RESSALVA

Atendendo solicitação do(a) autor(a), o texto completo desta tese será disponibilizado somente a partir de 05/09/2024.



UNESP - Universidade Estadual Paulista "Júlio de Mesquita Filho" Faculdade de Odontologia de Araraquara



Eric Hernán Coaguila Llerena

Root canal irrigation: physicochemical and biological properties of calcium hypochlorite and effects of the GentleWave system in endodontic treatment

Araraquara 2022



UNESP - Universidade Estadual Paulista "Júlio de Mesquita Filho" Faculdade de Odontologia de Araraquara



Eric Hernán Coaguila Llerena

Root canal irrigation: physicochemical and biological properties of calcium hypochlorite and effects of the GentleWave system in endodontic treatment

Thesis presented to São Paulo State University (Unesp), School of Dentistry, Araraquara, to obtain the PhD degree in Dentistry, in the area of Endodontics.

Supervisor: Gisele Faria, PhD

Araraquara 2022 L791r Coaguila Llerena, Eric Hernán Root canal irrigation: physicochemical and biological properties of calcium hypochlorite and effects of the GentleWave system in endodontic treatment / Eric Hernán Coaguila Llerena. -- Araraquara, 2022 126 p. Thesis (doctoral) – São Paulo State University (Unesp). School of Dentistry Supervisor: Gisele Faria 1. Biofilms. 2. Calcium hypochlorite. 3. High-throughput nucleotide sequencing. 4. Surface-active agents. 5. Materials testing. I. Title.

Sistema de geração automática de fichas catalográficas da Unesp. Biblioteca da Faculdade deOdontologia, Araraquara. Dados fornecidos pelo autor(a).

Essa ficha não pode ser modificada.

Eric Hernán Coaguila Llerena

Root canal irrigation: physicochemical and biological properties of calcium hypochlorite and effects of the GentleWave system in endodontic treatment

Judging Commission

Thesis for obtaining the PhD degree in Dentistry

- President and supervisor: Gisele Faria, PhD
- 2° Examiner: Ronald Ordinola-Zapata, PhD
- 3° Examiner: Giampiero Rossi-Fedele, PhD
- 4° Examiner: Mário Tanomaru-Filho, PhD

Araraquara, September 5, 2022

CURRICULAR DATA

Eric Hernán Coaguila Llerena

BIRTH DATE: October 27, 1984 – Arequipa – Peru

- FILIATION: Andrés Eloy Coaguila Rivera Irma Herminia Llerena Sierra
- 2002 2006 Graduated in Dentistry. Universidad Católica de Santa Maria, Peru.
- 2008 2009 Continuing education course in Oral Surgery and Radiology. Hospital Militar Regional, Arequipa, Peru.
- 2011 2012 Continuing education course in Endodontics. Colegio Odontológico del Perú, Región Arequipa, Peru.
- 2012 2014 Specialization in Endodontics. Universidad Peruana Cayetano Heredia, Peru.
- 2014 2014 International internship in Endodontics Nova Southeastern University, Fort Lauderdale, Florida - USA.
- 2015 2015 Professor at the Academic Department of Stomatology Clinic. Universidad Peruana Cayetano Heredia, Peru.
- 2016 2018 MSc in Endodontics. Faculdade de Odontologia de Araraquara – FOAr UNESP.
- 2021 2022 Sandwich doctorate. Division of Endodontics, School of Dentistry, University of Minnesota, MN, USA.
- 2018 2022 PhD in Endodontics (in progress). Faculdade de Odontologia de Araraquara – FOAr UNESP.

I dedicate this work:

To God, in whose omnipresence, infinite benevolence and unquestionable will, I know that there is no impossible. God gave me nothing I wanted, he gave me everything I needed.

To my family, because it is through them that I learned to remain humble in victory and elegant in defeat. I know that the tireless pursuit of the realization of my dreams and the consolidation of the principles that guide my life are a consequence of what I learned at home. And, without a doubt, it is through them that I learned that I should never give up because God provides enough strength.

ACKNOWLEDGMENTS

To my parents, Irma and Andrés; my brothers, Carlos and Daniel; to our new members, Milagros and Camila; our dear Paola (*in memorian*); and our unforgettable pet, Orson (*in memorian*). Also, the members of Coaguila and Llerena families. All of them are examples of life that I look up to, as well as being my inexhaustible source of inspiration, strength and perseverance.

To my supervisor, Gisele Faria, who guided me during six years, a cycle that I will always remember with great gratitude. I am privileged that the path taken during this time had constant doses of optimism, dedication, constant search for perfection and attention to detail that can only be achieved under her guidance. The fruit of this partnership brought academic production, learning, and personal development, but above that, true friendship.

To my supervisor during sandwich doctorate, Ronald Ordinola-Zapata, who believed in my potential to do an internship abroad and from whom I learned other academic paths that I had not explored. He gave me not only opportunities but his sincere friendship. My gratitude.

To the professors of the Discipline of Endodontics at FOAr, whose example is the paradigm in which I aspire to my professional path. Their wisdom is only comparable to the humbleness they show to their students.

To all the teachers that life gave me, whose teachings are always remembered with appreciation. All the professional aspirations I aspired to in life I know were and are being fulfilled through the training I received from them.

To my beloved friends that I made in Araraquara, both Brazilians and foreigners. I am absolutely sure that the bonds of fraternity will always be strong. Whether in the faculty, at home or anywhere, everything was better in their company. My gratitude.

To the School of Dentistry at Araraquara, FOAr, in the person of the Director, Edson Alves de Campos, PhD; and the Vice-Director, Patricia Petromilli Nordi Sasso Garcia, PhD.

To the Postgraduate Program in Dentistry, FOAr/UNESP, in the person of the Coordinator, Paulo Sergio Cerri, PhD; and the Vice-Coordinator, Morgana Rodrigues Guimarães-Stabili, PhD.

To the employees of FOAr/UNESP, always kind and willing to help. All of them are the point of support that leads to a proper functioning of our beloved institution. Their routine smile and kind words made the days better. My gratitude.

To the Regional Office of Support for Research and Internationalization / ERAPI, in the person of Renan Cesar Palomino, who always helped me in what was necessary in relation to the administrative processes of the scholarship. My gratitude.

To São Paulo Research Foundation (FAPESP) (Grant #2018/24662-6), for the essential financial support to perform this research.

This study was financed in part by the Coordenação de Aperfeiçoamento de Pessoal de Nível Superior – Brasil (CAPES) – Finance Code 001, and under the CAPES-PrInt Program, process nº 88887.310463/2018-00, Mobility nº 88887.570038/2020-00.

"O que dá o verdadeiro sentido ao encontro é a busca, e é preciso andar muito para se alcançar o que está perto" José Saramago*

^{*} Saramago J. Todos os nomes. Lisboa: Editorial Caminho; 1997.

Coaguila Llerena EH. Irrigação do canal radicular: propriedades físico-químicas e biológicas do hipoclorito de cálcio e efeitos do sistema GentleWave no tratamento endodôntico [tese de doutorado]. Araraquara: Faculdade de Odontologia da UNESP; 2022.

RESUMO

Parte 1: Na publicação 1, o objetivo foi avaliar as propriedades físico-químicas e a penetrabilidade nos túbulos dentinários do hipoclorito de cálcio [Ca(OCI)₂] a 2,5% associado aos surfactantes cloreto de benzalcônio (BAK), cetrimida (CTR), tween 80 (T80) e triton X-100 (TR100), na concentração micelar crítica (CMC), em comparação ao hipoclorito de sódio (NaOCI) a 2,5%. Foram avaliados tensão superficial, pH, conteúdo de cloro livre (FAC), íons cálcio livres (Ca2+) e a penetrabilidade nos túbulos dentinários. A adição de surfactantes reduziu a tensão superficial das soluções (p<0,05), e não alterou o pH e o FAC (p>0,05). Os surfactantes, principalmente BAK, aumentaram a disponibilidade de Ca^{2+} em $Ca(OCI)_2$ (p<0,05). $Ca(OCI)_2$ apresentou menor penetrabilidade nos túbulos dentinários que NaOCI (p<0.05) e a adição de surfactantes àquele não aumentou a sua penetrabilidade nos túbulos dentinários. Na publicação 2, o objetivo foi avaliar o mecanismo de citotoxicidade da solução de Ca(OCI)₂ em fibroblastos L929 e o seu efeito na biologia de osteoblastos-like (Saos-2), em comparação com NaOCI. Fibroblastos L929, expostos a Ca(OCI)₂ e NaOCI, foram avaliados quanto ao metabolismo celular, integridade dos lisossomos, tipo de morte, alterações no citoesqueleto e na ultraestrutura. O efeito das soluções sobre a atividade da fosfatase alcalina (ALP) foi determinada em Saos-2. O Ca(OCI)₂ promoveu maior viabilidade celular e menor porcentagem de apoptose e necrose do que o NaOCI (p<0,05). Ca(OCI)₂ e NaOCI diminuíram o metabolismo celular e a integridade dos lisossomos, induziram ruptura de microtúbulos e filamentos de actina, promoveram alterações do retículo endoplasmático rugoso e nas cristas mitocondriais, e não induziram a atividade da ALP. Concluiu-se que, embora a adição de surfactantes ao Ca(OCI)₂ não aumentou a sua penetrabilidade nos túbulos dentinários, eles promoveram menor tensão superficial, sem alterações nos valores de pH e FAC, além de maior disponibilidade de Ca²⁺ na solução. Adicionalmente, NaOCI e Ca(OCI)₂ apresentaram o mesmo mecanismo de citotoxicidade e não induziram atividade de ALP, porém, Ca(OCI)₂ foi menos citotóxico que NaOCI. Parte 2: Na publicação 3, o objetivo foi descrever, por meio de revisão de literatura, os efeitos do sistema GentleWave (GW) no tratamento endodôntico. GW mostrou resultados in vitro semelhantes ou melhores que a irrigação utrassônica passiva (PUI) sob diferentes aspectos. Clinicamente, embora GW tenha mostrado resultados promissores, ainda são necessários mais estudos. Na publicação 4, o objetivo foi avaliar a eficácia de remoção de biofilme multiespécie do GW e PUI. Molares inferiores humanos com configuração tipo II de Vertucci na raiz mesial foram inoculados com placa dental e incubados. As raízes mesiais foram instrumentadas até 20.06 (V-Taper) para o grupo GW, e até 35.04 (Vortex Blue) para o grupo PUI. Raspas de dentina foram obtidas pré e pós-tratamento para a análise por reação em cadeia da polimerase quantitativa (qPCR) e por sequenciamento do gene rRNA 16S (Next Generation Sequencing - NGS). Não houve diferença na remoção de biofilme entre GW e PUI (p>0,05). Pode-se concluir que mais pesquisas, principalmente clínicas, são necessárias para estabelecer se GW apresenta vantagens sobre outros métodos de irrigação.

Palavras chave: Biofilmes. Hipoclorito de cálcio. Hipoclorito de sódio. Permeabilidade da dentina. Sequenciamento de nucleotídeos em larga escala. Tensoativos. Teste de materiais.

Coaguila Llerena EH. Root canal irrigation: physicochemical and biological properties of calcium hypochlorite and effects of the GentleWave system in endodontic treatment [tese de doutorado]. Araraquara: Faculdade de Odontologia da UNESP; 2022.

ABSTRACT

Part 1: In publication 1, the aim was to assess the physicochemical properties and the penetration into dentinal tubules of calcium hypochlorite [Ca(OCI)₂] at 2.5% associated with the surfactants benzalkonium chloride (BAK), cetrimide (CTR), tween 80 (T80) and triton X-100 (TR100), at critical micellar concentration (CMC), compared to sodium hypochlorite (NaOCI) at 2.5%. Surface tension, pH, free available chlorine (FAC), free calcium ions (Ca²⁺) and penetration into dentinal tubules were evaluated. The addition of surfactants reduced the surface tension of the solutions (p<0.05), and did not change the pH and FAC (p>0.05). Surfactants, mainly BAK, increased the Ca²⁺ availability in Ca(OCI)₂ (p<0.05). Ca(OCI)₂ showed lower penetration into dentinal tubules than NaOCI (p<0.05) and the addition of surfactants to Ca(OCI)₂ did not increase its penetration into dentinal tubules. In publication 2, the aim was to assess the cytotoxicity mechanism of Ca(OCI)₂ solution on L929 fibroblasts and its effect on osteoblast-like (Saos-2) biology, compared to NaOCI. Cellular metabolism, lysosome integrity, type of cell death, changes in cytoskeleton and ultrastructure of L929 fibroblasts, exposed to Ca(OCI)₂ and NaOCI, were evaluated. The effect of the solutions on alkaline phosphatase (ALP) activity was determined in Saos-2. Ca(OCI)₂ promoted higher cell viability and lower percentage of apoptosis and necrosis than NaOCI (p<0.05). Ca(OCI)2 and NaOCI decreased cellular metabolism and lysosome integrity, induced disruption of microtubules and actin filaments, promoted changes in the rough endoplasmic reticulum and mitochondrial cristae, and did not induce ALP activity. It was concluded that, although the addition of surfactants to Ca(OCI)₂ did not increase its penetration into dentinal tubules, they promoted lower surface tension, without changes in pH and FAC values, in addition to higher Ca²⁺ availability. Additionally, NaOCI and Ca(OCI)₂ showed the same mechanism of cytotoxicity, and they did not induce ALP activity; however, Ca(OCI)₂ was less cytotoxic than NaOCI. Part 2: In publication 3, the aim was to describe, through a literature review, the effects of the GentleWave system (GW) in endodontic treatment. GW showed similar or better in vitro results than passive ultrasonic irrigation (PUI) in different aspects. Clinically, although GW has shown promising results, further studies are still needed. In publication 4, the aim was to assess the effectiveness of multispecies biofilm removal of GW and PUI. Human mandibular molars with Vertucci type II configuration in the mesial roots were inoculated with dental plaque and incubated. Mesial roots were instrumented up to 20.06 (V-Taper) for GW group, and up to 35.04 (Vortex Blue) for PUI group. Dentin shavings were obtained pre- and post-treatment for analysis by quantitative real-time Polymerase Chain Reaction (qPCR) and 16S ribosomal RNA gene sequencing (Next Generation Sequencing - NGS). There was no difference in biofilm removal between GW and PUI (p>0.05). It can be concluded that more research, mainly clinical, is needed to establish whether GW has advantages over other irrigation methods.

Keywords: Biofilms. Calcium hypochlorite. Sodium hypochlorite. Dentin permeability. High-throughput nucleotide sequencing. Surface-active agents. Materials testing.

SUMMARY

1 INTRODUCTION	11
2 PROPOSITION	17
3 PUBLICATIONS	
3.1 Publication 1	19
3.2 Publication 2	40
3.3 Publication 3	63
3.4 Publication 4	81
4 CONCLUSIONS	100
REFERENCES	101
APPENDICES	107
ATTACHMENTS	116

1 INTRODUCTION

The control of infection is critical for the success of endodontic treatment¹, and the irrigation solution and its métodos de agitação used in chemo-mechanical preparation has an important role to achieve this goal².

Sodium hypochlorite (NaOCI) is the most commonly used irrigating solution due to its antimicrobial activity and organic dissolution capacity^{3–5}, being considered as *"gold standard*"⁶. However, NaOCI negatively alters the mechanical properties of dentine, such as microhardness, elastic modulus, resistance to flexion and fatigue⁷, and can reduce the bond strength of root canal sealers⁸ and some dentine adhesive materials^{7,9}. Additionally, NaOCI does not provide adequate removal of the smear layer from the dentine surface¹⁰, and when interacts with ethylenediaminetetraacetic acid (EDTA) causes deleterious effects to root canal dentine¹¹. High concentrations of NaOCI are irritating when in contact with periapical tissues^{12,13} and have a pronounced negative effect on the survival and differentiation of apical papilla stem cells, which can hinder pulp regeneration/revascularization¹⁴. The research for alternative irrigating solutions is focused on substances that have an antimicrobial effect, of organic dissolution, as well as biocompatibility.

Calcium hypochlorite [Ca(OCI)₂], a chlorine disinfectant¹⁵, has been proposed as endodontic irrigant. It is available as granules powder and when prepared in aqueous solution, there is a release of hypochlorous acid and calcium hydroxide⁴: Ca(OCI)₂ + 2 H₂O \rightarrow 2HOCI + Ca(OH)₂. Compared with sodium hypochlorite (NaOCI + H₂O \rightarrow HOCI + NaOH), there is a higher generation of hypochlorous acid¹⁶. The generated calcium hydroxide [Ca(OH)₂] could favour the antimicrobial activity of Ca(OCI)₂ ¹⁶. In addition, calcium present in its composition, instead of sodium¹⁷, could be associated to a possible osteogenic induction and subsequent mineralization in periapical tissues. The Ca(OCI)₂ solutions are highly alkaline (pH around 11-12), and higher surface tension than NaOCI¹⁸. Ca(OCI)₂ solution had a stable FAC up to 30 days, which is comparable to NaOCI stabilized with sodium hydroxide^{18,19}. Studies have been performed assessing the Ca(OCI)₂ potential for tissue dissolution^{4,20}, efficacy against *Enterococcus faecalis*²⁰⁻²², effects on mechanical properties of root canal dentine^{23,24}, on composite resin microleakage¹⁷, and cytotoxicity²⁵⁻²⁸.

When used as irrigating solution in extracted teeth contaminated with *E. faecalis*, 2.5% Ca(OCI)₂ showed antimicrobial efficacy similar to 2.5% NaOCI²²,

regardless of the use of passive ultrasonic irrigation (PUI)²¹ or sonic activation using Vibringe system²⁹. Another study showed that 2.5% Ca(OCI)₂ was more effective against *E. faecalis* than 2.5% NaOCI when used during chemo-mechanical preparation with Reciproc R40 file³⁰. The 5% Ca(OCI)₂ has the same organic dissolution capacity as 5.25% NaOCI after 35 and 60 minutes of contact with the tissue⁴. This organic dissolution capacity gradually increases with time and with an increase in its concentration³¹. On the other hand, Ca(OCI)₂, does not have the ability to dissolve inorganic tissue and consequently does not remove the smear layer¹⁶.

Regarding the effects on root canal dentine surface, Ca(OCI)₂ changes dentine roughness in a similar manner to NaOCI²³. When used before an acetone-based adhesive system, Ca(OCI)₂ does not affect microleakage compared to NaOCI. However, the contact of Ca(OCI)₂ on dentine increases the amount of calcium and phosphorus ions, which can be beneficial for the mineralization process and for the formation of an amorphous calcium phosphate phase within the hybrid layer during bonding procedures¹⁷. Study showed that 6% NaOCI negatively affects the flexural strength, ultimate tensile strength and fracture resistance of dentine, while 6% Ca(OCI)₂ does not alter those properties²⁴.

Regarding Ca(OCI)₂ cytotoxicity, studies that used methyl-thiazole-tetrazolium - MTT^{26-28} or trypan blue and scratch assay²⁵ do not show consensus. A study found no difference between Ca(OCI)₂ at a high concentration (5%) and NaOCI at a low concentration (0.5%) in L929 fibroblasts, that is, Ca(OCI)₂ had more cytocompatibility than NaOCl²⁷, which was corroborated by a study that used both solutions at the same concentration, 2.5%²⁶. Another study that used 3T3 cells revealed that both Ca(OCl)₂ and NaOCl promoted similar cytotoxicity at 3h and 6h; however, at 24h, Ca(OCl)₂ promoted higher cell cytotoxicity²⁸. On the other hand, no differences were founded between Ca(OCl)₂ and NaOCl in 3T3 fibroblasts²⁵.

In teeth with pulp necrosis, microorganisms can penetrate areas that are difficult to clean mechanically such as isthmus, ramifications, lateral or accessory canals, apical deltas and dentinal tubules³². It has been reported that the biofilm of *E. faecalis* and *Porphyromonas gingivalis* can invade dentinal tubules up to 500 μ m deep³³. Thus, the depth that irrigants penetrate into dentinal tubules is an important factor in endodontic treatment³⁴. The penetration depth of NaOCI into dentinal tubules can be affected by concentration^{33,34}, contact time, temperature^{34,35}, agitation, use of gel form ³⁶ and surface tension of the irrigant^{35,37}.

Inside root canal, a high surface tension can hinder the penetration of the irrigating solution into dentinal tubules, regions of isthmus and anatomical irregularities, reducing its antibacterial effectiveness³⁸. It has been reported that a decreased surface tension can increase the penetration of the irrigating solution into inaccessible areas of the root canal system including dentinal tubules, improving the disinfection^{35,37,38}. Substances that reduce the surface tension of irrigants, which are called surfactants, including cetrimide (CTR), benzalkonium chloride (BAK), triton X-100 (TR100) and tween 80 - T80 has been associated to irrigating solutions^{36,39–42}.

CTR is a cationic surfactant that significantly reduces the surface tension of NaOCl³⁸ and has antimicrobial activity⁴³. Studies have shown that 0.2% CTR eradicated *E. faecalis* biofilm⁴³, improved the antimicrobial effect of 2% chlorhexidine against polymicrobial biofilm⁴² and did not affect the 2.5% NaOCl activity against *E. faecalis* biofilm⁴⁴.

BAK is a cationic surfactant widely used in the medical field as a preservative for eye solutions⁴⁵ that has been mixed with NaOCI. The addition of 0.008% BAK reduced the contact angle and surface free energy, and had no effect on FAC, cytotoxicity and antimicrobial effectiveness of 2.4% NaOCI⁴⁶. The mixture of 0.008% BAK with 6% NaOCI was more effective in eliminating *E. faecalis* from the root canal compared to 6% NaOCI alone⁴⁷.

T80 is a nonionic surfactant present in the composition of Biopure MTAD (Dentsply Sirona Endodontics, York, PA, USA)⁴⁸ and TR100, another nonionic surfactant, is present in Chlor-Xtra (Vista Dental Products, Racine, WI, USA)^{49,50}; in both cases manufacturers do not disclose their concentrations. A previous study shows that ChlorXtra has lower surface tension than NaOCI without surfactant³⁸. Another study revelead that the addition of 0.1% BAK, TR100 and T80 in 5% NaOCI reduced surface tension and did not change the pH and FAC in comparison to 5% NaOCI³⁹.

There is no consensus in the literature about the impact of surface tension on penetration of irrigating solutions into dentinal tubules^{35–37}. Two studies showed that NaOCI solutions with surfactant had higher penetration into dentinal tubules^{35,37}, while another study showed that this addition had no effect on the penetration of NaOCI³⁶. According to available literature, Ca(OCI)₂ had a higher surface tension than NaOCI at the same concentrations - 0.5%, 1%, 2.5% and 5.25%¹⁸. However, it is not known if Ca(OCI)₂ penetrates into dentinal tubules less than NaOCI. In

addition, it is not known if the addition of surfactants would decrease the surface tension and increase the penetration of Ca(OCI)₂ into dentinal tubules, without changing the pH and FAC.

For the selection of irrigating solution for root canal treatment it must be considered not only its antimicrobial effectiveness and organic dissolution capacity, but also its possible cytotoxic effects since it may come in contact with periapical tissues⁵¹, which may influence the prognosis of root canal treatment. This becomes even more critical in regenerative endodontics procedures, which are performed in immature teeth. This is because the irrigating solution also contacts the periapical tissues, which are essential for endodontic regeneration⁵². It is also necessary to consider the physicochemical properties of irrigating solutions such as pH, surface tension, FAC, as well as the ability to penetrate areas not touched by the instruments, which are fundamental factors for the disinfection of the root canal system.

As previously mentioned, the control of infection is necessary to promote the healing of periapical tissues affected by apical periodontitis⁵³. However, disinfection may be challenging when bacteria are organized in multispecies matrix-enclosed communities, called biofilms, especially in teeth with complex anatomies. These bacterial structures can colonize the canal walls⁵⁴, ramifications and isthmuses³².

To improve irrigation effectiveness, technologies such as ultrasonic activation have been used. Specifically, passive ultrasonic irrigation (PUI) improves the removal of debris, smear layer^{55,56}, and bacterial biofilm⁵⁷ from root canals. However, even with the use of ultrasonic activation associated with NaOCI, microorganisms, debris and even pulp tissue may remain in the isthmus, apical third⁵⁸, oval/flattened canals⁵⁹ and root canal curvatures⁶⁰. Another aspect to be considered is that if a small apical enlargement is performed during conventional chemical-mechanical preparation, a large amount of bacterial biofilm and necrotic tissue may remain in the root canal⁶¹.

The GentleWave (GW), a new system that combines multisonic and negative apical pressure (Sonendo Inc, Laguna Hills, CA, USA), was introduced on the US market in 2014, and represents a type of endodontic device developed for cleaning and disinfection of the root canal⁶². According to its manufacturer, GW can be used in situations that need only minimal instrumentation, instead of using conventional instrumentation⁶³.

GW uses high-speed fluid dynamics to deliver the irrigants into the root canal system without requiring the tip of the instrument to enter the root canals. The irrigant is delivered through a handpiece to the end of a nozzle placed in the sealed pulp⁶⁴, and the excess of irrigant is simultaneously removed from the chamber by the built-in vented suction through the handpiece into a waste canister inside the console⁶⁵. GW creates a powerful shear force, which causes hydrodynamic cavitation in the form of a cavitation cloud. The implosion of thousands of microbubbles creates an acoustic field of broadband frequencies that travels through the procedure fluid into the entire root canal system⁶⁶.

Regarding microbial reduction, a study using real-time PCR and bacterial cultures in molars, revealed that GW promoted higher reduction of total microbial DNA than CUI⁶⁷. In anterior teeth, GW promoted less bacterial reduction than PUI, as previously shown using next generation sequencing (NGS)⁶⁸.

Traditionally, culture methods have been used to assess the bacterial composition and decontamination of the root canal system ⁶⁹. This method allows a semi- or absolute quantification of culturable bacteria. However, a significant amount of microorganisms in the root canal space cannot be cultured under laboratory conditions. The development of The Human Genome Project ⁷⁰ allowed the subsequent development of databases (i.e., SILVA) for use in conjunction with NGS technologies ^{71,72}. NGS is a fifth-generation laboratory tool of microbiological analysis for the study of endodontic infections. This method provides vast information about bacterial communities and their profiles^{53,73}. To date, the decontamination efficacy of infected root canals irrigated with GW in molars has not been provided using a relevant infection model.

Studies have been conducted to assess other types of GW effects in endodontic treatment. The GW promoted a significantly fast rate of bovine muscle dissolution than conventional syringe irrigation (CSI), continuous ultrasonic irrigation (CUI) and negative-pressure irrigation⁶². The root canal treatment using GW was not associated with extrusion⁶⁵ since GW creates a negative pressure at the apical foramen, irrespective of canal instrumentation size⁶⁴. GW removed more debris than CSI in minimally and conventionally instrumented canals⁷⁴; as well as more debris than CUI, but no more than PUI in conventionally instrumented canals⁷⁵. Additionally, GW showed a greater removal of calcium hydroxide paste in comparison to CSI and PUI, in the root canals submitted to conventional instrumentation⁷⁶. A prospective

multicenter clinical study showed that the treatment of teeth with large periapical lesions with GW resulted in a 97.7% success rate after 12-month re-evaluation⁶⁶. The association of GW with 3% NaOCI without root canal instrumentation cleaned the organic matter (tissue remnants) and dentin debris even in irregular areas, especially in middle and apical thirds of premolars⁷⁷. Regarding the penetration of NaOCI into dentinal tubules, the use of GW promoted greater penetration of 3% NaOCI than PUI and CUI⁷⁸.

Currently, GW costs approximately \$80,000.00 per console, and \$50.00 to \$100.00 for a one-time use handpiece. However, several doubts have been raised in regard to GW: Is it worth investing in such high-cost equipment? Does it produce better results than conventional root canal treatment? What are the effects of GW on endodontic treatment? Has GW biofilm removal efficacy been evaluated using a relevant model?

4 CONCLUSIONS

Part 1:

-The addition of surfactants to Ca(OCI)₂ did not increase the penetration into dentinal tubules, but it did promote lower surface tension, without changing the pH or free available chlorine values, and higher availability of free calcium ions in Ca(OCI)₂ (Publication 1).

-Although Ca(OCI)₂ and NaOCI promoted the same cytotoxicity mechanism and did not stimulate ALP activity, Ca(OCI)₂ was less cytotoxic than NaOCI (Publication 2).

Part 2:

-Further research, mainly clinical, is needed to establish whether GentleWave has any advantages over other available irrigation methods (Publication 3).

-There was no difference between GentleWave and Passive Ultrasonic Irrigation (PUI) in the reduction of multispecies biofilm; bacterial reduction in mesial roots of mandibular molars prepared to 35.04 with PUI was similar to those prepared to 20.06 with a GentleWave (Publication 4).

REFERENCES*

- Rôças IN, Provenzano JC, Neves MAS, Siqueira JF. Disinfecting effects of rotary instrumentation with either 2.5% sodium hypochlorite or 2% chlorhexidine as the main irrigant: a randomized clinical study. J Endod. 2016; 42(6): 943–7.
- 2. Haapasalo M, Shen Y, Wang Z, Gao Y. Irrigation in endodontics. Br Dent J. 2014; 216(6): 299–303.
- del Carpio-Perochena A, Bramante CM, de Andrade FB, Maliza AGA, Cavenago BC, Marciano MA et al. Antibacterial and dissolution ability of sodium hypochlorite in different pHs on multi-species biofilms. Clin Oral Investig. 2015; 19(8): 2067–73.
- 4. Dutta A, Saunders WP. Comparative evaluation of calcium hypochlorite and sodium hypochlorite on soft-tissue dissolution. J Endod. 2012; 38(10): 1395–8.
- 5. Zehnder M. Root canal irrigants. J Endod. 2006; 32(5): 389–98.
- 6. Garcia F, Murray PE, Garcia-Godoy F, Namerow KN. Effect of aquatine endodontic cleanser on smear layer removal in the root canals of ex vivo human teeth. J Appl Oral Sci. 2010; 18(4): 403–8.
- Pascon FM, Kantovitz KR, Sacramento PA, Nobre-dos-Santos M, Puppin-Rontani RM. Effect of sodium hypochlorite on dentine mechanical properties. A review. J Dent. 2009; 37(12): 903–8.
- Neelakantan P, Sharma S, Shemesh H, Wesselink PR. Influence of irrigation sequence on the Adhesion of root canal sealers to dentin: a Fourier transform infrared spectroscopy and push-out bond strength analysis. J Endod. 2015; 41(7): 1108–11.
- Martinho FC, Carvalho CAT, Oliveira LD, Farias De Lacerda AJ, Xavier ACC, Gullo Augusto M et al. Comparison of different dentin pretreatment protocols on the bond strength of glass fiber post using self-etching adhesive. J Endod. 2015; 41(1): 83–7.
- Coaguila-Llerena H, Stefanini da Silva V, Tanomaru-Filho M, Guerreiro Tanomaru JM, Faria G. Cleaning capacity of octenidine as root canal irrigant: A scanning electron microscopy study. Microsc Res Tech. 2018; 81(6): 523–7.
- Tartari T, Bachmann L, Zancan RF, Vivan RR, Duarte MAH, Bramante CM. Analysis of the effects of several decalcifying agents alone and in combination with sodium hypochlorite on the chemical composition of dentine. Int Endod J. 2018; 51: e42–54.
- Farook SA, Shah V, Lenouvel D, Sheikh O, Sadiq Z, Cascarini L. Guidelines for management of sodium hypochlorite extrusion injuries. Br Dent J. 2014; 217(12): 679–84.

^{*} According to FOAr Academic Papers Guide, adapted from Vancouver Standards. Available on Library website: <u>http://www.foar.unesp.br/Home/Biblioteca/guia-de-normalizacao-atualizado.pdf</u>

- Coaguila-Llerena H, Denegri-Hacking A, Lucano-Tinoco L, Mendiola-Aquino C, Faria G. Accidental extrusion of sodium hypochlorite in a patient taking alendronate: a case report with an 8-year follow-up. J Endod. 2021; 47(12): 1947–52.
- 14. Martin DE, De Almeida JFA, Henry MA, Khaing ZZ, Schmidt CE, Teixeira FB et al. Concentration-dependent effect of sodium hypochlorite on stem cells of apical papilla survival and differentiation. J Endod. 2014; 40(1): 51–5.
- 15. Tyan K, Kang J, Jin K, Kyle AM. Evaluation of the antimicrobial efficacy and skin safety of a novel color additive in combination with chlorine disinfectants. Am J Infect Control. 2018; 46(11): 1254–61.
- 16. Görduysus M, Küçükkaya S, Bayramgil NP, Görduysus MÖ. Evaluation of the effects of two novel irrigants on intraradicular dentine erosion, debris and smear layer removal. Restor Dent Endod. 2015; 40(3): 216–22.
- Ferreira MB de C, Carlini Júnior B, Galafassi D, Gobbi DL. Calcium hypochlorite as a dentin deproteinization agent: microleakage, scanning electron microscopy and elemental analysis. Microsc Res Tech. 2015; 78(8): 676–81.
- Leonardo NGES, Carlotto IB, Luisi SB, Kopper PMP, Grecca FS, Montagner F. Calcium hypochlorite solutions: evaluation of surface tension and effect of different storage conditions and time periods over pH and available chlorine content. J Endod. 2016; 42(4): 641–5.
- Iqbal Q, Lubeck-Schricker M, Wells E, Wolfe MK, Lantagne D. Shelf-life of chlorine solutions recommended in Ebola virus disease response. PLoS One. 2016; 11(5): 1–12.
- 20. De Paula KB, Carlotto IB, Marconi DF, Ferreira MBC, Grecca FS, Montagner F. Calcium hypochlorite solutions an in vitro evaluation of antimicrobial action and pulp dissolution. Eur Endod J. 2019; 4(1): 15–20.
- De Almeida AP, Souza MA, Miyagaki DC, Dal Bello Y, Cecchin D, Farina AP. Comparative evaluation of calcium hypochlorite and sodium hypochlorite associated with passive ultrasonic irrigation on antimicrobial activity of a root canal system infected with Enterococcus faecalis: an in vitro study. J Endod. 2014; 40(12): 1953–7.
- 22. Dal Bello Y, Mezzalira GI, Jaguszewski LA, Hoffmann IP, Menchik VHS, Cecchin D et al. Effectiveness of calcium and sodium hypochlorite in association with reciprocating instrumentation on decontamination of root canals infected with Enterococcus faecalis. Aust Endod J. 2019; 45(1): 92–7.
- 23. Oliveira JS, Neto WR, De Faria NS, Fernandes FS, Miranda CES, Rached-Junior FJA. Quantitative assessment of root canal roughness with calciumbased hypochlorite Irrigants by 3D CLSM. Braz Dent J. 2014; 25(5): 409–15.
- Cecchin D, Soares Giaretta V, Granella Cadorin B, Albino Souza M, Vidal C de MP, Paula Farina A. Effect of synthetic and natural-derived novel endodontic irrigant solutions on mechanical properties of human dentin. J Mater Sci Mater Med. 2017; 28(9): 141.

- 25. Blattes GBF, Mestieri LB, Böttcher DE, Fossati ACM, Montagner F, Grecca FS. Cell migration, viability and tissue reaction of calcium hypochlorite based-solutions irrigants: An in vitro and in vivo study. Arch Oral Biol. 2017; 73: 34–9.
- 26. Coaguila-Llerena H, Rodrigues EM, Tanomaru-Filho M, Guerreiro-Tanomaru JM, Faria G. Effects of calcium hypochlorite and octenidine hydrochloride on L929 and human periodontal ligament cells. Braz Dent J. 2019; 30(3): 213–9.
- 27. Sedigh-Shams M, Gholami A, Abbaszadegan A, Yazdanparast R, Nejad MS, Safari A et al. Antimicrobial efficacy and cytocompatibility of calcium hypochlorite solution as a root canal irrigant: An in Vitro investigation. Iran Endod J. 2016; 11(3): 169–74.
- 28. Yilmaz Ş, Yoldas O, Dumani A, Guler G, Ilgaz S, Akbal E et al. Calcium hypochlorite on mouse embryonic fibroblast cells (NIH3T3) in vitro cytotoxicity and genotoxicity: MTT and comet assay. Mol Biol Rep. 2020; 47(7): 5377–83.
- 29. Dumani A, Guvenmez HK, Yilmaz S, Yoldas O, Kurklu ZGB. Antibacterial efficacy of calcium hypochlorite with vibringe sonic irrigation system on Enterococcus faecalis: an in vitro study. Biomed Res Int. 2016; 2016: 8076131.
- Souza MA, Tumelero Dias C, Zandoná J, Paim Hoffmann I, Sanches Menchik VH, Palhano HS et al. Antimicrobial activity of hypochlorite solutions and reciprocating instrumentation associated with photodynamic therapy on root canals infected with Enterococcus faecalis – An in vitro study. Photodiagnosis Photodyn Ther. 2018; 23(July): 347–52.
- Taneja S, Kumari M, Anand S. Effect of QMix, peracetic acid and ethylenediaminetetraacetic acid on calcium loss and microhardness of root dentine. J Conserv Dent. 2014; 17(2): 155–8.
- Ricucci D, Siqueira JF. Biofilms and apical periodontitis: study of prevalence and association with clinical and histopathologic findings. J Endod. 2010; 36(8): 1277–88.
- 33. Wong DTS, Cheung GSP. Extension of bactericidal effect of sodium hypochlorite into dentinal tubules. J Endod. 2014; 40(6): 825–9.
- 34. Zou L, Shen Y, Li W, Haapasalo M. Penetration of sodium hypochlorite into dentin. J Endod. 2010; 36(5): 793–6.
- 35. Palazzi F, Blasi A, Mohammadi Z, Del Fabbro M, Estrela C. Penetration of sodium hypochlorite modified with surfactants into root canal dentin. Braz Dent J. 2016; 27(2): 208–16.
- Faria G, Viola KS, Coaguila-Llerena H, Oliveira LRA, Leonardo RT, Aranda-García AJ et al. Penetration of sodium hypochlorite into root canal dentine: effect of surfactants, gel form and passive ultrasonic irrigation. Int Endod J. 2019; 52(3): 385–92.
- Coaguila-Llerena H, Barbieri I, Tanomaru-Filho M, Leonardo R de T, Ramos AP, Faria G. Physicochemical properties, cytotoxicity and penetration into dentinal tubules of sodium hypochlorite with and without surfactants. Restor Dent Endod. 2020; 45(4): e47.
- 38. Palazzi F, Morra M, Mohammadi Z, Grandini S, Giardino L. Comparison of the surface tension of 5.25% sodium hypochlorite solution with three new sodium hypochlorite-based endodontic irrigants. Int Endod J. 2012; 45(2): 129–35.

- 39. Guneser MB, Arslan D, Dincer AN, Er G. Effect of sodium hypochlorite irrigation with or without surfactants on the bond strength of an epoxy-based sealer to dentin. Clin Oral Investig. 2017; 21(4): 1259–65.
- Dragan O, Tomuta I, Casoni D, Sarbu C, Campian R, Frentiu T. Influence of mixed additives on the physicochemical properties of a 5.25% sodium hypochlorite solution: an unsupervised multivariate statistical approach. J Endod. 2018; 44(2): 280-285.e3.
- 41. Iglesias JE, Pinheiro LS, Weibel DE, Montagner F, Grecca FS. Influence of surfactants addition on the properties of calcium hypochlorite solutions. J Appl Oral Sci. 2019; 27: e20180157.
- 42. Ruiz-Linares M, Aguado-Pérez B, Baca P, Arias-Moliz MT, Ferrer-Luque CM. Efficacy of antimicrobial solutions against polymicrobial root canal biofilm. Int Endod J. 2017; 50(1): 77–83.
- 43. Baca P, Junco P, Arias-Moliz MT, González-Rodríguez MP, Ferrer-Luque CM. Residual and antimicrobial activity of final irrigation protocols on enterococcus faecalis biofilm in dentin. J Endod. 2011; 37(3): 363–6.
- Guerreiro-Tanomaru JM, Nascimento CA, Faria-Júnior NB, Graeff MSZ, Watanabe E, Tanomaru-Filho M. Antibiofilm activity of irrigating solutions associated with cetrimide. Confocal laser scanning microscopy. Int Endod J. 2014; 47(11): 1058–63.
- 45. Lee W, Lee S, Bae HW, Kim CY, Seong GJ. Efficacy and tolerability of preservative-free 0.0015% tafluprost in glaucoma patients: a prospective crossover study. BMC Ophthalmol. 2017; 17(1): 61.
- 46. Bukiet F, Couderc G, Camps J, Tassery H, Cuisinier F, About I et al. Wetting properties and critical micellar concentration of benzalkonium chloride mixed in sodium hypochlorite. J Endod. 2012; 38(11): 1525–9.
- 47. Baron A, Lindsey K, Sidow SJ, Dickinson D, Chuang A, McPherson JC. Effect of a benzalkonium chloride surfactant-sodium hypochlorite combination on elimination of Enterococcus faecalis. J Endod. 2016; 42(1): 145–9.
- 48. Singla MG, Garg A, Gupta S. MTAD in endodontics: an update review. Oral Surg Oral Med Oral Pathol Oral Radiol Endod. 2011; 112(3): e70–6.
- 49. Bukiet F, Soler T, Guivarch M, Camps J, Tassery H, Cuisinier F et al. Factors affecting the viscosity of sodium hypochlorite and their effect on irrigant flow. Int Endod J. 2013; 46(10): 954–61.
- 50. Giardino L, Mohammadi Z, Beltrami R, Poggio C, Estrela C, Generali L. Influence of temperature on the antibacterial activity of sodium hypochlorite. Braz Dent J. 2016; 27(1): 32–6.
- Yasuda Y, Tatematsu Y, Fujii S, Maeda H, Akamine A, Torabinejad M et al. Effect of MTAD on the differentiation of osteoblast-like cells. J Endod. 2010; 36(2): 260–3.
- Sabrah AHA, Yassen GH, Liu WC, Goebel WS, Gregory RL, Platt JA. The effect of diluted triple and double antibiotic pastes on dental pulp stem cells and established Enterococcus faecalis biofilm. Clin Oral Investig. 2015; 19(8): 2059–66.

- 53. Siqueira JF, Rôças IN. Present status and future directions: microbiology of endodontic infections. Int Endod J. 2022; 55(S3): 512–30.
- 54. Manoharan L, Brundin M, Rakhimova O, de Paz LC, Vestman NR. New insights into the microbial profiles of infected root canals in traumatized teeth. J Clin Med. 2020; 9(12): 3877.
- 55. De-Deus G, Belladonna FG, de Siqueira Zuolo A, Perez R, Carvalho MS, Souza EM et al. Micro-CT comparison of XP-endo Finisher and passive ultrasonic irrigation as final irrigation protocols on the removal of accumulated hard-tissue debris from oval shaped-canals. Clin Oral Investig. 2019; 23(7): 3087–93.
- 56. Urban K, Donnermeyer D, Schäfer E, Bürklein S. Canal cleanliness using different irrigation activation systems: a SEM evaluation. Clin Oral Investig. 2017; 21(9): 2681–7.
- 57. Ordinola-Zapata R, Bramante CM, Aprecio RM, Handysides R, Jaramillo DE. Biofilm removal by 6% sodium hypochlorite activated by different irrigation techniques. Int Endod J. 2014; 47(7): 659–66.
- 58. Neelakantan P, Devaraj S, Jagannathan N. Histologic assessment of debridement of the root canal isthmus of mandibular molars by irrigant activation techniques ex vivo. J Endod. 2016; 42(8): 1268–72.
- 59. Alves FRF, Almeida BM, Neves MAS, Moreno JO, Rôças IN, Siqueira JF. Disinfecting oval-shaped root canals: effectiveness of different supplementary approaches. J Endod. 2011; 37(4): 496–501.
- Neuhaus KW, Liebi M, Stauffacher S, Eick S, Lussi A. Antibacterial efficacy of a new sonic irrigation device for root canal disinfection. J Endod. 2016; 42(12): 1799–803.
- 61. Siqueira Junior JF, Rôças I das N, Marceliano-Alves MF, Pérez AR, Ricucci D. Unprepared root canal surface areas: causes, clinical implications, and therapeutic strategies. Braz Oral Res. 2018; 32(suppl 1): e65.
- 62. Haapasalo M, Wang Z, Shen Y, Curtis A, Patel P, Khakpour M. Tissue dissolution by a novel Multisonic Ultracleaning system and sodium hypochlorite. J Endod. 2014; 40(8): 1178–81.
- 63. GentleWave Datasheet, Sonendo, Inc., Laguna Hills, CA,USA. URL: https://www.sonendo.com (accessed on 01 February 2022).
- 64. Haapasalo M, Shen Y, Wang Z, Park E, Curtis A, Patel P et al. Apical pressure created during irrigation with the GentleWave[™] system compared to conventional syringe irrigation. Clin Oral Investig. 2016; 20(7): 1525–34.
- 65. Charara K, Friedman S, Sherman A, Kishen A, Malkhassian G, Khakpour M et al. Assessment of apical extrusion during root canal irrigation with the novel GentleWave system in a simulated apical environment. J Endod. 2016; 42(1): 135–9.
- 66. Sigurdsson A, Garland RW, Le KT, Rassoulian SA. Healing of periapical lesions after endodontic treatment with the GentleWave procedure: a prospective multicenter clinical study. J Endod. 2018; 44(3): 510–7.

- Zhang D, Shen Y, de la Fuente-Núñez C, Haapasalo M. In vitro evaluation by quantitative real-time PCR and culturing of the effectiveness of disinfection of multispecies biofilms in root canals by two irrigation systems. Clin Oral Investig. 2019; 23(2): 913–20.
- 68. Ordinola-Zapata R, Mansour D, Saavedra F, Staley C, Chen R, Fok AS. In vitro efficacy of a non-instrumentation technique to remove intracanal multispecies biofilm. Int Endod J. 2022; 55(5): 495–504.
- Siqueira JF, Rôças IN. A critical analysis of research methods and experimental models to study the root canal microbiome. Int Endod J. 2022; 55(S1): 46–71.
- 70. Venter JC, Adams MD, Myers EW, Li PW, Mural RJ, Sutton GG et al. The sequence of the human genome. Science (80-). 2001; 291(5507): 1304–51.
- 71. van Dijk EL, Jaszczyszyn Y, Naquin D, Thermes C. The third revolution in sequencing technology. Trends Genet. 2018; 34(9): 666–81.
- 72. Zhong Y, Xu F, Wu J, Schubert J, Li MM. Application of Next Generation Sequencing in laboratory medicine. Ann Lab Med. 2021; 41(1): 25–43.
- 73. Manoil D, Al-Manei K, Belibasakis GN. A systematic review of the root canal microbiota associated with apical periodontitis: lessons from Next-Generation Sequencing. Proteomics Clin Appl. 2020; 14(3): e1900060.
- Molina B, Glickman G, Vandrangi P, Khakpour M. Evaluation of root canal debridement of human molars using the GentleWave System. J Endod. 2015; 41(10): 1701–5.
- 75. Chan R, Versiani MA, Friedman S, Malkhassian G, Sousa-Neto MD, Leoni GB et al. Efficacy of 3 supplementary irrigation protocols in the removal of hard tissue debris from the mesial root canal system of mandibular molars. J Endod. 2019; 45(7): 923–9.
- Ma J, Shen Y, Yang Y, Gao Y, Wan P, Gan Y et al. In vitro study of calcium hydroxide removal from mandibular molar root canals. J Endod. 2015; 41(4): 553–8.
- 77. Wang Z, Shen Y, Haapasalo M. Root canal wall dentin structure in uninstrumented but cleaned human premolars: a scanning electron microscopic study. J Endod. 2018; 44(5): 842–8.
- 78. Vandrangi P. Evaluating penetration depth of treatment fluids into dentinal tubules using the GentleWave system. Dentistry. 2016; 06(03): 3–7.