



UNIVERSIDADE ESTADUAL PAULISTA
"JÚLIO DE MESQUITA FILHO"



**Programa de Pós-Graduação em Odontologia
Área de Estomatologia
FACULDADE DE ODONTOLOGIA DE ARAÇATUBA – UNESP**

TAMARA FERNANDES DE CASTRO

**CD4⁺, CD8⁺ AND FOXP3⁺ LYMPHOCYTES INFILTRATING ORAL
LEUKOPLAKIA: AN IMMUNOHISTOCHEMICAL ANALYSIS**

DISSERTAÇÃO DE MESTRADO

Araçatuba – SP

2020

TAMARA FERNANDES DE CASTRO

**CD4⁺, CD8⁺ and FOXP3⁺ lymphocytes infiltrating oral
leukoplakia: an immunohistochemical analysis**

Dissertação apresentada à Faculdade de Odontologia de Araçatuba, Universidade Estadual Paulista “Júlio de Mesquita Filho”, para obtenção do título de Mestre, pelo Programa de Pós-Graduação em Odontologia, área de concentração em Estomatologia.

Orientador: Prof. Tit. Glauco Issamu Miyahara

Co-orientador: Profa. Tit. Sandra Helena Penha de Oliveira

Araçatuba – SP

2020

Catálogo na Publicação (CIP)

Diretoria Técnica de Biblioteca e Documentação – FOA / UNESP

Castro, Tamara Fernandes de.

C355c CD4⁺, CD8⁺ and FOXP3⁺ lymphocytes infiltrating oral
leukoplakia: an immunohistochemical analysis / Tamara
Fernandes de Castro. – Araçatuba, 2020
40 f. : il.

Dissertação (Mestrado) – Universidade Estadual
Paulista, Faculdade de Odontologia, Araçatuba
Orientador: Prof. Glauco Issamu Miyahara
Coorientadora: Profa. Sandra Helena Penha de Oliveira

1. Leucoplasia 2. Linfócitos T 3. Imuno-histoquímica
I. Título

Black D6

CDD 617.63

Claudio Hideo Matsumoto CRB-8/5550

Dedicatória

Dedico este trabalho...

Aos meu pais, **Sebastião Soares de Castro** e **Maria Petronilha Fernandes de Castro** que nunca mediram esforços para meus sonhos se concretizarem,

Aos meus irmãos **Fausto Fernandes de Castro** e **Murilo Fernandes da Silva** por acreditarem em mim,

Aos meus avós, **Sunta Maria Fernandes** e **Mario Fernandes** pelo amor incondicional.

Agradecimento especiais

Ao meu orientador e atual diretor **Prof. Tit. Glauco Issamu Miyahara** pela confiança em mim empregada desde o meu egresso na universidade, pelas oportunidades proporcionadas, pelas orientações e conselhos. Você professor é um exemplo de competência, dedicação e profissionalismo.

À minha co-orientadora **Profa. Adj. Sandra Helena Penha de Oliveira** agradeço pela parceria com nossa equipe, você é um exemplo de dedicação, um exemplo de docente a ser seguido.

Ao professor **Prof. Assoc. Marcelo Macedo Crivelini**, pela paciência, ao manusear o microscópio, compreensão, em entender minhas limitações e disponibilidade, por sempre me encaixar em sua agenda. O senhor me estimulou com todo seu esforço a ser uma profissional melhor. Agradeço a dedicação que você tem pelo seu trabalho.

Ao **Prof. Ass. Dr. Daniel Galera Bernabé** pela dedicação, competência e todo conhecimento compartilhado. Eu admiro muito seu trabalho, você possui uma visão crítica e oportuna que amplia nosso campo de visão.

À **Profa. Ass. Dra. Leda Maria Pescinini Salzedas** pela oportunidade de trabalhar em conjunto no PAADES-B na disciplina de radiologia. A troca de experiências, sua didática e seu conceito sobre a área foi enriquecedor.

Ao **Prof. Adj. Éder Ricardo Biasoli**, muito obrigada pelos incentivos, pela disponibilidade, e saiba que sem sua prática clínica minha formação jamais seria a mesma.

Agradecimentos

A **Deus** pela dádiva da vida e por permitir que eu realizasse tanto sonhos nesta existência. Obrigada por me permitir errar, aprender, crescer e não desistir.

Ao meu pai **Sebastião Soares de Castro**, obrigada por me apoiar em cada decisão em minha vida. Obrigada por me fazer acreditar que a educação transforma as pessoas e que com determinação e coragem realizamos sonhos.

A minha mãe **Maria Petronilha Fernandes de Castro**, obrigada por tanto amor. Obrigada por cada palavra de estimulo-lo, por cada sorriso saudoso. Obrigada por esse jeito doce de ser.

Ao meu irmão, admirável e amigo **Fausto Fernandes de Castro**, meu muito obrigada. Você é um exemplo de docênte, de lealdade, de cumplicidade, de amor. Saiba que sem você essa vida não teria o menor sentido.

Ao meu irmão **Murilo Fernandes de Silva**, meu eterno amigo e cumplice. É com você as melhores histórias, os melhores momentos, saiba que você é essencial.

À minha amiga **Jéssica Araujo Figueira**, eu sou extremamente grata pela sua amizade. Seus conselhos e suas orientações foi de extrema valia para meu crescimento tanto pessoal quanto intelectual, sou sua fã.

Ao meu querido amigo **Saygo Tomo**, obrigada pelo convívio diário, pelas orientações, pelos trabalhos e apoios constantes. Saiba que você foi crucial para realização deste sonho e que sem você nada disso seria possível.

Ao meu namorado **Murilo Henrique Delamura Corrêa** por todo carinho, compreensão e apoio em tantos momentos difíceis desta caminhada. Obrigado pelo presente de cada dia, pelo seu sorriso e por saber me fazer feliz.

À **Giseli Mitsury Kayahara**, sou extremamente grata pelos ensinamentos do laboratório, pela paciência e compreensão. Você tem um coração enorme.

À minha IC, **Isabela Cerato** agradeço o apoio e motivação incondicional que ajudou a tornar este trabalho uma válida e agradável experiência de aprendizagem.

À minha amiga **Isabella Zanutto**, saiba que nem mesmo a distância é capaz de separar o eterno carinho que trago de ti.

Aos amigos da pós-graduação, **Bruna Amélia Serafim, Vítor Bonetti Valente, Daniela Brito Bastos, Daniela Cantielli, Ana Daniela Spinolla, Gabriela**

Lopes e Maria Clara Botelho obrigada por deixar meus dias mais felizes e a rotina mais leve.

Ao Departamento de Patologia e Propedêutica Clínica, representado **pela Prof.^a Ana Claudia Okamoto e Prof^a. Cristiane Furuse. Aos docentes, Prof.^a Ana Maria Soubhia, Prof.^a Leda Maria Salzedas e Prof.^a Renata Callestini Felipini**, gostaria de agradecer pela recepção no departamento, pela acessibilidade que tenho com todos. Aos funcionários, **Adriana de Paula e Robson Varlei Ranieri** por toda gentileza e auxílio prestado.

A toda equipe e quadro de funcionários do Centro de Oncologia Bucal, Unidade Auxiliar de Estrutura Simples, da Faculdade de Odontologia do Câmpus de Araçatuba – UNESP, **Jane Fátima Mendes Fernandes da Silva, Anne Cristina de Faria Cocato, Daniene Tesoni Cassavara Ribeiro, Regiane Mazzariolli Pereira Nogueira e Gabrielle Dias Duarte**, pelo convívio diário, pelas risadas e pelo acolhimento desde o meu egresso.

A equipe do laboratório de Endodontia, ao **Prof. Assoc. Luciano Tavares Angelo Cintra** e especialmente a mestranda **Cristiane Cantiga da Silva** por toda ajuda e disponibilidade. Cris você não mediu esforços para a realização das fotomicrografias do experimento, sou extremamente grata.

Às funcionárias da Seção de Pós-Graduação da FOA - UNESP, **Valéria Zagatto, Cristiane Lui e Lilian Mada** por toda a disponibilidade e toda a paciência.

Ao Programa de Pós-Graduação em Odontologia da Faculdade de Odontologia de Araçatuba – UNESP, na pessoa do coordenador, **Prof. Adj. André Luiz Fraga Briso**.

À Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES), pela bolsa de Demanda Social (DS) concedida a mim, que foi de extrema importância para minha manutenção durante o curso de mestrado.

À Fundação de Amparo à Pesquisa do Estado de São Paulo (FAPESP), pelo auxílio de pesquisa regular concedido à nossa equipe (Processo: 2016/12982-0), sem o qual a realização deste trabalho não seria possível.

Por fim, o meu profundo e sentido agradecimento a todas as pessoas que contribuíram para a concretização desta dissertação, estimulando-me intelectual e emocionalmente.

Epígrafe

*“Quando você sonha alto todos os passos parecem ser o primeiro
Então esse é só o primeiro passo mais uma vez
E a gente tem muito pra aprender”*

José Tiago Sabino Pereira

Projota

Castro TF. Linfócitos CD4⁺, CD8⁺ e FOXP3⁺ que se infiltram na leucoplasia oral: uma análise imunohistoquímica. [Dissertação]- Araçatuba: UNESP-Universidade Estadual Paulista; 2019.

Resumo

Objetivo: Avaliar a infiltração dos linfócitos CD4⁺, CD8⁺ e FOXP3⁺ e sua correlação com características sociodemográfica, clinicopatológicas e estilo de vida de pacientes com leucoplasias bucais. **Pacientes e métodos:** Oitenta pacientes com diagnóstico de leucoplasia bucal foram incluídos no estudo. Análises retrospectivas foram realizadas para verificar as características sociodemográficas, clinicopatológicas e estilo de vida dos pacientes. O infiltrado linfocitário foi caracterizado por imunohistoquímica com antígenos contra de CD4⁺, CD8⁺ e FOXP3⁺. **Resultados:** Dos 80 pacientes incluídos neste estudo, (60%) eram homens e a idade variou de 25 a 82 anos com idade média de 58,6 anos. Trinta e oito (47.5%) eram idosos, Trinta e dois (40%) eram adultos de meia idade e apenas dez (10%) adultos jovens. Sessenta e um dos pacientes eram fumantes (76.2%) e quarenta e seis eram etilistas (57.5%). Vinte e sete (35.5%) das lesões apresentaram algum grau de displasia epitelial. O grau de displasia epitelial apresentou correlação positiva com a intensidade do consumo do álcool ($p=0.008$). Houve correlação positiva entre os linfócitos CD4⁺ e CD8⁺ ($p=0.005$). **Conclusão:** O infiltrado linfocitário não foi relacionado com nenhuma característica clinicopatológica das leucoplasias bucais. Entretanto, o grau de displasia está relacionado ao estilo de vida dos pacientes.

Palavras-chave: Leucoplasia, Linfócito-T, Imunohistoquímica.

Castro TF. CD4⁺, CD8⁺ and FOXP3⁺ lymphocytes infiltrating oral leukoplakia: an immunohistochemical analysis. [dissertation]- Araçatuba: UNESP- São Paulo State University; 2019.

Abstract

Objective: To evaluate the infiltration of CD4⁺, CD8⁺ and FOXP3⁺ lymphocytes and their correlation with sociodemographic, clinicopathological and lifestyle characteristics of patients with oral leukoplakia. **Patients and methods:** Eighty patients diagnosed with oral leukoplakia were included in the study. Retrospective analyses were performed in order to verify the sociodemographic, clinicopathologic and lifestyle characteristics. The lymphocytic infiltrate characterization was performed by immunohistochemistry with antibodies against CD4⁺, CD8⁺, and FOXP3⁺ markers. **Results:** Of 80 patients included in the study, 60% were men, and their age ranged from 25 to 82 years, with a mean of 58.6. Thirty-eight patients (47.5%) were elderly, Thirty-two (40%) middle-aged, and only ten (10%) young adults. Sixty-one of the patients were smokers (76.2%) and forty-six were alcoholics (57.5%). Twenty-seven (35.5%) of the lesions presented some degree of dysplasia. The degree of epithelial dysplasia was correlated with the intensity of alcohol consumption ($p=0.008$). A positive correlation was found between CD4⁺ and CD8⁺ lymphocytes ($p=0.005$). **Conclusion:** The lymphocytic infiltrate of oral leukoplakia was not correlated with any clinicopathologic characteristic. However, the degree of epithelial dysplasia was correlated with the lifestyle of the patients.

Keywords: Leukoplakia; T-Lymphocytes; Immunohistochemistry

LIST OF FIGURES

Figure 1. The scores of CD4⁺, CD8⁺, and FOXP3⁺ in OL were detected by immunohistochemistry (400×). (A-C) The CD4⁺ scores. (D-F) The CD8⁺ scores. (G-I) The FOXP3⁺ scores. Arrows: indication of CD4/CD8/FOXP3 immunostaining.

LIST OF TABLES

Table 1. Sociodemographic, clinicopathologic and lifestyle characteristics, and CD4⁺, CD8⁺ and FOXP3⁺ score results of patients with oral leukoplakia.

Table 2. The correlation results between clinicopathologic and lifestyle characteristics and CD4⁺, CD8⁺ and FOXP3⁺ scores of patients with oral leukoplakia.

LIST OF ABBREVIATIONS

ED, Epithelial dysplasia

FOXP3, Forkhead Transcription Factor

LI, Lymphocytic Infiltrate

OPML, Oral potentially malignant lesions

OL, Oral leukoplakia

OSCC, Oral squamous cell carcinoma

T regs, Regulatory T cells

UNESP, Universidade Estadual Paulista Júlio de Mesquita Filho

SUMMARY

Introduction.....	18
Patients and Methods.....	20
Results	24
Discussion.....	29
References.....	34
Annex A.....	39
Annex B.....	40

CD4⁺, CD8⁺ and FOXP3⁺ lymphocytes infiltrating oral leukoplakia: an immunohistochemical analysis

Lymphocytic infiltrates in oral leukoplakia.

Authors: Tamara Fernandes de Castro¹; Saygo Tomo¹; Éder Ricardo Biasoli¹; Daniel Galera Bernabé¹; Marcelo Macedo Crivelini²; Sandra Helena Penha de Oliveira¹; Glauco Issamu Miyahara¹

¹Oral Oncology Center, São Paulo State University (Unesp), School of Dentistry, Araçatuba, São Paulo, Brazil.

²Department of Diagnostic and Surgery, São Paulo State University (Unesp), School of Dentistry, Araçatuba, São Paulo, Brazil.

Keywords:

Leukoplakia; T-Lymphocytes; Immunohistochemistry

Corresponding author:

miyahara@foa.unesp.br

Glauco Issamu Miyahara, DDS, PhD.

Oral Oncology Center, São Paulo State University (Unesp), School of Dentistry, 1193 José Bonifácio St, Araçatuba, SP 15050-015, + 55 18 36363275.

***Formatado de acordo com as normas do periódico Oral Diseases (ISSN: 1601-0825) (Anexo I).**

Introduction

Oral cancer is a global public health problem, responsible for 177,384 deaths worldwide in 2018, corresponding to 1.9 deaths per 100,000 inhabitants (Bray et al., 2018). Oral squamous cell carcinoma (OSCC) is the most common malignant disease of the head and neck, accounting for >90% of all malignancies in the mouth. It is one of the most common tumors worldwide, with approximately 300,000 incident cases each year (Ferlay et al., 2015; Stasikowska-Kanicka, Wągrowaska-Danilewicz, Danilewicz, 2018). In 2022, 11,180 new cases of oral cavity cancer in men and 4,010 new cases in women were estimated in Brazil (Inca, 2020). Despite therapeutic advances, 5-year survival remains below 50% in the last 50 years, the main reason being the late diagnosis (Mehrotra, Gupta, Singh, Ibrahim, 2006; Ganly, Patel, Shah, 2012; Singla, Singla, Zaheer, Rawat, Mandal, 2018). Thus, the early diagnosis of oral cancer plays a key role within the disease progression, treatment outcome, patient's quality of life and survival (Mehrotra et al., 2006; Ganly et al., 2012; Liu et al., 2013).

OSCC may develop from oral potentially malignant lesions (OPML), which are lesions with high risk of malignant evolution compared to other benign lesions and the normal oral mucosa (Mehrotra et al., 2006; Ganly et al., 2012; Liu et al., 2013), among which oral leukoplakia (OL) is the most frequent (Liu et al., 2010; Mehrotra et al., 2006; Liu et al., 2013; Bisht, Singh, Sikarwar, Darbari, 2013). According to the World Health Organization (WHO), OL is a white lesion that cannot be clinically or pathologically associated with any other disease (Warnakulasuriya, Johnson, Van der Waal, 2007; Liu et al., 2013; Bisht et al.,

2013). Clinicopathologic characteristics as advanced age, female gender, lesions larger than 2cm, non-homogeneous OL and the presence of epithelial dysplasia (ED), reflect a higher risk to malignant progression (Warnakulasuriya, Ariyawardana, 2016; Speight, Khurram, Kujan, 2018). Moreover, the severity of OL is proportional to the degree of ED (Stasikowska-Kanicka et al., 2018; Lodi, Sardella, Bez, Demarosi, Carrassi, 2006). From all the ED that OL may present, about 3-17% turn into OSCC (Liu et al., 2013; Tilakaratne, Sherriff, Morgan, Odell, 2011). The histological exam is mandatory for the diagnosis of OL and to assess the risk of malignant transformation of OL, which is based on the degree of ED observed (Lodi et al., 2006; Warnakulasuriya et al., 2007; Van Der Waal, 2014; Warnakulasuriya et al., 2016).

Recent studies show that the amount and type of lymphocytic infiltrate (LI) are related to the progression and survival of OSCC patients (Yagyu et al., 2017; Stasikowska-Kanicka et al., 2018; Kouketsu et al., 2019).

LI is related to the antitumor response and changes in LI quantity and type are associated with the prognosis and survival of patients with colorectal, ovarian and breast cancer (Zhang et al., 2003). The higher amounts of CD8⁺ cytotoxic T-cells infiltrating tumors has been associated with a better prognosis in several cancers (Chen et al., 2019), as it may be associated with a protective microenvironment (Chaves et al., 2019), whereas LI regulating T cells (Tregs) may promote tumor progression and high Tregs levels are linked to immunosuppression and poor prognosis (Liang et al., 2011; Sun et al., 2016; Stasikowska-Kanicka et al., 2018; Singla et al., 2018). This occurs, at least in part, because Treg cells play a crucial role in suppressing the anti-tumor immune system (Sakaguchi, 2010; Oleinika, 2013; Johnson et al., 2014), in which

Forkhead Transcription Factor (FOXP3) is a transcription factor and serves as the most specific marker for Treg CD4⁺CD25⁺FOXP3⁺ in human cancers and is involved in the development and function of Tregs cells (Sakaguchi, Miyara, Costantino, Hafler, 2010; Liang et al., 2011; Oleinika, Nibbs, Graham, Fraser, 2013; Sun et al., 2016; Stasikowska-Kanicka et al., 2017).

To understand the immunopathogenesis of OPML is essential for cancer research. This study aimed to assess the CD4⁺, CD8⁺ and FOXP3⁺ lymphocytes infiltrating OL by the immunohistochemistry method and evaluate their correlation with sociodemographic, clinicopathological and lifestyle characteristics of the patients.

Patients and Methods

Patients

This retrospective study included patients diagnosed with OL between 2011 and 2018 at the Oral Oncology Center, São Paulo State University (UNESP), School of Dentistry, Araçatuba, Brazil. This study was approved by the Research Ethics Committee in human studies (Protocol nº. 03876012.6.0000.5420). The inclusion criteria were patients with clinically and microscopically confirmed diagnosis of primary OL, and who agreed to participate in the study. The exclusion criteria were patients who received previous treatment for OL, or did not accept to participate in the study, and those diagnosed with non-homogeneous leukoplakia. A total of 80 patients were included. The corresponding

sociodemographic, clinicopathologic and lifestyle information was retrieved from the patients' medical records.

Clinicopathologic data collection

Sociodemographic, clinicopathologic, and lifestyle data of the patients were obtained from their individual records. The sociodemographic variables included age, sex, and race. The clinicopathologic variables included location, present or absent epithelial dysplasia (ED) and ED degree of the lesion, which was considered according to the grading proposed by the WHO (Warnakulasuriya et al., 2008) performed by a single pathologist for all cases involved in this study. Lifestyle factors included the status and exposure intensity to tobacco and alcohol. For analysis, the status of tobacco smoking and alcohol drinking were classified as current smoker/drinker, never smoker/drinker; and the intensity of these habits was classified as light, moderate and severe. The average number of cigarette equivalents smoked per day was categorically defined as light (≤ 19), moderate (20–39), and severe exposure intensity (≥ 40). Measurements of alcohol consumption were graded as light (0-2 drinks/day), moderate (3-4 drinks/day), and severe exposure intensity (>4 drinks/day) (Rapoport A, Kowalski, 1989).

Immunohistochemistry

So as to evaluate the presence of CD4⁺ and CD8⁺ lymphocytes in the OL, immunohistochemical staining was performed on 4µm tissue sections. The sections were deparaffinized and rehydrated. Epitope retrieval was performed by incubation in 10mM citrate buffer (pH= 6.0) for 20min at 55°C using a steamer. Incubations of 20 minutes with 3% hydrogen peroxide were carried out in order to block endogenous peroxidase activity. The primary antibodies anti-CD4 (clone EPR6855; Abcam, UK; 1:1500 dilution) and anti-CD8 (clone EP1150Y; Abcam, UK; 1:1500 dilution) were incubated for 3 hours. The sections were then incubated with the secondary antibody (Histofine MULTI; Nichirei Biosciences INC., JP) for 30 minutes. Immunostaining was performed immersing the sections in 3,3'-diaminobenzidine (DAB substrate kit; DAKO, USA) and subsequently were counterstained with Harris's hematoxylin, dehydrated, cleared and mounted.

In order to evaluate the presence of FOXP3⁺ lymphocytes, immunohistochemical staining was performed on 4µm tissue sections. They were then treated with 3% hydrogen peroxide in methanol for 30min at room temperature so as to block endogenous peroxidases. The sections were submitted to an antigenic recovery with 3 heating treatments (100°C) of 4 min, with intervals of 1 min, with sodium citrate buffer (pH 6.0) in a microwave oven (760W power). In order to block non-specific binding proteins, the sections were incubated in PBS (phosphate-buffered saline) containing 2% bovine serum albumin, for 30 min, at room temperature. The sections were then incubated with primary antibodies for FOXP3⁺ (clone 236A/E7; Abcam, UK; dilution 1:500), at 4°C, overnight. The sections were then incubated with the secondary antibody (Histofine MULTI; Nichirei Biosciences INC., JP) for 30 minutes. Immunostaining was performed

immersing the sections in 3,3'-diaminobenzidine (DAB substrate kit; DAKO, USA), which were subsequently counterstained with Harris's hematoxylin, dehydrated, cleared and mounted. Similarly produced sections of the tonsil were used as a positive control for all antibodies. The positive controls for the immunohistochemical staining were prepared using the same specific antibody concentration. The negative controls were produced by replacing the primary antibodies with the antibody diluent.

All samples inserted in parafin used for diagnostic purposes were deposited in the Oral Medicine Biobank, São Paulo State University (UNESP), School of Dentistry, Araçatuba, Brazil.

Morphometry

One examiner expert on microscopic observation and blinded to the ED information's and the clinical history of the patients assessed the immunohistochemical staining results. T cells were counted manually by means of the ImageJ Java 1.52a software (National Institutes of Health, Rockville, MD; <http://imagej.net/ImageJ>) in 7-10 high-magnification fields (400x), starting from the center of the histopathological section to 5 right fields and 5 left fields. Subepithelial CD4⁺, CD8⁺, and FOXP3⁺ cells were counted in the lamina propria between the interpapillary ridges and subepithelial areas. The average count was calculated and graded as 1+, 2+, or 3+, i.e., ≤5, 6-19, or ≥20 T cells for each lesion, respectively. (Zhang et al., 2003; Yagyu et al., 2017).

Statistical analysis

Statistical analysis was performed using GraphPad Prism® version 8.0 (GraphPad Software Inc, La Jolla, CA). Frequency distributions were provided for the clinicopathologic variables. For the correlation of clinical and pathological characteristics and CD4⁺, CD8⁺, and FOXP3⁺ scores, Spearman's rank correlation coefficient test was used. P-values < 0.05 were considered statistically significant.

Results

Patients' characteristics

The characteristics of the patients included in this study are summarized in Table 1. The study included 80 patients, 48 (60%) were men and 32 (40%) were women. The mean age was 58.6 years at initial diagnosis. Thirty-eight (47.5%) individuals were elderly (>60-years old), thirty-two (40%) middle-aged (45-59-years old) and ten (12.5%) young-adults (<45 years old). Fifty-seven patients were white (71.3%), while twenty-three (28.7%) were non-white. The OL most common involved site was buccal mucosa (n = 21; 26.2%).

Forty-nine (64.5%) patients did not exhibit ED, twenty-seven (35.5%) patients showed ED and from four patients these could not be obtained. Low, moderate, and severe dysplasia were identified in sixteen (59.3%), nine (33.3%), and two (7.4%) cases, respectively.

Regarding lifestyle, most patients were current smokers, amounting to sixty-one (76.3%), and nineteen (23.7%) nonsmokers. Thirty patients were considered light smokers (50.8%), eighteen (30.6%) moderate smokers, and only eleven (18.6%) heavy smokers. Forty-six (57.5%) patients with OL were current alcohol drinkers and thirty-four (42.5%) never drinkers. In respect to alcohol consumption, twenty patients were regarded as light drinkers (55.5%), thirteen (36.2%) moderate drinkers, and three (8.3%) heavy drinkers.

Seventy patients (87.4%) showed score 2 and 3 and only ten patients (12.5%) presented score 1 for CD4⁺ immunoreactivity. Fourteen (31.2%), twenty-seven (60%) and four patients (8.8%) exhibited 1, 2 and 3 scores, respectively, for CD8⁺ immunoreactivity. Thirty-two (71.2%), eleven (24.4%) and two patients (4.4%) displayed 1, 2 and 3 scores, respectively, for FOXP3⁺ immunoreactivity.

Table 1. Sociodemographic, clinicopathologic and lifestyle characteristics, and CD4+, CD8+ and FOXP3+ score results of patients with oral leukoplakia.

Variable	N	%
Sex		
Male	48	60.0%
Female	32	40.0%
Age		
Young	10	12.5%
Middle age	32	40.0%
Elderly	38	47.5%
Race		
White	57	71.3%
Non-white	23	28.7%
Location		
Tongue	17	21.2%
Gingiva	12	15.0%
Buccal mucosa	21	26.2%
Palate	12	15.0%
Lip	11	13.8%
Floor of mouth	7	8.8%
ED		
Present	27	35.5%
Absent	49	64.5%
No date	4	-
Degree of ED		
Low	16	59.3%
Moderate	9	33.3%
Severe	2	7.4%
Tobacco smoking status		
Smoker	61	76.3%
Nonsmoker	19	23.7%
Tobacco smoking intensity		
Light	30	50.8%
Moderate	18	30.6%
Heavy	11	18.6%
No date	2	-
Alcohol drinking status		
Drinker	46	57.5%
Nondrinker	34	42.5%
Alcohol drinking intensity		
Light	20	55.5%
Moderate	13	36.2%
Heavy	3	8.3%
No date	10	-
Score CD4		
1	10	12.5%
2	35	43.7%
3	35	43.7%
Score CD8		
1	14	31.2%
2	27	60.0%
3	4	8.8%
Score FOXP3		
1	32	71.2%
2	11	24.4%
3	2	4.4%
TOTAL	80	100%

ED: Epithelial dysplasia;

The correlative study revealed statistically significant correlations between the CD4⁺ and CD8⁺ scores of infiltrating cells. What is more, it was found a significant correlation between ED and alcohol drinking intensity (Table 2). Regarding dysplasia, tobacco smoking intensity, tobacco smoking status, and alcohol drinking status, no correlation was found between clinicopathologic and lifestyle characteristics and CD4⁺, CD8⁺ and FOXP3⁺ scores (Table 2).

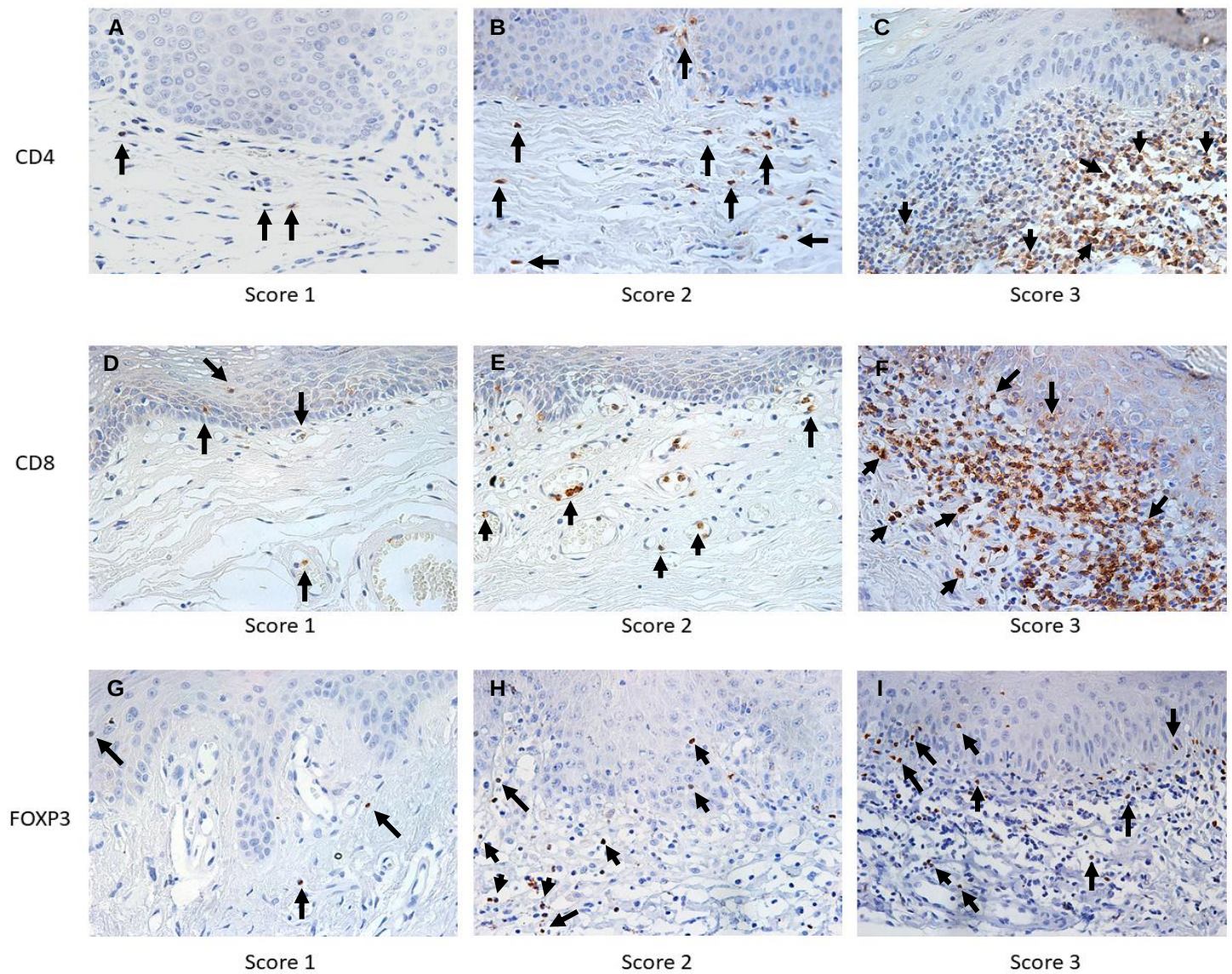
Table 2. The correlation results between clinicopathologic and lifestyle characteristics and CD4⁺, CD8⁺ and FOXP3⁺ scores of patients with oral leukoplakia.

Correlation between	R	p-value	CI
ED vs Tobacco smoking intensity	-0.129	0.330	-0.379 to 0.139
ED vs Tobacco smoking status	0.026	0.821	-0.201 to 0.250
ED vs Alcohol drinking intensity	0.035	0.838	-0.305 to 0.368
ED vs Alcohol drinking status	0.079	0.487	-0.149 to 0.299
ED vs CD4 score	-0.050	0.658	-0.273 to 0.177
ED vs CD8 score	-0.187	0.096	-0.396 to 0.040
ED vs FOXP3 score	-0.035	0.760	-0.258 to 0.192
Degree of ED vs Tobacco smoking intensity	0.017	0.945	-0.440 to 0.466
Degree of ED vs Tobacco smoking status	-0.033	0.871	-0.417 to 0.361
Degree of ED vs Alcohol drinking intensity	0.847	0.008*	0.516 to 0.957
Degree of ED vs Alcohol drinking status	-0.136	0.499	-0.499 to 0.268
Degree of ED dysplasia vs CD4 score	0.015	0.942	-0.377 to 0.402
Degree of ED vs CD8 score	-0.159	0.429	-0.516 to 0.246
Degree of ED vs FOXP3 score	-0.026	0.898	-0.411 to 0.367
Tobacco smoking intensity vs Tobacco smoking status	-0.122	0.356	-0.373 to 0.145
Tobacco smoking intensity vs Alcohol drinking intensity	0.136	0.472	-0.245 to 0.482
Tobacco smoking intensity vs Alcohol drinking status	-0.205	0.119	-0.444 to 0.06
Tobacco smoking intensity vs CD4 score	-0.067	0.614	-0.324 to 0.199
Tobacco smoking intensity vs CD8 score	0.082	0.537	-0.185 to 0.338
Tobacco smoking intensity vs FOXP3 score	0.199	0.131	-0.067 to 0.439
Tobacco smoking status vs alcoholism intensity	-0.149	0.385	-0.463 to 0.198
Tobacco smoking status vs Alcohol drinking status	0.411	1.45e-004	-0.204 to 0.583
Tobacco smoking status vs CD4 score	-0.035	0.759	-0.258 to 0.196
Tobacco smoking status vs CD8 score	0.008	0.947	-0.218 to 0.233
Tobacco smoking status vs FOXP3 score	-0.010	0.931	-0.235 to 0.216
Alcohol drinking intensity vs Alcohol drinking status	-0.082	0.634	-0.408 to 0.262
Alcohol drinking intensity vs CD4 score	-0.140	0.416	-0.455 to 0.207
Alcohol drinking intensity vs CD8 score	-0.104	0.548	-0.426 to 0.242
Alcohol drinking intensity vs FOXP3 score	0.067	0.698	-0.276 to 0.395
Alcohol drinking status vs CD4 score	-0.057	0.615	-0.279 to 0.171
Alcohol drinking status vs CD8 score	-0.005	0.964	-0.203 to 0.211
Alcohol drinking status vs FOXP3 score	-0.145	0.200	-0.359 to 0.083
CD4 score vs CD8 score	0.310	0.005*	0.090 to 0.500
CD4 score vs FOXP3 score	0.405	1904e-004	0.197 to 0.578
CD8 score vs FOXP3 score	0.199	0.076	-0.027 to 0.407

CI: confidence interval; ED: Epithelial dysplasia; r: correlation; *p<0.05

FOXP3⁺ was shown to be nuclear immunoreactive. CD4⁺ and CD8⁺ were clearly stained in the cell membrane of the infiltrating cells (Figure 1).

Figure 1. The scores of CD4⁺, CD8⁺, and FOXP3⁺ in OL were detected by immunohistochemistry (400×). (A-C) The CD4⁺ scores. (D-F) The CD8⁺ scores. (G-I) The FOXP3⁺ scores. Arrows: indication of CD4/CD8/FOXP3 immunostaining.



Discussion

Immunological factors have been associated with the development and progression of malignant and premalignant lesions. Therefore, it is imperative to investigate OPML antigenic repertoire, immunological environment, and immune modulation, and identify the regulators of its progression and regression, in order to provide information on the early stages of oncogenesis. Thus, understanding the immunopathogenesis of OL may provide new paths to OSCC immunotherapeutic strategies. In this retrospective study, we analyzed the sociodemographic, clinical-pathological and lifestyle data of patients with OL, and T-lymphocyte types that infiltrated the OL.

OL is the most common OPML and its risk of cancer progression ranges from 0.13 to 34% (Warnakulasuriya et al., 2016), and may depend on clinicopathologic characteristics as sex, age, location and etiology (Warnakulasuriya et al., 2016). According to Warnakulasuriya (2016), older female patients who have OL are more prone to develop malignancies than younger men. In our study, however, patients were older (47.5%) and men (60%). Moreover, the border of the tongue is known to have a higher risk for malignancy, especially from a pre-existing OL (Warnakulasuriya et al., 2016), which was not observed in our sample, since the most prevalent site of OL was the buccal mucosa (26.2%).

Not only is chronic tobacco smoking the main risk factor for OSCC but also for OL occurrence (Wu, Wang, Zhou, 2019). However, a higher risk for malignant progression of OL is attributed to non-smoker patients (Warnakulasuriya et al., 2007). In the present study, most patients were smokers

(76.2%) and drinkers (57.5%). These data, along with the location of most of the OLs included in this study may explain the low rate of ED observed in our study. However, it is important to notice that the ED degree increased with the severity of alcohol consumption.

A more intense LI was observed in OPML compared to OSCC (Johnson et al., 2014), suggesting that in the earlier stages of carcinogenesis, the cellular immune response is recruited to suppress the tumor progression. A hypothesis, still to be investigated, is that the successful malignant progression depends, at least in part, on the tumor cells in order to acquire the ability to escape the immunologic response. The CD4⁺ T-cell is known to be responsible for activating the cytotoxic T-cell CD8⁺ in the tumor microenvironment, reflecting a better outcome for patients with more intense infiltrates composed of these cells (Sato et al., 2005; Liang et al., 2011; Johnson et al., 2014; Stasikowska-Kanicka et al., 2018; Singla et al., 2018). On the other hand, the CD4⁺ FoxP3⁺ T-cells regulate the immune response by inactivating the CD8⁺ cytotoxic T-cells (Sun et al., 2016). Numerous studies suggest an association of high intratumoral CD8⁺ T-cell infiltration with a better clinical outcome, which may be related to a protective microenvironment in OPML and OSCC, corroborating the concept that reduced adaptive immunity facilitates malignant transformations and cancer metastases (Stasikowska-Kanicka et al., 2018; Chaves et al., 2019).

Stasikowska-Kanick et al., (2018) analyzed the amount of CD4⁺ and CD8⁺ lymphocytes in patients with OSCC metastasis (m+), OSCC without metastasis (m-), and OL with moderate to severe ED. A significant increase of intratumoral CD4⁺ cells in OSCC (m+) patients compared with patients with OSCC (m-) and OL was observed. On the other hand, an increase of the CD8⁺ LI was observed

in OL patients and OSCC patients (m-) in comparison with OSCC patients (m+). In the present study, we did not observe any correlation of the CD8⁺ LI with the degree of ED. However, the low rate of ED of our sample may have influenced this observation.

Furthermore, Chen et al. (2019) analyzed the amount of CD8⁺ cells in patients with OSCC and patients with OL. OSCC patients with infiltrating CD8⁺ cells have been shown to present better survival. Chaves et al. (2019) analyzed CD4⁺ and CD8⁺ cells in patients with OSCC, patients with OPML that transformed into OSCC, and patients with OPML that did not develop into OSCC. No significant differences in the CD4⁺ infiltrate among all groups was observed. However, reduced levels of CD8⁺ cells in OPML patients were found compared to OSCC patients and a significant increase in CD8⁺ cells was demonstrated in OPML that progressed to OSCC compared to OPML that did not progress into OSCC. In both studies, the CD8⁺ LI intensity was proportional to the degree of ED. Thus, the CD8⁺ IL present in the early OPML stages was considered low or nonexistent. Accordingly, no significant difference for CD8⁺ score was observed in our study, possibly due to the prevalent low degree of ED.

Sun et al. (2016) analyzed CD4⁺ lymphocyte status and the association with Foxp3 in OSCC, OL, and oral lichen planus patients. A gradual increase was shown in infiltrating CD4⁺ and FoxP3⁺ T-cells in parallel with the progression of ED in OL samples, which suggests that immune suppressor cells influence malignant tumor progression. Notwithstanding, Sun et al., (2016) did not present the location prevalence of OL, or the OL patients' habits, which may have influenced the results. In the present study, neither the CD4⁺ nor the FOXP3⁺ T

cells infiltrate were correlated with the degree of ED or any other clinicopathologic characteristic.

In our study, a correlation between CD4⁺ and CD8⁺ T cells was shown, which was expected from an adaptive immunological response (Johnson et al., 2014). However, no correlation between CD4⁺ T-cells and FoxP3⁺ was found. It is possible to assume that the number of altered cells was not sufficient to exacerbate the CD4⁺ immune response to such an extent to activate the FOXP3⁺ pathway, or the intensity of the CD4⁺ immune response was not sufficient to require FOXP3⁺ activation. This may have occurred due to the patients' profile in our sample, who did not show risk factors for malignant transformation. Therefore, long-term follow-up of these patients with OL, or the lack of a control group, either OSCC or healthy patients, are considered limitations of the present study. In conclusion, the lymphocyte inflammatory infiltrate, CD4⁺, CD8⁺, and FOXP3⁺, did not correlate with the available clinicopathological data; nonetheless, the intensity of alcohol consumption was correlated with the ED degree. Further investigations must be conducted to better understand the role of the LI on OL and its risk for malignancy. For that, the inclusion of a higher number of lesions with ED must be prioritized, long term follow-up data must be considered, and control groups of OSCC and healthy patients must be included. Moreover, epithelial antigens must be characterized in all the degrees of ED.

Conflict of interest statement

The authors declare that there is no potential conflict of interest regarding this work.

Founding

This work was funded by the Grant: 2016/12982-0 São Paulo Research Foundation (FAPESP).

References

Bray, F., Ferlay, J., Soerjomataram, I., Siegel, R.L., Torre, L.A., Jemal, A. (2018). Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin.* 68 (6):394-424. doi: 10.3322/caac.21492.

Bisht, R.S., Singh, A.K., Sikarwar, V., Darbari, A. (2013). Study over the clinical picture and histopathology of leukoplakia and to establish the correlation between causative factors in the patients of Garhwal hill region. *Natl J Maxillofac Surg.* 4 (2):177-180.

Chaves, A.L.F., Silva, A.G., Maia, F.M., Lopes, G.F.M., de Paulo, L.F.B., Muniz, L.V., Dos Santos, H.B., de Azambuja Ribeiro, R.I.M. (2019) Reduced CD8⁺ T cells infiltration can be associated to a malignant transformation in potentially malignant oral epithelial lesions. *Clin Oral Investig.* 23(4):1913-1919. DOI: 10.1007/s00784-018-2622-8.

Chen, X.J., Tanm Y.Q., Zhang, N., He, M.J., Zhou, G. (2019). Expression of programmed cell death-ligand 1 in oral squamous cell carcinoma and oral leukoplakia is associated with disease progression and CD8⁺ tumor-infiltrating lymphocytes. *Pathol Res Pract.* 215 (6): 152418. DOI: 10.1016 / j.prp.2019.04.010.

Ferlay, J., Soerjomataram, I., Dikshit, R., Eser, S., Mathers, C., Rebelo, M., Parkin, D.M., ... Bray, F. (2015). Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. *Int J Cancer.* 136(5):359-386.

Ganly, I., Patel, S., Shah, J. (2012). Early-stage squamous cell cancer of the oral tongue--clinicopathologic features affecting the outcome. *Cancer*. 1;118(1):101-11.

Estimativa 2019: incidência de câncer no Brasil / Instituto Nacional de Câncer José Alencar Gomes da Silva – Rio de Janeiro: INCA, 2020.

Johnson, S.D., De Costa, A-M.A., Young, M.R.I. (2014). Effect of the Premalignant and Tumor Microenvironment on Immune Cell Cytokine Production in Head and Neck Cancer. *Cancers*. 6(2):756–70.

Juretić, M., Cerović, R., Belušić-Gobić, M., Brekalo Pršo, I., Kqiku, L., Špalj, S., Pezelj-Ribarić, S. (2013). Salivary levels of TNF-alpha and IL-6 in patients with oral premalignant and malignant lesions. *Folia Biol*. 59, (2), 99-102.

Kouketsu, A., Sato, I., Oikawa, M., Shimizu, Y., Saito, H., Takahashi, T., Kumamoto, H. (2019). Expression of immunoregulatory molecules PD-L1 and PD-1 in oral cancer and precancerous lesions: A cohort study of Japanese patients. *J Craniomaxillofac Surg*. 47(1):33-40. DOI: 10.1016/j.jcms.2017.04.013.

Liu, W., Wang, Y.F., Zhou, H.W., Shi, P., Zhou, Z.T., Tang, G.Y. (2010) Malignant transformation of oral leukoplakia: a retrospective cohort study of 218 Chinese patients. *BMC Cancer*. 10(1):1-6.

Liu, W., Wu, L., Shen, X.M., Shi, L.J., Zhang, C.P., Xu, L.Q., Zhou, Z.T. (2013). Expression patterns of cancer stem cell markers ALDH1 and CD133 correlate with a high risk of malignant transformation of oral leukoplakia. *Int J Cancer*. 15;132(4):868-74.

Liang, Y.J., Liu, H.C., Su, Y.X., Zhang, T.H., Chu, M., Liang, L.Z., Liao, G.Q. (2011). Foxp3 expressed by tongue squamous cell carcinoma cells correlates with clinicopathologic features and overall survival in tongue squamous cell carcinoma patients. *Oral Oncol.* 47(7):566–570.

Laishram, D., Rao, K., Devi, H.S.U., Priya, N.S., Smitha, T., Sheethal, H.S. (2017). Mast cells and angiogenesis in malignant and premalignant oral lesions: An immunohistochemical study. *J Oral Maxillofac Pathol.* 21(2):229-238. DOI: 10.4103/jomfp.JOMFP_111_15.

Lodi, G., Sardella, A., Bez, C., Demarosi, F., Carrassi, A. Interventions for treating oral leukoplakia. *Cochrane Database Syst Rev.* 2006;18;(4):CD001829.doi:10.1002/14651858.CD001829.pub3

Mehrotra, R., Gupta, A., Singh, M., Ibrahim, R. (2006). Application of cytology and molecular biology in diagnosing premalignant or malignant oral lesions. *Mol Cancer*; 5:11. DOI:10.1186/1476-4598-5-11

Oleinika, K., Nibbs, R.J., Graham, G.J., Fraser, A.R. (2013). Suppression, subversion, and escape: the role of regulatory T cells in cancer progression. *Clin Exp Immunol.* 171(1):36-45.

Rapoport, A., Kowalski, L.P. O diagnóstico clínico em cabeça e pescoço. In: Brandão LC, Ferraz AR, eds. *Cirurgia de cabeça e pescoço. São Paulo: Roca*, 1989:5e10.

Sakaguchi, S., Miyara, M., Costantino, C.M., Hafler, D.A. (2010). FOXP3+ regulatory T cells in the human immune system. *Nat Rev Immunol.* 10(7):490-500.

Singla, S., Singla, G., Zaheer, S., Rawat, D.S., Mandal, A.K. (2018). Expression of p53, epidermal growth factor receptor, c-erbB2 in oral leukoplakias and oral squamous cell carcinomas. *J Cancer Res Ther.*14(2):388-393.

Stasikowska-Kanicka, O., Wągrowaska-Danilewicz, M., Danilewicz, M.(2017) Immunohistochemical Analysis of Foxp3⁺, CD4⁺, CD8⁺ Cell Infiltrates and PD-L1 in Oral Squamous Cell Carcinoma. *Pathol Oncol Res.* 24(3):497-505. DOI: 10.1007/s12253-017-0270-y.

Stasikowska-, O., Wągrowaska-Danilewicz, M., Danilewicz, M. (2018). T cells are involved in the induction of macrophage phenotypes in oral leukoplakia and squamous cell carcinoma-a preliminary report. *J Oral Pathol Med.* 47(2):136-143.

Speight, P.M., Khurram, S.A., Kujan, O. (2018). Oral potentially malignant disorders: risk of progression to malignancy. *Oral Surg Oral Med Oral Pathol Oral Radiol.* 125(6):612-627.

Sun, Y., Liu, N., Guan, X., Wu, H., Sun, Z., Zeng, H. (2016). Immunosuppression Induced by Chronic Inflammation and the Progression of Oral Squamous Cell Carcinoma. *Mediators Inflamm.* 2016:5715719. DOI: 10.1155/2016/5715719.

Tilakaratne, W.M., Sherriff, M., Morgan, P.R., Odell, E.W. (2011). Grading oral epithelial dysplasia: analysis of individual features. *J Oral Pathol Med.* 40(7):533-40.

Van der Waal, I. (2014). Oral potentially malignant disorders: is malignant transformation predictable and preventable? *Med Oral Patol Oral Cir Bucal.* 19 (4): e386-90. DOI:10.4317/medoral.20205

Warnakulasuriya, S., Johnson, N., Van der Waal, I. (2007). Nomenclature and classification of potentially malignant disorders of the oral mucosa. *J Oral Pathol Med.* 36(10):575-580.

Warnakulasuriya, S., Reibel, J., Bouquot, J., Dabelsteen, E. (2008). Oral epithelial dysplasia classification systems: predictive value, utility, weaknesses and scope for improvement. *J Oral Pathol Med.* 37 (3):127-133.

Warnakulasuriya, S., Ariyawardana, A. (2016). Malignant transformation of oral leukoplakia: a systematic review of observational studies. *J Oral Pathol Med.* 45(3):155-66.

Wu, W., Wang, Z., Zhou, Z. (2019). Risk factors associated with malignant transformation in patients with oral leukoplakia in a chinese population: a retrospective study. *J Oral Maxillofac Surg.* 9. pii: S0278-2391(19)30945-0. DOI: 10.1016/j.joms.2019.08.002.

Yagyuu, T., Hatakeyama, K., Imada, M., Kurihara, M., Matsusue, Y., Yamamoto, K., Obayashi, C., Kirita, T. (2017). Programmed death ligand 1 (PD-L1) expression and tumor microenvironment: Implications for patients with oral precancerous lesions. *Oral Oncol.* 68: 36-43. DOI: 10.1016 / j.oraloncology.2017.03.006.

Zhang, L., Conejo-Garcia, J.R., Katsaros, D., Gimotty, P.A., Massobrio, M., Regnani, G., Makrigiannakis, A., ... Coukos, G. (2003). Intratumoral T cells, recurrence, and survival in epithelial ovarian cancer. *N Engl J Med.* 16;348(3):203-13. doi:10.1056/NEJMoa020177

Anexo A

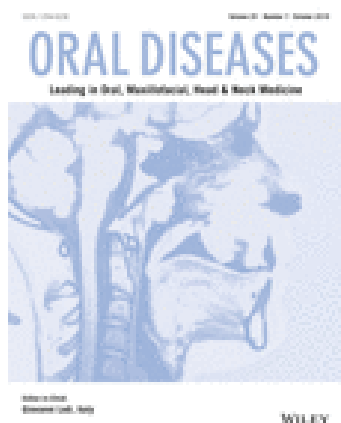
Parecer consubstanciado do CEP – FOA/UNESP

	UNIVERSIDADE ESTADUAL PAULISTA "JULIO DE MESQUITA FILHO" Campus de Araçatuba
COMITÊ DE ÉTICA EM PESQUISA	
 CERTIFICADO	
 Certificamos que o Projeto <i>"Detecção do HPV por nPCR em leucoplasias bucais: Estudo caso-controle"</i> , sob a responsabilidade do Pesquisador GLAUCO ISSAMU MIYAHARA , está de acordo com os Princípios Éticos em Pesquisa e foi aprovado em 27/05/2011, de acordo com o Processo FOA-01034/2011.	
Araçatuba, 06 de junho de 2011 ¹ .	
	
ALESSANDRA MARCONDES ARANEGA Vice-Coordenadora do CEP	
ANA/mbr.	Faculdade de Odontologia e Curso de Medicina Veterinária Rua José Bonifácio, 1193 CEP 16015-050 Araçatuba - SP Tel (18) 3636-3234 E-mail: cep@foa.unesp.br

Anexo – B

Periódico de interesse para submissão

Oral Diseases



Edited By: Giovanni Lodi

Impact factor: 2.625

ISI Journal Citation Reports © Ranking: 2018:17/91

Online ISSN:1601-0825

Qualis CAPES: A1.