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Journal of Environmental Chemical Engineering





Metallo-stannosilicate heterogeneous catalyst for biodiesel production using edible, non-edible and waste oils as feedstock



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ARTICLE INFO

Keywords: Sol-gel chemistry Metallo-stannosilicate Biofuels Biomass Edible and non-edible lipid feedstocks

ABSTRACT

This study reports the synthesis, physicochemical characterization by X-ray powder diffraction (XRD), scanning electron microscopy (SEM) and solid-state MAS NMR (¹H, ²³Na, ²⁹Si and ¹¹⁹Sn MAS NMR) of a metallo-stannosilicate and its application as a heterogeneous catalyst for biodiesel production via ethanolysis and methanolysis routes using edible, non-edible an waste oils as feedstocks. XRD characterization of the metallo-stannosilicate has revealed a typical structure of mixed octahedral-pentahedral-tetrahedral material with an orthorhombic unit cell (a = 14.522 Å, b = 12.469 Å, and c = 10.654 Å). ¹¹⁹Sn single-pulse MAS NMR results presented two resonance lines, at δ_1 = -683.8 ppm and δ_2 = -690.9 ppm, which were assigned to the tin octahedrally coordinated in the metallo-stannosilicate structure. Heterogenous catalytic studies in the transesterification of refined edible, non-edible, and waste oils have resulted in high yields of FAMEs and FAEEs (fatty acid methyl and ethyl esters), nevertheless the highest FAMES (98.2%) and FAEEs (96.6%) yields were obtained for non-edible microalgae oil extracted from the genetically modified heterotrophic algal strain Prototheca moriformis. Catalytic studies also using non-edible macaw palm oil (Acrocomia aculeata) with a high content of free fatty acids (FFAs) demonstrated that the catalyst could simultaneously perform esterification and transesterification reactions. These findings indicate the potential application of this novel metallo-stannosilicate in the production of renewable and environmentally clean biofuels, like biodiesel, using different sources of lipids feedstocks, notably those that do not compete with food production.

1. Introduction

The search for alternative fuels has stimulated and intensified the efforts of the scientific community to identify viable ways of producing biofuels to replace or reduce the use of fossil fuels. Among the ecofriendly green technology, biodiesel is an attractive option. It is a sustainable and renewable form of energy, less harmful to the environment in comparison to petroleum diesel [1]. Chemically, biodiesel consists of a mixture of fatty acid alkyl esters (FAAEs), and at the industrial scale, it is predominantly produced using transesterification reactions using refined vegetable oils as sources of triacylglycerides, together with short-chain alcohols (methanol or ethanol) and homogeneous catalysts. There are clear potential environmental benefits of replacing petroleum diesel with biodiesel. Nevertheless, there are several drawbacks associated with the choice of the triacylglyceride feedstocks and the catalysts that preclude a faster development of this industry.

Concerning the available triacylglyceride sources, the current use of edible vegetable oils as lipid feedstocks for biodiesel production, instead of their exclusive use as foods, has raised ethical and economic questions, and this controversy has been the main driving force behind scientific efforts to identify alternative triacylglyceride sources that do not compete with food production [2,3]. Regarding catalyst technology, homogeneous catalysts such as sodium hydroxide (NaOH), potassium hydroxide (KOH) and sodium methoxide (NaOCH₃) are currently the main catalysts agents employed in the biodiesel industry. Their advantages include high biodiesel yields under mild conditions and in relatively short reaction time [4]. Nevertheless, homogeneous catalysts also have drawbacks: (a) Due to its hygroscopic nature, homogeneous

https://doi.org/10.1016/j.jece.2018.08.047 Received 28 April 2018; Received in revised form 26 July 2018; Accepted 22 August 2018 Available online 23 August 2018 2213-3437/ © 2018 Elsevier Ltd. All rights reserved.

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catalyst (NaOH, KOH) can be very hazardous for the environment, as compared to heterogeneous catalyst, (b) low quality feedstock vegetable oils with high contents of water and free fatty acids (FFAs) lead to the formation of soap, consequent loss of oil and problems of product separation during homogeneous catalytic transesterification. During homogeneous catalytic transesterification, the glycerol produced is of low quality and requires lengthy process and distillation for purification. All these processing increases the cost of the end products (biodiesel and glycerin). Moreover, the homogeneous base catalyzed transesterification process encountered problems to handle multiple feedstocks [5–7]. Therefore, from an economic, ethical, and environmental perspective, it is important to advance the development of more efficient catalysts that can be safely employed in industrial transesterification reactions using low-quality triacylglyceride feedstocks [8].

Although not currently predominantly employed on an industrial scale for biodiesel production using the transesterification process, heterogeneous catalysts could, in principle, be attractive alternatives to the homogeneous catalysts, since they are less sensitive to triacylglyceride feedstocks with high contents of FFAs and water [4,9]. Some of the advantages of the heterogeneous catalytic transesterification are: (a) the catalyst can be recycled (reused), (b) there is no or very less amount of wastewater produced during the process and (c) separation of the catalysts from biodiesel and glycerol is relatively easy, because the methanol or ethanol does not mix with the heterogeneous catalysts after the transesterification reaction [5]. There are several reports in the literature showing heterogeneous catalytic transesterification as a viable catalytic alternative for industrial large scale of biodiesel using batch or continuous processes [8,10]. Several excellent reviews have described a plethora of inorganic, organic and composite materials employed as heterogeneous catalysts in laboratory scale for biodiesel production: alkali metal oxides (MgO,CaO, SrO, BaO), transition metal oxides and their derivatives (TiO₂/SO₄²⁻, ZrO₂/SO₄² and Na₂MoO₄), mixed oxides and their derivatives (KF/Zn(Al)O, Zn(Al)O, KF/Zn(Al)O, MgO-MgAl₂O₄, KNO₃/ZrO₂ and SO₄²⁻/SnO₂), ion exchange resins type acid heterogeneous catalyst, carbon-based heterogeneous catalysts, boron group base heterogeneous catalyst, waste material based heterogeneous catalysts and enzyme-based heterogeneous catalyst [4,9,11–15]. Although microporous crystalline solid materials, built up of tetrahedrally coordinated (e.g., Si, Al, P) atoms have been studied as heterogeneous catalysts for transesterification reactions, heterogeneous catalysts based on mixed octahedral-pentahedral-tetrahedral microporous (OPT) siliceous frameworks have not yet been reported in the literature for this purpose.

Chukanov et al. [16] and Rocha et al. [17] have reviewed the steady progress in methods of synthesis and investigation of physical properties and crystal-chemical features of silicate microporous materials with transition elements (so-called zeolite-like amphoterosilicates). Comprehensive research into synthetic OPT materials was initiated by the seminal contributions of the titanosilicates ETS-10 and ETS-4 [18], tantalo and niobiosilicates [19,20] and stannosilicates [21,22]. Materials of this type are promising ion exchangers, sorbents, catalysts or catalyst carriers and, accordingly, can be used in chromatography, catalysis, water purification, among another process. Unlike common zeolites being aluminosilicates, frameworks of amphoterosilicates with transition metals are built of both tetrahedral fragments and "strong" cations (Ti, Nb, Zr, Ta, Sn, W, Fe, Mn, Zn, etc) with coordination numbers 5 or 6.

Tin-based homogeneous and heterogeneous catalyst have been used in triacylglyceride transesterification, mainly with refined edible oils as feedstocks and methanol as a solvent in the reaction, typically providing high yields of fatty acid methyl esters (FAMEs) [23–30]. Nevertheless, a careful analysis of the literature reveals a lack of technical information concerning the application of OPT tin-based catalysts for biodiesel production using edible, non-edible vegetable oils or various waste cooking and degummed oils as triacylglyceride feedstocks. Therefore, in order to address this gap, the present work describes the synthesis and the spectroscopic characterization of an OPT tin-based heterogeneous catalyst which was tested in the transesterification reaction of different sources of edible oils (soybean, corn, canola, sunflower), non-edible oils (microalgae, palm, macaw palm) and also waste cooking oils as lipid feedstocks. Both the methanolysis and ethanolysis routes were employed. Optimization of the transesterification reactions by taking into consideration relevant factors such as temperature, pressure, and the turnover (reuse) of the catalyst were also performed. The experimental herein reported indicate the potential of this OPT metallo-stannosilicate being employed as a viable heterogeneous catalyst for the production of biodiesel using both methanolysis and ethanolysis route and a variety of different kinds of lipid feedstocks, notably those that do not compete with food production.

2. Materials and methods

2.1. Chemicals

The stannosilicate synthesis and the catalytic reactions employed the following chemicals: Ludox HS-40 colloidal silica (40 wt. % SiO₂ in water), tin chloride dihydrate (SnCl2•2H2O, 98%), and sodium hydroxide (NaOH) were purchased from Sigma-Aldrich (Steinheim, Germany). Anhydrous methyl and ethyl alcohols (99.8%) were purchased from Dinâmica Química (Diadema, Brazil). Refined and degummed palm oils (Elaeis guineensis) were obtained from Agropalma (Belém, Brazil). Refined soybean oil (Glycine max), corn oil (Zea mays), rapeseed oil (Brassica napus), sunflower oil (Helianthus annuus), and degummed soybean oil were acquired from Cargill (Sao Paulo, Brazil). Refined microalgae oil extracted from the genetically modified heterotrophic algal strain Prototheca moriformis and produced according to Trimbur et al. [31] was obtained from Solazyme Company (Orindiúva, Brazil). Degummed macaw palm oil (Acrocomia aculeata) was acquired from UBCM Company (Montes Claros, Brazil). The FFAs contents (wt. %) and acid values (mg KOH g^{-1}) were measured for all the lipid feedstocks used in this study, according to the AOCS Ca 5a-40 (2009).

2.2. Hydrothermal synthesis of the stannosilicate

The stannosilicate was synthesized using experimental procedures adapted from the literature [21,22]. In a typical synthesis, 21.1 g of sodium hydroxide (NaOH) was dissolved in 15 mL of distilled water (solution A). In the next step, 54.9 g of Ludox HS-40 colloidal silica (40 wt. % SiO₂ in water) was added to solution A and the reaction mixture was kept under stirring for 30 min (500 rpm) at 25 °C, resulting in the formation of a homogeneous gel (solution B). Subsequently, 12.8 g of stannous chloride (SnCl₂·2H₂O) was added to solution B, under moderate stirring (500 rpm) for 60 min at 25 °C, resulting in a homogeneous gel. The final sol-gel solution was transferred to a 125 mL Teflon-lined stainless steel autoclave (Parr Instruments Co., Illinois, USA) and kept at 200 °C from 2 days to 42 days. Afterward, the reactor was cooled down and the solid product was collected by filtration, washed with distilled water, and dried at 100 °C for 12 h, prior to its physicochemical characterization and application in the catalytic reactions.

2.3. Physicochemical characterization of the stannosilicate

The as-synthesized stannosilicate was characterized by X-ray diffraction (XRD) using a Miniflex instrument (Rigaku, Tokyo, Japan) operated at 40 kV, 15 mA, with Ni-filtered Cu-K α radiation ($\lambda = 1.5418$ Å), in the 2 θ range from 3 to 80°, at a goniometer scan rate of 2° (2 θ) min⁻¹. Scanning electron microscope (SEM) and energy-dispersive X-ray spectrometry (EDS) analyses were performed using an Inspect F50 microscope (FEI Instruments, Oregon, USA) using the facilities of the Brazilian National Laboratory of Nanotechnology (LNNano, Electron

Microscopy Laboratory, Campinas, Brazil). The electron microscope was equipped with a Schottky field emission source, the probe current was 200 mA, and the electron beam acceleration voltage ranged between 0.2 and 30 kV. Atomic force microscope analyzes (AFM) were also made in the Brazilian National Laboratory of Nanotechnology (LNNano, Laboratory of Surface Science, Campinas, Brazil) using a Park NX-10 microscope (Park Systems, Santa Clara, USA) with scanning of $100 \,\mu\text{m} \times 100 \,\mu\text{m}$ and z-axis detector with amplitude of $15 \,\mu\text{m}$, fitted with a PPP-NCHR probe operated at 320 KHz resonance frequency and constant force of 42 N/m. Surface area measurements at 77 K were performed on a Micromeritics ASAP 2020 (Micrometrics Instrument Corporation, Norcross, USA) using the facilities of the BAM, Federal Institute for Materials Research and Testing (Division 1.3. Structural Analysis, Berlin, Germany). Before the measurement, all samples were degassed at temperature of 300 °C and pressure of 5×10^{-5} mbar for 3 h, and isotherms were processed by the BET (Brunauer-Emmett-Teller) method.

Solid state magic angle spinning NMR (MAS NMR) experiments were performed using the facilities of the BAM, Federal Institute for Materials Research and Testing (Division 1.3, Structural Analysis, Berlin, Germany). The stannosilicate was characterized by ¹H, ²³Na, ²⁹Si, and ¹¹⁹Sn solid-state MAS NMR. All the experiments were carried out at room temperature, with the samples filled into zirconia rotors equipped with Kel-F caps (Bruker, Wissembourg, France). The ¹H and ²³Na MAS NMR experiments were performed using a 600 MHz Avance spectrometer (Bruker, Wissembourg, France) with B₀ of 14.1 T, using 4 mm (¹H nucleus) and 2.5 mm (²³Na nucleus) MAS probes. The ²⁹Si and ¹¹⁹Sn single-pulse MAS NMR experiments were performed using a Bruker 400 MHz spectrometer ($B_0 = 9.4 \text{ T}$) with 7 mm MAS probes. The data were analyzed and processed using the TopSpin 3.0 software. For removing the ¹H background signals of the MAS probe, the EASY method was employed [32,33] using a MAS frequency of 12.5 kHz, 90° pulse duration of 3 us, recycle delay of 10 s, and accumulation of 16 scans. For removing all ¹H backgrounds, an additional ¹H EASY spectrum of an empty rotor is needed, which is subtracted from the ¹H EASY spectrum of the sample. The ¹H chemical shifts were referenced to TMS, using adamantane as a secondary reference ($\delta = 1.78$ ppm).

The ²³Na single-pulse MAS NMR spectrum was obtained using radiofrequency pulses at a Larmor frequency of 158.7 MHz, MAS frequency of 25 kHz, 90° pulse duration of 3.4 µs, recycle delay of 10 s, 64 scans, 2048 data points, and spectral width of 50 kHz. The ²³Na chemical shifts were reported relative to NaCl as a secondary reference ($\delta = 7.3$ ppm). The ²⁹Si single-pulse MAS NMR spectra were obtained using radiofrequency pulses at a Larmor frequency of 79.5 MHz, MAS frequency of 6.5 kHz, 90° pulse duration of 7.7 µs, and recycle delay varying from 2 s to 900 s. The ²⁹Si chemical shifts were reported relative to kaolin as a secondary reference ($\delta = -91.5$ ppm). The ¹¹⁹Sn single-pulse MAS NMR spectra were obtained using radiofrequency pulses at a Larmor frequency of 6.5 kHz, 90° pulse duration of 3.0 µs, and recycle delay varying from 2 s to 900 s. The ¹¹⁹Sn chemical shifts were reported relative to SnO₂ as a secondary reference ($\delta = -604$ ppm) [34].

2.4. Transesterification reactions of the refined edible, non-edible, and waste oils

Syntheses of FAMEs and FAEEs using the transesterification reactions were performed using the refined soybean oil (FFAs content = 0.2 wt. %, acid value = $0.4 \text{ mg KOH g}^{-1}$), palm oil (FFAs content = 0.3 wt. %, acid value = $0.6 \text{ mg KOH g}^{-1}$), corn oil, canola oil, sunflower oil, and microalgae oil (these last four triacylglyceride feedstocks presented FFAs contents < 0.1 wt. % and acid values < 0.2 mgg $^{-1}$), employing both the methanolysis and ethanolysis routes. A typical transesterification reaction, as exemplified for refined soybean oil as feedstock, was performed by transferring 20.0 g of the oil to a 100 mL open glass reactor equipped with a reflux condenser and a magnetic stirrer, immersed in a thermostatic bath. Anhydrous methanol or ethanol was added to give alcohol: oil molar ratio of 1:30. The appropriate amount of catalyst (3%, relative to the mass of oil) was added and the reaction mixture was heated at 100 °C for 12 h, under magnetic stirring (900 \pm 100 rpm). Afterward, the mixture was filtered and the excess alcohol (methanol or ethanol) was removed using a rotary evaporator (Rotavapor R-210, Büchi, Flawil, Switzerland). The glycerol formed as a by-product was easily separated by using a High-Speed refrigerated centrifuge (model CR22 N, Hitachi Koki Co., Ltd., Hitachikina-Ibaraki, Japan) for 10 min at 13,400 \times g.

FAMEs and FAEEs were also produced by transesterification of the non-edible and waste oils: degummed macaw palm oil (FFAs content = 55.8 wt. %, acid value = $111.0 \text{ mg KOH g}^{-1}$), degummed soybean oil (FFAs content = 3.6 wt. %, acid value = $7.2 \text{ mg KOH g}^{-1}$), degummed palm oil (FFAs content = 4.9 wt.%, acid value = 9.7 mg KOH g^{-1}), waste soybean cooking oil (FFAs content = 1.5 wt.%, acid value = $3.0 \text{ mg KOH g}^{-1}$) and waste palm cooking oil (FFAs content = 0.8 wt.%, acid value = 1.6 mg KOH g^{-1}). The reactions were carried out in a closed steel pressure reactor (Parr Instrument Company, Illinois, USA) equipped with a mechanical stirrer set at 700 \pm 100 rpm. The reaction temperature (150 \pm 5 °C) and the residence time (6 h) for these reactions resulted in maximum autogenic pressures of 8-10 bar (methanolysis reaction) and 6-8 bar (ethanolysis reaction). Reusability experiments were performed using refined and waste cooking soybean oils (Glycine max). These experiments were performed under two conditions catalytic procedures: (a) the catalyst was immediately reused after the transesterification reaction without any treatment and (b) after each reaction cycle, the catalyst was subjected to a calcination treatment (air atmosphere at temperature of 500 °C for 3 h, rate of 3 °C min⁻¹). The purpose of the calcination was to eliminate the glycerol byproduct from the catalytic sites of the stannosilicate, since its presence could hinder the transesterification reaction and consequently decrease the FAMEs yield [9,35].

2.5. Surface acidity measurements by FTIR of pyridine adsorption and TPD-NH3

Surface acidity was measured by Fourier Transform Infrared (FTIR) spectroscopy of pyridine adsorption and temperature-programmed desorption of ammonia (TPD-NH₃) using the facilities of the Humboldt-Universität zu Berlin (Institut für Chemie, Berlin, Germany). For the FTIR measurements, 75 mg of the sample was preheated at 150 °C for 2 h, under N2 flow. Afterward, 30 µL of pyridine (Sigma-Aldrich, Steinheim, Germany) was chemisorbed on the material at 150 °C for 15 min and the FTIR spectra was recorded at room temperature using an MTEC cell and an FTIR system 2000 (Perkin-Elmer, Massachusetts, USA). Spectra of the samples before pyridine adsorption reactions were also measured as the background results. For the acidity measurement by TPD-NH₃, the samples were pretreated at 500 °C for 2 h. Afterward, ammonia was adsorbed onto the surface of the samples at 100 °C and its desorption (10 °C min⁻¹ up to 500 °C) was monitored by FTIR detection of the band at 930 cm^{-1} . The amount of ammonia was determined by reaction with a solution of sulfur acid (H₂SO₄) and titration with sodium hydroxide (NaOH).

2.6. Determination of the FAMEs and FAEEs contents by gas chromatography

Chromatographic analyses were performed with a PerkinElmer Clarus 580 gas chromatograph (Perkin-Elmer, Massachusetts, USA) equipped with a flame ionization detector (GC-FID) using a Restek-Carbowax capillary column (30 m long \times 0.25 mm internal diameter). Gas chromatography analyses of the FAMEs and FAEEs in the biodiesel samples were performed using a sample injection volume of 1 mL, he-lium as the carrier gas at a flow rate of 2 mL min⁻¹ and pressure of 83 kPa, and injector and detector temperatures of 250 °C. The initial

oven temperature was at 50 °C (for 1 min), followed by an increase to 250 °C at a rate of 5 °C min⁻¹. In a typical procedure, standard stock solutions of ethyl myristate (C14:0) and ethyl nervonate (C24:1) were prepared at concentrations of 20 mg mL⁻¹, and an ethylic non-adecanoate standard stock solution was prepared at a concentration of 10 mg mL⁻¹. The ethyl esters (C14:0 and C24:1) were used to identify the peak integration range and the ethylic ester (C19:0) was used as an internal standard (IS). The FAMEs and FAEEs contents were obtained by integrating the peak areas from C14:0 to C24:1 and subtracting the nonadecanoate peak area, according to the following equation:

$$C = \frac{\sum A - A_{IS}}{A_{IS}} x \frac{C_{IS} - V_{IS}}{m} x 100\%$$

where,

 ΣA = sum of the areas of all peaks in the range from C14:0 to C24:0;

 $A_{IS} = C19:0$ internal standard peak area;

 C_{IS} = concentration (mg mL⁻¹) of the C19:0 solution;

 V_{IS} = volume of the C19:0 solution added to the sample;

m = mass of the sample (mg).

3. Results and discussion

3.1. Physicochemical characterization of the stannosilicate synthesized

OPTs microporous silicates are in general prepared under hydrothermal conditions ranging from ca. 120 to 230 °C and times varying between a few hours and ca. 30 days. In this study, the synthesis of the metallo-stannosilicate was investigated at 200 °C with a time interval ranging from 2 to 42 days. The XRD pattern of the as-synthesized stannosilicate, as a result of different reaction times, are displayed in Fig.1. It can be observed that at 2 days at 200 °C the main phases of the material are formed, although it shows poor crystallinity. However, after 4 days of synthesis, the material is completely formed and it kept its crystallinity, and no other impurity or concurrent phase is formed, even when the synthesis was kept for 42 days at 200 °C. The XRD pattern, for the sample prepared at 42 days at 200 °C, detailed in the inset of Fig. 1, shows that the as-synthesized stannosilicate corresponded to a crystalline material, with the main Bragg reflections in the 20 range 7.5-40°, characteristic of a microporous mixed OPT (Octahedral-Pentahedral-Tetrahedral) framework silicate structure [17,36].

Indexation of the main diffraction peaks of the sample prepared at 200 °C for 21 days using TREOR software [37] resulted in an orthorhombic unit cell with a = 14.522 Å, b = 12.469 Å, and c = 10.654 Å. The material obtained differed from other main stannosilicates reported in the literature, such as the stannosilicates Sn-A, Sn-B and Sn-L



Fig. 2. BET-N₂ adsorption isotherm of the stannosilicate.

reported by Corcoran et al. [21,22], the stannosilicate family AV-10, AV-11, AV-13 and AV-14 reported by Rocha et al., [38–41] and the stannosilicate EMS-2, described by Millini et al. [42]. The XRD pattern showed no evidence of tin oxide or a sodium stannate formation, with no diffraction peaks attributable to a SnO_2 phase (JCPDS 41–1445) or Na_2SnO_3 '3H₂O (JCPDS 01–1115).

Chemical analysis by Energy dispersive X-ray spectroscopy (EDS) showed atomic compositions of Si = 15.5 \pm 0.8%. Sn = 3.7 \pm 0.1% and Na = 13.3 \pm 1.8%, resulting in Si:Sn = 4.2 and Na:Sn = 3.6 ratios. BET-N2 adsorption isotherm of the stannosilicates synthesized, showed in Fig. 2, and the calculated microporous volume of $0.048 \text{ cm}^3/$ g, indicated that the porous of this material are small, probably a consequence of its dense structure. The material presents a low surface area of $14.8 \text{ m}^2/\text{g}$ when compared to $41.6 \text{ m}^2/\text{g}$ for the small-pore framework sodium stannosilicate AV-10 [38] and with values reported in the literature for SnO₂ nanoparticles synthesized by different methods [43-45]. The line broadening in the powder XRD pattern of the stannosilicate could be due to the small particles size of the sample: SEM and AFM images of the stannosilicates obtained at 7, 21 and 42 days at 200 °C indicated that the sample is formed by small plates aggregates, as shown in Fig. 3 (particle size ranging from 100 nm to 300 nm).



Fig. 1. XRD patterns of the stannosilicate synthesized at different reaction times and indexed Bragg reflections for the stannosilicate obtained at 42 days/200 °C (inset).



Fig. 3. Scanning electron microscopy (SEM) images of stannosilicates synthesized at (a) 7 days, (b) 21 days, (c) 42 days and (d) Atomic force microscopy (AFM) image.



Fig. 4. Characteristic ²⁹Si single-pulse MAS NMR spectra obtained for stannosilicate synthesized at different reaction times. Detailed deconvoluted spectrum of the stannosilicate prepared at 42 days/200 °C.

 29 Si single-pulse MAS NMR studies of the stannosilicate obtained at 4, 7, 21 and 42 days at 200 °C was performed and the spectra are shown in Fig.4. Although the XRD data revealed that the same material was obtained at 4–42 days at 200 °C, analysis of the 29 Si single-pulse MAS NMR spectra revealed a more complex local symmetry disorder is occurring. The deconvoluted 29 Si single-pulse MAS NMR spectrum of the

sample prepared at 42 days at 200 °C is shown in the inset of Fig. 4. ²⁹Si environment assignment was based on the seminal research for ²⁹Si solid-state MAS NMR of aluminosilicates structures [46,47]. Nine resonance lines were present, with the following chemical shift values: $\delta_1 = -76.5 \text{ ppm}, \delta_2 = -78.1 \text{ ppm}, \delta_3 = -80.3 \text{ ppm}, \delta_4 = -81.4 \text{ ppm}, \delta_5 = -83.6 \text{ ppm}, \delta_6 = -84.7 \text{ ppm}, \delta_7 = -85.5 \text{ ppm}, \delta_8 =$



Fig. 5. (a) ¹H and (b) ²³Na single-pulse MAS NMR spectra of the stannosilicate (*spinning sidebands, MAS frequency = 6.5 KHz).

-97.1 ppm, and $\delta_9 = -109.7$ ppm. These chemical shifts differed from those described in literature, as the obtained for the AV-10 stannosilicate [38], where the spectrum showed two chemical shifts, at δ_1 = -87.0 ppm and $\delta_2 = -88.7$ ppm, with relative intensities of 1:2, associated with the Si(2Si, 2Sn) environment. Rocha et al. [40] reported a broad resonance ²⁹Si spectrum for AV-13 material, with chemical shift value at $\delta = -87.1$ ppm, which calls for the presence of a single Si (2Si,2Sn) site. Corcoran et al. [21,22] reported ²⁹Si chemical shifts for the three stannosilicate phases prepared by them, at $\delta_1 = -88.0$ ppm and $\delta_2 = -90.3 \text{ ppm}$ (Sn-A), $\delta_1 = -89.0 \text{ ppm}$ (Sn-B), and $\delta_1 =$ -82.6 ppm and $\delta_2 = -87.2$ ppm (Sn-L). Millini et al. [42] reported three well-resolved ²⁹Si peaks for the EMS-2 stannosilicate: δ_1 = -90 ppm, $\delta_2 = -99$ ppm and $\delta_3 = -112$ ppm. Based on the reported ²⁹Si chemical shift values and the XRD diffraction results, it is evident that the stannosilicate obtained in this study is different from those reported in the literature.

Considering that the catalytic studies were systematically done with the stannosilicate obtained at 21 days at 200 °C, a more detailed analysis of ¹H, ²³Na and ¹¹⁹Sn single-pulse MAS NMR spectra of this sample was performed. The ¹H MAS NMR spectrum of the stannosilicate (Fig. 5a) showed only one broad resonance line at $\delta_2 = 4.8$ ppm, associated with the presence of structural water molecules [47,48]. The ²³Na single-pulse MAS NMR spectrum (Fig. 5b) presented a chemical shift at around $\delta = 7.4$ ppm, which could be attributed to the formation of NaCl, as reported by Hunger et al. [49]. The role of sodium in the synthesis of stannosilicates and tin zeolites has been discussed in the literature and one of the conclusions reached is that interactions among the sodium and tin cations can lead to the formation of sodium stannate as an impure phase [50], however XRD analysis showed no evidence of this impure phase (JCPDS 01-1115). Attempts to assign the resonance lines with greater accuracy were unsuccessful, due to the difficult task of performing the spectrum deconvolution (Fig. 5b), considering the quadrupolar nature of the sodium (²³Na) nucleus.

The ¹¹⁹Sn single-pulse MAS NMR spectrum (Fig. 6) presented two resonance lines, at $\delta_1 = -683.8$ ppm and $\delta_2 = -690.9$ ppm, which could be assigned to tin species in the Sn⁴⁺ oxidation state, indicating that the tin atoms were octahedrally coordinated in the stannosilicate structure [34,38,51]. The ¹¹⁹Sn single-pulse MAS NMR analysis indicated that Sn²⁺ was oxidized to Sn⁴⁺ during the hydrothermal crystallization. It is entirely possible that Sn²⁺ acted as a reducing agent during the course of the hydrothermal crystallization. Sn²⁺ is a strong reducing agent and in the presence of a strongly alkaline solution it can cause the formation of tetrahydroxystannate species, according to the following chemical reaction: Sn²⁺ + 4OH⁻ = (Sn(OH)₄)²⁻.

3.2. Transesterification reactions of edible, non-edible and waste oils

Transesterifications reactions were performed according to the

experimental conditions described in Section 2.4 using the stannosilicate at 21 days at 200 °C; however, experimental evidence acquired throughout this study lead to the conclusion that depending on the triacylglycerides feedstocks, two types of reactors were a need to obtain the best FAMEs or FAEEs yields. For refined edible and non-edible vegetable oils (average FFAs content ≤ 0.3 wt. %, and acid value $\leq 0.6 \text{ mg g}^{-1}$), a typical reactor (an open glass reactor equipped with a reflux condenser and a magnetic stirrer, immersed in a thermostatic bath) was used, while for degummed vegetable and waste cooking oils (average FFAs content ≤ 55.8 wt. %, and acid value $\leq 111 \text{ mg g}^{-1}$) was utilized a closed steel pressure reactor (Parr Instrument Company, Illinois, USA) equipped with a mechanical stirrer.

The FAMEs and FAEEs yields obtained in the transesterification reactions of the refined edible and non-edible vegetable oils are shown in Table 1. The catalytic conversions of the refined soybean, corn, canola, and sunflower oils exceeded 90.0% for FAMEs and FAEEs. A lower FAEEs yield of 82.6% was obtained with the refined palm oil. It is interesting to note the higher FAMEs and FAEEs yields obtained using microalgae oil, compared to the other triacylglyceride feedstocks. According to several critical reviews [52-55], biodiesel production from oil crops, waste cooking oil, and animal fat cannot satisfy even a small fraction of the existing demand for transport fuels, and microalgae appear to be the only source of renewable biodiesel capable of meeting the global transport fuel demand [52,53]. To the best of our knowledge, there is no report in the scientific or patent literature reporting the use of heterogeneous catalyst based on metallo-stannosilicate on the transesterification of microalgae oils using both the methanolysis or ethanolysis processes.

Due to the fact that the transesterification reactions of the non-edible and waste oils were performed in a closed steel reactor (150 \pm 5 °C for 6 h), the generated autogenous pressure derived from the closed nature of the container was similar for both the methanolysis (8-10 bar) and ethanolysis (6-8 bar) reactions. The FAMEs and FAEEs yields obtained for these reactions are displayed in Table 2. Transesterification with degummed soybean oil (3.6 wt. % of FFAs, acid value = $7.2 \text{ mg KOH g}^{-1}$) resulted in a FAMEs yield of 82.0%, while the yield of FAEEs was about 20.3%. The same pattern was repeated with degummed palm oil (FFAs content of 4.9 wt. %, acid value = 9.7mg KOH g^{-1}) with yields of 59.5% and 10.6% for FAMEs and FAEEs, respectively. The FAMEs and FAEEs yields for degummed macaw palm oil (FFAs content = 55.8 wt.%, acid value = $111.0 \text{ mg KOH g}^{-1}$) were 65.5% and 30.9% respectively, and the results provide clear evidence of the potential of the stannosilicate for simultaneous use as a heterogeneous catalyst in both esterification and transesterification reactions involving low-quality raw materials and by methanolysis and ethanolysis reactions. Although several reports are describing simultaneous esterification and transesterification of vegetable oils using tin-based catalysts [25,28-30,56-58], none of the studies were performed with



Fig. 6. ¹¹⁹Sn single-pulse MAS NMR spectrum of the stannosilicate.

degummed macaw palm oil as feedstocks.

Table 3 summarizes the studies reported in the literature that have used tin-based catalysts in methanolysis and ethanolysis reactions for biodiesel production, and a comparison of the FAMEs and FAEEs obtained in this study. Serra et al. [25] used commercial Lewis acid Sn(IV) compounds in the transesterification of refined soybean oil and obtained a maximum FAMEs yield of around 23.0%, while here the stannosilicate provided a FAMEs yield of 99.5%. Abreu et al. [23] tested several catalysts based on transition metals, including Sn $(C_6H_5O_3)_2(H_2O)_2$, $Zn(C_6H_5O_3)_2(H_2O)_2$, $Pb(C_6H_5O_3)_2(H_2O)_2$, and Hg $(C_6H_5O_3)_2(H_2O)_2$, in the transesterification of refined soybean oil and found that the tin-based catalyst was most active: the FAMEs yield was 93.8%, closed to the values obtained using the stannosilicate in the present work. A similar FAMEs yield (89.2%) was reported by Casas et al. [28] who employed tin(II) acetate $(Sn(C_6H_5O_3)_2(H_2O)_2)$ in the transesterification.

Solid acids catalysts have been established as being other alternatives to the heterogeneous alkaline, unrecyclable-homogeneous acid and alkaline catalysts [59-62], and since the seminal work of Chavan et al. [63] and Matsuhashi et al. [64] on the esterification of ß Keto ester, tin-based catalysts was related to the strong acidity on its surface. In the case of the metallo-stannosilicate reported in this study, the experimental results for FTIR of pyridine adsorption and temperatureprogrammed desorption of ammonia (TPD-NH₃) revealed a very weak acidity (Fig.7). Characteristic IR-bands of adsorbed pyridine related to the strength and quantity of both Lewis and Bronsted acid sites were not observed on the stannosilicate surface. According to the literature, the bands 1453 cm⁻¹ and 1622 cm⁻¹ are associated with the interaction between the pyridine molecule and Lewis sites, the bands 1545 cm⁻¹ and 1645 cm⁻¹ are associated with Bronsted sites, while 1493 cm⁻¹ and 1580 cm^{-1} are associated to both Lewis and Bronsted sites [65]. The low amount of ammonia desorbed on the catalyst surface = 1.1 $\times \ 10^{-2} \, \mu mol \, g^{-1})$ corroborates the low acidity of this stannosilicate.

It is quite possible that the catalytic activity of the synthesized stannosilicate is associated with the basicity induced by the sodium

Table 2

FAMEs and FAEEs yields obtained for the transesterification reactions with waste cooking and degummed vegetable oils. Reactions were performed in a closed steel pressure reactor; temperature = 150 °C; oil:alcohol molar ratio = 1:30; reaction time = 6 h; 3% of catalyst (relative to the mass of oil).

Feedstocks	Soybean oil (<i>Glycine max</i>)		Palm oi (Elaeis g	il guineensis)	Macaw palm oil (<i>Acrocomia</i>	
Waste		Degummed	Waste Degummed		uculeulu)	
FAMEs yield	99.0	82.0	91.2	59.5	66.5 ^a	
FAEEs yield (%)	70.2	20.3	71.3	10.6	30.9 ²	

Methanolysis reaction (autogenic pressure = 8-10 bar)Ethanolysis reaction (autogenic pressure = 6-8 bar).

^a Final free fatty acid (FFAs) content = $23.5 \text{ wt.}\%^2$ Final free fatty acid (FFAs) content = 32.1 wt.%.

species present in the crystalline structure of this material. The weak acidity measured by temperature-programmed desorption of ammonia (TPD-NH₃) (Fig.7b) and the ²³Na single-pulse MAS NMR spectrum (Fig. 5b) showing evidence of the formation of polymeric sodium species, corroborate this assumption. Several studies have also reported the effect of the basicity of sodium-based catalysts [66–70]. Zhang et al. [67] described the synthesis of a sodium stannate with a high concentration of basic sites and high catalytic activity in transesterification reactions of a cyclic carbonate with methanol. Marciniuk et al. [68] reported the application of sodium titanate as a basic catalyst in the transesterification of ethyl acetate with ethanol. Guo et al. [70] synthesized a sodium silicate (19.3–22.8 wt. % of Na₂O) used as a heterogeneous catalyst in the transesterification of soybean oil, achieving a FAMEs yield of around 100% under experimental conditions similar to those used in the present work.

Reuse experiments were performed with the refined and waste cooking soybean oil as the lipid feedstock in the methanolysis route. The results obtained with the refined soybean oil (Fig. 8a) showed that

Table 1

FAMEs and FAEEs yields obtained for the transesterification reactions with refined vegetable oils. Reactions were performed using an open glass reactor at atmospheric pressure; temperature = 100 °C; oil:alcohol molar ratio = 1:30; reaction time = 12 h; 3% of catalyst (relative to the mass of oil).

Refined oils	Palm oil	Soybean oil	Corn oil	Canola oil	Sunflower oil	Microalgae oil
FAMEs yield (%)	99.5	99.5	96.0	97.6	95.8	98.2
FAEEs yield (%)	82.6	90.2	93.8	95.0	94.8	96.6

Table 3

Examples of	tin-based	heterogeneous	catalysts	used ir	1 the	transesterification	reactions	of vegetable	oils into	biodiesel.

Feedstock	Catalyst	Alcohol	Alcohol:oil	Catalyst loading (wt.%)	Temperature (°C)	Time (h)	Yield (%)	Reference
Refined	Stannosilicate	Methanol	30:1	3	100	12	99.5	Present work
soybean oil	Stannosilicate	Ethanol	30:1	3	100	12	90.2	Present work
	Sn(3-hydroxy-2-methyl-4-pyrone) ₂ (H ₂ O) ₂	Methanol	400:100:1	_	60	5	93.8	[23]
	Dibutyltin dilaurate	Methanol	400:100:1	_	80	10	43.1	[24]
	Dibutyltin diacetate	Methanol	400:100:1	_	60	4	23.0	[25]
	Dibutyltin diacetate	Methanol	400:100:1	_	150	4	79.0	[25]
	SnO ₂ /SiO ₂	Methanol	24:1	5	180	5	81.7	[26]
Refined	Stannosilicate	Methanol	30:1	3	100	12	99.5	Present work
palm oil	Stannosilicate	Ethanol	30:1	3	100	12	82.6	Present work
	Stannosilicate	Ethanol	30:1	3	150	6	99.5	Present work
	Sn(3-hydroxy-2-methyl-4-pyrone) ₂ (H ₂ O) ₂	Methanol	400:100:1	_	60	1	16.2	[27]
Refined	Stannosilicate	Methanol	30:1	3	100	12	95.8	Present work
sunflower oil	Stannosilicate	Ethanol	30:1	3	100	12	94.8	Present work
	Tin(II) acetate	Methanol	30:1	0.15	100	1	46.4	[28]
	Tin(II) acetate	Methanol	30:1	0.15	150	1	89.2	[28]
Waste	Stannosilicate	Methanol	30:1	3	150	6	99.0	Present work
soybean cooking	Stannosilicate	Ethanol	30:1	3	150	6	70.2	Present work
oil	SO4 ²⁻ /SnO2-SiO2	Methanol	15	6	150	1.5	88.2	[30]
	SO4 ²⁻ /SnO2-SiO2	Methanol/ethanol	9-6:1	6	150	1	81.4	[29]

the calcined stannosilicate maintained a FAMEs yield of around 90.0% after four reaction cycles, while the yield achieved with the non-calcined catalyst decreased sharply to 38.7% after three reaction cycles. Meanwhile, the results for the waste cooking oil using the calcined catalyst (Fig. 8b) indicated FAMEs yields of \geq 90.0% in the first three cycles, after which the yield progressively decreased to 35.2% in the sixth cycle. The findings reported here using the synthesized stannosilicate are novel and demonstrate the considerable potential of this material in the field of biodiesel research.

palm oil (*Acrocomia aculeate*) with a high content of FFAs indicated that the catalyst could simultaneously perform esterification and transesterification reactions. The experimental results obtained with this new material indicated its potential for being used as a heterogeneous catalyst in the production of clean, renewable and environmentally friendly biofuels with different kinds of lipid feedstocks, notably those that do not compete with food production.

Acknowledgments

4. Conclusions

A metallo-stannosilicate was obtained by hydrothermal synthesis and fully characterized by powder XRD, electron microscopy (SEM) and solid-state MAS NMR (¹H, ²³Na, ²⁹Si and ¹¹⁹Sn MAS NMR). High yields of FAMEs and FAEEs were obtained using the metallo-stannosilicate as heterogeneous catalyst in the transesterification reactions of refined edible, non-edible, and waste oils. In comparison with several non-edible lipid feedstocks, the highest FAEEs yields were obtained non-edible microalgae oil extracted from the genetically modified heterotrophic algal *Prototheca moriformis*. Catalytic tests using non-edible macaw We thank the State of São Paulo Research Foundation (FAPESP) and National Council for Scientific and Technological Development (CNPq) for the grant awards #11/51851-5 and #406761/2013-2, respectively. D.A.S. thanks CAPES for his fellowship (Proc. n^o BEX 2428/15-6). We acknowledge Dr. Franziska Emmerling and Mr. Carsten Prinz (BAM, Division 1.3. Structural Analysis) for the BET-N₂ measurements, Prof. Dr. Erhard Kemnitz and Dr. Kerstin Scheurell (Humboldt Universität zu Berlin, Institut für Chemie) for the FTIR-Pyridine and TPD-NH₃ measurements. We appreciate the assistance of Dr. Jefferson Bettini, Dr. Carlos A. O. Ramirez and Dr. Carlos A. Costa (Brazilian National Laboratory of Nanotechnology, LNNano) for the SEM and AFM







Fig. 8. Surface acidity measurements of the stannosilicate: (a) FTIR spectroscopy of pyridine adsorption and (b) temperature-programmed desorption of ammonia (TPD-NH₃).

microscopy experiments.

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