

**GEOGRAPHICAL PHENOTYPIC VARIATION AND
REPRODUCTIVE SYSTEM OF A DISTYLOUS
RUBIACEAE IN THE BRAZILIAN CERRADO**

MARIA JÚLIA WALDEMARIN DORIA

Dissertação apresentada ao Instituto
de Biociências, Câmpus de
Botucatu, UNESP, para obtenção do
título de Mestre no Programa de
Pós-Graduação em Ciências
Biológicas Botânica, Área de
concentração Morfologia e
Diversidade Vegetal.

BOTUCATU – SP

2020

UNIVERSIDADE ESTADUAL PAULISTA
"Júlio de Mesquita Filho"
INSTITUTO DE BIOCÊNCIAS DE BOTUCATU

GEOGRAPHICAL PHENOTYPIC VARIATION AND
REPRODUCTIVE SYSTEM OF A DISTYLOUS
RUBIACEAE IN THE BRAZILIAN CERRADO

MARIA JÚLIA WALDEMARIN DORIA

FELIPE WANDERLEY DE AMORIM
ORIENTADOR

CAMILA KISSMANN
CO-ORIENTADORA

Dissertação apresentada ao Instituto de Biotecnologia, Câmpus de Botucatu, UNESP, para obtenção do título de Mestre no Programa de Pós-Graduação em Ciências Biológicas Botânica, Área de concentração Morfologia e Diversidade Vegetal.

BOTUCATU – SP
2020

FICHA CATALOGRÁFICA ELABORADA PELA SEÇÃO TÉC. AQUIS. TRATAMENTO DA INFORM.
DIVISÃO TÉCNICA DE BIBLIOTECA E DOCUMENTAÇÃO - CÂMPUS DE BOTUCATU - UNESP
BIBLIOTECÁRIA RESPONSÁVEL: ROSEMEIRE APARECIDA VICENTE-CRB 8/5651

Doria, Maria Júlia Waldemarin.

Geographical phenotypic variation and reproductive system of a distylous Rubiaceae in the brazilian cerrado / Maria Júlia Waldemarin Doria. - Botucatu, 2020

Dissertação (mestrado) - Universidade Estadual Paulista "Júlio de Mesquita Filho", Instituto de Biociências de Botucatu

Orientador: Felipe Wanderley de Amorim

Coorientador: Camila Kissmann

Capes: 20303025

1. Rubiaceae. 2. Polinização. 3. Plantas - Reprodução.
4. Plantas dos cerrados.

Palavras-chave: Biologia da polinização; Distilia; Ecologia.

Dedico minha dissertação primeiramente a minha mãe, Lourdes, que por sua força, determinação e suporte me possibilitou essa enorme conquista. Ao meu pai, Antônio, por todo apoio, carinho e zelo que me permitiram trilhar meu caminho.

A toda minha família por todo o apoio.

A minha companheira Karla Polo, por todo o amor, companheirismo e apoio incondicional nessa jornada e em todas as outras que compartilharemos.

A minha melhor amiga Danielle Yamauchi, que me ensinou muito sobre a vida e sobre amizade. Obrigada por sempre estar ao meu lado.

Dedico a todos meus amigos, especialmente a Gabriel Fellipe, Fábio Henrique, Bruno Salata, Francislene Martins e todos os “Pererecos”.

A meu orientador Felipe Amorim, por acreditar em mim mesmo quando eu não consigo e por todo carinho e paciência.

A minha terapeuta, Karina Pani, pelo excelente auxílio profissional durante as maiores tempestades.

A todos que de alguma forma acrescentaram em minha vida e minha formação acadêmica.

AGRADECIMENTOS

Ao Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq) - Brasil pelo apoio no presente trabalho.

Ao meu orientador Felipe W. Amorim por todo o acolhimento e orientação que possibilitaram a existência desse trabalho.

Aos colegas de trabalho Pedro A. Barbosa, Pablo Oliveira, Natalia Kano, Eduardo Dal Farra, Larissa Silva, Isabella Castro, Caio S. Ballarin, Leandro Hachuy Filho, Priscila S.Veiga, Diego Polizello e Murilo M. Giffu pela colaboração científica e técnica na elaboração e condução do trabalho.

A Amábilio Camargo e toda equipe da Expedição Veadeiros da Embrapa Cerrados, e a Hélder Consolaro e alunos por toda enorme contribuição na coleta de dados.

A Vinícius Lourenço Garcia de Brito pela grande colaboração científica, auxílio nas análises das cores e discussões.

E por fim a minha universidade casa, a Universidade Estadual Paulista “Júlio de Mesquita Filho” - UNESP e ao Instituto de Biociências de Botucatu – IBB sem os quais nenhuma etapa desse trabalho ou de minha formação pessoal ou profissional teria sido concretizada.

CONTENTS

ABSTRACT 7

INTRODUCTION 8

MATERIAL AND METHODS 12

RESULTS 16

DISCUSSION..... 23

REFERENCES..... 26

1 **ABSTRACT**

2

3 *Palicourea rigida* is a Rubiaceae species widely distributed across the Brazilian Cerrado. The
4 species presents distyly, a type of floral dimorphism comprised of two floral morphs that are
5 associated with a genetic heteromorphic self-incompatibility that enables the reproduction only
6 after pollen flow between the distinct floral morphs. *Palicourea rigida* is typically pollinated by
7 hummingbirds and has flowers varying in the color spectrum between orange and red. The
8 species, however, presents a phenotypic variation across its distribution, and this variation
9 includes floral size, color, and life form. In the southernmost distribution of the Cerrado
10 vegetation, *P. rigida* presents a phenotypic variation with a markedly different life form, flower
11 size, and color. In this sense, considering the phenotypic variation in *Palicourea rigida* populations
12 both in life form and floral traits, as well as its dependence on pollinators for sexual reproduction, in this
13 study we described the changes in the floral morphology of different populations of *P. rigida* and tested
14 if they are followed by alterations of the basal features of distyly, *i.e.*, breakdown of the heteromorphic
15 self-incompatibility system, reciprocal herkogamy and isoplethy. Additionally, due to the change in
16 flower color and size, we discussed the hypothesis of a potential pollinator shift from hummingbirds
17 (ornithophily) to bees (melitophilily) in the dwarf with yellow flowered population as a possible outcome
18 of pollinator-mediated selection. For this, we perform controlled pollination experiments in the
19 southernmost dwarf population, analyzed the floral morphology, and evaluate floral color and pollinator
20 perception. Our results showed that the basic features of distyly were maintained in the
21 southernmost dwarf population, and flowers were significantly smaller, with a larger stigmatic
22 lobe in both floral morphs. Our results reinforce the role of inaccuracy in legitimate pollen
23 deposition, where the higher inaccuracy in high organs seems to be responsible for lower pollen
24 deposition at that level. Besides that, we found a pollinator shift from hummingbirds to bees in
25 the population, even with no cognitive difference for bees when exposed to *P. rigida* yellow
26 and reddish flowers.

27

28 INTRODUCTION

29

30 Environmental heterogeneity is an important factor affecting the phenotype of species
31 with broad distribution, because of the influence it plays on the local microevolutionary
32 processes (Liao *et al.* 2016). Phenotypic variations may be the result of either the population
33 response to local selective pressures under which they are subjected, *i.e.*, local adaptation, or
34 the ability of a given genotype to express different phenotype according to the local
35 environmental conditions, which allows acclimation, *i.e.*, phenotypic plasticity (Liao *et al.*
36 2016, de Villemereuil *et al.* 2018). Environmental conditions play a key role in the phenotypic
37 plasticity and in local adaptation of species, which may affect traits related to survival and
38 reproduction of species, such as plant phenology, floral morphology, variation and specificity
39 of pollinators systems, as well as growth habits in plants (Ollerton *et al.* 2006; Etterson *et al.*
40 2016; Petry *et al.* 2016). Therefore, species with a broad distribution are subject to different
41 selective pressures throughout their geographical range and may have different responses to the
42 environmental conditions (Liao *et al.* 2016).

43 The Cerrado (the Neotropical savanna of Brazil) is one of the largest biomes in South
44 America and is composed of a complex of plant formations. Like other savanna biomes
45 worldwide, such as those in Africa and Australia, the Cerrado is also affected by environmental
46 conditions, such as climate, soil, and fire events as well as by several anthropic disturbances
47 (Ribeiro & Walter 2008). The effect of these factors can influence the structure, spatial
48 distribution, and floristic composition of these habitats (Oliveira 2000; Ribeiro & Walter 2008).
49 Recent studies on the Brazilian Cerrado and in species under different environmental pressures
50 have shown contrasting forms of variations and anomalies in the common features of
51 heterostylous (distylous or tristylous species) plant populations, such as the variations in the
52 level of reciprocal herkogamy, self-compatibility, anisoplethy and in the frequency of
53 monomorphic and homostylous populations in typically distylous species (Baker 1955; Bawa
54 & Beach 1983; Sobrevila *et al.* 1983; Richards & Koptur 1993; Paillet *et al.* 1998; Faivre &
55 McDade 2001; Coelho & Barbosa 2003; Consolaro *et al.* 2005; Toledo *et al.* 2007; Consolaro
56 *et al.* 2009; Silva & Segura 2015).

57 *Palicourea rigida* Kunth. (Rubiaceae) is a distylous species, with a broad distribution in
58 the Brazilian Cerrado. In the core distribution of *P. rigida* in the Cerrado of central Brazil, the
59 species is a treelet (with a phanerophytic growth form) of about 1-1.5 meters in height (Fig. 1
60 A). Flower color varies from orange to reddish and the species is typically pollinated by
61 hummingbirds (Fig. 2 A, but see Machado *et al.* 2010; Justino *et al.* 2011; Maruyama 2016).
62 However, in the southernmost distribution of the Cerrado vegetation, *P. rigida* presents a
63 notoriously phenotypic variation with a markedly reduced size (Fig. 1 C). In this region, growth

64 form changes from phanerophyte to hemicryptophyte, *i.e.*, the species present underground
65 stems with shoot apical meristems born at soil level, and plants are dwarfs. Flowers and
66 inflorescences are also notoriously smaller and the color is quite distinct, being yellowish in
67 tone (Fig. 2 B), a color spectrum which is associated with melittophily (Chittka & Waser 1997;
68 Lunau *et al.* 1996; Dyer *et al.* 2016; Papiorek *et al.* 2016). Also, *P. rigida* also presents another
69 extreme of the phenotype variation in which plants are trees reaching more than 2 meters in
70 height, and flowers are reddish (Fig. 2 C).

71 Heterostyly, which is observed in *P. rigida*, is a floral polymorphism genetically
72 determined that are present in at least 28 families of angiosperms and is a basal feature in the
73 genus *Palicourea* (Vuilleumier 1967; Taylor 1997; Barrett & Shore 2008). Heterostylous
74 species populations are comprised of plants with different floral morphs, which can be either
75 distylous (two floral morphs) or tristylous (three floral morphs). Distylous species produce pin
76 (long-style) and thrum (short-style) flowers, and the populations, in general, present a morph
77 (*i.e.* pin : thrum) ratio of 1:1 due to the genetic control. Pin flowers have the stigma at a higher
78 level than the anthers, and thrum flowers have the anthers at a higher level than the stigma.
79 This conformation allows the level of low and high organs to be reciprocal between floral
80 morphs (reciprocal herkogamy). Tristylous species, in turn, produce short-, mid- and long-style
81 morphs that differ reciprocally in the heights at which stigmas and anthers are positioned within
82 flowers (Lloyd & Webb 1992; Barrett 2013). This morphology is associated with a
83 heteromorphic self-incompatibility system, which only enables fertilization in pollination
84 crosses between plants of different morphs. As a result, heterostyly is expected to promote
85 disassortative pollen transfer between different morphs optimizing cross-pollinations (Barrett
86 2019). Thus, due to the heteromorphic self-incompatibility system, isoplethy, and the reciprocal
87 herkogamy, the reproduction of heterostylous species relies on legitimate cross-pollinations
88 (*i.e.*, intermorph cross-pollinations) mediated by biotic pollinators (Ganders 1979; Barrett
89 2013).

90 Although heterostyly is followed by several basal features, variations within
91 heterostylous populations are frequent and may arise as a consequence of pollinator availability
92 and behavior, genetic mutations or recombination, environmental pressure or disturbances
93 (Sobrevila *et al.* 1983; Washitani *et al.* 1994; Consolaro *et al.* 2011), as well as reduced
94 population size (Kéry *et al.* 2000), clonal reproduction (Barrett 1977), geographic isolation or
95 habitat fragmentation (Van Rossum *et al.* 2006). These factors may lead to the breakdown of
96 the heteromorphic self-incompatibility system and the loss of heterostyly in distinct populations
97 of the same species, often leading to the evolution of homostylous populations, or even to an
98 alternative outcrossing system (Baker 1955; Charlesworth & Charlesworth 1979, Barrett and
99 Shore 1987; Coelho & Barbosa 2003; Consolaro *et al.* 2005; Toledo *et al.* 2007; Barrett &

100 Shore 2008; Consolaro *et al.* 2009, Consolaro *et al.* 2011, Bramow *et al.* 2013; Rodrigues *et al.*
101 2013; Costa *et al.* 2017; Barrett 2019). Under these circumstances, heterostyly may represent a
102 reproductive constraint for some plant populations, due to the absence of mates or actual
103 pollinators. As a result, many species may suffer from a limitation in the pollination services
104 (Sobrevila *et al.* 1983, Washitani *et al.* 1994; Bramow 2013).

105 Although abiotic conditions may affect floral morphology (Etterson *et al.* 2016),
106 pollinator-mediated selection can also play a crucial role in the evolution of floral traits
107 (Stebbins 1970; Agrawal 2001; Kearns *et al.* 1998; Klein *et al.* 2007; Aguilar *et al.* 2006; Menz
108 *et al.* 2011; Schiestl & Johnson, 2013). Stebbins (1970) postulates the principle of the most
109 effective pollinator, in which an animal group with a high frequency of visits and effectiveness
110 as pollinators, can mediate the selection of floral traits that will increase plant fitness in a given
111 location where the species has evolved. In this sense, some floral attributes subjected to
112 phenotypic variation can be selected throughout the plant distribution due to a higher
113 attractiveness of a given pollinator group, or a higher plant-pollinator morphological matching,
114 which increases plant fitness, and can be fixed in the population (Anderson & Johnson 2008;
115 Pauw *et al.* 2009). Whereas bees are excluded from flowers on the red spectrum since they lack
116 visual conspicuity due to the characteristics of their visual system (Chittka & Waser 1997;
117 Lunau *et al.* 1996; Dyer *et al.* 2016; Papiorek *et al.* 2016), and hummingbirds are associated
118 with red flowers (Faegri & van der Pijl 1979; Lunau *et al.* 2011; Bergamo *et al.* 2016; Camargo
119 *et al.* 2018), flowers in other color spectrum are more conspicuous to bees, improving bee
120 pollination.

121 In this sense, considering the phenotypic variation in *Palicourea rigida* populations
122 either in life form and floral morphology, as well as its dependence on pollinators for sexual
123 reproduction, in this study we described the changes in the floral morphology of different
124 populations of *P. rigida* and tested if they are followed by alterations of the basal features of
125 distyly, *i.e.*, breakdown of the heteromorphic self-incompatibility system, reciprocal herkogamy
126 and isoplethy. Additionally, due to the change in flower color and size, we discussed the
127 hypothesis of a potential pollinator shift from hummingbirds (ornithophily) to bees
128 (melitophily) in the dwarf population presenting yellow flowers as a possible outcome of
129 pollinator-mediated selection. This population is located at the southernmost distribution of
130 *Palicourea rigida* in the Brazilian Cerrado, which presents consistent morphological alterations
131 from the typical plant populations in the core region of the distribution of the species. To better
132 understand the southernmost population, we also compare it with two populations, one
133 presenting the usual phenotypic features described for the species, and a second representing
134 another extreme of the phenotype distribution of *Palicourea rigida*, where the plants are trees
135 with long-tubed reddish flowers.



136

137 **Figure 1.** Phenotypic variation of *Palicourea rigida* populations across its distribution in the Brazilian
 138 Cerrado: (A) Clube Caça e Pesca de Uberlândia, Uberlândia - MG; (B) Conservation Unit of Botucatu,
 139 Botucatu - SP, at the southernmost limit of the species distribution; (C) Chapada dos Veadeiros National
 140 Park, Alto Paraíso de Goiás - GO.



141

142 **Figure 2.** Phenotypic variation of *Palicourea rigida* flowers and inflorescences across its distribution in
 143 different sites of the Brazilian Cerrado: (A) Chapada dos Veadeiros National Park, Alto Paraíso de
 144 Goiás - GO; (B) Clube Caça e Pesca de Uberlândia, Uberlândia - MG; and (C) Conservation Unit of
 145 Botucatu, Botucatu - SP, at the southernmost limit of the species distribution.

146

147 MATERIAL AND METHODS

148

149 SPECIES CHARACTERIZATION AND STUDY SITES - *Palicourea rigida* is a Rubiaceae
 150 species distributed in the Brazilian Cerrado and disjunct areas of amazon savannah, found in
 151 37% of the Cerrado areas until now (Taylor, 1997, Ratter *et al.* 2003). Flowers producing
 152 nectar are receptive for only 1 day and are grouped into thyrsoid inflorescence with
 153 zygomorphic flowers and the plant has characteristic large coriaceous leaves (Robbrecht 1988,
 154 Machado 2010). The blooming season occurs during the rainy season, varying throughout the
 155 distribution range of the species (Robbrecht 1988; Machado *et al.* 2010).

156 This work was conducted in three study sites between the years of 2017 and 2019. The
 157 first site, where plants are dwarfs, is a Cerrado area in the Conservation Unit of Botucatu
 158 (BTU), municipality of Botucatu, São Paulo state, Brazil (22°56'S, 48°27'O). The climate is
 159 warm-temperate with two distinct seasons, a cold dry winter (from March to September), and a
 160 hot wet summer (from October to April), Cwa according to the Köppen classification, and with

161 an annual rainfall of 1336 mm (Köppen 1948; Alvares *et al.* 2013). The main study site is an
162 integral protection conservation unit that comprehends a Cerrado vegetation characterized by
163 shrub-tree vegetation, with sparsely distributed trees and treelets, and scattered shrubs and sub-
164 shrubs (Ribeiro e Walter, 1998). The total area comprehends 33.8 hectares, located at an
165 altitude of 860 meters a.s.l.. The second site, where the population presents the typical
166 morphology of the species, is located at Serra de Caldas Novas State Park (Caldas), Caldas
167 Novas, Goiás State, Brazil (17°43'S, 48°40'O) and has a total area of 12,500 hectares. Finally,
168 the third site is located at Chapada dos Veadeiros National Park (CV), Alto Paraiso de Goiás,
169 Goiás State, Brazil (47°30'S, 14°02'O) with an area of 240.611 ha, where *P. rigida* plants are
170 trees. Serra de Caldas Novas State Park and Chapada dos Veadeiros National Park comprehend
171 a Cerrado *strictu sensu*, with seasonal tropical climate with summer rains, Aw. according to
172 Köppen classification, and with annual rainfall averaging about 1500 mm (Alvares *et al.* 2013).

173 FLORAL MORPHOMETRIC AND RECIPROCAL HERCOGAMY- To test the reciprocal
174 hercogamy and also to evaluate the floral phenotypic variation among sites, we measured
175 flowers from the three studied populations. For this, we collected 4-5 flowers per plant,
176 totaling 1127 flowers (fixed in 70% alcohol solution) distributed in 238 plants sampled in three
177 populations: 61 plants in Caldas, 100 in Chapada, and 77 in BTU. In these flowers we
178 measured the length of the following structures using a digital caliper: 1) corolla tube length; 2)
179 diameter of the corolla; 3) anther length; 4) anther height; 5) stigma length; 6) stigma height; 7)
180 distance between stigma and anther heights. To avoid overestimation of the anther height, we
181 measured from the bottom of the corolla until 2/3 of the anther length, and to avoid the stigma
182 height overestimation we measured from the basis of the corolla until the middle of the stigma
183 length. We used an analysis of variance (ANOVA) with Tukey's posthoc test for multiple
184 comparisons among pairs of means to evaluate the phenotypic variances of floral verticiles
185 among populations considering their floral morphs (Zuur *et al.* 2007).

186 To assess the hercogamy in the populations of *P. rigida*, we use the reciprocity index of
187 Armbruster (2017), which are the most updated analyses for heterostyly measure. The index
188 uses the measure of heights of the reproductive organs, anther, and stigma, to enable the
189 estimation of the ability to achieve an inter-morph cross-pollinations and the relation of this
190 ability to reproductive fitness (Armbruster *et al.* 2017).

191 The Armbruster's index assesses the adaptative inaccuracy (I), which measures the
192 degree of maladaptation of a floral morph on a fitness scale that regards both the mean and
193 variance of the phenotypic values of the morph (Armbruster *et al.* 2004; Hansen *et al.* 2006,
194 Armbruster *et al.* 2017). We calculate the adaptive inaccuracy of low (pin anthers and thrum
195 stigma) and high organs (thrum anthers and pin stigma), and the total inaccuracy (sum of both
196 inaccuracies) to better evaluate the floral differences between the populations and floral morphs

197 and the maladaptive bias, which is a result of the mismatch of the means of the anthers and
198 stigma (the square of the departure of the trait mean from the optimum). Besides, we calculated
199 the total mean inaccuracy (Mean^2) standardized by the squared mean of all anther and stigma
200 heights in each population, which will be most used and will facilitate comparisons across
201 different populations. In this test, the closer to zero the value obtained, the lower the
202 inaccuracy, and the greater the reciprocity of sexual organs (Armbruster *et al.* 2017).

203 **MATING SYSTEM AND ISOPLETY-** To evaluate the reproductive system of the
204 population of *P. rigida* in the southernmost distribution (BTU), we covered inflorescences with
205 nylon mesh bags to avoid interference of floral visitors, and conducted the following controlled
206 pollination experiments using 4 to 5 flowers per plant: 1) hand self-pollination, where we tested
207 for self-incompatibility (n = 87 flowers from 19 individuals); 2) intra-morph cross-pollination,
208 *i.e.*, crosses between distinct individuals of the same flower morph (n = 98 flowers in 19
209 plants); 3) inter-morph cross-pollinations, *i.e.*, crosses between distinct individuals of different
210 flower morphs, (n = 98 flowers in 19 plants); 4) autogamy, flowers without pollination covered
211 in nylon mesh bags (n = 181 flowers from 37 individuals); 5) Control, flowers in natural
212 conditions naturally exposed to the floral visitors (n = 9866 in 79 plants, 4582pin and 5184
213 thrum).

214 Cross pollinations were carried out in both directions, using pollen of thrum flowers
215 (anthers at a higher level than stigma) to pollinate the pin ones and vice versa. After the
216 experiments, the flowers were isolated again to analyze the fruit set, which was assessed 30-60
217 days after pollination treatments. Also, we used a two-sample *t*-test to assess whether the
218 proportion of fruits formed in natural conditions (control) differed between the floral morphs.
219 The index of self-incompatibility (ISI) was calculated as the ratio between the proportion of
220 fruits formed after hand self-pollination and hand cross-pollination. The index indicates that
221 species with values under 0.25 are self-incompatible, values between 0.25 and 0.75 indicate
222 self-incompatibility with some level of self-compatibility (pseudocompatibility), and values
223 above 0.75 indicate self-compatibility (Bullock 1985, Lloyd & Schoen 1992). We also used an
224 index to measure the reproductive efficacy (IRE) of the population. This index was calculated
225 by dividing the average number of fruits per flowers hand-self-pollinated by the same
226 following Inter-morph cross-pollination (*sensu* Zapata & Arroyo 1978). Index values above
227 0.66 indicate high reproductive efficiency of species due to a pollen flow promoted by
228 pollinators (Zapata & Arroyo 1978) and values similar to or less than 0.25 indicate pollen
229 limitation (Sobrevilla & Arroyo 1982). The reproductive system of Caldas population was
230 assessed by Machado *et al.* (2010) and Maruyama *et al.* (2016).

231 To test for isoplethy, we quantify the number of individuals of each morph (*i.e.*, pin or
232 thrum) in all studied populations. The occurrence of isoplethy in the populations was assessed

233 with a Chi-square goodness of fit test, where the populations were considered isoplethic when
234 they have equal proportions for each morph. All analyses were performed using R software
235 (Version 4.0.2; R. Core Team, 2020)

236

237 FLORAL COLOR, BEE PERCEPTION AND POLLINATION BIOLOGY- To assess the
238 differences in flower color between different populations, we modeled the color of flowers
239 according to the bumblebee visual system using newly opened flowers from 8 different
240 individuals in BTU and 14 from Caldas (n= 1 - 5 flowers per individual). From each flower, we
241 measured the spectral reflectance of the corolla (tip and base) by using a Jazz spectrometre
242 (Ocean Optics S2000, Ocean Optics, Dunedin, FL) relative to a calibrated white DH 2000-CAL
243 (Ocean Optics) and black (light absence) sources. We assumed a standard green background
244 (Chittka and Kevan 2005), a daylight illumination (D65; Wyszecki and Stiles 1982) and the
245 spectral sensitivity of *Bombus terrestris* as surrogate, since the plant species are often visited by
246 the sister species *Bombus morio*. We used the color hexagon model (Chittka 1992; Lunau *et al.*
247 2011) to predict the color discrimination capacity of bees to the plant species. We established
248 the discrimination threshold of 0.09 hexagon units based on previous behavioral experiments
249 (Dyer *et al.* 2008).

250 To estimate the ability of bumblebees to discriminate flower colors, we use linear
251 mixed-effect models analysis (LMMs). The dependent variables were the mean Euclidian
252 distance in hexagon units between BTU and Caldas, measured from the corolla tip or base, in
253 both populations, we use as fixed factors the paired Euclidian distances measured, and the
254 values of x- and y-axis of the color hexagon as the response variable. Individual plants were
255 included as a random factor. To perform the LMM, we used the *lme4* package version 1.1-21
256 (Bates *et al.* 2019) and the *lmer* function. To determine the significance of each fixed factor
257 (morphs and populations) and their interaction, a type II analysis of variance was conducted
258 using the *Anova* function in the *car* package version 3.0-2 (Fox *et al.* 2019). As a reference,
259 bumblebees can distinguish correctly by 60% between colors with 0.09 hexagon units of
260 perceptual distance (Dyer, 2006). From these color loci, we also calculated the green contrast
261 using the same color hexagon model (Chittka *et al.* 1994; Lunau *et al.* 2011). The green
262 contrast is measured as the distance between the target color locus and the central point of the
263 color hexagon representing the locus of the standard green background (Chittka, 1992).

264 To determine the pollinators of the BTU population, we performed 31 hours of focal
265 observations between 07:00h and 18:00h, being 12 hours in 2017 and 18 hours in 2019.
266 Observations were made in sessions of 30 minutes per plant during the flowering period of the
267 species and floral visitors were photographed for identification. The pollinators of Caldas and

268 Chapada were assessed by Silva 1995, Machado *et al.* (2010), and Maruyama *et al.* (2016), but
 269 we also have conducted about 15 hours of observations at the Chapada site to record the floral
 270 visitors and the actual pollinators.

271 All analyses were performed using R software (Version 4.0.2; R. Core Team, 2020)

272

273

274 RESULTS

275 FLORAL MORPHOMETRIC AND RECIPROCAL HERKOGAMY - Floral morphology
 276 differed among the three analyzed populations in several parameters, varying among morphs
 277 and populations compared (Table 1, Fig. 3). The BTU population has the smallest mean corolla
 278 length in both floral morphs, while Chapada has the highest, BTU is also different in corolla
 279 length with both populations and pin and thrum morphs ($F=4.6314$, $P < 0.05$). Considering
 280 corolla diameter, BTU has also the smallest diameter in both morphs, while Chapada has the
 281 largest, BTU also differs from Chapada in both morphs ($F=0.4736$, $P < 0.05$), while when
 282 compared with Caldas, the diameter is only different in thrum morph ($F=0.4736$, $P < 0.05$).
 283 Regarding reproductive structures (Fig. 4), BTU has the smallest stigma heights in pin and
 284 thrum flowers and is also different from all populations in both floral morphs ($F=1.3811$, $P <$
 285 0.05). The anther height, BTU is only different in thrum morphs with both populations
 286 ($F=9.3463$, $P > 0.05$). Floral dimorphism remained in all studied populations, with all
 287 measured organs differing between thrum and pin morphs within populations, with exception
 288 of anther length in the BTU population (Table 1).

289 Considering the Inaccuracy index (Armbruster *et al.* 2017), the standardized inaccuracy
 290 (Mean²) of the BTU population is the highest (6.766) and the Caldas have the lowest
 291 inaccuracy (6.370). When we detach low and high organs inaccuracy, Chapada presents a
 292 contrasting difference between their low and high inaccuracy (Table 2), whereas BTU and
 293 Caldas present a more paired index. The highest Inaccuracy proportion in low organs was
 294 found in Caldas and for high organs in the Chapada population. Chapada presented the lowest
 295 inaccuracy proportion of low organs and Caldas has the lowest inaccuracy for high organs. The
 296 maladaptive bias has shown that BTU presented the higher maladaptation, while Chapada
 297 presented the lowest.

298

299 MATING SYSTEM AND ISOPLETY - Pollination treatments showed that the
 300 heteromorphic self-incompatibility in BTU remained, with only fruit formation after inter-
 301 morph cross-pollination treatments (Table 3), and as a result, the ISI and the IRE in the
 302 population were equal to zero, showing complete self-incompatibility (Table 3). Population

303 fruit set in natural conditions was 37.98%, but thrum flowers presented a higher fruit set
304 (41.53%) than pin flowers (33.98%; $t = 2.5196$, $p = 0.01$).

305 However, the Chapada deviated from the 1:1 ratio between pin and thrum plants ($\chi^2 =$
306 5.76; $P = 0.0164$), where 38 are pin and 62 are thrum.

307

308 *FLORAL COLOR, BEE PERCEPTION AND POLLINATION BIOLOGY* - Considering the
309 *Palicourea rigida* flower's color discrimination by the bees between the populations, both
310 corolla's tip and base were fully visible to the bee visual system (Fig 6). Although, we have
311 found difference in tip color comparing both populations (Fig 7; $t = -3.2334$, $P < 0.05$), and also
312 within population, comparing tip with base of the same population (BTU $t = -2.8249$ and
313 Caldas $t = -8.4804$, $P < 0.05$). We also found that more than 95% of the bee receptors exited
314 were green receptors, considering both parts of the corolla analyzed.

315 Considering color contrast, BTU presents a very low contrast between tip and base ($t =$
316 -1.5015 , $P > 0.05$), and Caldas population presented a much higher contrast between both
317 corolla's parts analyzed ($t = 15.852$, $P < 0.05$). When we compared the contrast within
318 populations, the tip ($t = -4.4471$, $P < 0.05$) and base ($t = -2.994$, $P < 0.05$) also differed between
319 each other.

320 *Palicourea rigida* in BTU were visited by bees, wasps, and butterflies, but bees were
321 the main pollinators of this population. We recorded a total of 121 visits in 31 hours of
322 observations, of which 79 were from bees of the genus *Bombus*, *Xylocopa*, and *Centris*, 22
323 visits were made by small-sized bees (Meliponini and Halictidae), 15 from *Epicharis cockerelli*
324 and five by butterflies. Large-sized bees were the main pollinators of *P. rigida* in the BTU
325 population, particularly, *Bombus morio* which was the most frequent flower visitor performing
326 57.5% of the visits (Fig. 5A). In Chapada, we recorded bees (*Bombus*, *Xylocopa*, *Centris*, and
327 *Trigona*) and hummingbirds visiting the *P. rigida* flowers (Fig. 5B). The bees were observed
328 foraging all day long, but due to the long floral tube, most of them acted only as nectar robbers
329 and occasional pollinators. Hummingbirds, in turn, were quite frequent, performing about five
330 visits per hour per plant, visiting many flowers in each foraging bout.

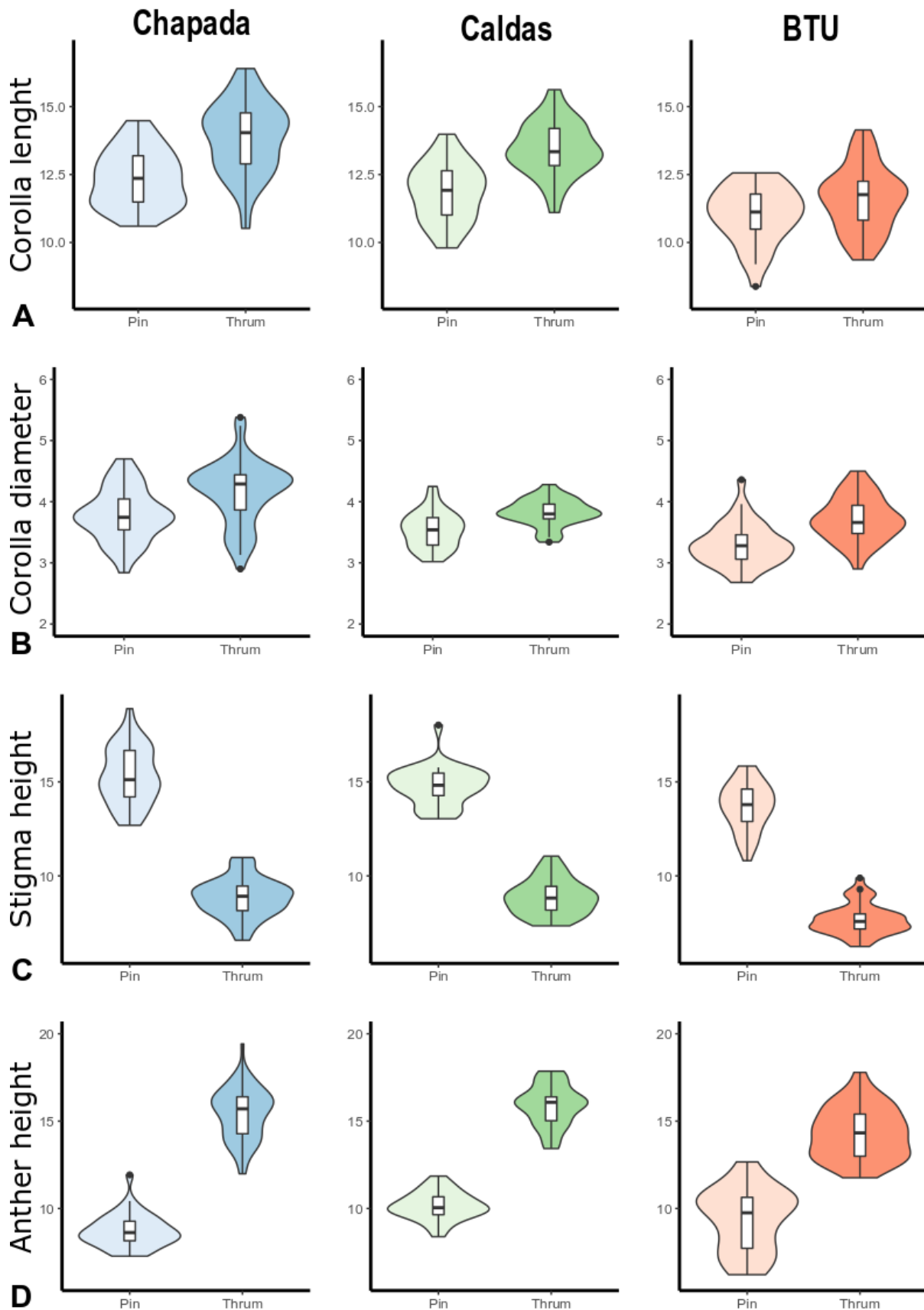
331

332

333 **Table 1.** Floral morphometrics of *Palicourea rigida* in Chapada dos Veadeiros National Park, Alto
 334 Paraíso de Goiás - GO (Chapada); Serra de Caldas Novas State Park, Caldas Novas - GO (Caldas); and
 335 at Conservation Unit of Botucatu, Botucatu - SP (BTU) (mean \pm standard deviation). N = number of
 336 individuals analyzed. Means followed by different letters indicate a statistical difference at $P < 0.05$
 337 between morphs in each population. P values of t-test analysis for comparison among floral morphs.
 338

Measured items (mm)	Chapada		Caldas		BTU	
	thrum N=62	pin N=38	thrum N=30	pin N=31	thrum N=41	pin N=36
Corolla length	13.86 \pm 1.31a	12.35 \pm 1.03b	13.45 \pm 0.98a	11.85 \pm 1.06b	11.60 \pm 1.18a	11.04 \pm 1.02b
Corolla diameter	4.15 \pm 0.26a	3.78 \pm 0.18b	3.83 \pm 0.22a	3.54 \pm 0.30b	3.71 \pm 0.36a	3.30 \pm 0.35b
Anther length	3.48 \pm 0.11a	3.28 \pm 0.11b	3.80 \pm 0.29a	3.38 \pm 0.29b	3.07 \pm 0.31a	2.96 \pm 0.28a
Anther height	15.42 \pm 2.15a	8.70 \pm 0.89b	14.79 \pm 1.20a	10.15 \pm 0.620b	14.31 \pm 2.25a	9.47 \pm 3.36b
Stigmatic lobe length	4.20 \pm 1.01a	0.96 \pm 0.05b	4.01 \pm 0.54a	0.72 \pm 0.22b	4.54 \pm 0.83a	3.31 \pm 0.45b
Stigma height	8.85 \pm 1.04a	15.31 \pm 2.28b	8.87 \pm 0.87a	14.79 \pm 1.05b	7.68 \pm 0.66a	13.69 \pm 1.59b

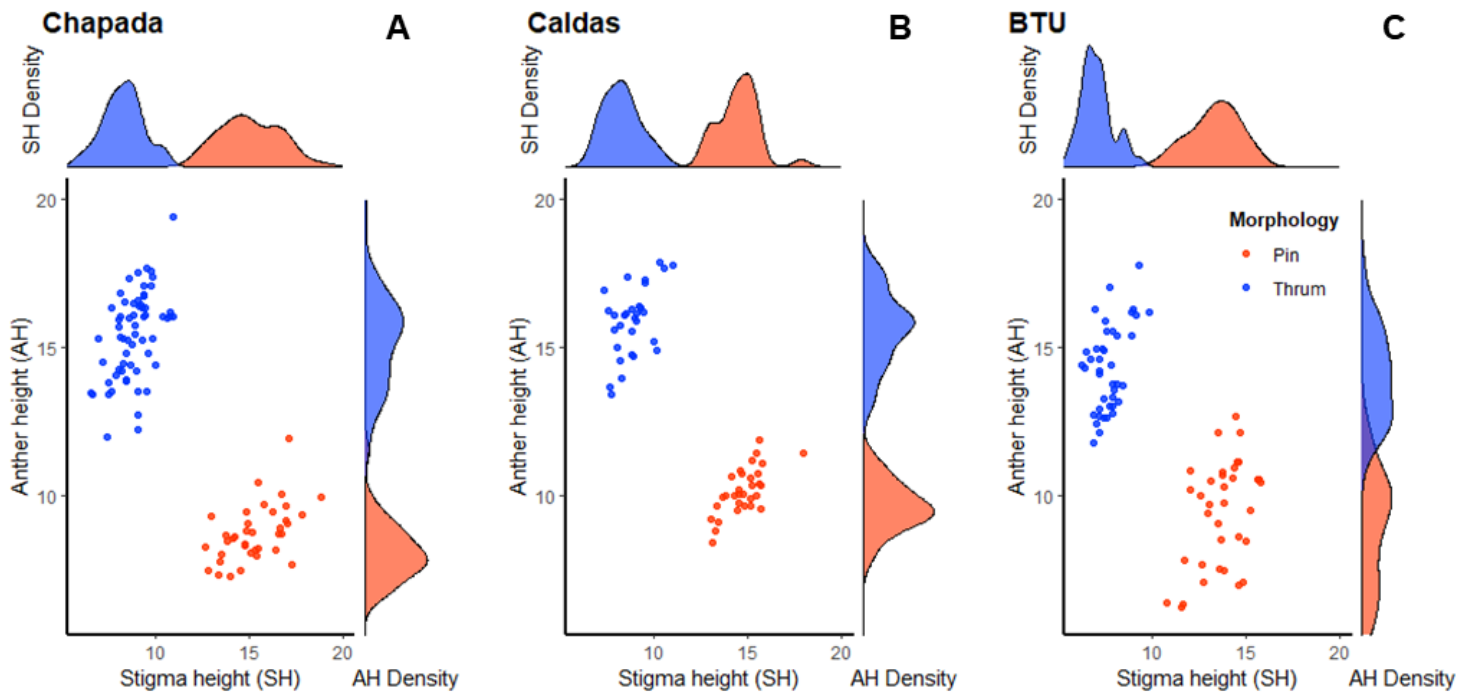
339



340

341 **Figure 3.** Variation of *Palicourea rigida* floral morphology across its distribution in the Brazilian
 342 *Cerrado* at Chapada dos Veadeiros National Park, Alto Paraíso de Goiás – GO (A); Serra de Caldas
 343 Novas State Park, Caldas Novas - GO (B); and Conservation Unit of Botucatu, Botucatu – SP (C). The
 344 measurements are in millimeters. Colors represent the density of the distribution of each population.

345



346

347 **Figure 4.** Distribution of anther and stigma heights across its distribution in the Brazilian Cerrado in
 348 different population of *Palicourea rigida* across its distribution in the Brazilian Cerrado: A) Chapada
 349 dos Veadeiros National Park, Alto Paraíso de Goiás - GO (Chapada), B) Serra de Caldas Novas State
 350 Park, Caldas Novas - GO (Caldas) and at C) Conservation Unit of Botucatu, Botucatu -SP (BTU).
 351 Height and length are in millimeters.

352

353

354 **Table 2.** Summary statistics of heterostyly reciprocity indexes calculated of *Palicourea rigida*
 355 populations in Chapada dos Veadeiros National Park, Alto Paraíso de Goiás - GO (Chapada), Serra de
 356 Caldas Novas State Park, Caldas Novas - GO (Caldas) and at Conservation Unit of Botucatu, Botucatu
 357 - SP (BTU).

Populations	Inaccuracy Low	Inaccuracy High	Total inaccuracy	Mean ²	Maladaptive bias
Chapada	1.950 (30.46%)	4.452 (69.54%)	6.402 (100%)	4.392	0.032
Caldas	3.138 (49.26%)	3.232 (50.74%)	6.370 (100%)	4.158	2.4305
BTU	3.138 (46.38%)	3.628 (53.62%)	6.766 (100%)	4.381	2.8265

358

359

360

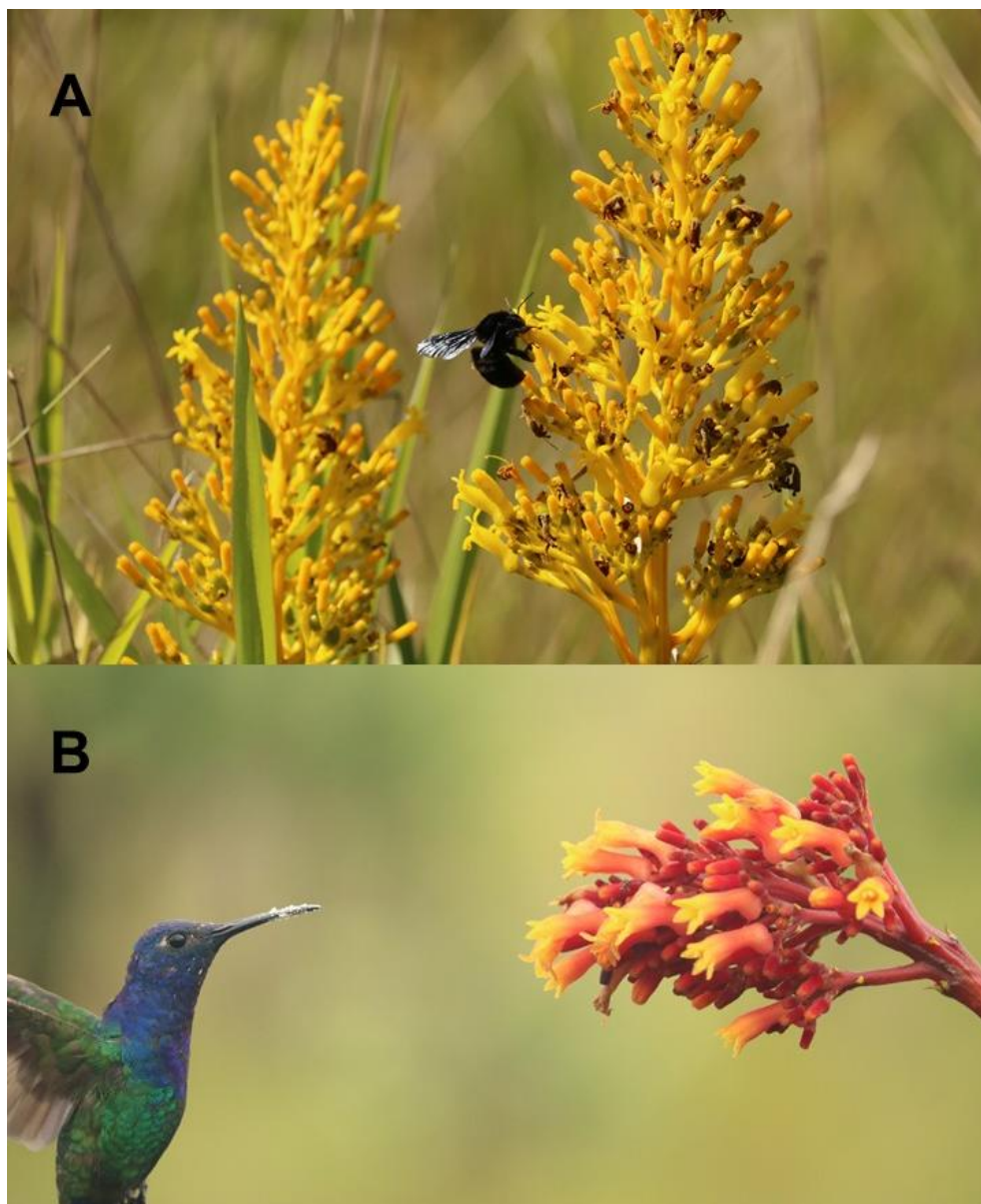
361

362

Table 3. Fruit set of *Palicourea rigida* after experimental pollination treatments in a Cerrado area in Conservation Unit of Botucatu, Botucatu - SP. T: thrum; P: pin. () = number of formed fruits.

Pollination treatments		Flowers	Fruits set (%)
Hand-self-pollination	T	48	0 (0)
	P	39	0 (0)
Inter-morph cross-pollination	T	48	4 (8.3)
	P	50	6 (12)
Intra-morph cross-pollination	T	52	0 (0)
	P	46	0 (0)
Autogamy	T	70	0 (0)
	P	111	0 (0)
Control	T	5184	2153 (41.5)
	P	4582	1556 (33.9)

363



364

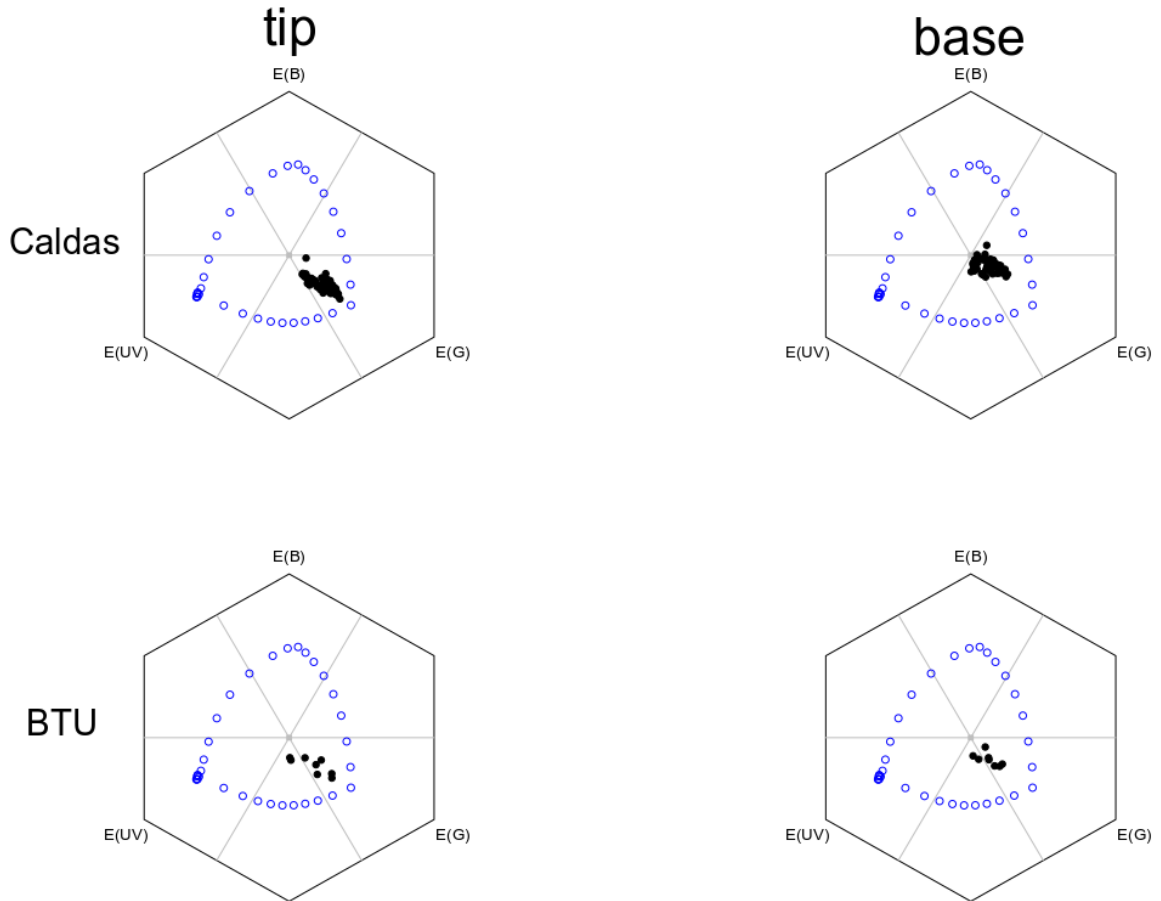
365

366

367

Figure 5. Pollinators visiting *Palicourea rigida* flowers **A-** *Bombus morio* visiting pin flowers at Conservation Unit of Botucatu, Botucatu - SP. **B-** *Eupetomena macroura* visiting pin flowers at Chapada dos Veadeiros National Park, Alto Paraíso de Goiás - GO

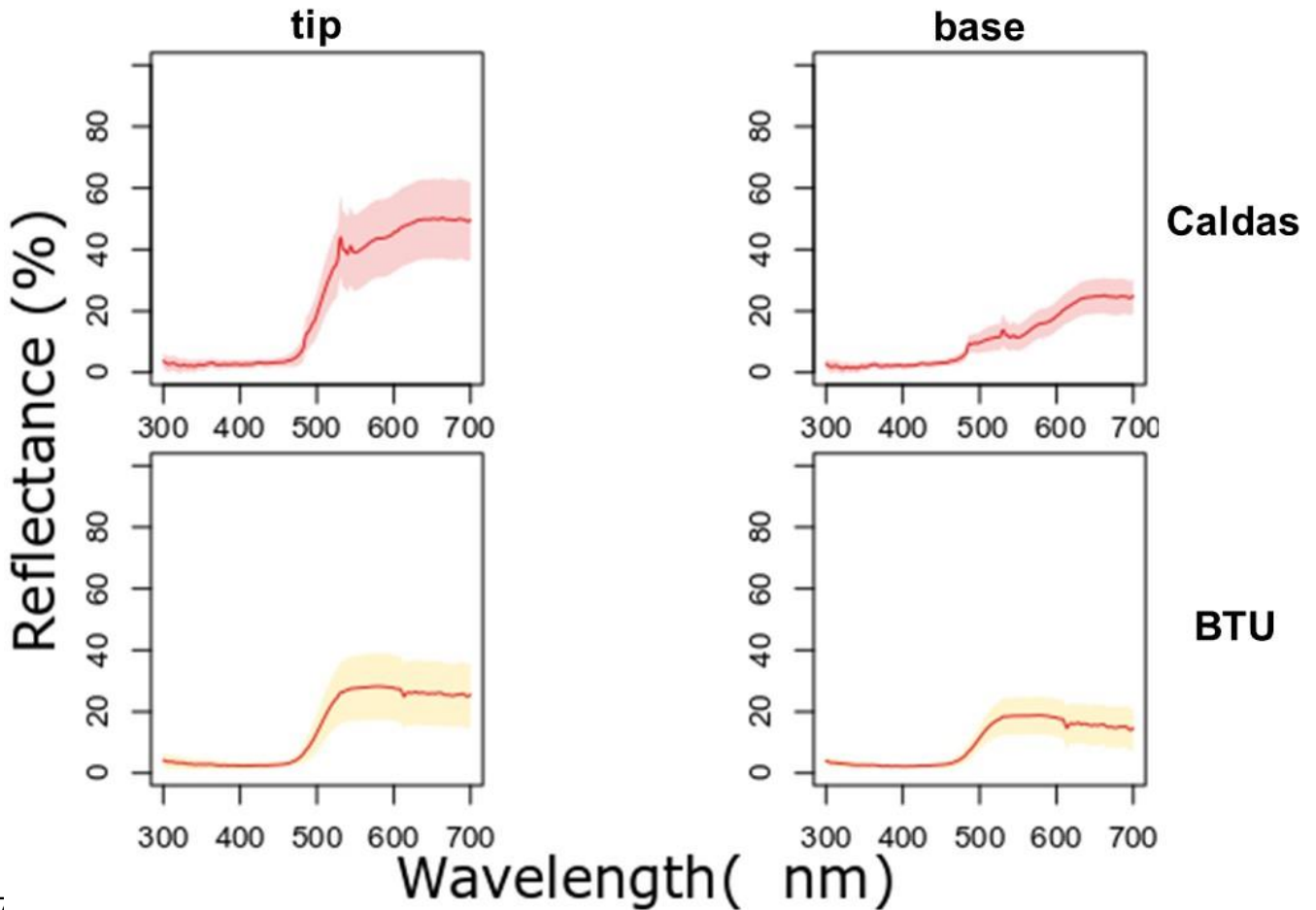
368



369

370 **Figure 6.** Color hexagon representation for bee vision of *Palicourea rigida* flowers, tip or base in two
 371 populations, Caldas and Botucatu. The grey central point in the visual spaces represents the achromatic
 372 center, and the vertices represent each bee photoreceptor: ultraviolet E(UV), blue E(B), and green E(G),
 373 the grey line represents the maximum sensitivity of each photoreceptor to a monochromatic light and
 374 the blue dots represents the bee visual detection. The excitation of ultraviolet, blue, and green
 375 photoreceptor types is indicated with respective black points in the hexagon.

376



377
378
379
380
381

Figure 7. Spectral relative reflectance of *Palicourea rigida* flowers. The red line represents the mean reflectance, and the shaded area represents the standard variation on reflectance.

382 DISCUSSION

383 Our results show that the basal characteristics of distyly remained in the dwarf-
384 population at the southernmost distribution of *Palicourea rigida* in Brazil, despite the
385 contrasting morphology observed in this population in relation to the populations occurring in
386 central Brazil. Despite that, the dwarf population presents the smallest corolla length, as well
387 as, the smallest anther and stigma heights. The heteromorphic self-incompatibility and
388 isoplethy were also maintained, which preserve the basal characteristic of heterostyly, which
389 means that the species relies strictly on pollinators for reproduction. The efficiency of
390 pollination depends on how functional pollinators groups get high- and low-level pollen
391 deposited on different body parts, which can differ according to the morphology and length of
392 buccal parts, body size and fit with the flower morphology (Ornelas et al., 2004; Massinga et
393 al., 2005; Armbruster et al., 2006; Zhu et al., 2015; Deschepper et al., 2018). This population is
394 visited and pollinated mainly by large-sized bees, such as *Bombus morio*, which have a much
395 smaller buccal length when compared to hummingbirds, which act as the main pollinator of
396 other *P. rigida* populations in Brazil. Thus, according to the Stebbins (1970) postulation of the
397 principle of the most effective pollinator, an animal group with a high frequency of visits and

398 effectiveness as pollinator can mediate the selection of floral traits that will increase plant
399 fitness in a given location where the species has evolved. Hence, we suggest that the pollinator-
400 mediated selection by bees had led to a decrease in the corolla tube length of the dwarf
401 population of *P. rigida*. This kind of pollinator shift (hummingbird pollination to bee
402 pollination), is relatively common, and the evidence for multiple reverse transitions from
403 presumed ancestral hummingbird pollination to more derived bee or insect pollination are well
404 documented in another plant family (Tripp & Manos 2008).

405 Furtado (2019) analyzed the inaccuracy of seven *Palicourea rigida* populations in the
406 central distribution of the species in Brazil, and the dwarf population analyzed in this study
407 presented an inaccuracy lower than the five populations studied. Also, taking into account the
408 populations we have analyzed the dwarf population presents an inaccuracy lower than seven
409 populations, in a total of 10, so far studied in Brazil. This shows that the dwarf population
410 presents accurate reciprocity higher than most populations already analyzed. This can be
411 corroborated by the high fruit set in natural conditions found in the dwarf population, which are
412 within the expected for the hummingbird-pollinated populations (Machado *et al.* 2010),
413 showing that bee pollination is as efficient as hummingbird pollination in this species. Hence,
414 bees are very effective pollinators of the dwarf population, suggesting the occurrence of a
415 pollinator shift in this southernmost population of *Palicourea rigida*.

416 Despite that, when comparing the floral morphology with the populations from Caldas
417 and Chapada we noticed a difference between the main morphological parameters involved in
418 reproduction, such as corolla size, diameter, and height of reproductive organs. The dwarf
419 population has the smallest flower tube, with the lowest position of the reproductive organs,
420 however, presents the largest stigmatic lobe of pin flowers among all populations, which is 3.4
421 and 4.6 times larger than the Chapada and Caldas populations, respectively. Raupp *et al.* (2020)
422 have found that the *P. rigida* flowers in three populations studied presented greater reciprocity
423 in low organs, which in association to the larger stigmas of thrum flowers, enable a greater
424 legitimate pollen deposition on thrum flowers. The factors that could affect pin stigmas from
425 receiving a higher amount of legitimate pollen are the shorter length of the stigmatic lobes, as
426 well as the lower levels of reciprocity found in high organs (Furtado *et al.* 2020; Raupp *et al.*
427 2020). The average length previously found of pin stigmas was (1.10 mm), and in thrum ones
428 (3.38 mm), which presents a ratio of 3.7 times larger (Raupp *et al.* 2020). Considering the
429 populations that we analyzed, Chapada has the thrum stigma four times larger than the pin
430 flowers, and in Caldas, this rate was 4.6 times larger, while in the dwarf population (BTU) this
431 ratio is only 1.37. The dwarf population presents a mean pin stigma length of 3.31 mm and the
432 average length of thrum was 4.54 mm. Thereby, the larger stigmatic lobe found in BTU, as a
433 consequence, can increase the legitimate pollen deposition rate, once the larger stigmatic lobes,

434 larger the surface for pollen reception (Furtado *et al.* 2020). With that, since the species rely on
435 legitimate crosses for sexual reproduction (Ganders 1979; Barrett 2013), a greater legitimate
436 pollen deposition could proportionate a higher reproduction rate in a population.

437 Our results show that even though the BTU population presents a much larger stigmatic
438 lobe in comparison to the other studied populations (Machado *et al.* 2010), plants producing pin
439 flowers do not have a higher fruit set such as we suggested. These results suggest that the lower
440 levels of reciprocity in high organs (stigmas of pin flowers and anthers of the thrum ones)
441 founded in all *P. rigida* populations analyzed so far, seems to more likely to cause low
442 legitimate pollen deposition in high stigmas, leading to lower fruit set (see also Furtado *et al.*
443 2020, Raupp *et al.* 2020). Our results show that even with a larger stigmatic surface, the
444 reproduction rate of pin flowers in the dwarf population was not significantly higher when
445 compared with other *P. rigida* populations with a smaller stigmatic surface. Therefore, the level
446 of inaccuracy observed in the high organs seems to play a more important role for fruit
447 formation of pin flowers, than the size of the stigmatic lobes.

448 Moreover, regarding floral color and bee perception of *P. rigida* flowers, although the
449 dwarf population presents a different corolla's color compared with Caldas population, such
450 difference seems to do not play a significant role from the perspective of the bee visual system
451 model used in this study (*i.e.* the *Bombus terrestris* visual system), once the flowers seem to be
452 equally visible to the bee visual system model used, and the receptors that are excited when
453 exposed to *P. rigida* flowers were the same in both populations, suggesting the flowers are
454 essentially the same between populations, even the dwarf population presenting a marked
455 yellowish color from the human visual system perspective (Chittka 1992; Lunau *et al.* 2011).
456 Considering the color contrast, the dwarf population presented no contrast between tip and
457 base, so flowers are homogeneously yellowish, while the Caldas population presents a
458 contrasting color pattern between the tip and the base of the corolla. In this sense, reddish color
459 may be an attribute selected by hummingbirds in the populations pollinated by these animals,
460 while in the dwarf population in BTU, which is pollinated by bees, the yellowish color could
461 represent an economy in term of pigment production. However, further analyses are needed to
462 better understand the role of pollinators on this marked difference in the flower color of
463 different populations of *P. rigida* across the Brazilian Cerrado.

464 Thereby, our results show a *Palicourea rigida* pollinated by bees, highlighting a
465 pollinator-shift in the dwarf population of the species in its southernmost distribution, where
466 bees are the main pollinators of the area and promote the reproductive success that maintains
467 the heterostyly in the population. We suggest that the smaller size of *P. rigida* flowers observe
468 in the dwarf population is a result of pollinator-mediated selection of floral traits driven by
469 bees. We also elucidate the role of stigmatic surface in the legitimate pollen deposition, where

470 the lower legitimate pollen deposition on high-level stigmas seems to be due to the unevenness
471 of reciprocity between the low- and high organs level inaccuracy, rather than the smaller stigma
472 in high organs. Thereby, our work helps to better understand the reproduction of distylous
473 species and the role of pollinators in phenotypic variant species.

474

475 REFERENCES

476

477 Agrawal, A.A. (2001). Phenotypic Plasticity in the Interactions and Evolution of Species.
478 *Science*, 294, 321–326.

479 Aguilar, R., Ashworth, L., Galetto, L. & Aizen, M.A. (2006). Plant reproductive susceptibility
480 to habitat fragmentation: review and synthesis through a meta-analysis. *Ecol Letters*, 9, 968–
481 980.

482 Alvares, C.A., Stape, J.L. & Sentelhas, P.C. (2013). Kooppen’s climate classification map for
483 Brazil. *Meteorol. Z.*, 18.

484 Anderson, B. & Johnson, S.D. (2007). The geographical mosaic of coevolution in a plant-
485 pollinator mutualism: Coevolution between mutualists. *Evolution*, 62, 220–225.

486 Armbruster, S, Pérez-Barrales R, Arroyo J, Edwards ME, Vargas P. 2006. Three-dimensional
487 reciprocity of floral morphs in wild flax (*Linum suffruticosum*): a new twist on heterostyly.
488 *New Phytol* 171: 581-590.

489 Armbruster, W.S. (2004). Floral integration, modularity, and precision: distinguishing complex
490 adaptations from genetic constraints. *Phenotypic Integration: Studying the Ecology and*
491 *Evolution of Complex Phenotypes*, 23–49.

492 Armbruster, W.S., Bolstad, G.H., Hansen, T.F., Keller, B., Conti, E. & Pélabon, C. (2017). The
493 measure and mismeasure of reciprocity in heterostylous flowers. *New Phytol*, 215, 906–917.

494 Baker, H.G. (1955). Self-Compatibility and Establishment After “Long-Distance” Dispersal.
495 *Evolution*, 9, 347.

496 Barrett, S.C.H. & Shore, J.S. (1987). Variation and evolution of breeding systems in the
497 *Turnera ulmifolia* L. complex (Turneraceae). *Evolution*, 41, 340–354.

498 Barrett, S.C.H. & Shore, J.S. (2008). New Insights on Heterostyly: Comparative Biology,
499 Ecology and Genetics. In: *Self-Incompatibility in Flowering Plants*. Springer Berlin
500 Heidelberg, Berlin, Heidelberg, pp. 3–32.

501 Barrett, S.C.H. (1977). Tristyly in *Eichhornia crassipes* (Mart.) Solms (Water Hyacinth).
502 *Biotropica*, 9, 230.

- 503 Barrett, S.C.H. (2019). ‘A most complex marriage arrangement’: recent advances on
504 heterostyly and unresolved questions. *New Phytol*, 224, 1051–1067.
- 505 Barrett, S.C.H. (Ed.). (1992). *Evolution and Function of Heterostyly*. Monographs on
506 Theoretical and Applied Genetics. Springer Berlin Heidelberg, Berlin, Heidelberg.
- 507 Bates D., Maechler M., Bolker B., Walker S. (2015). Fitting Linear Mixed-Effects Models
508 Using lme4. *J of Stat Soft*, 67(1), 1-48. doi:10.18637/jss.v067.i01.
- 509 Bawa, K.S. & Beach, J.H. (2020). Self-Incompatibility Systems in the Rubiaceae of a Tropical
510 Lowland Wet Forest. *American J of Bot*, 70, 9.
- 511 Bergamo, P. J., Rech, A. R., Brito, V. L., & Sazima, M. (2016). Flower colour and visitation
512 rates of *Costus arabicus* support the ‘bee avoidance’ hypothesis for red-reflecting
513 hummingbird-pollinated flowers. *Funct Ecol*, 30(5), 710-720.
- 514 Bramow, C., Hartvig, I., Larsen, S.B. & Philipp, M. (2013). How a heterostylous plant species
515 responds to life on remote islands: a comparative study of the morphology and reproductive
516 biology of *Waltheria ovata* on the coasts of Ecuador and the Galápagos Islands. *Evol Ecol*, 27,
517 83–100.
- 518 Brito, V.L.G., Weynans, K., Sazima, M. & Lunau, K. (2015). Trees as huge flowers and
519 flowers as oversized floral guides: the role of floral color change and retention of old flowers in
520 *Tibouchina pulchra*. *Front. Plant Sci.*, 6.
- 521 Bullock, S.H. (1985). Breeding Systems in the Flora of a Tropical Deciduous Forest in Mexico.
522 *Biotropica*, 17, 287–301.
- 523 Camargo, M.G.G., Lunau, K., Batalha, M.A., Brings, S., Brito, V.L.G. & Morellato, L.P.C.
524 (2019). How flower colour signals allure bees and hummingbirds: a community-level test of the
525 bee avoidance hypothesis. *New Phytol*, 222, 1112–1122.
- 526 Charlesworth, B. & Charlesworth, D. (1979). The Maintenance and Breakdown of Distyly. *The*
527 *American Nat*, 114, 499–513.
- 528 Chittka, L. & Waser, N.M. (1997). Why red flowers are not visible to bees. *Israel J Plant Sci*,
529 45, 169–183.
- 530 Chittka, L. (1992). The colour hexagon: a chromaticity diagram based on photoreceptor
531 excitations as a generalized representation of colour opponency. *J Comp Physiol A*, 170.
- 532 Chittka, L., and Kevan, P.G. (2005). “Flower colours as advertisement”, in Practical Pollination
533 Biology, eds A. Dafni, P.G. Kevan, and B.C. Husband (*Cambridge: Enviroquest Ltd.*),157–230

- 534 Chittka, L., Shmida, A., Troje, N. & Menzel, R. (1994). Ultraviolet as a component of flower
535 reflections, and the colour perception of hymenoptera. *Vision Research*, 34, 1489–1508.
- 536 Coelho, C.P. & Barbosa, A.A.A. (2003). Biologia reprodutiva de *Palicourea macrobotrys* Ruiz
537 & Pavon (Rubiaceae): um possível caso de homostilia no gênero *Palicourea* Aubl. *Rev. bras.*
538 *Bot.*, 26.
- 539 Consolaro, H., Silva, E.B. da & Oliveira, P.E. de. (2005). Variação floral e biologia reprodutiva
540 de *Manettia cordifolia* Mart. (Rubiaceae). *Rev. bras. Bot.*, 28.
- 541 Consolaro, H., Silva, S.C.S. & Oliveira, P.E. (2011). Breakdown of distyly and pin-
542 monomorphism in *Psychotria carthagenensis* Jacq. (Rubiaceae): Distyly in *Psychotria*
543 *carthagenensis*. *Plant Species Biology*, 26, 24–32.
- 544 Consolaro, H., Toledo, R.D.P., Ferreguti, R.L., Hay, J. & Oliveira, P.E. de. (2009). Distilia e
545 homostilia em espécies de *Palicourea* Aubl. (Rubiaceae) do Cerrado do Brasil Central. *Rev.*
546 *bras. Bot.*, 32, 677–689.
- 547 Costa, J., Castro, S., Loureiro, J. & Barrett, S.C.H. (2017). Experimental insights on the
548 function of ancillary pollen and stigma polymorphisms in plants with heteromorphic
549 incompatibility: Disassortative pollination in heteromorphic plants. *Evolution*, 71, 121–134.
- 550 de Villemereuil, P., Mouterde, M., Gaggiotti, O.E. & Till-Bottraud, I. (2018). Patterns of
551 phenotypic plasticity and local adaptation in the wide elevation range of the alpine plant *Arabis*
552 *alpina*. *J Ecol*, 106, 1952–1971.
- 553 Deschepper P, Brys R, Jacquemyn H. 2018. The impact of flower morphology and pollinator
554 community composition on pollen transfer in the distylous *Primula veris*. *Botanical Journal of*
555 *the Linnean Society* 186: 414-424.
- 556 Dyer, A.G. (2006). Discrimination of Flower Colours in Natural Settings by the Bumblebee
557 species *Bombus terrestris* (Hymenoptera: Apidae). *entomologia*, 28, 257–268.
- 558 Dyer, A.G., Boyd-Gerny, S., Shrestha, M., Lunau, K., Garcia, J.E., Koethe, S., *et al.* (2016).
559 Innate colour preferences of the Australian native stingless bee *Tetragonula carbonaria* Sm. *J*
560 *Comp Physiol A*, 202, 603–613.
- 561 Dyer, A.G., Spaethe, J. & Prack, S. (2008). Comparative psychophysics of bumblebee and
562 honeybee colour discrimination and object detection. *J Comp Physiol A*, 194, 617–627.
- 563 Etterson, J.R. & Mazer, S.J. (2016). How climate change affects plants' sex lives. *Science*, 353,
564 32–33.

- 565 F. Hansen, T., J. R. Carter, A. & Pélabon, C. (2015). On Adaptive Accuracy and Precision in
566 Natural Populations. *The American Naturalist*.
- 567 Faegri, K. & Pijl, L. van der. (1979). *The principles of pollination ecology*. Pergamon
568 international library of science, technology, engineering, and social studies. 3d rev. ed.
569 Pergamon Press, Oxford; New York.
- 570 Faivre, A.E. & McDade, L.A. (2001). Population-level variation in the expression of
571 heterostyly in three species of Rubiaceae: does reciprocal placement of anthers and stigmas
572 characterize heterostyly? *Am. J. Bot.*, 88, 841–853.
- 573 Fox J. and Weisberg S. (2019). An {R} Companion to Applied Regression, Third Edition.
574 Thousand Oaks CA: Sage.
- 575 Furtado, M. T., Matias, R., Consolaro, H., & Perez-Barrales, R. (2020). Do reciprocal
576 herkogamy and pollinators affect the legitimate pollen flow in distylous species? *Botanical*
577 *Journal of the Linnean Society*.
- 578 Furtado, MT. (2019). *Biologia da polinização de Palicourea AUBL. E Psychotria L.*
579 *(RUBIACEAE): Variações e funcionalidade da hercogamia recíproca e dos polinizadores*. PhD
580 Tesis. Universidade de Brasília
- 581 Ganders, F.R. (1979). The biology of heterostyly. *New Zealand Journal of Botany*, 17, 607–
582 635.
- 583 Kearns, C.A., Inouye, D.W. & Waser, N.M. (1998). Endangered mutualisms: The Conservation
584 of Plant-Pollinator Interactions. *Annu. Rev. Ecol. Syst.*, 29, 83–112.
- 585 Kery, M., Matthies, D. & Spillmann, H.-H. (2000). Reduced fecundity and offspring
586 performance in small populations of the declining grassland plants *Primula veris* and *Gentiana*
587 *lutea*. *J Ecology*, 88, 17–30.
- 588 Klein, A.-M., Vaissière, B.E., Cane, J.H., Steffan-Dewenter, I., Cunningham, S.A., Kremen, C.,
589 *et al.* (2007). Importance of pollinators in changing landscapes for world crops. *Proc. R. Soc. B*,
590 274, 303–313.
- 591 Liao, H., D'Antonio, C.M., Chen, B., Huang, Q. & Peng, S. (2016). How much do phenotypic
592 plasticity and local genetic variation contribute to phenotypic divergences along environmental
593 gradients in widespread invasive plants? A meta-analysis. *Oikos*, 125, 905–917.
- 594 Lloyd, D.G. & Schoen, D.J. (1992). Self- and Cross-Fertilization in Plants. I. Functional
595 Dimensions. *International J of Plant Sci*, 153, 358–369.

- 596 Lloyd, D.G. (1992). Self- and Cross-Fertilization in Plants. II. The Selection of Self-
597 Fertilization. *International J of Plant Sci*, 153, 370–380.
- 598 Lunau, K., Papiorek, S., Eltz, T. & Sazima, M. (2011). Avoidance of achromatic colours by
599 bees provides a private niche for hummingbirds. *J of Exp Biology*, 214, 1607–1612.
- 600 Lunau, K., Wacht, S. & Chittka, L. (1996). Colour choices of naive bumble bees and their
601 implications for colour perception. *J Comp Physiol A*, 178.
- 602 Machado, A. de O., Silva, A.P., Consolaro, H., Barros, M.A.G. e & Oliveira, P.E. (2010).
603 Breeding biology and distyly in *Palicourea rigida* H. B. & K. (Rubiaceae) in the Cerrados of
604 Central Brazil. *Acta Bot. Bras.*, 24, 686–696.
- 605 Maruyama, P.K., Justino, D.G. & Oliveira, P.E. (2016). Does intraspecific behavioural
606 variation of pollinator species influence pollination? A quantitative study with hummingbirds
607 and a Neotropical shrub. *Plant Biol J*, 18, 913–919.
- 608 Massinga PH, Johnson SD, Harder LD. 2005. Heteromorphic Incompatibility and Efficiency of
609 Pollination in Two Distylous *Pentanisia* Species (Rubiaceae). *Annals of botany* 95: 389-399
- 610 Menz, M.H.M., Phillips, R.D., Winfree, R., Kremen, C., Aizen, M.A., Johnson, S.D., *et al.*
611 (2011). Reconnecting plants and pollinators: challenges in the restoration of pollination
612 mutualisms. *Trends in Plant Science*, 16, 4–12.
- 613 Oliveira, P.E. & Gibbs, P.E. (2000). Reproductive biology of woody plants in a Cerrado
614 community of Central Brazil. *Flora*, 195, 311–329.
- 615 Ollerton, J., Johnson, S. D., Hingston, A. B., Waser, N. M., & Ollerton, J. (2006). Geographical
616 variation in diversity and specificity of pollination systems (pp. 283-308). *Chicago: University*
617 *of Chicago Press*.
- 618 Ornelas JF, Jiménez L, González C, Hernández A. 2004. Reproductive ecology of distylous
619 *Palicourea padifolia* (Rubiaceae) in a tropical montane cloud forest. I. Hummingbirds'
620 effectiveness as pollen vectors. *American Journal of Botany* 91: 1052-1060.
- 621 Pailler, T., Humeau, L., Figier, J. & Thompson, J.D. (1998). Reproductive trait variation in the
622 functionally dioecious and morphologically heterostylous island endemic *Chassalia*
623 *corallioides* (Rubiaceae). *Biological J of the Linnean Society*, 64, 297–313.
- 624 Papiorek, S., Junker, R.R., Alves-dos-Santos, I., Melo, G.A.R., Amaral-Neto, L.P., Sazima, M.,
625 *et al.* (2016). Bees, birds and yellow flowers: pollinator-dependent convergent evolution of UV
626 patterns. *Plant Biol J*, 18, 46–55.

- 627 Pauw, A., Stofberg, J. & Waterman, R.J. (2009). Flies and flowers in Darwin's race. *Evolution*,
628 63, 268–279.
- 629 Petry, W.K., Soule, J.D., Iler, A.M., Chicas-Mosier, A., Inouye, D.W., Miller, T.E.X., *et al.*
630 (2016). Sex-specific responses to climate change in plants alter population sex ratio and
631 performance. *Science*, 353, 69–71.
- 632 Pigliucci, M. & Preston, K. (2004). *Phenotypic Integration: Studying the Ecology and*
633 *Evolution of Complex Phenotypes*. Oxford University Press.
- 634 Ratter, J.A., Bridgewater, S. & Ribeiro, J.F. (2003). Analysis of the floristic composition of the
635 Brazilian Cerrado vegetation III: Comparison of the woody vegetation of 376 areas. *Edin. Jnl*
636 *of Bot.*, 60.
- 637 Raupp, P.P., Matias, R., Furtado, M.T. & Consolaro, H. (2020). The role of distyly in pollen
638 flow of the hummingbird-pollinated *Palicourea rigida* (Rubiaceae). *Flora*, 271, 151681.
- 639 Ribeiro, J. F., & Walter, B. M. T. (2008). As principais fitofisionomias do bioma Cerrado.
640 *Cerrado: ecologia e flora*, 1, 151-212.
- 641 Richards, J.H. & Koptur, S. (n.d.). Floral Variation and Distyly in *Guettarda scabra*
642 (Rubiaceae), 11.
- 643 Robbrecht, E. (Nationale P. van B. (1988). Tropical woody Rubiaceae. Characteristic features
644 and progressions. Contributions to a new subfamilial classification. *Opera Botanica Belgica*
645 (Belgium).
- 646 Rodrigues, E.B. & Consolaro, H. (2013). Atypical distyly in *Psychotria goyazensis* Mull. Arg.
647 (Rubiaceae), an intramorph self-compatible species. *Acta Bot. Bras.*, 27, 155–161.
- 648 Rohde, K., Papiorek, S. & Lunau, K. (2013). Bumblebees (*Bombus terrestris*) and honeybees
649 (*Apis mellifera*) prefer similar colours of higher spectral purity over trained colours. *J Comp*
650 *Physiol A*, 199, 197–210.
- 651 Schiestl, F.P. & Johnson, S.D. (2013). Pollinator-mediated evolution of floral signals. *Trends in*
652 *Ecology & Evol*, 28, 307–315.
- 653 Silva, A.P. (1995). Biologia reprodutiva e polinização de *Palicourea rigida*.
- 654 Silva, C.A. & Segura, J.A.L. (2015). Reproductive Biology and Herkogamy of *Psychotria elata*
655 (Rubiaceae), a Distylous Species of the Tropical Rain Forests of Costa Rica. *AJPS*, 06, 433–
656 444.

- 657 Sobrevila, C., Ramirez, N. & de Enrech, N.X. (1983). Reproductive Biology of *Palicourea*
658 *fendleri* and *P. petiolaris* (Rubiaceae), Heterostylous Shrubs of a Tropical Cloud Forest in
659 Venezuela. *Biotropica*, 15, 161.
- 660 Sobrevila, Claudia, and Mary T. Kalin Arroyo. (1992). Breeding systems in a montane tropical
661 cloud forest in Venezuela. *Plant Systematics and Evol* 140.1: 19-37.
- 662 Spaethe, J., Tautz, J. & Chittka, L. (2001). Visual constraints in foraging bumblebees: Flower
663 size and color affect search time and flight behavior. *Proceedings of the National Academy of*
664 *Sciences*, 98, 3898–3903.
- 665 Stebbins, G.L. (1970). Adaptive Radiation of Reproductive Characteristics in Angiosperms, I:
666 Pollination Mechanisms. *Annu. Rev. Ecol. Syst.*, 1, 307–326.
- 667 Taylor, C.M. (1997). Conspectus of the Genus *Palicourea* (Rubiaceae: Psychotrieae) with the
668 Description of Some New Species from Ecuador and Colombia. *Annals of the Missouri*
669 *Botanical Garden*, 84, 224.
- 670 Toledo, R., Carvalho, A., Hashimoto, D., Rodrigues, M., Ferreguti, R., Consolaro, H., *et al.*
671 (2007). Caracterização Morfológica de Quatro Espécies de *Palicourea Aubl.* (Rubiaceae) do
672 Cerrado do Brasil Central, 5, 3.
- 673 Tripp, E. A., & Manos, P. S. (2008). Is floral specialization an evolutionary dead-end?
674 Pollination system transitions in *Ruellia* (Acanthaceae). *Evolution: International Journal of*
675 *Organic Evol*, 62(7), 1712-1737.
- 676 Van Rossum, F., De Sousa, S.C. & Triest, L. (2006). Morph-specific differences in
677 reproductive success in the distylous *Primula veris* in a context of habitat fragmentation. *Acta*
678 *Oecologica*, 30, 426–433.
- 679 Vuilleumier, B.S. (n.d.). The Origin and Evolutionary Development of Heterostyly in the
680 Angiosperms, 18.
- 681 Washitani, I., Osawa, R., Namai, H. & Niwa, M. (1994). Patterns of Female Fertility in
682 Heterostylous *Primula Sieboldii* under Severe Pollinator Limitation. *The Journal of Ecology*,
683 82, 571.
- 684 Zapata, T.R. & Arroyo, M.T.K. (1978). Plant Reproductive Ecology of a Secondary Deciduous
685 Tropical Forest in Venezuela. *Biotropica*, 10, 221–230.
- 686 Zhu XF, Jiang XF, Li L, Zhang ZQ, Li QJ. 2015. Asymmetrical disassortative pollination in a
687 distylous primrose: the complementary roles of bumblebee nectar robbers and syrphid flies.
688 *Scientific Reports* 5: article number 7721.

689 Zuur, A.F., Ieno, E.N. & Smith, G.M. (2007). *Analysing ecological data*. Statistics for biology
690 and health. Springer, New York ; London.