesis, characterization, and computational study of a new dimethoxy-chalcone. *J Mol Model* 20:. <https://doi.org/10.1007/s00894-014-2526-8>
83. Felipe WQ, Barbosa IR, Oliveira AA, et al (2025) Antifungal effects of thiosemicarbazone-chalcones on *Aspergillus*, *Candida* and *Sporothrix* strains. *Arch Microbiol* 207:. <https://doi.org/10.1007/s00203-024-04229-4>
84. Silva WA, Kleber C, Andrade Z, et al (2013) Biological and Structure-Activity Evaluation of Chalcone Derivatives against Bacteria and Fungi
85. Emeri FT de AS de, Rosalen PL, Paganini ÉR, et al (2019) Antimicrobial activity of nitrochalcone and pentyl caffeate against hospital pathogens results in decreased microbial adhesion and biofilm formation. *Biofouling* 35:129–142. <https://doi.org/10.1080/08927014.2019.1574763>
86. Garcia MAR, Theodoro RS, Sardi JCO, et al (2021) Design, synthesis and antibacterial activity of chalcones against MSSA and MRSA planktonic cells and biofilms. *Bioorg Chem* 116:. <https://doi.org/10.1016/j.bioorg.2021.105279>
87. de Almeida Romagna R, dos Santos RB, de Cassia Ribeiro Gonçalves R, Kitagawa RR (2024) Synthesis of Chalcones, Screening In silico and In vitro and Evaluation of *Helicobacter pylori* Adhesion by Molecular Docking. *Curr Pharm Des* 30:3350–3366. <https://doi.org/10.2174/0113816128327090240821101355>
88. Ribeiro Lima FR, Figueiredo LC de, Oliveira Braga AR, et al (2025) Antimicrobial and anti-biofilm activity of a mucoadhesive hydrogel functionalized with aminochalcone on titanium surfaces and in *Galleria mellonella* model: In vitro and in vivo study. *Microb Pathog* 200:. <https://doi.org/10.1016/j.micpath.2025.107286>
89. Satokata AAC, Souza JH, Silva LLO, et al (2022) Chalcones with potential antibacterial and antibiofilm activities against periodontopathogenic bacteria. *Anaerobe* 76:. <https://doi.org/10.1016/j.anaerobe.2022.102588>
90. Morão LG, Lorenzoni ASG, Chakraborty P, et al (2020) Investigating the modes of action of the antimicrobial chalcones BC1 and T9A. *Molecules* 25:. <https://doi.org/10.3390/molecules25204596>
91. Rocha JE, de Freitas TS, Xavier JC, et al (2023) ADMET study, spectroscopic characterization and effect of synthetic nitro chalcone in combination with norfloxacin, ciprofloxacin, and ethidium bromide against *Staphylococcus aureus* efflux pumps. *Fundam Clin Pharmacol* 37:163–173. <https://doi.org/10.1111/fcp.12830>

92. Hemília de Souza Nunes P, Sampaio de Freitas T, Esmeraldo Rocha J, et al (2022) Potentiation of antibiotic activity, and efflux pumps inhibition by (2E)-1-(4-aminophenyl)-3-(4-fluorophenyl)prop-2-en-1-one. *Fundam Clin Pharmacol* 36:1066–1082. <https://doi.org/10.1111/fcp.12785>
93. Siqueira MMR, Freire P de TC, Cruz BG, et al (2021) Aminophenyl chalcones potentiating antibiotic activity and inhibiting bacterial efflux pump. *European Journal of Pharmaceutical Sciences* 158:. <https://doi.org/10.1016/j.ejps.2020.105695>
94. Xavier J da C, de Almeida-Neto FWQ, Rocha JE, et al (2021) Spectroscopic analysis by NMR, FT-Raman, ATR-FTIR, and UV-Vis, evaluation of antimicrobial activity, and in silico studies of chalcones derived from 2-hydroxyacetophenone. *J Mol Struct* 1241:. <https://doi.org/10.1016/j.molstruc.2021.130647>
95. Freitas TS De, Xavier JDC, Pereira RLS, et al (2020) Direct antibacterial and antibiotic resistance modulatory activity of chalcones synthesized from the natural product 2-hydroxy-3,4,6-trimethoxyacetophenone. *FEMS Microbiol Lett* 367:. <https://doi.org/10.1093/femsle/fnaa124>
96. Ferraz CAN, Tintino SR, Teixeira AMR, et al (2020) Potentiation of antibiotic activity by chalcone (E)-1-(4'-aminophenyl)-3-(furan-2-yl)-prop-2-en-1-one against gram-positive and gram-negative MDR strains. *Microb Pathog* 148:. <https://doi.org/10.1016/j.micpath.2020.104453>
97. de Sousa JN, de Andrade Ferreira Barreto J, de Alcântara Oliveira FA, et al (2024) ADMET study and inhibition of *Staphylococcus aureus* efflux pumps by a synthetic p-aminochalcone. *Results Chem* 7:. <https://doi.org/10.1016/j.rechem.2024.101449>
98. Palanco AC, Lacorte Singulani J De, Costa-Orlandi CB, et al (2017) Activity of 3'-hydroxychalcone against *Cryptococcus gattii* and toxicity, and efficacy in alternative animal models. *Future Microbiol* 12:1123–1134. <https://doi.org/10.2217/fmb-2017-0062>
99. Rocha Garcia MA, Sardi J de CO, dos Santos MB, et al (2025) Synthesis and evaluation of the antifungal and antibiofilm potential of aminochalcones. *Arch Microbiol* 207:. <https://doi.org/10.1007/s00203-025-04244-z>
100. Nogueira Bandeira P, Oliveira R, Fontenele S, et al In Vitro Antifungal Activity Against *Trychophyton Rubrum* of p-Aminochalcones and 3'-Methoxy-4'-Hydroxy Chalcone

101. Tristão TC, Campos-Buzzi F, Corrêa R, et al (2012) Antimicrobial and cytotoxicity potential of acetamido, amino and nitrochalcones. *Arzneimittel-Forschung/Drug Research* 62:590–594. <https://doi.org/10.1055/s-0032-1327610>
102. Da Silva PT, Lopes LMA, Xavier JDC, et al (2020) Cytotoxic and antifungal activity of chalcones synthesized from natural acetophenone isolated from *Croton anisodontus*. *Revista Virtual de Quimica* 12:712–723. <https://doi.org/10.21577/1984-6835.20200057>
103. Marques BC, Santos MB, Anselmo DB, et al (2019) Methoxychalcones: Effect of Methoxyl Group on the Antifungal, Antibacterial and Antiproliferative Activities. *Med Chem (Los Angeles)* 16:881–891. <https://doi.org/10.2174/1573406415666190724145158>
104. Maschio-Lima T, Lemes TH, Marques MDR, et al (2025) Synergistic activity between conventional antifungals and chalcone-derived compound against dermatophyte fungi and *Candida* spp. *International Microbiology* 28:265–275. <https://doi.org/10.1007/s10123-024-00541-7>
105. Bila NM, Costa-Orlandi CB, Vaso CO, et al (2021) 2-Hydroxychalcone as a Potent Compound and Photosensitizer Against Dermatophyte Biofilms. *Front Cell Infect Microbiol* 11:. <https://doi.org/10.3389/fcimb.2021.679470>
106. Melo WCMA, Santos MB dos, Marques B de C, et al (2017) Selective photoinactivation of *Histoplasma capsulatum* by water-soluble derivatives chalcones. *Photodiagnosis Photodyn Ther* 18:232–235. <https://doi.org/10.1016/j.pdpdt.2017.03.001>
107. Medina-Alarcón KP, Singulani JDL, Dutra LA, et al (2020) Antifungal activity of 2'-hydroxychalcone loaded in nanoemulsion against *Paracoccidioides* spp. *Future Microbiol* 15:21–33. <https://doi.org/10.2217/fmb-2019-0095>
108. Lauro R, de Souza Fernandes L, Sayuri Ogasawara L, et al (2024) Bioprospecting, Synergistic Antifungal and Toxicological Aspects of the Hydroxychalcones and Their Association with Azole Derivates against *Candida* spp. for Treating Vulvovaginal Candidiasis. <https://doi.org/10.3390/pharmaceutics>
109. Brasil AMV, Lopes JRDF, Chagas AFDS, Franco AMR (2024) Therapeutic potential of hesperidin methyl chalcone in the experimental treatment of cutaneous leishmaniasis. *Acta Amazon* 54:. <https://doi.org/10.1590/1809-4392202203181>

110. Miranda-Sapla MM, Tomiotto-Pellissier F, Assolini JP, et al (2019) trans-Chalcone modulates *Leishmania amazonensis* infection in vitro by Nrf2 overexpression affecting iron availability. *Eur J Pharmacol* 853:275–288. <https://doi.org/10.1016/j.ejphar.2019.03.049>
111. Alonso L, Menegatti R, Gomes RS, et al (2020) Antileishmanial activity of the chalcone derivative LQFM064 associated with reduced fluidity in the parasite membrane as assessed by EPR spectroscopy. *European Journal of Pharmaceutical Sciences* 151:. <https://doi.org/10.1016/j.ejps.2020.105407>
112. de Mello TFP, Bitencourt HR, Pedroso RB, et al (2014) Leishmanicidal activity of synthetic chalcones in *Leishmania (Viannia) braziliensis*. *Exp Parasitol* 136:27–34. <https://doi.org/10.1016/j.exppara.2013.11.003>
113. Bello ML, Chiaradia LD, Dias LRS, et al (2011) Trimethoxy-chalcone derivatives inhibit growth of *Leishmania braziliensis*: Synthesis, biological evaluation, molecular modeling and structure-activity relationship (SAR). *Bioorg Med Chem* 19:5046–5052. <https://doi.org/10.1016/j.bmc.2011.06.023>
114. Gomes KS, da Costa-Silva TA, Oliveira IH, et al (2019) Structure-activity relationship study of antitrypanosomal chalcone derivatives using multivariate analysis. *Bioorg Med Chem Lett* 29:1459–1462. <https://doi.org/10.1016/j.bmcl.2019.04.020>
115. Marcovicz C, Camargo G dos A, Scharr B, et al (2022) Schistosomicidal evaluation of synthesized bromo and nitro chalcone derivatives. *J Mol Struct* 1258:. <https://doi.org/10.1016/j.molstruc.2022.132647>
116. Trein MR, Rodrigues e Oliveira L, Rigo GV, et al (2019) Anti-*Trichomonas vaginalis* activity of chalcone and amino-analogues. *Parasitol Res* 118:607–615. <https://doi.org/10.1007/s00436-018-6164-4>
117. Leite FF, de Sousa NF, de Oliveira BHM, et al (2023) Anticancer Activity of Chalcones and Its Derivatives: Review and In Silico Studies. *Molecules* 28
118. Siqueira E da S, Concato VM, Tomiotto-Pellissier F, et al (2021) Trans-chalcone induces death by autophagy mediated by p53 up-regulation and β -catenin down-regulation on human hepatocellular carcinoma HuH7.5 cell line. *Phytomedicine* 80:. <https://doi.org/10.1016/j.phymed.2020.153373>
119. Leal ALAB, Pinheiro DP, Barros-Nepomuceno FWA, et al (2021) Structural and spectroscopic analysis and evaluation of cytotoxic activity of 2-hydroxychalcones

- against human cancer cell lines. *J Mol Struct* 1245:..
<https://doi.org/10.1016/j.molstruc.2021.131135>
120. Costa A, Chiaradia-Delatorre LD, Dos Santos Bubniak L, et al (2014) Apoptotic effect of synthetic 2',4',5'-trimethoxychalcones in human K562 and Jurkat leukemia cells. *Medicinal Chemistry Research* 23:4301–4319.
<https://doi.org/10.1007/s00044-014-1002-4>
 121. Winter E, Chiaradia LD, De Cordova CAS, et al (2010) Naphthylchalcones induce apoptosis and caspase activation in a leukemia cell line: The relationship between mitochondrial damage, oxidative stress, and cell death. *Bioorg Med Chem* 18:8026–8034. <https://doi.org/10.1016/j.bmc.2010.09.025>
 122. Michelini LJ, Castro MRC, Custodio JMF, et al (2018) A novel potential anticancer chalcone: Synthesis, crystal structure and cytotoxic assay. *J Mol Struct* 1168:309–315. <https://doi.org/10.1016/j.molstruc.2018.05.010>
 123. De Vasconcelos A, Campos VF, Nedel F, et al (2013) Cytotoxic and apoptotic effects of chalcone derivatives of 2-acetyl thiophene on human colon adenocarcinoma cells. *Cell Biochem Funct* 31:289–297.
<https://doi.org/10.1002/cbf.2897>
 124. Pinheiro S, Pessôa JC, Pinheiro EMC, et al (2020) 2H-1,2,3-Triazole-chalcones as novel cytotoxic agents against prostate cancer. *Bioorg Med Chem Lett* 30:..
<https://doi.org/10.1016/j.bmcl.2020.127454>
 125. Custodio JMF, Custodio JMF, Perez CN, et al (2020) On the: In silico and in vitro anticancer activity of sulfonamide chalcones: Potential JNKK3 inhibitors. *New Journal of Chemistry* 44:3294–3309. <https://doi.org/10.1039/c9nj05612b>
 126. Fogaça TB, Martins RM, Begnini KR, et al (2017) Apoptotic effect of chalcone derivatives of 2-acetylthiophene in human breast cancer cells. *Pharmacological Reports* 69:156–161. <https://doi.org/10.1016/j.pharep.2016.10.003>
 127. Cabral BLS, da Silva ACG, de Ávila RI, et al (2017) A novel chalcone derivative, LQFM064, induces breast cancer cells death via p53, p21, KIT and PDGFRA. *European Journal of Pharmaceutical Sciences* 107:1–15.
<https://doi.org/10.1016/j.ejps.2017.06.018>
 128. Neves MP, Cravo S, Lima RT, et al (2012) Solid-phase synthesis of 2'-hydroxychalcones. Effects on cell growth inhibition, cell cycle and apoptosis of human tumor cell lines. *Bioorg Med Chem* 20:25–33.
<https://doi.org/10.1016/j.bmc.2011.11.042>

129. Santos MB, Pinhanelli VC, Garcia MAR, et al (2017) Antiproliferative and pro-apoptotic activities of 2'- and 4'-aminochalcones against tumor canine cells. *Eur J Med Chem* 138:884–889. <https://doi.org/10.1016/j.ejmech.2017.06.049>
130. Gomes KS, Coelho JA, Tesser PEH, et al (2026) Activity–Selectivity of Flavonoid Derivatives in Endometriotic Cells. *ACS Omega* 11:10703–10711. <https://doi.org/10.1021/acsomega.5c12546>
131. Winter E, Devantier Neuenfeldt P, Chiaradia-Delatorre LD, et al (2014) Symmetric bis-chalcones as a new type of breast cancer resistance protein inhibitors with a mechanism different from that of chromones. *J Med Chem* 57:2930–2941. <https://doi.org/10.1021/jm401879z>
132. Zeraik ML, Pauli I, Dutra LA, et al (2021) Identification of a Prenyl Chalcone as a Competitive Lipoyxygenase Inhibitor: Screening, Biochemical Evaluation and Molecular Modeling Studies. *Molecules* 26:2205. <https://doi.org/10.3390/molecules26082205>
133. Ventura T, Calixto S, Abraham-Vieira B, et al (2015) Antimycobacterial and Anti-Inflammatory Activities of Substituted Chalcones Focusing on an Anti-Tuberculosis Dual Treatment Approach. *Molecules* 20:8072–8093. <https://doi.org/10.3390/molecules20058072>
134. Santos MB dos, Carvalho Marques B, Miranda Ayusso G, et al (2021) Chalcones and their B-aryl analogues as myeloperoxidase inhibitors: In silico, in vitro and ex vivo investigations. *Bioorg Chem* 110:. <https://doi.org/10.1016/j.bioorg.2021.104773>
135. Zeraik ML, Ximenes VF, Regasini LO, et al (2012) 4'-Aminochalcones As Novel Inhibitors of the Chlorinating Activity of Myeloperoxidase. *Curr Med Chem* 19:5405–5413. <https://doi.org/10.2174/092986712803833344>
136. Piva M, Yaekashi KM, Pereira TGO, et al (2026) Trans-Chalcone alleviates overt pain-like behavior by targeting the activation of nociceptive neuron TRPV1 and TRPA1 channels. *Inflammopharmacology*. <https://doi.org/10.1007/s10787-025-02099-w>
137. Sulis PM, Bittencourt Mendes AK, Fernandes TA, et al (2023) Signal transduction of the insulin secretion induced by the chalcone analogue, (E)-3-(phenyl)-1-(3,4,5-trimethoxyphenyl)prop-2-en-1-one, and its role in glucose and lipid metabolism. *Biochimie* 212:85–94. <https://doi.org/10.1016/j.biochi.2023.04.006>

138. Xavier JC, Ferreira MKA, Silva AW, et al (2021) Anxiolytic-like and Anticonvulsant Effect in Adult Zebrafish (*Danio rerio*) through GABAergic System and Molecular Docking Study of Chalcone Derived from Natural Products. *Biointerface Res Appl Chem* 11:14021–14031. <https://doi.org/10.33263/BRIAC116.1402114031>
139. de Matos IAF, Fernandes NAR, Cirelli G, et al (2023) Chalcone T4 Inhibits RANKL-Induced Osteoclastogenesis and Stimulates Osteogenesis In Vitro. *Int J Mol Sci* 24:7624. <https://doi.org/10.3390/ijms24087624>
140. Arroio TR, Caires FJ, Almeida G de O, et al (2026) Transition-Metal-Free One-Pot Synthesis of (Hetero)chalcones with Cysteine Protease Inhibitory Activity. *ACS Omega* 11:7616–7626. <https://doi.org/10.1021/acsomega.5c08888>
141. Lipinski CA (2004) Lead- and drug-like compounds: the rule-of-five revolution. *Drug Discov Today Technol* 1:337–341. <https://doi.org/10.1016/j.ddtec.2004.11.007>
142. Veber DF, Johnson SR, Cheng H-Y, et al (2002) Molecular Properties That Influence the Oral Bioavailability of Drug Candidates. *J Med Chem* 45:2615–2623. <https://doi.org/10.1021/jm020017n>
143. Egan WJ, Merz , Kenneth M., Baldwin JJ (2000) Prediction of Drug Absorption Using Multivariate Statistics. *J Med Chem* 43:3867–3877. <https://doi.org/10.1021/jm000292e>
144. Ghose AK, Viswanadhan VN, Wendoloski JJ (1999) A Knowledge-Based Approach in Designing Combinatorial or Medicinal Chemistry Libraries for Drug Discovery. 1. A Qualitative and Quantitative Characterization of Known Drug Databases. *J Comb Chem* 1:55–68. <https://doi.org/10.1021/cc9800071>
145. Muegge I, Heald SL, Brittelli D (2001) Simple Selection Criteria for Drug-like Chemical Matter. *J Med Chem* 44:1841–1846. <https://doi.org/10.1021/jm015507e>
146. Brenk R, Schipani A, James D, et al (2008) Lessons Learnt from Assembling Screening Libraries for Drug Discovery for Neglected Diseases. *ChemMedChem* 3:435–444. <https://doi.org/10.1002/cmdc.200700139>
147. Baell JB, Holloway GA (2010) New Substructure Filters for Removal of Pan Assay Interference Compounds (PAINS) from Screening Libraries and for Their Exclusion in Bioassays. *J Med Chem* 53:2719–2740. <https://doi.org/10.1021/jm901137j>

148. Hann MM, Keserü GM (2012) Finding the sweet spot: the role of nature and nurture in medicinal chemistry. *Nat Rev Drug Discov* 11:355–365. <https://doi.org/10.1038/nrd3701>
149. Teague SJ, Davis AM, Leeson PD, Oprea T (1999) The Design of Leadlike Combinatorial Libraries. *Angew Chem Int Ed* 38:3743–3748. [https://doi.org/10.1002/\(SICI\)1521-3773\(19991216\)38:24<3743::AID-ANIE3743>3.0.CO;2-U](https://doi.org/10.1002/(SICI)1521-3773(19991216)38:24<3743::AID-ANIE3743>3.0.CO;2-U)
150. Martin YC (2005) A Bioavailability Score. *J Med Chem* 48:3164–3170. <https://doi.org/10.1021/jm0492002>
151. Ertl P, Schuffenhauer A (2009) Estimation of synthetic accessibility score of drug-like molecules based on molecular complexity and fragment contributions. *J Cheminform* 1:8. <https://doi.org/10.1186/1758-2946-1-8>
152. Fukunishi Y, Kurosawa T, Mikami Y, Nakamura H (2014) Prediction of Synthetic Accessibility Based on Commercially Available Compound Databases. *J Chem Inf Model* 54:3259–3267. <https://doi.org/10.1021/ci500568d>
153. Daina A, Michielin O, Zoete V (2017) SwissADME: a free web tool to evaluate pharmacokinetics, drug-likeness and medicinal chemistry friendliness of small molecules. *Sci Rep* 7:42717. <https://doi.org/10.1038/srep42717>
154. Echeverría J, Opazo J, Mendoza L, et al (2017) Structure-Activity and Lipophilicity Relationships of Selected Antibacterial Natural Flavones and Flavanones of Chilean Flora. *Molecules* 22:608. <https://doi.org/10.3390/molecules22040608>
155. Waring MJ (2010) Lipophilicity in drug discovery. *Expert Opin Drug Discov* 5:235–248. <https://doi.org/10.1517/17460441003605098>
156. Gackowski M, Koba M, Pluskota R, et al (2021) Pharmacological classification of anticancer drugs applying chromatographic retention data and chemometric analysis. *Chemical Papers* 75:265–278. <https://doi.org/10.1007/s11696-020-01301-3>
157. Doak BC, Over B, Giordanetto F, Kihlberg J (2014) Oral Druggable Space beyond the Rule of 5: Insights from Drugs and Clinical Candidates. *Chem Biol* 21:1115–1142. <https://doi.org/10.1016/j.chembiol.2014.08.013>
158. Cragg GM, Newman DJ (2013) Natural products: A continuing source of novel drug leads. *Biochimica et Biophysica Acta (BBA) - General Subjects* 1830:3670–3695. <https://doi.org/10.1016/j.bbagen.2013.02.008>

159. Thapa P, Upadhyay SP, Singh V, et al (2023) Chalcone: A potential scaffold for NLRP3 inflammasome inhibitors. *European Journal of Medicinal Chemistry Reports* 7:100100. <https://doi.org/10.1016/j.ejmcr.2022.100100>
160. Sangpheak K, Tabtimmai L, Seetaha S, et al (2019) Biological Evaluation and Molecular Dynamics Simulation of Chalcone Derivatives as Epidermal Growth Factor-Tyrosine Kinase Inhibitors. *Molecules* 24:1092. <https://doi.org/10.3390/molecules24061092>
161. Murray CJL, Ikuta KS, Sharara F, et al (2022) Global burden of bacterial antimicrobial resistance in 2019: a systematic analysis. *The Lancet* 399:629–655. [https://doi.org/10.1016/S0140-6736\(21\)02724-0](https://doi.org/10.1016/S0140-6736(21)02724-0)
162. World Health Organization (2020) Global 2020 Antibacterial Agents in Clinical and Preclinical Development: An Overview and Analysis; WHO: Geneva, Switzerland, 2020. In: <https://www.who.int/publications/i/item/9789240021303/>
163. Costerton JW, Stewart PS, Greenberg EP (1999) Bacterial Biofilms: A Common Cause of Persistent Infections. *Science* (1979) 284:1318–1322. <https://doi.org/10.1126/science.284.5418.1318>
164. Newman DJ, Cragg GM (2020) Natural Products as Sources of New Drugs over the Nearly Four Decades from 01/1981 to 09/2019. *J Nat Prod* 83:770–803. <https://doi.org/10.1021/acs.jnatprod.9b01285>
165. Dai J, Han R, Xu Y, et al (2020) Recent progress of antibacterial natural products: Future antibiotics candidates. *Bioorg Chem* 101:103922. <https://doi.org/10.1016/j.bioorg.2020.103922>
166. Rossiter SE, Fletcher MH, Wuest WM (2017) Natural Products as Platforms To Overcome Antibiotic Resistance. *Chem Rev* 117:12415–12474. <https://doi.org/10.1021/acs.chemrev.7b00283>
167. Wahab S, Annadurai S, Abullais SS, et al (2021) Glycyrrhiza glabra (Licorice): A Comprehensive Review on Its Phytochemistry, Biological Activities, Clinical Evidence and Toxicology. *Plants* 10:2751. <https://doi.org/10.3390/plants10122751>
168. Shen F, Tang X, Wang Y, et al (2015) Phenotype and expression profile analysis of *Staphylococcus aureus* biofilms and planktonic cells in response to licochalcone A. *Appl Microbiol Biotechnol* 99:359–373. <https://doi.org/10.1007/s00253-014-6076-x>

169. Liu X, Xiong Y, Shi Y, et al (2022) In vitro activities of licochalcone A against planktonic cells and biofilm of *Enterococcus faecalis*. *Front Microbiol* 13:. <https://doi.org/10.3389/fmicb.2022.970901>
170. Tsukiyama R-I, Katsura H, Tokuriki N, Kobayashi M (2002) Antibacterial Activity of Licochalcone A against Spore-Forming Bacteria. *Antimicrob Agents Chemother* 46:1226–1230. <https://doi.org/10.1128/AAC.46.5.1226-1230.2002>
171. Qiu J, Feng H, Xiang H, et al (2010) Influence of subinhibitory concentrations of licochalcone A on the secretion of enterotoxins A and B by *Staphylococcus aureus*. *FEMS Microbiol Lett* 307:135–141. <https://doi.org/10.1111/j.1574-6968.2010.01973.x>
172. Qiu J, Jiang Y, Xia L, et al (2010) Subinhibitory concentrations of licochalcone A decrease alpha-toxin production in both methicillin-sensitive and methicillin-resistant *Staphylococcus aureus* isolates. *Lett Appl Microbiol* 50:223–229. <https://doi.org/10.1111/j.1472-765X.2009.02783.x>
173. Hao H, Hui W, Liu P, et al (2013) Correction: Effect of Licochalcone A on Growth and Properties of *Streptococcus suis*. *PLoS One* 8:. <https://doi.org/10.1371/annotation/59a7568c-915d-4d9c-b912-f93af72acdcb>
174. Haraguchi H, Tanimoto K, Tamura Y, et al (1998) Mode of antibacterial action of retrochalcones from *Glycyrrhiza inflata*. *Phytochemistry* 48:125–129. [https://doi.org/10.1016/S0031-9422\(97\)01105-9](https://doi.org/10.1016/S0031-9422(97)01105-9)
175. Wu S-C, Yang Z-Q, Liu F, et al (2019) Antibacterial Effect and Mode of Action of Flavonoids From Licorice Against Methicillin-Resistant *Staphylococcus aureus*. *Front Microbiol* 10:. <https://doi.org/10.3389/fmicb.2019.02489>
176. Wang Z, Cao Y, Paudel S, et al (2013) Concise synthesis of licochalcone C and its regioisomer, licochalcone H. *Arch Pharm Res* 36:1432–1436. <https://doi.org/10.1007/s12272-013-0222-3>
177. Kim CG, Jeon J-H, Seo YH, Jun J-G (2014) Facile Synthesis of Licochalcone C. *Bull Korean Chem Soc* 35:1996–1998. <https://doi.org/10.5012/bkcs.2014.35.7.1996>
178. Dovhaniuk N, Blahun OP, Sosunovych B, et al (2023) Regioselective and Scalable Total Synthesis of Licochalcone C and Related Licoagrochalcones. *European J Org Chem* 26:. <https://doi.org/10.1002/ejoc.202201226>

179. Kajiyama K, Demizu S, Hiraga Y, et al (1992) Two prenylated retrochalcones from *Glycyrrhiza inflata*. *Phytochemistry* 31:3229–3232. [https://doi.org/10.1016/0031-9422\(92\)83481-D](https://doi.org/10.1016/0031-9422(92)83481-D)
180. Fergestad ME, Stamsås GA, Morales Angeles D, et al (2020) Penicillin-binding protein PBP2a provides variable levels of protection toward different β -lactams in *Staphylococcus aureus* RN4220. *Microbiologyopen* 9:. <https://doi.org/10.1002/mbo3.1057>
181. Fishovitz J, Hermoso JA, Chang M, Mobashery S (2014) Penicillin-binding protein 2a of methicillin-resistant *Staphylococcus aureus*. *IUBMB Life* 66:572–577. <https://doi.org/10.1002/iub.1289>
182. Ávila HP, Smânia E de FA, Monache FD, Smânia A (2008) Structure–activity relationship of antibacterial chalcones. *Bioorg Med Chem* 16:9790–9794. <https://doi.org/10.1016/j.bmc.2008.09.064>
183. Li X-Z, Nikaido H (2004) Efflux-Mediated Drug Resistance in Bacteria. *Drugs* 64:159–204. <https://doi.org/10.2165/00003495-200464020-00004>
184. Poole K (2005) Efflux-mediated antimicrobial resistance. *Journal of Antimicrobial Chemotherapy* 56:20–51. <https://doi.org/10.1093/jac/dki171>
185. Pagès J-M, Amaral L (2009) Mechanisms of drug efflux and strategies to combat them: Challenging the efflux pump of Gram-negative bacteria. *Biochimica et Biophysica Acta (BBA) - Proteins and Proteomics* 1794:826–833. <https://doi.org/10.1016/j.bbapap.2008.12.011>
186. Kuete V, Ngameni B, Tangmouo JG, et al (2010) Efflux Pumps Are Involved in the Defense of Gram-Negative Bacteria against the Natural Products Isobavachalcone and Diospyrone. *Antimicrob Agents Chemother* 54:1749–1752. <https://doi.org/10.1128/AAC.01533-09>
187. Kusters JG, van Vliet AHM, Kuipers EJ (2006) Pathogenesis of *Helicobacter pylori* Infection. *Clin Microbiol Rev* 19:449–490. <https://doi.org/10.1128/CMR.00054-05>
188. Xu X, Xu L, Yuan G, et al (2018) Synergistic combination of two antimicrobial agents closing each other's mutant selection windows to prevent antimicrobial resistance. *Sci Rep* 8:7237. <https://doi.org/10.1038/s41598-018-25714-z>
189. Gaur R, Gupta VK, Pal A, et al (2015) In vitro and in vivo synergistic interaction of substituted chalcone derivatives with norfloxacin against methicillin resistant

Staphylococcus aureus. RSC Adv 5:5830–5845.
<https://doi.org/10.1039/C4RA10842F>

190. Božić DD, Milenković M, Ivković B, Ćirković I (2014) Antibacterial activity of three newly-synthesized chalcones & synergism with antibiotics against clinical isolates of methicillin-resistant Staphylococcus aureus. Indian J Med Res 140:130–7
191. Götz F (2002) *Staphylococcus* and biofilms. Mol Microbiol 43:1367–1378.
<https://doi.org/10.1046/j.1365-2958.2002.02827.x>
192. Wu H, Moser C, Wang H-Z, et al (2015) Strategies for combating bacterial biofilm infections. Int J Oral Sci 7:1–7. <https://doi.org/10.1038/ijos.2014.65>
193. Schwarz S, Shen J, Kadlec K, et al (2016) Lincosamides, Streptogramins, Phenicols, and Pleuromutilins: Mode of Action and Mechanisms of Resistance. Cold Spring Harb Perspect Med 6:a027037.
<https://doi.org/10.1101/cshperspect.a027037>
194. Cheesman M, Ilanko A, Blonk B, Cock I (2017) Developing new antimicrobial therapies: Are synergistic combinations of plant extracts/compounds with conventional antibiotics the solution? Pharmacogn Rev 11:57.
https://doi.org/10.4103/phrev.phrev_21_17
195. Wiedemann I, Benz R, Sahl H-G (2004) Lipid II-Mediated Pore Formation by the Peptide Antibiotic Nisin: a Black Lipid Membrane Study. J Bacteriol 186:3259–3261. <https://doi.org/10.1128/JB.186.10.3259-3261.2004>
196. Browne N, Heelan M, Kavanagh K (2013) An analysis of the structural and functional similarities of insect hemocytes and mammalian phagocytes. Virulence 4:597–603. <https://doi.org/10.4161/viru.25906>
197. Allegra E, Titball RW, Carter J, Champion OL (2018) Galleria mellonella larvae allow the discrimination of toxic and non-toxic chemicals. Chemosphere 198:469–472. <https://doi.org/10.1016/j.chemosphere.2018.01.175>
198. Pereira TC, De Barros PP, Fugisaki LR de O, et al (2018) Recent Advances in the Use of Galleria mellonella Model to Study Immune Responses against Human Pathogens. Journal of Fungi 4:128. <https://doi.org/10.3390/jof4040128>
199. WANG P, YUAN X, WANG Y, et al (2015) Licochalcone C induces apoptosis via B-cell lymphoma 2 family proteins in T24 cells. Mol Med Rep 12:7623–7628.
<https://doi.org/10.3892/mmr.2015.4346>

200. Lee S-O, Joo SH, Lee J-Y, et al (2024) Licochalcone C Inhibits the Growth of Human Colorectal Cancer HCT116 Cells Resistant to Oxaliplatin. *Biomol Ther (Seoul)* 32:104–114. <https://doi.org/10.4062/biomolther.2023.167>
201. Abdel-Wahab A-HA, Effat H, Mahrous EA, et al (2021) A Licorice Roots Extract Induces Apoptosis and Cell Cycle Arrest and Improves Metabolism via Regulating MiRNAs in Liver Cancer Cells. *Nutr Cancer* 73:1047–1058. <https://doi.org/10.1080/01635581.2020.1783329>
202. Oh H, Seo J, Lee M, et al (2018) Licochalcone C induced apoptosis in human oral squamous cell carcinoma cells by regulation of the JAK2/STAT3 signaling pathway. *J Cell Biochem* 119:10118–10130. <https://doi.org/10.1002/jcb.27349>
203. Kwak A-W, Choi J-S, Liu K, et al (2020) Licochalcone C induces cell cycle G1 arrest and apoptosis in human esophageal squamous carcinoma cells by activation of the ROS/MAPK signaling pathway. *Journal of Chemotherapy* 32:132–143. <https://doi.org/10.1080/1120009X.2020.1721175>
204. Ye Z, Yang J, Feng Y, et al (2016) First Enantioselective Synthesis of Brosimacutins H and I. *Chinese Journal of Organic Chemistry* 36:547. <https://doi.org/10.6023/cjoc201508021>
205. Rikimaru K, Wakabayashi T, Abe H, et al (2012) Structure–activity relationships and key structural feature of pyridyloxybenzene-acylsulfonamides as new, potent, and selective peroxisome proliferator-activated receptor (PPAR) γ Agonists. *Bioorg Med Chem* 20:3332–3358. <https://doi.org/10.1016/j.bmc.2012.03.036>
206. Guo Y, Wang Y, Li H, et al (2018) Novel Nitric Oxide Donors of Phenylsulfonylfuroxan and 3-Benzyl Coumarin Derivatives as Potent Antitumor Agents. *ACS Med Chem Lett* 9:502–506. <https://doi.org/10.1021/acsmedchemlett.8b00125>
207. Songthammawat P, Wangngae S, Matsumoto K, et al (2018) Bioinspired Diastereoconvergent Synthesis of the Tricyclic Core of Palodesangrens via Diels–Alder Reaction, LiAlH_4 -Mediated Isomerization, and Acid-Mediated Cyclization. *J Org Chem* 83:5225–5241. <https://doi.org/10.1021/acs.joc.8b00668>
208. Leandro LF, Cardoso MJO, Silva SDC, et al (2014) Antibacterial activity of *Pinus elliottii* and its major compound, dehydroabietic acid, against multidrug-resistant strains. *J Med Microbiol* 63:1649–1653. <https://doi.org/10.1099/jmm.0.081711-0>

209. CLSI M07 (2012) Methods for Dilution Antimicrobial Susceptibility Tests for Bacteria That Grow Aerobically. Approved Standard. 9th ed. Clinical and Laboratory Standards Institute (CLSI): Wayne, PA, USA, 2012; Volume 32.
210. Palomino J-C, Martin A, Camacho M, et al (2002) Resazurin Microtiter Assay Plate: Simple and Inexpensive Method for Detection of Drug Resistance in *Mycobacterium tuberculosis*. *Antimicrob Agents Chemother* 46:2720–2722. <https://doi.org/10.1128/AAC.46.8.2720-2722.2002>
211. Alves JA, Abrão F, Silva Moraes T da, et al (2020) Investigation of *Copaifera* Genus as a New Source of Antimycobacterial Agents. *Future Sci OA* 6:. <https://doi.org/10.2144/fsoa-2020-0018>
212. White RL, Burgess DS, Manduru M, Bosso JA (1996) Comparison of three different in vitro methods of detecting synergy: time-kill, checkerboard, and E test. *Antimicrob Agents Chemother* 40:1914–1918. <https://doi.org/10.1128/AAC.40.8.1914>
213. Wei G-X (2006) Effect of MUC7 peptides on the growth of bacteria and on *Streptococcus mutans* biofilm. *Journal of Antimicrobial Chemotherapy* 57:1100–1109. <https://doi.org/10.1093/jac/dkl120>
214. Vieira RGL, Moraes T da S, Silva L de O, et al (2018) In vitro studies of the antibacterial activity of *Copaifera* spp. oleoresins, sodium hypochlorite, and peracetic acid against clinical and environmental isolates recovered from a hemodialysis unit. *Antimicrob Resist Infect Control* 7:14. <https://doi.org/10.1186/s13756-018-0307-3>
215. Parai D, Banerjee M, Dey P, Mukherjee SK (2020) Reserpine attenuates biofilm formation and virulence of *Staphylococcus aureus*. *Microb Pathog* 138:103790. <https://doi.org/10.1016/j.micpath.2019.103790>
216. Saising J, Dube L, Ziebandt A-K, et al (2012) Activity of Gallidermin on *Staphylococcus aureus* and *Staphylococcus epidermidis* Biofilms. *Antimicrob Agents Chemother* 56:5804–5810. <https://doi.org/10.1128/AAC.01296-12>
217. Dilarri G, Zamuner CF, Mendes CR, et al (2021) Evaluating the potential of electrolysed water for the disinfection of citrus fruit in packinghouses. *J Sci Food Agric* 101:2584–2591. <https://doi.org/10.1002/jsfa.10888>
218. Savietto A, Polaquini CR, Kopacz M, et al (2018) Antibacterial activity of monoacetylated alkyl gallates against *Xanthomonas citri* subsp. *citri*. *Arch Microbiol* 200:929–937. <https://doi.org/10.1007/s00203-018-1502-6>

219. Martins PMM, Lau IF, Bacci M, et al (2010) Subcellular localization of proteins labeled with GFP in *Xanthomonas citri* ssp. *citri*: targeting the division septum. *FEMS Microbiol Lett* 310:76–83. <https://doi.org/10.1111/j.1574-6968.2010.02047.x>
220. Król E, de Sousa Borges A, da Silva I, et al (2015) Antibacterial activity of alkyl gallates is a combination of direct targeting of FtsZ and permeabilization of bacterial membranes. *Front Microbiol* 6:. <https://doi.org/10.3389/fmicb.2015.00390>
221. Megaw J, Thompson TP, Lafferty RA, Gilmore BF (2015) *Galleria mellonella* as a novel in vivo model for assessment of the toxicity of 1-alkyl-3-methylimidazolium chloride ionic liquids. *Chemosphere* 139:197–201. <https://doi.org/10.1016/j.chemosphere.2015.06.026>
222. Girard M, Nelson CB, Picot V, Gubler DJ (2020) Arboviruses: A global public health threat. In: *Vaccine*. Elsevier Ltd, pp 3989–3994
223. World Health Organization (2025) WHO guidelines for clinical management of arboviral diseases: dengue, chikungunya, Zika and yellow fever. In: <https://iris.who.int/server/api/core/bitstreams/634a55a5-327e-459b-a633-0650fe8ad6c9/content>
224. McMichael AJ, Woodruff RE, Hales S (2006) Climate change and human health: present and future risks. *The Lancet* 367:859–869. [https://doi.org/10.1016/S0140-6736\(06\)68079-3](https://doi.org/10.1016/S0140-6736(06)68079-3)
225. Espinal MA, Andrus JK, Jauregui B, et al (2019) Emerging and reemerging aedes-transmitted arbovirus infections in the region of the americas: Implications for health policy. *Am. J. Public Health* 109:387–392
226. Beckham JD, Tyler KL (2015) Arbovirus Infections. *Continuum (N Y)* 21:1599–1611. <https://doi.org/10.1212/CON.0000000000000240>
227. Vasconcelos PFC, Calisher CH (2016) Emergence of Human Arboviral Diseases in the Americas, 2000-2016. *Vector-Borne and Zoonotic Diseases* 16:295–301
228. Hotez PJ, Murray KO (2017) Dengue, West Nile virus, chikungunya, Zika—and now Mayaro? *PLoS Negl. Trop. Dis.* 11
229. World Health Organization - Global Arbovirus Initiative (2024) Preparing for the next pandemic tackling mosquito-borne viruses with epidemic and pandemic potential. In: <https://iris.who.int/server/api/core/bitstreams/2a55ec8b-bbf5-46c9-a5bb-696f15196d49/content>

230. Lima-Camara TN (2016) Emerging arboviruses and public health challenges in Brazil. *Rev Saude Publica* 50:. <https://doi.org/10.1590/S1518-8787.2016050006791>
231. Musso D, Rodriguez-Morales AJ, Levi JE, et al (2018) Unexpected outbreaks of arbovirus infections: lessons learned from the Pacific and tropical America. *Lancet Infect. Dis.* 18:e355–e361
232. Gould E, Pettersson J, Higgs S, et al (2017) Emerging arboviruses: Why today? *One Health* 4:1–13
233. Ministério da Saúde - Secretaria da Vigilância em Saúde (2020) Monitoramento dos casos de arboviroses urbanas transmitidas pelo *Aedes Aegypti* (dengue, chikungunya e zika), semanas epidemiológicas 1 a 46.
234. Goh VSL, Mok CK, Chu JJH (2020) Antiviral natural products for arbovirus infections. *Molecules* 25
235. Teixeira RR, Pereira WL, Da Silveira Oliveira AFC, et al (2014) Natural products as source of potential dengue antivirals. *Molecules* 19:8151–8176
236. Da Silveira Oliveira AFC, Teixeira RR, De Oliveira AS, et al (2017) Potential antivirals: Natural products targeting replication enzymes of dengue and Chikungunya viruses. *Molecules* 22
237. Koehn FE, Carter GT (2005) The evolving role of natural products in drug discovery. *Nat. Rev. Drug Discov.* 4:206–220
238. Li JW-H, Vederas JC (2009) Drug Discovery and Natural Products: End of an Era or an Endless Frontier? *Science* (1979) 325:161–165. <https://doi.org/10.1126/science.1168243>
239. Cataneo AHD, Ávila EP, Mendes LA de O, et al (2021) Flavonoids as Molecules With Anti-Zika virus Activity. *Front. Microbiol.* 12
240. Kiat TS, Pippen R, Yusof R, et al (2006) Inhibitory activity of cyclohexenyl chalcone derivatives and flavonoids of fingerroot, *Boesenbergia rotunda* (L.), towards dengue-2 virus NS3 protease. *Bioorg Med Chem Lett* 16:3337–3340. <https://doi.org/10.1016/j.bmcl.2005.12.075>
241. Almeida e Sá FH, Silva ARN, de Oliveira TJS, et al (2023) A chalcone identified by in silico and in vitro assays possesses high larvicidal activity against *Aedes aegypti*. *Acta Trop* 238:. <https://doi.org/10.1016/j.actatropica.2022.106791>

242. Maria Pia GD, Sara F, Mario F, Lorenza S (2018) Biological Effects of Licochalcones. *Mini-Reviews in Medicinal Chemistry* 19:647–656. <https://doi.org/10.2174/1389557518666180601095420>
243. Kang CB, Keller TH, Luo D (2017) Zika Virus Protease: An Antiviral Drug Target. *Trends Microbiol.* 25:797–808
244. Nitsche C (2019) Proteases from dengue, West Nile and Zika viruses as drug targets. *Biophys. Rev.* 11:157–165
245. Roy A, Lim L, Srivastava S, et al (2017) Solution conformations of Zika NS2B-NS3pro and its inhibition by natural products from edible plants. *PLoS One* 12:. <https://doi.org/10.1371/journal.pone.0180632>
246. Lima CS, Mottin M, de Assis LR, et al (2021) Flavonoids from *Pterogyne nitens* as Zika virus NS2B-NS3 protease inhibitors. *Bioorg Chem* 109:. <https://doi.org/10.1016/j.bioorg.2021.104719>
247. Li Y, Zhang Z, Phoo WW, et al (2018) Structural Insights into the Inhibition of Zika Virus NS2B-NS3 Protease by a Small-Molecule Inhibitor. *Structure* 26:555-564.e3. <https://doi.org/10.1016/j.str.2018.02.005>
248. Lu K, Chu J, Wang H, et al (2013) Regioselective iodination of flavonoids by N-iodosuccinimide under neutral conditions. *Tetrahedron Lett* 54:6345–6348. <https://doi.org/10.1016/j.tetlet.2013.09.051>
249. Wang H, Yan Z, Lei Y, et al (2014) Concise synthesis of prenylated and geranylated chalcone natural products by regiospecific iodination and Suzuki coupling reactions. *Tetrahedron Lett* 55:897–899. <https://doi.org/10.1016/j.tetlet.2013.12.044>
250. Miyaura' N, Suzuki' Nt A (1995) *Palladium-Catalyzed Cross-Coupling Reactions of Organoboron Compounds* Contents I. Introduction II. Synthesis of Organoboron Reagents A Synthesis of Organoboron Compounds B. Hydroboration of Alkenes and Alkynes C. Hydroboration of Terminal Alkynes D. Miscellaneous Methods
251. Marinho M dos S, Zhang YN, Cassani NM, et al (2024) Development and validation of Mayaro virus with luciferase reporter genes as a tool for antiviral assays. *Heliyon* 10:. <https://doi.org/10.1016/j.heliyon.2024.e33885>
252. Cassani NM, Santos IA, Grosche VR, et al (2023) Roles of Bothrops jararacussu toxins I and II: Antiviral findings against Zika virus. *Int J Biol Macromol* 227:630–640. <https://doi.org/10.1016/j.ijbiomac.2022.12.102>

253. Santos IA, Shimizu JF, de Oliveira DM, et al (2021) Chikungunya virus entry is strongly inhibited by phospholipase A2 isolated from the venom of *Crotalus durissus terrificus*. *Sci Rep* 11:. <https://doi.org/10.1038/s41598-021-88039-4>
254. ISO 10993-5:2009 (2009) Biological evaluation of medical devices Part 5: Tests for in vitro cytotoxicity. In: <https://www.iso.org/standard/36406.html>.
255. Charles R. Anderson, Wilbur G. Downs, George H. Wattley, et al (1998) The American journal of tropical medicine and hygiene. *Am J Trop Med Hyg* 6:1012–1016
256. De Oliveira Mota MT, Ribeiro MR, Vedovello D, Nogueira ML (2015) Mayaro virus: A neglected arbovirus of the Americas. *Future Virol.* 10:1109–1122
257. Acosta-Ampudia Y, Monsalve DM, Rodríguez Y, et al (2018) Mayaro: an emerging viral threat? *Emerg. Microbes Infect.* 7
258. Santiago FW, Halsey ES, Siles C, et al (2015) Long-Term Arthralgia after Mayaro Virus Infection Correlates with Sustained Pro-inflammatory Cytokine Response. *PLoS Negl Trop Dis* 9:. <https://doi.org/10.1371/journal.pntd.0004104>
259. Andreolla AP, Borges AA, Bordignon J, Duarte dos Santos CN (2022) Mayaro Virus: The State-of-the-Art for Antiviral Drug Development. *Viruses* 14
260. Paschoalino M, Marinho M dos S, Santos IA, et al (2023) An update on the development of antiviral against Mayaro virus: from molecules to potential viral targets. *Arch. Microbiol.* 205
261. Ferreira PG, Ferraz AC, Figueiredo JE, et al (2018) Detection of the antiviral activity of epicatechin isolated from *Salacia crassifolia* (Celastraceae) against Mayaro virus based on protein C homology modelling and virtual screening. *Arch Virol* 163:1567–1576. <https://doi.org/10.1007/s00705-018-3774-1>
262. Salles TS, Meneses MDF, Yamamoto KA, et al Chemical composition and anti-Mayaro virus activity of *Schinus terebinthifolius* fruits. <https://doi.org/10.1007/s13337>
263. Camini FC, da Silva TF, da Silva Caetano CC, et al (2018) Antiviral activity of silymarin against Mayaro virus and protective effect in virus-induced oxidative stress. *Antiviral Res* 158:8–12. <https://doi.org/10.1016/j.antiviral.2018.07.023>
264. dos Santos AE, Kuster RM, Yamamoto KA, et al (2014) Quercetin and quercetin 3-O-glycosides from *Bauhinia longifolia* (Bong.) Steud. show anti-Mayaro virus activity. *Parasit Vectors* 7:130. <https://doi.org/10.1186/1756-3305-7-130>

265. Ferraz AC, Moraes T de FS, Nizer WS da C, et al (2019) Virucidal activity of proanthocyanidin against Mayaro virus. *Antiviral Res* 168:76–81. <https://doi.org/10.1016/j.antiviral.2019.05.008>
266. Lima JCB, Barbosa JARG (2023) Interaction models between peptide substrate and Alphavirus Protease nsP2 of Chikungunya and Mayaro and implications to the mechanism of action. *J Biomol Struct Dyn* 41:10851–10858. <https://doi.org/10.1080/07391102.2022.2158941>
267. Faye O, Freire CCM, Iamarino A, et al (2014) Molecular Evolution of Zika Virus during Its Emergence in the 20th Century. *PLoS Negl Trop Dis* 8:36. <https://doi.org/10.1371/journal.pntd.0002636>
268. Wikan N, Smith DR (2016) Zika virus: History of a newly emerging arbovirus. *Lancet Infect. Dis.* 16:e119–e126
269. Baud D, Gubler DJ, Schaub B, et al (2017) An update on Zika virus infection. *The Lancet* 390:2099–2109
270. Pierson TC, Diamond MS (2018) The emergence of Zika virus and its new clinical syndromes. *Nature* 560:573–581
271. Gaudry A, Bos S, Viranaicken W, et al (2018) The flavonoid isoquercitrin precludes initiation of Zika virus infection in human cells. *Int J Mol Sci* 19:. <https://doi.org/10.3390/ijms19041093>
272. Cataneo AHD, Kuczera D, Koishi AC, et al (2019) The citrus flavonoid naringenin impairs the in vitro infection of human cells by Zika virus. *Sci Rep* 9:. <https://doi.org/10.1038/s41598-019-52626-3>
273. Ramos PRP da S, Mottin M, Lima CS, et al (2022) Natural Compounds as Non-Nucleoside Inhibitors of Zika Virus Polymerase through Integration of In Silico and In Vitro Approaches. *Pharmaceuticals* 15:. <https://doi.org/10.3390/ph15121493>
274. Lee J Le, Loe MWC, Lee RCH, Chu JJH (2019) Antiviral activity of pinocembrin against Zika virus replication. *Antiviral Res* 167:13–24. <https://doi.org/10.1016/j.antiviral.2019.04.003>
275. Lin SC, Chen MC, Liu S, et al (2019) Phloretin inhibits Zika virus infection by interfering with cellular glucose utilisation. *Int J Antimicrob Agents* 54:80–84. <https://doi.org/10.1016/j.ijantimicag.2019.03.017>

276. Phoo WW, Zhang Z, Wirawan M, et al (2018) Structures of Zika virus NS2B-NS3 protease in complex with peptidomimetic inhibitors. *Antiviral Res* 160:17–24. <https://doi.org/10.1016/j.antiviral.2018.10.006>
277. Phoo WW, Li Y, Zhang Z, et al (2016) Structure of the NS2B-NS3 protease from Zika virus after self-cleavage. *Nat Commun* 7:. <https://doi.org/10.1038/ncomms13410>
278. Lei J, Hansen G, Nitsche C, et al (2016) Crystal structure of zika virus ns2b-ns3 protease in complex with a boronate inhibitor. *Science* (1979) 353:503–505. <https://doi.org/10.1126/science.aag2419>
279. Mirza MU, Alanko I, Vanmeert M, et al (2022) The discovery of Zika virus NS2B-NS3 inhibitors with antiviral activity via an integrated virtual screening approach. *European Journal of Pharmaceutical Sciences* 175:. <https://doi.org/10.1016/j.ejps.2022.106220>
280. Starvaggi J, Previti S, Zappalà M, Ettari R (2024) The Inhibition of NS2B/NS3 Protease: A New Therapeutic Opportunity to Treat Dengue and Zika Virus Infection. *Int. J. Mol. Sci.* 25
281. Majerová T, Novotný P, Krýsová E, Konvalinka J (2019) Exploiting the unique features of Zika and Dengue proteases for inhibitor design. *Biochimie* 166:132–141
282. Mukhametov A, Newhouse EI, Aziz NA, et al (2014) Allosteric pocket of the dengue virus (serotype 2) NS2B/NS3 protease: In silico ligand screening and molecular dynamics studies of inhibition. *J Mol Graph Model* 52:103–113. <https://doi.org/10.1016/j.jmgm.2014.06.008>
283. Brecher M, Li Z, Liu B, et al (2017) A conformational switch high-throughput screening assay and allosteric inhibition of the flavivirus NS2B-NS3 protease. *PLoS Pathog* 13:. <https://doi.org/10.1371/journal.ppat.1006411>
284. Othman R, Kiat TS, Khalid N, et al (2008) Docking of noncompetitive inhibitors into dengue virus type 2 protease: Understanding the interactions with allosteric binding sites. *J Chem Inf Model* 48:1582–1591. <https://doi.org/10.1021/ci700388k>
285. Andrade MA, Mottin M, Sousa BK de P, et al (2023) Identification of novel Zika virus NS3 protease inhibitors with different inhibition modes by integrative experimental and computational approaches. *Biochimie* 212:143–152. <https://doi.org/10.1016/j.biochi.2023.04.004>

286. Fernandes RS, Noske GD, Gawriljuk VO, et al (2021) High-throughput antiviral assays to screen for inhibitors of Zika virus replication. *Journal of Visualized Experiments*. <https://doi.org/10.3791/62422>
287. Lim H jung, Nguyen TTH, Kim NM, et al (2017) Inhibitory effect of flavonoids against NS2B-NS3 protease of ZIKA virus and their structure activity relationship. *Biotechnol Lett* 39:415–421. <https://doi.org/10.1007/s10529-016-2261-6>
288. Nunes DA de F, Santos FR da S, da Fonseca STD, et al (2022) NS2B-NS3 protease inhibitors as promising compounds in the development of antivirals against Zika virus: A systematic review. *J. Med. Virol.* 94:442–453
289. Lin WW, Huang YJ, Wang YT, et al (2023) Development of NS2B-NS3 protease inhibitor that impairs Zika virus replication. *Virus Res* 329:. <https://doi.org/10.1016/j.virusres.2023.199092>
290. Tian H, Ji X, Yang X, et al (2016) The crystal structure of Zika virus helicase: basis for antiviral drug design. *Protein Cell* 7:450–454
291. Godoy AS, Giroud C, Mesquita NCMR, et al (2024) High-throughput crystallographic fragment screening of Zika virus NS3 Helicase
292. Coutard B, Decroly E, Li C, et al (2014) Assessment of Dengue virus helicase and methyltransferase as targets for fragment-based drug discovery. *Antiviral Res* 106:61–70. <https://doi.org/10.1016/j.antiviral.2014.03.013>
293. Lo MC, Aulabaugh A, Jin G, et al (2004) Evaluation of fluorescence-based thermal shift assays for hit identification in drug discovery. *Anal Biochem* 332:153–159. <https://doi.org/10.1016/j.ab.2004.04.031>
294. Huynh K, Partch CL (2015) Analysis of protein stability and ligand interactions by thermal shift assay. *Curr Protoc Protein Sci* 79:28.9.1-28.9.14. <https://doi.org/10.1002/0471140864.ps2809s79>
295. Lim SP, Noble CG, Shi P-Y (2015) The dengue virus NS5 protein as a target for drug discovery. *Antiviral Res* 119:57–67. <https://doi.org/10.1016/j.antiviral.2015.04.010>
296. Godoy AS, Lima GMA, Oliveira KIZ, et al (2017) Crystal structure of Zika virus NS5 RNA-dependent RNA polymerase. *Nat Commun* 8:. <https://doi.org/10.1038/ncomms14764>
297. Yokokawa F, Nilar S, Noble CG, et al (2016) Discovery of Potent Non-Nucleoside Inhibitors of Dengue Viral RNA-Dependent RNA Polymerase from a Fragment Hit

- Using Structure-Based Drug Design. *J Med Chem* 59:3935–3952. <https://doi.org/10.1021/acs.jmedchem.6b00143>
298. Lim SP, Noble CG, Seh CC, et al (2016) Potent Allosteric Dengue Virus NS5 Polymerase Inhibitors: Mechanism of Action and Resistance Profiling. *PLoS Pathog* 12:. <https://doi.org/10.1371/journal.ppat.1005737>
 299. Guo Y, Wang Y, Li H, et al (2018) Novel Nitric Oxide Donors of Phenylsulfonylfuroxan and 3-Benzyl Coumarin Derivatives as Potent Antitumor Agents. *ACS Med Chem Lett* 9:502–506. <https://doi.org/10.1021/acsmedchemlett.8b00125>
 300. Souza GB, Santos TAC, Silva APS, et al (2024) Synthesis of chalcone derivatives by Claisen-Schmidt condensation and in vitro analyses of their antiprotozoal activities. *Nat Prod Res* 38:1326–1333. <https://doi.org/10.1080/14786419.2022.2140337>
 301. Pozzetti L, Ibba R, Rossi S, et al (2022) Total Synthesis of the Natural Chalcone Lophirone E, Synthetic Studies toward Benzofuran and Indole-Based Analogues, and Investigation of Anti-Leishmanial Activity. *Molecules* 27:. <https://doi.org/10.3390/molecules27020463>
 302. Kajiyama K, Demizu S, Hiraga Y, et al (1992) TWO PRENYLATED RETROCHALCONES FROM GLYCY~RHIZ~ 1NFLAZX
 303. Matkovic R, Bernard E, Fontanel S, et al (2019) The Host DHX9 DExH-Box Helicase Is Recruited to Chikungunya Virus Replication Complexes for Optimal Genomic RNA Translation. *J Virol* 93:. <https://doi.org/10.1128/jvi.01764-18>
 304. Espósito DLA, da Fonseca BAL (2015) Complete genome sequence of Mayaro virus (Togaviridae, Alphavirus) strain BeAr 20290 from Brazil. *Genome Announc* 3:. <https://doi.org/10.1128/genomeA.01372-15>
 305. Donald CL, Brennan B, Cumberworth SL, et al (2016) Full Genome Sequence and sfRNA Interferon Antagonist Activity of Zika Virus from Recife, Brazil. *PLoS Negl Trop Dis* 10:. <https://doi.org/10.1371/journal.pntd.0005048>
 306. da Conceição PJP, Ayusso GM, Carvalho T, et al (2025) In Vitro Evaluation of the Antiviral Activity of Polyphenol (-)-Epigallocatechin-3-Gallate (EGCG) Against Mayaro Virus. *Viruses* 17:. <https://doi.org/10.3390/v17020258>
 307. Cao X, Li Y, Jin X, et al (2016) Molecular mechanism of divalent-metal-induced activation of NS3 helicase and insights into Zika virus inhibitor design. *Nucleic Acids Res* 44:10505–10514. <https://doi.org/10.1093/nar/gkw941>

308. Berman HM (2000) The Protein Data Bank. *Nucleic Acids Res* 28:235–242. <https://doi.org/10.1093/nar/28.1.235>
309. Jones G, Willett P, Glen RC, et al (1997) Development and validation of a genetic algorithm for flexible docking 1 Edited by F. E. Cohen. *J Mol Biol* 267:727–748. <https://doi.org/10.1006/jmbi.1996.0897>
310. Krauer F, Riesen M, Reveiz L, et al (2017) Zika Virus Infection as a Cause of Congenital Brain Abnormalities and Guillain–Barré Syndrome: Systematic Review. *PLoS Med* 14:e1002203. <https://doi.org/10.1371/journal.pmed.1002203>
311. de Araújo TVB, Ximenes RA de A, Miranda-Filho D de B, et al (2018) Association between microcephaly, Zika virus infection, and other risk factors in Brazil: final report of a case-control study. *Lancet Infect Dis* 18:328–336. [https://doi.org/10.1016/S1473-3099\(17\)30727-2](https://doi.org/10.1016/S1473-3099(17)30727-2)
312. Musso D, Ko AI, Baud D (2019) Zika Virus Infection — After the Pandemic. *New England Journal of Medicine* 381:1444–1457. <https://doi.org/10.1056/NEJMr1808246>
313. Mottin M, Borba JVB, Braga RC, et al (2018) The A–Z of Zika drug discovery. *Drug Discov Today* 23:1833–1847. <https://doi.org/10.1016/j.drudis.2018.06.014>
314. Souza RRM, Orrico-Ferreira CM, Rocha DJPG, et al (2026) Recent advances in drug discovery for Zika virus therapeutics. *Virology* 619:110889. <https://doi.org/10.1016/j.virol.2026.110889>
315. Roy A, Lim L, Srivastava S, et al (2017) Solution conformations of Zika NS2B-NS3pro and its inhibition by natural products from edible plants. *PLoS One* 12:e0180632. <https://doi.org/10.1371/journal.pone.0180632>
316. Lim H, Nguyen TTH, Kim NM, et al (2017) Inhibitory effect of flavonoids against NS2B-NS3 protease of ZIKA virus and their structure activity relationship. *Biotechnol Lett* 39:415–421. <https://doi.org/10.1007/s10529-016-2261-6>
317. Simmler C, Lankin DC, Nikolić D, et al (2017) Isolation and structural characterization of dihydrobenzofuran congeners of licochalcone A. *Fitoterapia* 121:6–15. <https://doi.org/10.1016/j.fitote.2017.06.017>
318. Ayabe S-I, Yoshikawa T, Kobayashi M, Furuya T (1980) BIOSYNTHESIS OF A RETROCHALCONE, ECHINATIN: INVOLVEMENT OF O-METHYLTRANSFERASE TO LICODIONE". Pergamon Press Ltd
319. Kajiyama K, Hiraga Y, Takahashi K, et al (1993) Flavonoids and Isoflavonoids of Chemotaxonomic Significance from *Glycyrrhiza pallidiflora* (Leguminosae)

320. Malaník M (2025) Natural retrochalcones: rare compounds with diverse biological activities. *Phytochemistry Reviews*. <https://doi.org/10.1007/s11101-025-10103-y>
321. Ozanigue PR, Helena AL, Menezes R de P, et al (2024) Synthesis, Antibacterial Effects, and Toxicity of Licochalcone C. *Pharmaceuticals* 17:. <https://doi.org/10.3390/ph17050634>
322. Yoon G, Jung Y Do, Cheon SH (2005) Cytotoxic Allyl Retrochalcone from the Roots of *Glycyrrhiza inflata*. *Chem Pharm Bull (Tokyo)* 53:694–695. <https://doi.org/10.1248/cpb.53.694>
323. Liu Z, Yoon G, Cheon SH (2010) An enantioselective total synthesis of (S)-(-)-licochalcone E: determination of the absolute configuration. *Tetrahedron* 66:3165–3172. <https://doi.org/10.1016/j.tet.2010.02.089>
324. Yoon G, Liu Z, Jeong HJ, Cheon SH (2009) Total synthesis of licochalcone E. *Bull Korean Chem Soc* 30:2959–2961. <https://doi.org/10.5012/bkcs.2009.30.12.2959>
325. Li J, Mi Y, He J, Deng X (2010) A short and efficient synthesis of licochalcone E. *Synlett* 2289–2292. <https://doi.org/10.1055/s-0030-1258029>
326. Na Y, Cha J-H, Yoon H-G, Kwon Y (2009) A Concise Synthesis of Licochalcone E and Its Regio-Isomer, Licochalcone F. *Chem Pharm Bull (Tokyo)* 57:607–609. <https://doi.org/10.1248/cpb.57.607>
327. Yoon G, Oak MH, Lee JO, Cheon SH (2010) Semisynthesis of licochalcone E and biological evaluation as vasorelaxant agents. *Bull Korean Chem Soc* 31:1085–1087. <https://doi.org/10.5012/bkcs.2010.31.04.1085>
328. Liu Z, Wang Z, Yoon G, Cheon SH (2011) Stereoselective total synthesis of (+)-licochalcone e. *Arch Pharm Res* 34:1269–1276. <https://doi.org/10.1007/s12272-011-0805-9>
329. Yu SJ, Cho IA, Kang KR, et al (2017) Licochalcone-E induces caspase-dependent death of human pharyngeal squamous carcinoma cells through the extrinsic and intrinsic apoptotic signaling pathways. *Oncol Lett* 13:3662–3668. <https://doi.org/10.3892/ol.2017.5865>
330. Zhang Q, Zhang J, Liu B, Wei J (2022) Licochalcone E inhibits *trxR1* expression, alters *Nrf2/STAT6* signal, and induces antitumor effects in vitro against human SH-SY5Y and SK-N-BE(2) neuroblastoma cells. *Environ Toxicol* 37:1173–1184. <https://doi.org/10.1002/tox.23474>

331. Kwon SJ, Park SY, Kwon GT, et al (2013) Licochalcone E present in licorice suppresses lung metastasis in the 4T1 mammary orthotopic cancer model. *Cancer Prevention Research* 6:603–613. <https://doi.org/10.1158/1940-6207.CAPR-13-0012>
332. Yoon G, Kang BY, Cheon SH (2007) Topoisomerase I Inhibition and Cytotoxicity of Licochalcones A and E from *Glycyrrhiza inflata*
333. Jung CHANG H, Yoon G, Sun PARK J, et al (2290) Induction of Apoptosis by the Licochalcone E in Endothelial Cells via Modulation of NF-k kB and Bcl-2 Family Fig. 1. Structure of Retrochalcones
334. Lee HN, Cho HJ, Lim DY, et al (2013) Mechanisms by which licochalcone e exhibits potent anti-inflammatory properties: Studies with phorbol ester-treated mouse skin and lipopolysaccharide-stimulated murine macrophages. *Int J Mol Sci* 14:10926–10943. <https://doi.org/10.3390/ijms140610926>
335. Cho YC, Lee SH, Yoon G, et al (2010) Licochalcone E reduces chronic allergic contact dermatitis and inhibits IL-12p40 production through down-regulation of NF- κ B. *Int Immunopharmacol* 10:1119–1126. <https://doi.org/10.1016/j.intimp.2010.06.015>
336. Muto E, Okada T, Yamanaka T, et al (2023) Licochalcone E, a β -Amyloid Aggregation Inhibitor, Regulates Microglial M1/M2 Polarization via Inhibition of CTL1-Mediated Choline Uptake. *Biomolecules* 13:. <https://doi.org/10.3390/biom13020191>
337. Cao Y, Si Y, Li M, et al (2021) Licochalcone E improves insulin sensitivity in palmitic acid-treated HepG2 cells through inhibition of the NLRP3 signaling pathway. *Int Immunopharmacol* 99:. <https://doi.org/10.1016/j.intimp.2021.107923>
338. Park HG, Bak EJ, Woo GH, et al (2012) Licochalcone E has an antidiabetic effect. *Journal of Nutritional Biochemistry* 23:759–767. <https://doi.org/10.1016/j.jnutbio.2011.03.021>
339. Guo Z, Niu X, Xiao T, et al (2015) Chemical profile and inhibition of α -glycosidase and protein tyrosine phosphatase 1B (PTP1B) activities by flavonoids from licorice (*Glycyrrhiza uralensis* Fisch). *J Funct Foods* 14:324–336. <https://doi.org/10.1016/j.jff.2014.12.003>
340. Han J, Wang D, Li D, et al (2017) Licochalcone e protects against carbon tetrachloride-induced liver toxicity by activating peroxisome proliferator-activated

receptor gamma. Mol Med Rep 16:5269–5276.
<https://doi.org/10.3892/mmr.2017.7268>

341. Kim SS, Lim J, Bang Y, et al (2012) Licochalcone E activates Nrf2/antioxidant response element signaling pathway in both neuronal and microglial cells: Therapeutic relevance to neurodegenerative disease. *Journal of Nutritional Biochemistry* 23:1314–1323. <https://doi.org/10.1016/j.jnutbio.2011.07.012>
342. Han JY, Park SH, Yang JH, et al (2014) Licochalcone suppresses LXR α -induced hepatic lipogenic gene expression through AMPK/sirt1 pathway activation. *Toxicol Res* 30:19–25. <https://doi.org/10.5487/TR.2014.30.1.019>
343. Huang W, Ding L, Zhang N, et al (2021) Flavonoids from *Eucommia ulmoides* and their in vitro hepatoprotective activities. *Nat Prod Res* 35:3584–3591. <https://doi.org/10.1080/14786419.2020.1715402>
344. Zeng F, Shao S, Zou Z, et al (2024) Multi-omics revealed antibacterial mechanisms of licochalcone A against MRSA and its antimicrobial potential on pork meat. *Food Chem X* 24:101893. <https://doi.org/10.1016/j.fochx.2024.101893>
345. Zhou T, Deng X, Qiu J (2012) Antimicrobial activity of licochalcone E against *Staphylococcus aureus* and its impact on the production of staphylococcal alpha-toxin. *J Microbiol Biotechnol* 22:800–805. <https://doi.org/10.4014/jmb.1112.12020>
346. Jeon JH, Kim MR, Jun JG (2011) Concise synthesis of licochalcone a through water-accelerated [3,3]-sigmatropic rearrangement of an aryl prenyl ether. *Synthesis (Stuttg)* 370–376. <https://doi.org/10.1055/s-0030-1258381>
347. Li Y, Zhang Z, Phoo WW, et al (2018) Structural Insights into the Inhibition of Zika Virus NS2B-NS3 Protease by a Small-Molecule Inhibitor. *Structure* 26:555-564.e3. <https://doi.org/10.1016/j.str.2018.02.005>
348. Phoo WW, Zhang Z, Wirawan M, et al (2018) Structures of Zika virus NS2B-NS3 protease in complex with peptidomimetic inhibitors. *Antiviral Res* 160:17–24. <https://doi.org/10.1016/j.antiviral.2018.10.006>
349. Andrade MA, Mottin M, Sousa BK de P, et al (2023) Identification of novel Zika virus NS3 protease inhibitors with different inhibition modes by integrative experimental and computational approaches. *Biochimie* 212:143–152. <https://doi.org/10.1016/j.biochi.2023.04.004>

350. Majerová T, Novotný P, Krýsová E, Konvalinka J (2019) Exploiting the unique features of Zika and Dengue proteases for inhibitor design. *Biochimie* 166:132–141. <https://doi.org/10.1016/j.biochi.2019.05.004>
351. Song W, Qiao X, Chen K, et al (2017) Biosynthesis-Based Quantitative Analysis of 151 Secondary Metabolites of Licorice To Differentiate Medicinal *Glycyrrhiza* Species and Their Hybrids. *Anal Chem* 89:3146–3153. <https://doi.org/10.1021/acs.analchem.6b04919>
352. Li W, Asada Y, Yoshikawa T (2000) Flavonoid constituents from *Glycyrrhiza glabra* hairy root cultures. *Phytochemistry* 55:447–456. [https://doi.org/10.1016/S0031-9422\(00\)00337-X](https://doi.org/10.1016/S0031-9422(00)00337-X)
353. Xiao Z-P, Wei W, Huang S, et al (2014) Selective Acylation of Phenols in Boron Trifluoride Diethyl Etherate Solution and the Mechanistic Implication. *Asian Journal of Chemistry* 26:8039–8042. <https://doi.org/10.14233/ajchem.2014.17019>
354. Miralles N, Alam R, Szabó KJ, Fernández E (2016) Transition-Metal-Free Borylation of Allylic and Propargylic Alcohols. *Angew Chem Int Ed* 55:4303–4307. <https://doi.org/10.1002/anie.201511255>
355. Pozzetti L, Ibba R, Rossi S, et al (2022) Total Synthesis of the Natural Chalcone Lophirone E, Synthetic Studies toward Benzofuran and Indole-Based Analogues, and Investigation of Anti-Leishmanial Activity. *Molecules* 27:463. <https://doi.org/10.3390/molecules27020463>
356. Batista MN, Braga ACS, Campos GRF, et al (2019) Natural Products Isolated from Oriental Medicinal Herbs Inactivate Zika Virus. *Viruses* 11:49. <https://doi.org/10.3390/v11010049>
357. Faria NR, Azevedo R do S da S, Kraemer MUG, et al (2016) Zika virus in the Americas: Early epidemiological and genetic findings. *Science* (1979) 352:345–349. <https://doi.org/10.1126/science.aaf5036>