

## An eco-designed paper-based algal biosensor for nanoformulated herbicide optical detection

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### Abstract

In this study we reported the development of a paper-based algal biosensor for the optical detection of nanoencapsulated-atrazine, a forefront nanoformulated herbicide with a high effective post-emergence herbicidal activity. In particular, the unicellular green photosynthetic algae *Chlamydomonas reinhardtii* was immobilised on a paper substrate soaked with an agar thin film and placed in a glass optical measurement cell, obtaining a totally environmental-friendly device. Nanoencapsulated-atrazine was detected by following the variable fluorescence ( $1-V_f$ ) parameter, which decreased inversely proportional to the herbicide concentrations, in a range between 0.5 and 200 nM, indicating a linear relationship in the measured dose-response curves and a detection limit of 4 pM. Interference studies resulted in a very slight interference in presence of 0.5 ppm copper and 10 ppb arsenic at safety limits, as well as a slight matrix effect and a satisfactory recovery value of  $96 \pm 5 \%$  for 75 nM nanoencapsulated-atrazine in tap water. Stability studies were also performed obtaining a good storage stability up to 3 weeks. Results demonstrated the suitability of the proposed paper-based optical biosensor as a valid support in smart agriculture for on site, environmentally friendly, cost effective and sensitive nanoencapsulated-atrazine analysis.

**Keywords:** paper-based algal biosensor, *Chlamydomonas reinhardtii*, chlorophyll fluorescence, nanoencapsulated-atrazine, sustainable agriculture

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## 1. Introduction

The increasing demographic pressure, climate change, and industrial globalization of last decades represent nowadays few main global challenges, being factors that generate a huge impact on the environment and people well-being. Among human actions, agriculture is one of the main cause of environmental pollution since farming activities have been oriented toward intensive agricultural practices. Indeed, there is the indiscriminate use of pesticides that contribute to soil and water pollution, with important consequences on different ecosystems. For this reason, most countries have established maximum residue levels (MRL) in food products to limit pesticide use and exposure to such dangerous compounds (European Union Legislation, <http://europa.eu.int/eurlx/en/indx.html>). Their accepted concentrations are 0.1 µg/L for each single pollutant and 0.5 µg/L for total pesticides (Drinking water Directive, Council Directive 98/83/EC).

As a consequence, consumer demand for more sustainable agricultural practices and waste minimisation has become imperative in modern agrifood sector. In this context, smart agriculture entails the exploitation of different multifarious approaches based on a more efficient energy and environmentally friendly cross-cutting technologies, including i) novel nanoformulations to increase the dispersion and wettability of agricultural pesticides/fertilisers and ii) advanced diagnostic tools for the analysis of ground/drinking water to assess water matrix pollution levels [1,2]. Nanoherbicides are gaining momentum in the last years as novel formulations for smart agriculture, being a more sustainable route for weed growth control with minimised release of toxic residues in soil and environment, owing to a smarter delivery system of the active ingredients [3,4]. In addition, some preliminary results indicated that the nanoformulation can impact on the fate of herbicides, which are usually undergo biotic or abiotic degradation [5].

Among different pesticides, atrazine represents a case study herbicide, being widely used for pre- and post-emergence control of weeds in agriculture and acting as an inhibitor of photosynthesis by specific binding to photosystem II. After soil treatment, this herbicide can reach surface or subsurface waters during irrigation or rain events, contributing to the pollution [6,7]. Moreover, the slow degradation of this herbicide under natural conditions results in high persistence in the environment and consequently in the contamination of soil, water resources, and food [8]. Recently many regulations have been defined also for atrazine, establishing a limits between 0.6 and 2 µg/L in surface water matrices (Directive 2013/39/EU) [9].

A more proficient use of atrazine can be achieved by the use of nanoscale carriers, which reduce the amount of herbicide to be applied but increasing its efficiency, with minimised impact on the environment. Recently, Oliveira and co-workers developed nanostructured carrier systems for

atrazine as a promising strategy to minimize its contamination of natural resources [10]. In detail, the authors prepared nanocapsules of poly( $\epsilon$ -caprolactone) (PCL), a biodegradable aliphatic polyester nontoxic to humans and environment, and loaded them with atrazine to obtain a nanoencapsulated herbicide. This nanoencapsulated-atrazine (nanoatrazine) shows a huge potential as a forefront nanoformulation for applications in smart agriculture, being able to provide higher effective herbicidal activity and lower toxic impact on the environment.

However, as stated by Kah and co-workers, “*the current level of knowledge does not appear to allow a fair assessment of the benefits and/or risks that will result from the use of some nanopesticides*” [5]. Moreover, any reliable data evidenced that these novel formulations can be persistent in the different environmental compartments, thus generating detriment to water matrices of agrifood and human health interest. For this reason, effective biosensing systems to reveal the pollution level of different environments, as surface and drinking waters, caused by these novel nanoformulated herbicides are mandatory, with analytical features custom-shaped on the structure of these compounds. **To date, some paper-based biosensors are reported in the literature based on different bioreceptors towards different toxic compounds []**; conversely biosensors or bioassays have been not specifically designed for the detection of nanoformulated pesticides. In this scenario, we present for the first time the development of a biosensor to sense nanoherbicides for applications in smart agriculture.

In particular, we described a paper-based optical biosensor consisting of whole cells of the green photosynthetic algae *Chlamydomonas reinhardtii* immobilised on a paper substrate for the detection of nanoencapsulated-atrazine. Paper is gaining momentum in the design of biosensors for many application fields owing to its enormous advantages, compared to traditional 3D scaffolds made of inorganic materials, polymers, metals, and composite materials, in terms of cost-effectiveness, simple synthesis procedures, and environmental sustainability. In addition, the high porosity, flexibility, tunability, amenability to high-throughput analysis, biocompatibility and thus suitability of paper towards cell growth, make this substrate an old material performing forefront tricks [11,12]. ***Chlamydomonas reinhardtii* algae was herein combined with paper as substrate for the first time thanks to its ability to reveal photosynthetic herbicides by means of a simple and effective mechanism. Indeed, variations of algal chlorophyll fluorescence emission occur in the presence of similar toxic compounds, due to a specific interaction between the photosynthetic herbicide and the plastoquinone Q<sub>B</sub> binding niche present in the D1 protein of the Photosystem II of the algae [19-23].** The rationale to design a paper-based algal biosensor for the detection of nanoatrazine is to furnish a pioneering analytical tool for application in smart agriculture encompassing the fundamentals of sustainability of the recent sustainable agricultural practices. Indeed, the combination of a Generally Recognized as Safe (GRAS) organism, as *Chlamydomonas* is recognised by the FDA [13], with an

environmentally-friendly material as paper opens novel directions towards the development of innovative biosensor concepts entailing the basic principles of sustainability of the intelligent farming. A similar biosensing configuration can be used as a disposable system for the optical monitoring of nanoencapsulated-atrazine and further wasted as biodegradable material without impacting on the environment.

## **2. Experimental section**

### *2.1 Chemicals*

Copper ( $\text{Cu}^{2+}$ ) and arsenic ( $\text{AsIII}$ ) were purchased from Carlo Erba. Tris-acetate-phosphate, poly( $\epsilon$ -caprolactone), Polysorbate 80 (Tween 80), acetone and sorbitane monostearate (Span 60) were purchased from Sigma (Aldrich Corporation, USA).

### *2.2 PCL nanocapsules preparation and characterization*

PCL nanocapsules were prepared by the interfacial deposition of pre-formed polymer method [14]. This technique involves the mixing of an organic phase consisting of polymer, surfactant (Span 60), oil (Myritol 318), and the active compound (herbicide) into an aqueous phase containing Tween 80. The nanocapsules formed were stored in amber flasks at a temperature of 25 °C. The technique of photon correlation spectroscopy and microelectrophoresis were used to determine the hydrodynamic diameter and zeta potential of the nanoparticles, respectively. The samples (nanocapsules and nanoatrazine) were diluted 100-fold with deionized water (Milli-Q) and analysed using a ZetaSizer ZS 90 (Malvern®) at a fixed angle of 90° and temperatures of 25°C. In addition, the hydrodynamic diameter and nanoparticle concentration/mL were analysed using the Nanoparticle tracking analysis (NTA). For NTA, the samples were diluted 5000-fold and analysed in a volumetric cell using a NanoSight LM10 equipment (CMOS camera and NanoSight software - version 3.2). All measurements were performed in triplicate. The nanoparticle morphology was measured using the Easy Scan 2 Basic AFM instrument (Nanosurf, Switzerland). For the AFM analyses, the nanoparticles suspensions were diluted 500-fold and then dripped onto a silicon plate and left to dry for 7 days. In order to scan the samples, the instrument was operated in contact mode using a TapA1-G cantilevers (BudgetSensors, Bulgaria). The images were processed using the Gwyddion 2.45 software. The size distribution of the nanoparticles was measured using a total of 100 nanoparticles.

### 2.3 *C. reinhardtii* IL strain and growth culture conditions

For all the experiments an IL strain of *C. reinhardtii* was exploited, a mutant containing an intronless *psbA* gene encoding for the D1 protein of Photosystem II [15,16]. Algae cultures were photoheterotrophically grown in Tris-acetate-phosphate (TAP) medium pH 7.2 prepared according to Harris protocol [17], under continuous light at 50  $\mu\text{mol photons/m}^2/\text{s}$  (OSRAM Fluora) and with stirring at 150 rpm at 25 °C (Certomat BS-1 incubator, Sartorius).

### 2.4 Measurement of culture growth, photosynthetic activity, and chlorophyll contents

All experiments were performed exploiting cell cultures in early mid-exponential growth phase, with  $\text{Abs}_{750} \sim 0.5$  O.D., corresponding to  $8 \pm 2 \times 10^5$  cells/mL, and 8  $\mu\text{g/mL}$  chlorophyll content. Cell culture growth was evaluated spectrophotometrically by quantifying the absorbance (O.D.) at 750 nm wavelength. Cell number was assessed by a Bio-Rad TC-10 automated counter (Hemel Hempstead, UK). Pigment content was calculated by the Lichtenthaler method once extracted with 80% acetone [18].

The algae photosynthetic profile was estimated by recording the chlorophyll fluorescence induction curves (Kautsky curves) in the absence and in the presence of nanencapsulated atrazine by a Plant Efficiency Analyzer (PEA, Hansatech Instr. Ltd., Kings Lynn, Norfolk, UK), at room temperature after 10 min of dark adaptation and with a 3 s saturating pulse excitation light ( $600 \text{ W/m}^2$ ) provided by an array of six red light emitting diodes (650 nm peak). Kautsky curves are described by a polyphasic fluorescence rise in the time, called *OJIP* curves, where O represents the minimal dark-adapted fluorescence level (where all  $\text{Q}_\text{A}$  are oxidised) and P the maximal level (where all PSII quinone acceptors are fully reduced). Using the difference in the fluorescence signal between these two distinct states it is possible to evaluate PSII functionality via the parameter  $F_V/F_M$ , which represent the maximum fluorescence yield of PSII photochemical reaction and is calculated according to the equation (1):

$$(1) \quad F_V/F_M = (F_M - F_0)/F_M$$

where  $F_V$  represents the maximum variable fluorescence calculated as  $F_M - F_0$ ,  $F_M$  corresponds to the maximum fluorescence emission and  $F_0$  is the minimum fluorescence emission.  $F_V/F_M$  values

decrease in an analyte concentration dependent manner where the target analyte is able to impact on the integrity of the photosynthetic apparatus thus inhibiting photosynthetic reactions.

Moreover, the intermediate points J and I are a direct indication of transient changes in the plastoquinone redox state within the PSII reaction centre, as the accumulation of Q<sub>A</sub> in reduced state and the consequent filling up of the plastoquinone pool with electrons [19-22]. Any change in the PSII electron transfer efficiency directly causes variations in the fluorescence intensity of the inflection points, as a consequence of different stress conditions, as for example the presence of pollutants. In particular, the variable fluorescence parameter 1-V<sub>J</sub> considers all the parameters involved in the fluorescence emission profile and provides a more specific response directly correlated to the herbicide-D1 binding niche interaction [23]. This parameter is calculated according to the equation (2):

$$(2) \quad 1-V_J = 1 - (F_{2ms} - F_0) / (F_M - F_0)$$

where F<sub>0</sub>, F<sub>M</sub>, F<sub>2ms</sub> are respectively the initial fluorescence, the maximum fluorescence and the fluorescence measured after 2 ms.

### 2.5 *C. reinhardtii* immobilisation and instrument set-up

*C. reinhardtii* cell cultures in early mid-exponential growth phase, with Abs<sub>750</sub> ~ 0.5 O.D., corresponding to  $8 \pm 2 \times 10^5$  cells/mL, and 8 µg/mL chlorophyll content, were exploited for the immobilisation on agar hydrogel/paper substrate. Approximately  $4 \pm 0.5 \times 10^6$  cells were harvested by means of 5 min centrifugation at  $2000 \times g$  and 15 °C. The cell pellet was washed, re-suspended in Tris-acetate-phosphate (TAP) medium pH 7.2 to a final volume of 200 µL, and placed on a cellulose substrate (diameter 1 cm) soaked into an 1.5 % (w/v) agar hydrogel made in Tris-acetate-phosphate (TAP) medium pH 7.2 to form a film. The paper substrate was then accommodated into the measurement cell of the PEA instrument for fluorescence analysis. Storage stability analyses were performed for a period of 21 days by monitoring the parameter F<sub>V</sub>/F<sub>M</sub>.

### 2.6 Calibration curves and interference studies

*C. reinhardtii* IL cultures in triplicates were incubated with 200 nM nanoencapsulated atrazine for 24 hours under continuous light ( $50 \mu\text{mol photons m}^{-2} \text{s}^{-1}$ ), at room temperature and continuous stirring (150 rpm). Control cultures were grown under physiological conditions and carried out in parallel in all trials. Photosynthetic activity of *C. reinhardtii* IL cell cultures were monitored according to the Section 2.3, performing a 15 min incubation with nanoencapsulated atrazine in a concentration range between 0.5 and 200 nM. Interference studies were performed after a 15 min incubation with solutions of 0.5 ppm copper ( $\text{Cu}^{2+}$ ), 10 ppb arsenic ( $\text{AsIII}$ ), and 100 nM nanoencapsulated atrazine. Matrix effect was evaluated using a 1:2 dilution of tap water in Tris-acetate-phosphate (TAP) medium pH 7.2.

## 2.7 Statistical analyses

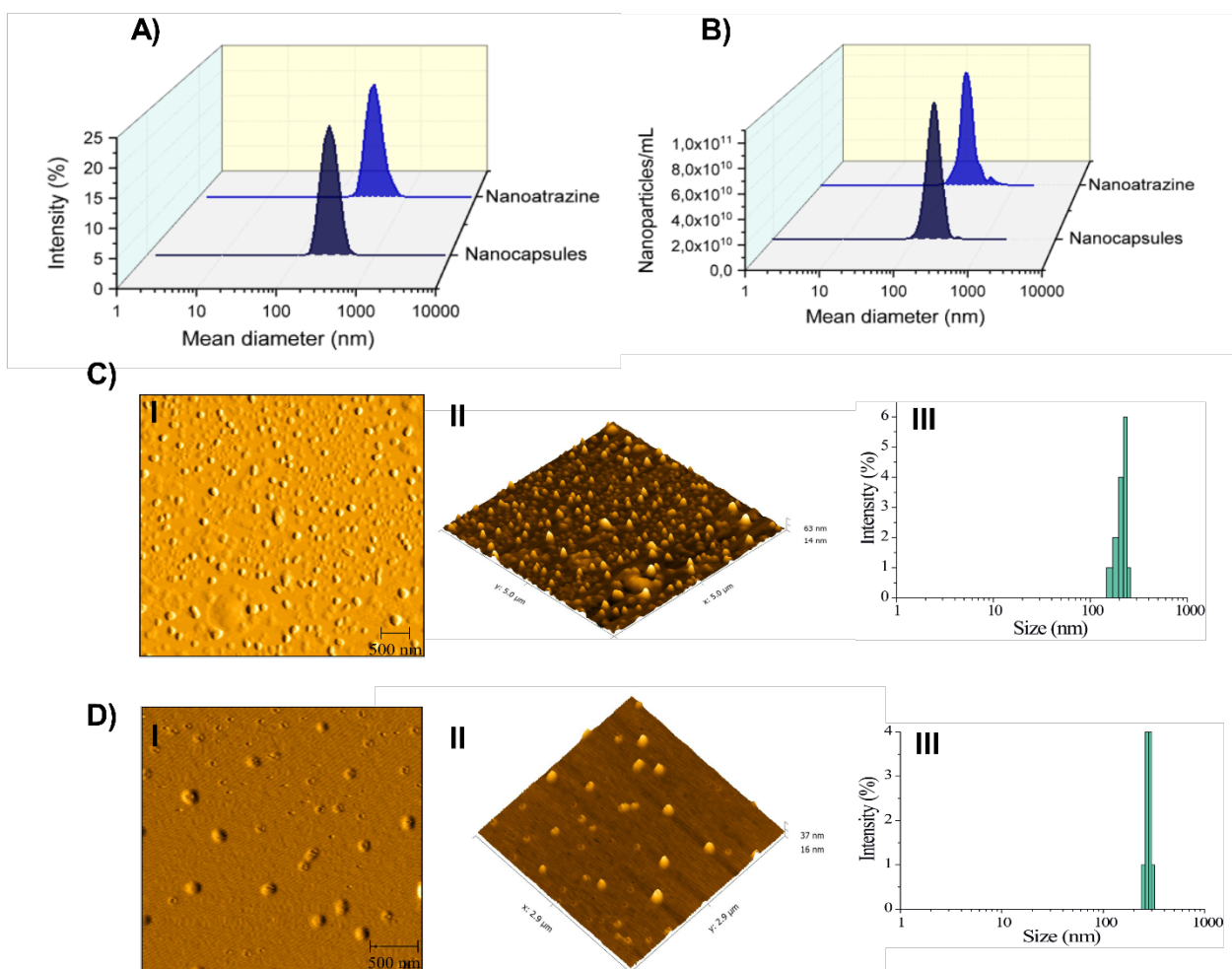
Data from three separated experiments were analysed and mean values  $\pm$  standard deviation reported. For a given experiment, each condition was tested in triplicate. Detection limits were calculated on the basis of 99 % confidence interval, which, assuming the normal distribution, corresponds to  $2.6 \times$  standard error of the measurements ( $\sigma$ ), exploiting the modified relationship for the Langmuir absorption isotherm,  $\text{LOD} = 2.6 \times \sigma \times I_{20} / (100 - 2.6 \times \sigma)$  [24].

## 3. Results and discussion

Both empty poly( $\epsilon$ -caprolactone) nanocapsules and loaded with atrazine (nanoatrazine) have been previously characterized and showed similar results as stated in [14]. However, here, the physicochemical properties of both nanocapsules and nanocapsules loaded with atrazine (nanoatrazine) were determined and the results are showed in Figure 1. Results obtained by DLS measurement (Figure 1A) showed that the poly( $\epsilon$ -caprolactone) nanocapsules presented a mean size diameter of  $250 \pm 2 \text{ nm}$  e polidisperisty index of  $0.046 \pm 0.004$ , while for nanoatrazine, the results showed a mean size diameter of  $335 \pm 3 \text{ nm}$  e polidispersity index of  $0.157 \pm 0.09$ . The zeta potential of nanocapsules and nanoatrazine showed a negative zeta potential of  $-27 \text{ mV}$ . The mean diameter was also measured by NTA (Figure 1 B). For nanocapsules and nanoatrazine the mean size diameter and concentration of nanoparticles were  $204 \pm 1 \text{ nm}$  ( $1.23 \pm 0.06 \times 10^{13}$  nanoparticles/mL) and  $218 \pm 4 \text{ nm}$  ( $1.17 \pm 0.05 \times 10^{13}$  nanoparticles/mL) respectively. In addition, the micrographs of nanocapsules and nanoatrazine analysed by AFM are showed in Figure 1C, D, respectively. According to these images, it can be seen that the nanoparticles showed a spherical shape. AFM

images presented a narrow size distribution ( $211 \pm 22$  nm for nanocapsules and  $270 \pm 13$  nm for nanoatrazine), in agreement to the results obtained by DLS and NTA.

Figure 1: **Physicochemical characterization of the nanocapsules and nanoatrazine:** **A)** size distribution measured by Dynamic Light Scattering. All analyses were performed in triplicate, at 25 °C. The mean diameter for nanocapsules was  $250 \pm 2$  nm and for nanoatrazine was  $335 \pm 3$  nm; **B)** Size distribution measured by NTA. All analyses were performed in triplicate, at 25 °C. The mean diameter for nanocapsules was  $204 \pm 1$  nm and for nanoatrazine was  $218 \pm 4$  nm; **C)** AFM micrographs of nanocapsules (I - 2D AFM micrographs; II - 3D AFM micrographs and III - histogram size distribution obtained from AFM micrographs) and **D)** AFM micrographs of nanoatrazine (I - 2D AFM micrographs; II - 3D AFM micrographs and III - histogram size distribution obtained from AFM micrographs). The AFM results show the spherical morphology of nanoparticles that presented an average diameter of  $211 \pm 22$  nm for nanocapsules and  $270 \pm 13$  nm for nanoatrazine.





With the aim to characterise the physiological condition of the algae in the presence of both poly( $\epsilon$ -caprolactone) nanocapsules and nanoencapsulated-atrazine, their effect was investigated on algal growth, photosynthetic activity, and chlorophyll content. In this context, atrazine was exploited as reference herbicide to strength the validity of nanoencapsulated-atrazine analysis. Once optimised all the experimental conditions, algae were immobilised on a paper substrate soaked with an agar hydrogel to form a sort of thin film for algae absorption. The paper-based algal biosensor was challenged for nanoatrazine detection in both standard solutions and real samples. Also interference studies, matrix effect, recovery studies, and stability studies were optimised. In particular, the analytical performances of this device were evaluated in tap water, a vulnerable matrix of environmental and human health interest, which can be subject to pesticide pollution due to agricultural practices.

### 3.1. Effect of nanoformulated herbicides on algae physiological parameters

The physiological parameters of *C. reinhardtii* were evaluated in the presence of poly( $\epsilon$ -caprolactone) nanocapsules, nanoencapsulated-atrazine, and atrazine at a 200 nM concentration, by following algae growth, photosynthetic activity, and chlorophyll content at specific incubation times.

In detail, cell number, chlorophyll *a* fluorescence, and the total chlorophyll content were measured. Growth data indicated that PCL nanocapsules, nanoencapsulated-atrazine, and atrazine were not able to inhibit algal proliferation, indicating any influence on cell grow and emphasising any cytotoxic effects on *C. reinhardtii* algae in terms of altered vital processes and variations in the physiological parameters (e.g. cell duplication) (Figure 2A).

The photochemical efficiency of PSII was evaluated by means of Kautsky curves as described in the Section 2.3 of “Methods”, following both the maximum fluorescence yield of Photosystem II  $F_V/F_M$  and the variable fluorescence parameter  $1-V_J$  during an analysed period of 24 h. In Figure 2B and C, the fluorescence parameters  $F_V/F_M$  and  $1-V_J$  values were reported, calculated by each Kautsky curve, and plotted as function of time.

In particular, any effect was registered regarding the maximum fluorescence yield  $F_V/F_M$  which remains constant during time, while the inhibitory effect of both nanoatrazine and atrazine was underlined by the decrease of the variable fluorescence  $1-V_J$ , being able to cause variations of the  $F_V$  parameter (intimately connected with the  $Q_A^-$  accumulation). Results indicated that both herbicides were not able to completely block the photosynthetic activity of the algae, while affecting the transient changes in the redox state of the electron carriers within the PSII reaction centre, such as the

accumulation of  $Q_A$  in the reduced state and the further filling up of the membrane plastoquinone pool with electrons.

The content of photosynthetic total chlorophyll *a* and *b* (Chl*a* and Chl*b*) also indicated any showed effect of PCL nanocapsules, nanoatrazine and atrazine, evidencing a similar pigment production profile after 4 and 24 h with respect to control cultures (Figure 2D).

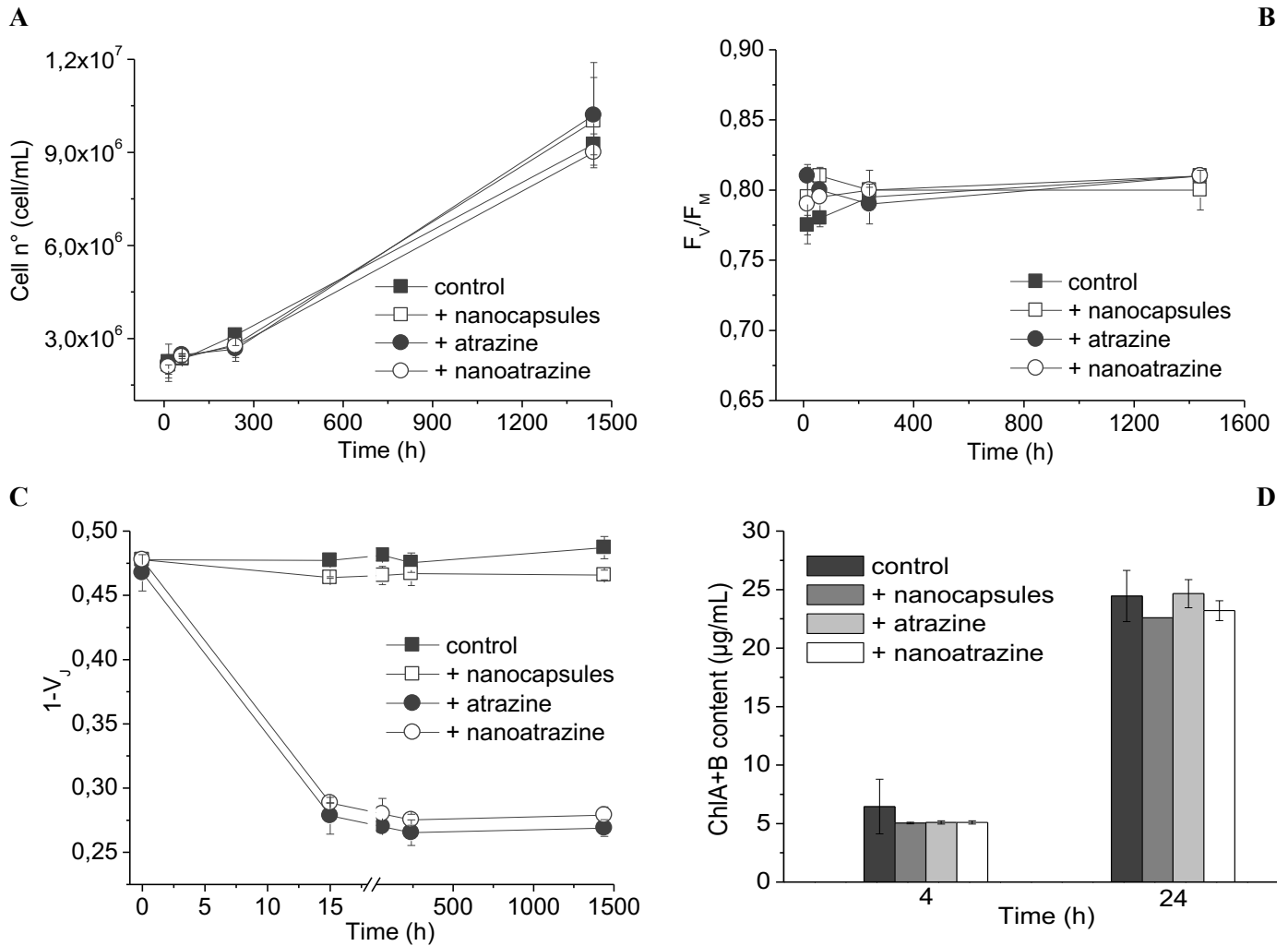


Figure 2. *C. reinhardtii* physiological characterisation. Effect of 200 nM atrazine and nanoencapsulated-atrazine on *C. reinhardtii* following: A) cell culture growth reported as cell number, B) maximum fluorescence yield  $F_V/F_M$  calculated on each Kautsky curve, C) variable fluorescence  $1-V_J$  calculated on each Kautsky curve, and D) chlorophyll content. Incubation time: 24 hours under continuous light ( $50 \mu\text{mol photons m}^{-2} \text{s}^{-1}$ ). Average values  $\pm$  SE (n=3).

### 3.3 Development of the paper-based algal biosensor

Once evaluated the effect of nanoencapsulated-atrazine on the different algal processes, the algae were immobilised on paper substrate and the assembled biosensor was optimized for nanoherbicide detection.

### 3.3.1 Immobilisation of *C. reinhardtii* on paper substrate

*C. reinhardtii* whole-cells were immobilised on a paper substrate to provide a low-cost and sensitive analytical tool encompassing the fundamentals of sustainability proposed by the smart agriculture. Indeed, the combination of algae, recognised by the Food and Drug Administration (FDA) as a Generally Recognized as Safe (GRAS) organism [13], with an eco-friendly material, opens novel routes towards the basic principles of sustainability of the smart agriculture.

In detail, a *C. reinhardtii* cell culture in early mid-exponential growth phase, with  $\text{Abs}_{750} \sim 0.5$  O.D., corresponding to  $(8 \pm 2) \times 10^5$  cells/ml and 8  $\mu\text{g/ml}$  chlorophyll content, was immobilised on a paper substrate (diameter 1 cm) soaked into an 1.5 % (w/v) agar hydrogel made of Tris-acetate-phosphate (TAP) medium pH 7.2 to form a thin agar film for algae absorption. The paper substrate was then accommodated into a glass measurement cell of the PEA instrument for fluorescence analysis, with the aim to evaluate the bioreceptor response towards nanoatrazine by monitoring the variable fluorescence parameter  $1-V_J$  calculated on the Kautsky curves, as well as its shelf lifetime by following the maximum fluorescence yield  $F_v/F_m$ . Figure 3 reports a scheme of the proposed paper-based algal biosensor.

Beside the sustainability reason as above described, the main role of paper substrate was to fabricate an eco-friendly disposable dish where algae were absorbed and stored in a culture media-like environment retaining a good photosynthetic activity for up to 3 weeks. In case of herbicide analysis, the paper dish absorbed with algae was used as a disposable biosensor for the optical monitoring of nanoencapsulated-atrazine and further wasted as biodegradable material without impacting on the ecosystems. Moreover, the paper dish is a flexible support that can be easily customised and integrated in the measurement cell of the optical transduction system, thus allowing the construction of a reusable system where only the paper dish is wasted.

The so configured paper-based algal biosensor was challenged for nanoencapsulated-atrazine analysis in standard solutions and real samples. Also interference and recovery studies were performed, as described in the following sections.

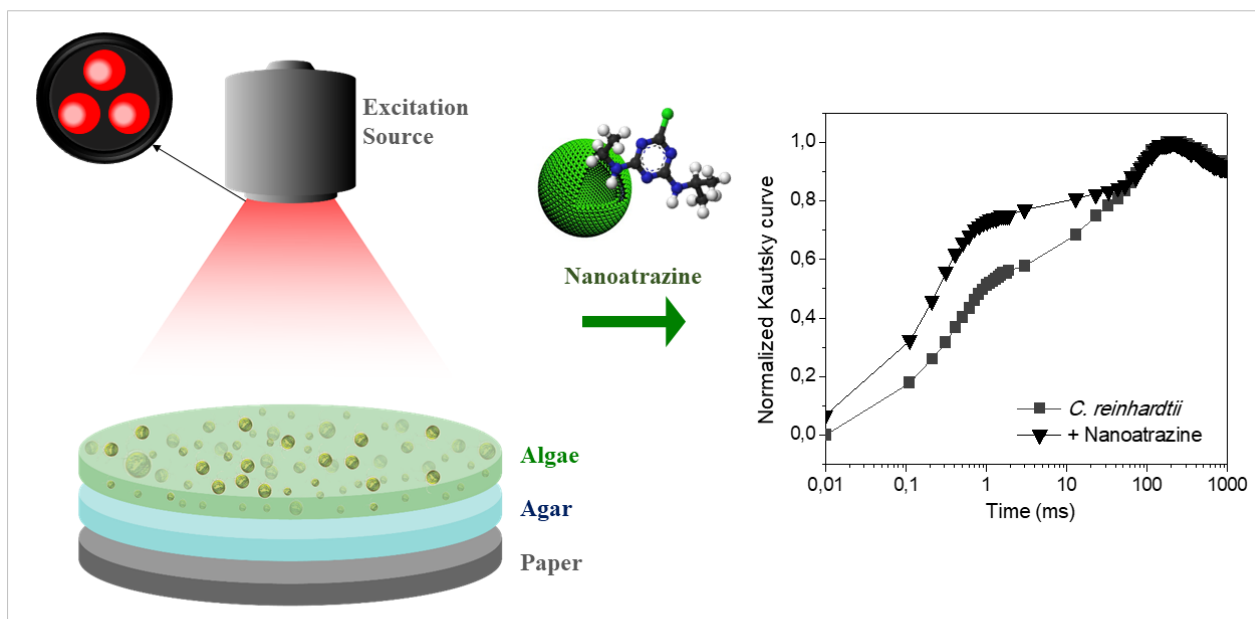


Figure 3. Paper-based algal biosensor scheme. *C. reinhardtii* cells are immobilised on paper soaked into an agar hydrogel for the detection of nanoencapsulated-atrazine.

### 3.3.2. Optimisation of nanoencapsulated-atrazine analysis in standard solutions

The paper-based algal biosensor was optimized for nanoencapsulated-atrazine detection by following the variable fluorescence parameter  $1-V_J$  calculated from the algae fluorescence response described by the Kautsky curve. Thus, cell cultures at mid-point of exponential growth phase, entrapped in an agar hydrogel, and immobilised on paper were incubated for 15 min with standard solutions of nanoatrazine in a concentration range from 10 to 150 nM. Results evidenced that  $1-V_J$  parameter decreased proportionally to the increase of nanoherbicide concentrations, allowing the construction of a calibration curve using the linear regression described by the equation  $y = 0.503(\pm 0.005) - 0.00117(\pm 0.00004) x$ , with an  $R^2 = 0.9809$ . A detection limit of 4 pM was obtained for  $I_{20}$  value (calculated as  $2.6 \times \sigma \times I_{20}/100 - 2.6 \times \sigma$ ). Figure 4 reports the paper-based algal biosensor response towards nanoencapsulated-atrazine.

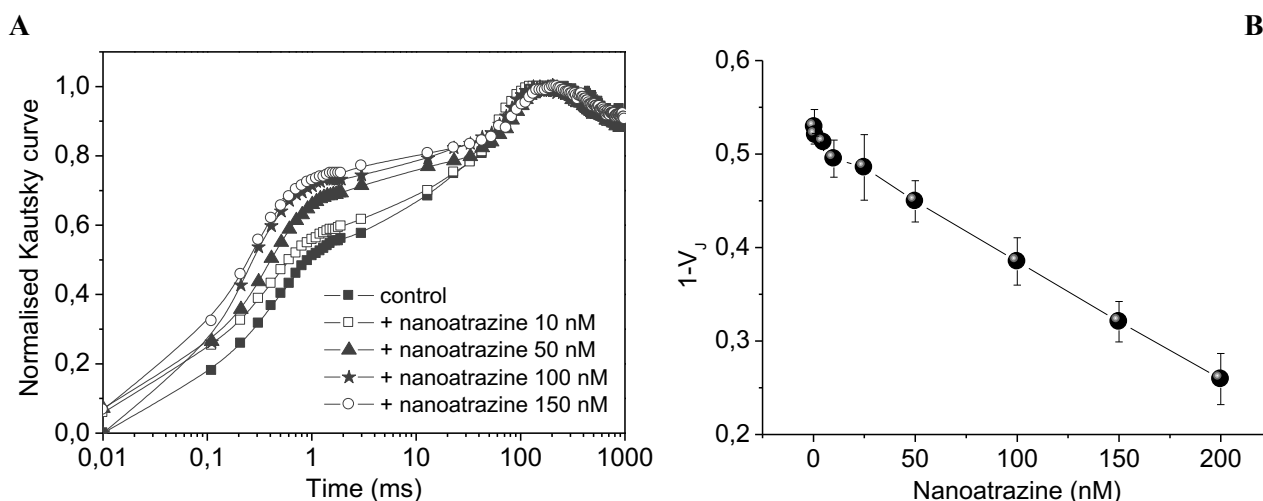


Figure 4. Paper-based algal biosensor analytical performances. A) Kautsky curve of *C. reinhardtii* in the presence of increasing amount of nanoencapsulated-atrazine. B) Variable fluorescence  $1-V_j$  calculated on each curve (incubation time 15 min). Average values  $\pm$  SE (n=3).

### 3.3.4. Interference studies

With the aim to challenge the capability of the proposed paper-based algal biosensor in drinking water samples, **some chemicals including copper and arsenic were tested as interferences at legal level established by World Health Organization (WHO) in drinking water.** In detail, *C. reinhardtii* was incubated for 15 min with standard solutions of 0.5 ppm copper ( $\text{Cu}^{2+}$ ), 10 ppb arsenic ( $\text{AsIII}$ ), and 100 nM nanoencapsulated-atrazine as well as with a solution spiked with all the above listed compounds, to evaluate the synergistic effects of different chemicals in a mixture. Results reported in Figure 5 highlighted that the interfering species did not affect the analysis of nanoencapsulated-atrazine at the exploited concentrations.

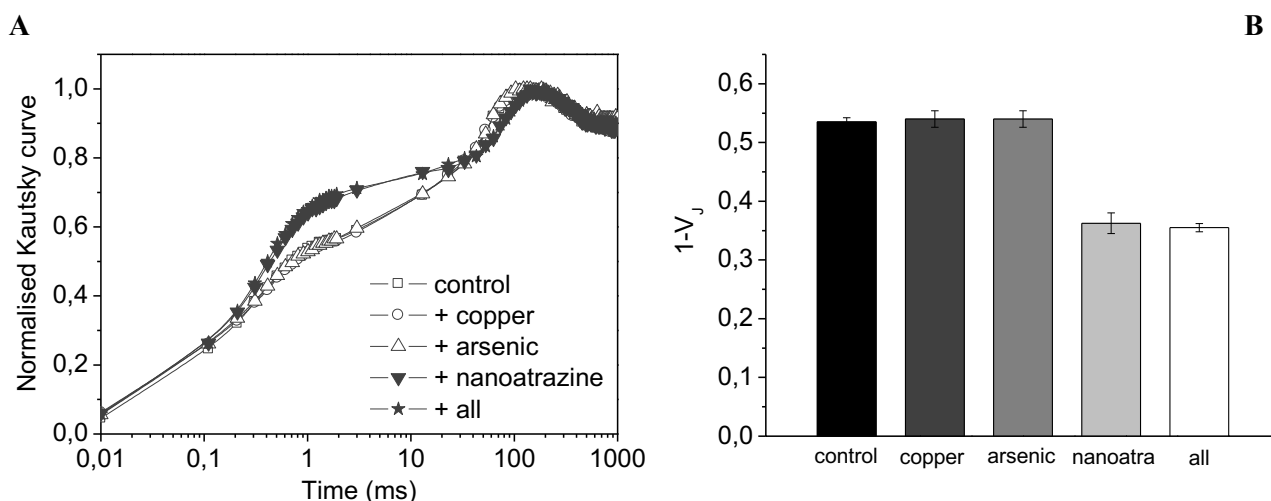


Figure 5. Paper-based algal biosensor interference study. A) Kautsky curve of *C. reinhardtii* as control and incubated for 15 min with 0.5 ppm copper, 10 ppb arsenic, 100 nM nanoatrazine, and with the mixture of all chemicals. B) Variable fluorescence 1-V<sub>j</sub> calculated on each curve (incubation time 15 min). Average values ± SE (n=3).

### 3.3.3. Matrix effect, recovery, and stability studies

To investigate the suitability of the proposed paper-based algal biosensor in real samples, matrix effect and recovery studies were performed. To estimate matrix effect, a *C. reinhardtii* cell culture was diluted 1:2 (v:v) in tap water fortified with nanoencapsulated-atrazine in a concentration range from 10 and 150 μM. The conditions of the cell cultures resulting from the dilution were in agreement with the optimised conditions set up for all the previous experimental sessions (Section 2.3 of Methods). A calibration curve was constructed calculating the 1-V<sub>j</sub> parameter (Figure 6A), obtaining a linear range of detection in the 10 - 150 nM range, described by the equation  $y = 0.403(\pm 0.004) - 0.00127(\pm 0.00006) \times x$  ( $R^2 = 0.999$ ). A detection limit of 4 pM (calculated as  $2.6 \times \sigma \times I_{20} / (100 - 2.6 \times \sigma)$ ) was assessed. The slopes of the calibration curves obtained in standard solutions and real samples were very similar with a ratios of 1.08, indicating a very slightly dependence from the tap water matrix. The calibration curve obtained for the paper-based algal biosensor challenged in real samples was further used to calculate the recovery values of the tap water samples. A recovery value of  $96 \pm 5$  % was obtained for 75 nM of nanoencapsulated-atrazine, highlighting a satisfactory capability to detect nanoatrazine also in real samples (Table 1).

The shelf life time of the paper-based algal biosensor was also evaluated by storing algae immobilised on the paper substrate and located in the glass measurement cell under continuous light ( $50 \mu\text{mol photons m}^{-2} \text{s}^{-1}$ ) at RT for 1 month. Assessing the photosynthetic activity expressed as  $F_v/F_m$

(maximum fluorescence yield of PSII), the paper-based algal biosensor was able to retain a good photosynthetic activity (70% activity) for up to 3 weeks (Figure 6B).

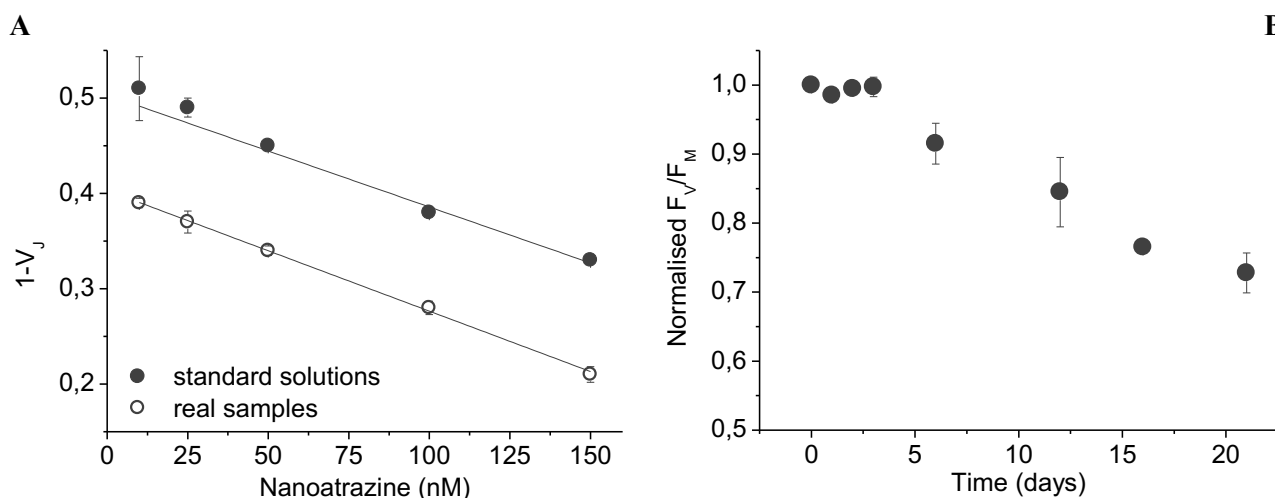


Figure 6. A) Paper-based algal biosensor performances in real sample: Kautsky curve of *C. reinhardtii* in the presence of increasing amount of nanoencapsulated-atrazine in standard solutions and real samples (tap water). B) Paper-based algal biosensor storage stability: photosynthetic activity expressed as  $F_v/F_M$  (maximum fluorescence yield of PSII) (incubation time 15 min). Average values  $\pm$  SE (n=3).

Sample	Fortified nanoatrazine (nM)	Revealed nanoatrazine (nM)	Recovery %
Tap water	75	72 $\pm$ 4	96 $\pm$ 5

Table 1. Recovery values for algae bioassay and paper algal biosensor from tap water (n=3).

The proposed paper-based algal biosensor demonstrated to be able of revealing nanoencapsulated-atrazine with detection limit in the picomolar range (4 pM corresponding to 80 ng/L), meeting the requirements of atrazine law limits between 0.6 and 2  $\mu$ g/L (Directive 2013/39/EU) [9]. The biosensor was also challenged in tap water, a vulnerable matrix that can be subject to pesticide pollution due to agricultural practices, indicating its suitability for herbicide analysis in real samples and without interferences from compounds which can be present at safety limits provided by World Health Organization (WHO), as well as with very slight matrix effect and satisfactory recovery values. In addition, being this biosensor based on a whole-cell bioreceptor, no sample pre-treatment was required. The biosensor is also capable to meet the requirements of sustainability, being a totally eco-designed device composed of biodegradable materials (both bioreceptor and substrate) without

impacting on the environment. Moreover, it represents a simple and cost-effective analytical tool for application in smart agriculture where the use of novel nanoformulations (e.g. fertilisers/pesticides encapsulated on nano-delivery systems) is gaining momentum and against whom novel implements need to be further developed.

Furthermore, taking into account these promising results, the proposed biosensor represents a competitive tool towards similar photosynthesis-based analytical systems reported in literature, owing to its very high sensitivity and stability, as well as its eco-designed configuration. Some of the most representative examples are summarized in Table 2, based on different sources of photosynthetic elements including whole cells, thylakoids and PSII particles from different photosynthetic organisms, as well as different transducers as optical and electrochemical ones.



Bioreceptor	Transducer	Support	Detection limit		Storage stability	Response time	Real matrix	Ref.
			M	g/L				
<i>Chlamydomonas reinhardtii</i> whole cells	Chlorophyll fluorescence	Agar medium on a silicon septum	$9.95 \times 10^{-10}$ M	21 µg/L	3 weeks at RT	15 min	n.d.	[23]
<i>Chlamydomonas reinhardtii</i> whole cells	Chlorophyll fluorescence	Alginate and silica hydrogel on plastic substrate	0.1 µM	21 µg/L	n.d.	40 min	n.d.	[25]
<i>Chlorella vulgaris</i> whole cells	Fibre optic	Glass microfiber membrane	5 µM	1 mg/L	1 week	30 min	n.d.	[26]
<i>Chlorella vulgaris</i> whole cells	Fibre optic	Quartz microfibre filter	-	0.25 µg/L	4 weeks	10 min	n.d.	[27]
<i>Chlorella vulgaris</i> whole cells	Amperometric	Screen-printed electrode in polycarbonate	1 µmol dm <sup>-3</sup>	0.2 g/L	4 weeks at 4 °C retaining 80% of activity	4 min	n.d.	[28]
<i>Chlorella pyrenoidosa</i> whole cells	Amperometric	Screen-printed electrode in polycarbonate	0.7 µM	0.1 mg/L	n.d.	n.d.	n.d.	[29]
PSII from <i>Synechococcus elongatus f. thermalis</i>	Amperometric	Screen-printed electrode in polycarbonate	$1.6 \times 10^{-9}$ M	34 µg/L	n.d.	n.d.	n.d.	[30]
Thylakoids from <i>Spinacia oleracea</i> , <i>Senecio vulgaris</i>	Amperometric	Screen-printed electrode in ceramic	12.7 µM	2.7 mg/L	Half-life of 16.7 hours	30 min	River water with recovery value of 56%	[31]

Thylakoids from spinach	Amperometric	Screen-printed electrodes in ceramic	1.15 nM	24 µg/L	24 hours at 4 °C retaining 50% of activity	Few minutes	n.d.	[32]
Thylakoids from spinach	Amperometric	Screen-printed electrodes in polyester	-	2.8 µg/L	Some days at 4 °C	n.d.	River water with low reproducibility and recovery %	[33]
<i>Chlamydomonas reinhardtii</i> whole cells	<i>Chlorophyll fluorescence</i>	<i>Removable algal paper filter</i>	<i>4 pM</i>	<i>80 ng/L</i>	<i>4 weeks at RT retaining 70% of activity</i>	<i>15 min</i>	<i>Tap water with recovery value of 96±5%</i>	<i>Our paper-based algal biosensor</i>

Legend: n.d. not determined.

Table 2. Examples of photosynthesis-based analytical systems for atrazine detection (safety limit for atrazine is 0.6 - 2 µg/L, Directive 2013/39/EU).

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