

RESSALVA

Atendendo solicitação do(a)
autor(a), o texto completo desta tese
será disponibilizado somente a partir
de 04/12/2026.

**UNIVERSIDADE ESTADUAL PAULISTA (UNESP)
FACULDADE DE CIÊNCIAS AGRÁRIAS E VETERINÁRIAS
CÂMPUS DE JABOTICABAL**

**GENOTYPIC VARIABILITY OF COWPEA (*Vigna Unguiculata*
(L.) Walp) AND SELENIUM UPTAKE FOR DETERMINING
AGRONOMIC BIOFORTIFICATION STRATEGIES TO
COMBAT HIDDEN HUNGER**

M.Sc. MARIA GABRIELA DANTAS BERETA LANZA

Biosystems Engineer, Master in Agronomy

**UNIVERSIDADE ESTADUAL PAULISTA (UNESP)
FACULDADE DE CIÊNCIAS AGRÁRIAS E VETERINÁRIAS
CÂMPUS DE JABOTICABAL**

**GENOTYPIC VARIABILITY OF COWPEA (*Vigna unguiculata*
(L.) Walp) AND SELENIUM UPTAKE FOR DETERMINING
AGRONOMIC BIOFORTIFICATION STRATEGIES TO
COMBAT HIDDEN HUNGER**

M.Sc. Maria Gabriela Dantas Bereta Lanza

Supervisor: Prof. Ph.D. André Rodrigues dos Reis

Thesis presented to the School of Agricultural and
Veterinary Studies (FCAV) – UNESP as part of the
requirements to obtain the title of Doctor in
Agronomy (Crop Production).

2024

L297g

Lanza, Maria Gabriela Dantas Bereta

Genotypic Variability Of Cowpea (*Vigna Unguiculata* (L.) Walp)
And Selenium Uptake For Determining Agronomic Biofortification
Strategies To Combat Hidden Hunger / Maria Gabriela Dantas Bereta
Lanza. -- Jaboticabal, 2025
278 p.

Tese (doutorado) - Universidade Estadual Paulista (UNESP),
Faculdade de Ciências Agrárias e Veterinárias, Jaboticabal
Orientador: André Rodrigues dos Reis

1. Nutrição de Plantas. 2. Fisiologia Vegetal. 3. Feijão-caupi. 4.
Marcha de absorção. 5. Selênio. I. Título.

CERTIFICADO DE APROVAÇÃO

TÍTULO DA TESE: GENOTYPIC VARIABILITY OF COWPEA (*Vigna unguiculata* (L.) Walp) AND SELENIUM UPTAKE FOR DETERMINING AGRONOMIC BIOFORTIFICATION STRATEGIES TO COMBAT HIDDEN HUNGER

AUTORA: MARIA GABRIELA DANTAS BERETA LANZA

ORIENTADOR: ANDRÉ RODRIGUES DOS REIS

Aprovada como parte das exigências para obtenção do Título de Doutora em Agronomia (Produção Vegetal), pela Comissão Examinadora:

Prof. Dr. ANDRÉ RODRIGUES DOS REIS (Participação Virtual)
Departamento de Engenharia de Biosistemas / FCE UNESP Tupa

Documento assinado digitalmente
 **ANDRÉ RODRIGUES DOS REIS**
Data: 19/12/2024 20:27:36-0300
Verifique em <https://validar.iti.gov.br>

Prof. Dr. JOSÉ LAVRES JUNIOR (Participação Virtual)
Laboratório de Isótopos Estáveis / CENA USP Piracicaba/SP

Documento assinado digitalmente
 **JOSE LAVRES JUNIOR**
Data: 20/12/2024 14:57:11-0300
Verifique em <https://validar.iti.gov.br>

Prof. Dr. ELCIO FERREIRA DOS SANTOS (Participação Virtual)
Departamento de Agronomia / Instituto Federal de Mato Grosso do Sul (IFMS) - Nova Andradina/MS

Documento assinado digitalmente
 **ELCIO FERREIRA DOS SANTOS**
Data: 20/12/2024 17:24:46-0300
Verifique em <https://validar.iti.gov.br>

Prof. Dr. LEANDRO BORGES LEMOS (Participação Virtual)
Departamento de Produção Vegetal / FCAV UNESP Jaboticabal

Documento assinado digitalmente
 **LEANDRO BORGES LEMOS**
Data: 20/12/2024 09:53:07-0300
Verifique em <https://validar.iti.gov.br>

Pesquisador. Dr. MAURISRAEL DE MOURA ROCHA (Participação Virtual)
Centro de Pesquisa Agropecuária do Meio-Norte / EMBRAPA - Teresina/PI



Documento assinado digitalmente

MAURISRAEL DE MOURA ROCHA

Data: 20/12/2024 07:23:34-0300

Verifique em <https://validar.iti.gov.br>

Jaboticabal, 04 de dezembro de 2024

DADOS CURRICULARES DO AUTOR

MARIA GABRIELA DANTAS BERETA LANZA - Nascida no município de Tupã, São Paulo, no dia 6 de março de 1997. Em 2020, graduou-se em Engenharia de Biossistemas pela Universidade Estadual Paulista (UNESP), Câmpus de Tupã. Durante a graduação foi bolsista de iniciação científica da Fundação de Amparo à Pesquisa do Estado de São Paulo (Processos nº 2015/26817-9 e 2018/15490-7). Como bolsista FAPESP, conquistou a Bolsa Estágio de Pesquisa no Exterior (BEPE) na University of Nottingham (Inglaterra) em 2019. Em março de 2020, ingressou no Mestrado do Programa de Pós-Graduação em Agronomia (Produção Vegetal), da Universidade Estadual Paulista, Câmpus Jaboticabal, sob a orientação do Prof. Dr. André Rodrigues dos Reis e co-orientação do Prof. Dr. Hudson Wallace Pereira de Carvalho do Centro de Energia Nuclear na Agricultura da Universidade de São Paulo (ESALQ, USP). No ano de 2021 foi aprovada no Doutorado do Programa de Pós-Graduação em Agronomia (Produção Vegetal), UNESP, Câmpus Jaboticabal. Em 2018, foi co-autora do artigo **(1)** *“Physiological, biochemical, and ultrastructural characterization of selenium toxicity in cowpea plants”* publicado na Environmental And Experimental Botany (FI = 4.5). Em 2020 foi co-autora dos artigos **(2)** *“Selenium toxicity stress-induced phenotypical, biochemical and physiological responses in rice plants”* e **(3)** *“Selenium toxicity in upland field-grown rice”* publicados na Ecotoxicology And Environmental Safety (FI = 6.2). Em 2021 publicou **(4)** *“Assessment of selenium spatial distribution using μ -XFR in cowpea plants”* na Ecotoxicology And Environmental Safety (FI = 6.2) e **(5)** *“Roles of selenium in mineral plant nutrition: ROS scavenging responses against abiotic stresses”* na Plant Physiology And Biochemistry (FI = 6.1), o qual foi indicado na lista dos artigos mais citados desde Julho de 2021. Em 2022 foi co-autora de **(6)** *“Physiological Roles of Nickel on Antioxidant and Nitrogen Metabolism Increasing the Yield of Sugarcane Plants”* publicado na Journal of Soil Science and Plant Nutrition (FI = 3.4). Em 2023, foi co-autora **(7)** *“Physiological and biochemical roles of ascorbic acid on mitigation of abiotic stresses in plants”* publicado na Plant Physiology And Biochemistry (IF = 6.1); **(8)** *“Phosphorus and Selenium Interaction Effects on Agronomic Biofortification of Cowpea Plants”* publicado na Journal Of Soil Science And Plant Nutrition (IF = 3.4); **(9)** *“Selenium Increases Ureide Metabolism and Yield of Cowpea Plants”* publicado na Gesunde Pflanzen (FI = 2.4). De acordo com o banco de dados da Web of Science, sua produção científica soma 529 citações e resulta em índice H 7 (06/10/2024).

“Life is not easy for any of us. But what of that? We must have perseverance and above all confidence in ourselves. We must believe that we are gifted for something and that this thing must be attained.”

- Marie Curie

Physicist, chemist, and winner of the 1903 and 1911 Nobel Prize in Physics

May 14, 1927, Vassar College, Poughkeepsie, Nova York.

DEDICO

A Deus, por colocar em meu coração sonhos que minhas forças humanas jamais seriam capazes de realizar, e me conceder paciência, sabedoria e resiliência para suportar o processo. *“Porque Dele, por Ele e para Ele são todas as coisas. A Ele a glória por toda a eternidade. Amém!” (Romanos, 11-36).*

Ao meu pai, Marcos Gregório Lanza (*em memória*), que dedicou sua vida para oferecer a melhor educação a nós, seus filhos, e que do céu observa sua filha se tornando a primeira Doutora da família. Obrigada por me ensinar que na simplicidade da vida de um homem comum se encontram os verdadeiros e maiores títulos: o caráter, o amor e a amizade. E que na saudade mora a admiração e o respeito, lições que nunca se apagarão com o passar do tempo. Com todo o meu amor, e saudades eternas.

A minha mãe, Ana Paula Dantas Bereta Lanza, meu exemplo de força e determinação, e minha inspiração de mulher que enfrenta todas as dificuldades da vida com resiliência, paciência e doçura. A você mãe, que esteve presente em todos os dias de minha formação acadêmica, acompanhou todas as minhas chegadas e partidas, e nunca se conformou até a chegada desse dia. Você que acreditou na minha capacidade de forma incansável, hoje compartilha o mérito de todo título atribuído a sua filha. Com todo o meu amor, e admiração de sua filha e melhor amiga.

Ao meu irmão, Matheus Dantas Gregório Lanza, por me atribuir a responsabilidade de maior honra a um ser humano: ser exemplo. A você que me acompanhou em todos os percursos da minha jornada, e que sempre me ofereceu um olhar de confiança e admiração. Que sempre me aplaudiu mais forte do que todos, e que nunca desistiu dos meus sonhos, mesmo quando eu mesmo havia desistido. A você todo o meu respeito, orgulho e gratidão.

A todos os demais membros da minha família, em especial a minha avó Iraide Gregório Lanza por todo amor e cuidado. Aos meus padrinhos (Roseli Maestre e Edvaldo da Costa), tios/tias (Elaine, Garba, Regina, José, Sandro, Ana Lúcia, Milena, Márcio, Deise, Milena e Marcos), primos/primas (Gabriel, Fernando e Guilherme), da Família Dantas (Adriana, Alessandra, Luciana, Júlia, Davi, Maria Clara, João Paulo, Maria Paula, Geovana e Guilherme) e Lanza (Guilherme, Vitor e Leticia).

A Erica Dantas Brasil (*em memória*) e sua filha Gaby, por todo carinho, acolhimento e suporte em minha experiência na Inglaterra. Sou grata pelo privilégio de aprender com vocês

que *“There are flowers everywhere for those who are willing to see them”*. Carrego comigo os ensinamentos de uma mulher forte, determinada e muito querida que um dia me disse que esse dia chegaria. Cuidarei dos seus conselhos como você cuidava das rosas que nasciam nos jardins da Bramwell Drive em Nottingham (UK), com a esperança de que o inverno é passageiro, e a primavera sempre retornará.

Ao meu orientador, André Rodrigues dos Reis, por acompanhar a minha jornada científica desde a graduação, e por me permitir ser parte do seu legado científico. Agradeço o apoio, amizade, incentivo e todas as oportunidades concedidas para o meu crescimento profissional. Agradeço o exemplo de persistência e resiliência, e serei eternamente uma grande admiradora da sua história e legado.

As minhas amigas Barbara, Lais, Mayra, Bruna, Tamara e Andressa por serem minha inspiração de mulheres fortes e determinadas, que me apoiam em cada passo do meu caminho e me oferecem a honra de compartilhar o cotidiano da vida com a mágica da amizade, sinceridade, amor, lealdade e companheirismo.

A todas as mulheres que lutam diariamente pelo desenvolvimento da ciência, em especial as mulheres da área de STEM (Ciência, Tecnologia, Engenharia e Matemática). Que esse seja mais um trabalho desenvolvido por uma mulher na ciência brasileira, e que inspire outras meninas e mulheres a buscarem esse lugar.

A todos os indivíduos que fazem parte da minha rotina de trabalho como Desenvolvimento de Mercado em Nutrição e Fisiologia de Plantas, que tocam a minha vida diariamente, e me ensinam que o verdadeiro significado do aprendizado está em tornar a vida do outro melhor, e assim fazer uma sociedade mais justa e humana, principalmente para todas as mulheres que movimentam o agronegócio mundial.

ACKNOWLEDGMENTS

First, to God, to whom I owe everything in my life.

I would like to thank my supervisor, Prof. PhD. André Rodrigues dos Reis, for accompanying and guiding me throughout my academic career. Thank you for trusting my work, for your friendship, and for being one of my main supporters in science.

Thanks to EMBRAPA's researcher PhD. Maurisrael de Moura Rocha provides the 20 genotypes of cowpeas used in this research. I also thank Prof. PhD. Adriana Lima Moro, from the University of Western São Paulo (UNOESTE), for granting us access to the Infrared gas analyzer (IRGA). These partnerships were essential for the development of the project and provided us with an exchange of knowledge, which is fundamental for the advancement of science.

I am grateful to Prof. PhD. Hudson Wallace Pereira de Carvalho for helping me to expand my project and develop solid work. And for granting me access to the X-ray equipment. I also thank all the members of the Nuclear Instrumentation Laboratory of the Center for Nuclear Energy in Agriculture of the University of São Paulo (ESALQ, USP), especially Gabriel Sgarbiero Montanha and Eduardo de Almeida, for their reception and for helping me to develop the methods and analytical and nuclear techniques.

I thank all the members of the Agricultural Physiology Research Group (GEFA) for their teamwork, learning, cooperation, and friendship, especially Lara Caroline, Matheus, Maycon, Andressa, Júlio, Vinicius, Nandhara, Marcos e Carolina. I am very proud to belong to a multidisciplinary group, as the knowledge and expertise of each professional made my work unique and with a broad vision. I also appreciate the trust in my work. Each member will always be recognized in my heart for the ethics, humanity, and professionalism lived daily during these years.

I'm glad for the opportunity to cross paths with all the amazing people I've befriended during my time at the University of Nottingham, under the supervision of Prof. Dr. Martin Broadley, and with the help of the entire Plant Science department, especially Lolit Wilson and Kenneth Davis, Sonia Khokar, Bill Broadley and Promise Muleya. I also thank my Brazilian friends who supported me during my journey in the UK Janaina e Carolina. I am grateful for all the support received from Erica Brasil (*in memory*) and Gaby, for welcoming me into their home and accepting me as part of their family during my months in Nottingham.

I would like to thank the São Paulo Research Foundation (FAPESP) for the financial support of scientific carrier, and the entire civil society that contributes to the development of science in Brazil. And appreciate the efforts and work of all the staff of UNESP (Campus Tupã and Jaboticabal), who gave me all the support necessary to perform my work.

I thank the members of the Graduate Program in Agronomy (Vegetable Production) at the Faculty of Agrarian and Veterinary Sciences, Campus de Jaboticabal, for all the empathy, patience, and support offered to me during my PhD, course.

I also thank the Universidade Estadual Paulista (UNESP), Faculdade de Ciências e Engenharia Tupã, for allowing me to use their facilities to conduct my work even with the limitations imposed by the pandemic. The Academic Support Assistant Priscilla Caparroz de Moraes (Biology Laboratory), Yasmin Saegusa Tadayozzi (Chemistry Laboratory), and Jéssica Aparecida Serafim (Multi-User Equipment Laboratory). And to all the servers that contributed directly or indirectly to the development of my project.

I would like to thank all the family (Parents - Ana Paula, Marcos Gregório (*in memory*); Brother - Matheus Lanza; Grandfather - Iraide Gregório; Godparents - Roseli Maestro e Edvaldo da Costa and all the uncles, aunts, and cousins who supported me with great patience and encouragement along this path. I also thank my friends Barbara, Mayra, Bruna, Lais, Tamara, and Andressa for following my journey and for being faithful and encouraging friends.

TABLE OF CONTENT

CHAPTER 1: SELENIUM BIOFORTIFICATION: IMPROVE HUMAN NUTRITION AND RESILIENCE TO ECONOMIC CRISES.....	1
1. INTRODUCTION.....	3
2. PAST REVIEW AND CURRENT STATUS OF AGRONOMIC BIOFORTIFICATION	4
3. SOCIOECONOMIC ASPECTS OF AGRONOMIC BIOFORTIFICATION	5
4. SELENIUM METABOLISM IN PLANTS	6
5. BENEFITS OF BIOFORTIFICATION PRACTICES I: REDUCTION OF PHYTIC ACID	8
6. BENEFITS OF BIOFORTIFICATION PRACTICES II: STRESS MITIGATION	9
7. BENEFITS OF BIOFORTIFICATION PRACTICES III: HUMAN HEALTH.....	11
8. FUTURES PERSPECTIVES	13
9. CONCLUSION.....	14
10. REFERENCES	14
 CHAPTER 2 – GENOTYPIC VARIATION IN PHENOLOGICAL PHASES OF COWPEA CULTIVARS: IMPLICATIONS FOR GRAIN YIELD AND NITROGEN USE EFFICIENCY	51
1. INTRODUCTION.....	53
2. MATERIALS AND METHODS	54
2.1 Experimental area.....	54
2.2 Experimental Setup	54
2.3 Sample collection and yield estimative	54
2.4 Digestion and Mineral Analysis	55
2.5 Gas exchange and water use efficiency	55
2.6 Analysis of Nitrogenous Compounds.....	55
2.7 Profile of Sugars and Flavonoids in HPLC.....	55
2.8 Leaf and root morphology	56
2.9 Scanning Electron Microscopy.....	57
2.10 Statistical analysis	57
2.11 Phenological Phases Determination	57
2.11.1 Vegetative Phase:	57
2.11.2 Reproductive Phase:	58
3. RESULTS	58
3.1 Phenological and Biochemical characterization of Brazilian cowpea genotypes.....	58
3.2 Photosynthetic characterization of Brazilian cowpea genotypes	61
3.3 Phenological Stages of Brazilian cowpea genotypes	62

3.3.1 Vegetative Stage.....	63
3.3.2 Reproductive Stage.....	64
3.4 Nitrate reductase, urea and urease enzymes accumulation in cowpea genotypes	65
4. DISCUSSION	66
5. CONCLUSION	69
6. FIGURES	70
7. SUPPLEMENTARY FIGURES	84
8. REFERENCES	99

CHAPTER 3 - SELENIUM ACCUMULATION AT DIFFERENT PHENOLOGICAL STAGES OF COWPEA PLANTS: PHYSIOLOGICAL AND BIOCHEMICAL RESPONSES 106

1. INTRODUCTION	108
2. MATERIAL AND METHODS	109
2.1 Experimental área.....	109
2.2 Experiment design and installation.....	110
2.3 Metabolites analyses	111
2.3.1 Total sugars and sucrose.....	111
2.3.2 Ureides concentration.....	112
2.3.3 Amino acids and Storage Protein.....	112
2.3.4 Extraction and phytic acid concentration in grains.	113
2.3.5 Digestion and mineral analysis of leaf and seed samples	113
2.3.6 Statistical analysis	114
3 RESULTS	114
3.1 Phenological phases of cowpea	114
3.2 Uptake and accumulation of Se	115
3.3 Nitrogen compounds: ureides and amino acids	115
3.4 Determination of sucrose and total sugars	116
3.5 Grain yield, proteins, and phytate.....	116
4 DISCUSSION	116
5 CONCLUSION	118
7 REFERENCES	127
8 SUPPLEMENTARY MATERIAL	155

CHAPTER 4 - ASSESSMENT OF SELENIUM SPATIAL DISTRIBUTION USING M-XFR IN COWPEA (*Vigna unguiculata* (L.) Walp.) PLANTS: INTEGRATION OF PHYSIOLOGICAL AND BIOCHEMICAL RESPONSES 158

1. INTRODUCTION	160
2. MATERIALS AND METHODS	161

2.1 Experimental area.....	161
2.2 Chemical analysis of soil of plants	162
2.3 Histological analysis	163
2.4 Scanning electron microscopy and X-ray fluorescence spectroscopy (XRF)	164
2.5 Analysis of photosynthetic pigments.....	164
2.6 Lipid peroxidation and hydrogen peroxide determination	165
2.7 Determination of total sugar and sucrose	165
2.8 Yield.....	166
2.9 Statistical analysis	166
3. RESULTS	166
3.1 Ultrastructural and nutrient analysis of distribution by μ -XRF.....	166
3.2 Photosynthetic pigments, photoassimilates, and oxidative stress	167
3.3 Nutritional analysis, seed element distribution and yield	168
3. DISCUSSION	168
4. CONCLUSIONS.....	171
5. FIGURES	172
6. REFERENCES	179
7. SUPPLEMENTARY MATERIAL	207

CHAPTER 5: AGRONOMIC BIOFORTIFICATION OF COWPEA PLANTS WITH SELENIUM: A KEY RESOURCE FOR FOOD SECURITY AND ALLEVIATION OF MALNUTRITION	211
1. INTRODUCTION	213
2. MATERIALS AND METHODS	214
2.1 Experimental area and land details.....	214
2.2 Experimental Setup	215
2.3 Sample collection and yield estimative	216
2.4 Digestion and Mineral Analysis	216
2.5 Daily Se Intake.....	216
2.5 Statistical analysis	217
3. RESULTS	218
3.1 Selenium concentration.....	218
3.1.1 Leaves	218
3.1.2 Seeds	218
3.2 Yield.....	219
3.3 Selenium Daily Intake	219
3.4 Nutrient content in biofortified cowpea.....	220

3.4.1 Macronutrients	220
3.4.2 Micronutrients	221
4. DISCUSSION	221
5. CONCLUSION	225
6. FIGURES	226
7. REFERENCES	233

GENOTYPIC VARIABILITY OF COWPEA (*Vigna unguiculata* (L.) Walp) AND SELENIUM UPTAKE FOR DETERMINING AGRONOMIC BIOFORTIFICATION STRATEGIES TO COMBAT HIDDEN HUNGER

RESUMO – O aumento da produtividade de grãos tem apresentado relação inversa como conteúdo nutricional das plantas, ocasionando o aumento da fome oculta. O Feijão-caupi é rico em proteínas, e fonte de alimentação com baixo valor agregado para países emergentes. Em hipótese, a determinação de genótipos mais responsivos de feijão-caupi a aplicação de Se por meio da marcha de absorção de nutrientes proporciona estratégias de biofortificação agronômica. O experimento I foi conduzido com 20 genótipos de feijão-caupi para determinação da variação genotípica nos parâmetros de produção e na eficiência do uso de nitrogênio. No experimento II foram aplicadas via solo e foliar as doses de Se (0, 10, 25, 50, 100 e 150 g ha⁻¹) na fonte selenato de sódio e avaliados as respostas fisiológicas e morfológicas dos tratamentos. No experimento III, o genótipo responsivo do experimento I e dose-resposta do experimento II foram utilizados para determinar a aplicação no feijão-caupi e avaliado a marcha de absorção de Se nos estádios fenológicos da culturas para determinar a curva de acúmulo de nutrientes. Ao final dos experimentos, foi avaliado o acúmulo de nutrientes (essenciais, benéficos e antinutrientes) nos grãos de feijão-caupi em resposta a aplicação de Selênio para a alimentação humana. Foram realizadas análises de fotossíntese (A), condutância estomática (GS), clorofila (Cl), concentração interna de CO₂ (E), metabolismo antioxidante (malonaldeído, peróxido de hidrogênio, enzimas antioxidantes), metabolismo do nitrogênio (ureia, urease e nitrato redutase), perfil de açúcares e flavonoides (HPLC), proteínas de reserva (albumina, globulina, prolamina e glutelina) e concentração de elementos. Os resultados obtidos nesse estudo poderão gerar contribuições significativas para a seleção de genótipos de feijão-caupi responsivos aos programas de biofortificação agronômica e doses seguras de selênio para o combate das deficiências nutricionais por meio de estratégias agronômicas.

Palavras-chave: Nutrição de Plantas, Fisiologia Vegetal, Feijão-caupi, Marcha De Absorção, Selênio.

GENOTYPIC VARIABILITY OF COWPEA (*Vigna unguiculata* (L.) Walp) AND SELENIUM UPTAKE FOR DETERMINING AGRONOMIC BIOFORTIFICATION STRATEGIES TO COMBAT HIDDEN HUNGER.

ABSTRACT – The increase in grain yield has shown an inverse relationship with the nutritional content of plants, leading to an increase in hidden hunger. Cowpea is rich in proteins and serves as a low-value food source for emerging countries. Hypothetically, determining more responsive cowpea genotypes to selenium (Se) application through nutrient uptake gait provides agronomic biofortification strategies. The experiment I conducted with 20 cowpea genotypes to determine genotypic variation in production parameters and nitrogen use efficiency. In Experiment II, Se doses (0, 10, 25, 50, 100, and 150 g ha⁻¹) were applied via soil and foliar routes using sodium selenate, and the physiological and morphological responses of the treatments will be evaluated. In Experiment III, the responsive genotype from Experiment I and the dose-response from Experiment II were used to determine Se application in cowpea and evaluate Se uptake gait during the phenological stages of the crop to determine the nutrient accumulation curve. At the end of the experiments, the accumulation of nutrients (essential, beneficial, and antinutrients) in cowpea grains in response to Se application for human consumption was evaluated. Analyses of photosynthesis (A), stomatal conductance (GS), chlorophyll (Cl), internal CO₂ concentration (E), antioxidant metabolism (malondialdehyde, hydrogen peroxide, antioxidant enzymes), nitrogen metabolism (urea, urease, and nitrate reductase), sugar and flavonoid profiles (HPLC), storage proteins (albumin, globulin, prolamin, and glutelin), and element concentration were performed. The results obtained from this study may generate significant contributions to the selection of responsive cowpea genotypes for agronomic biofortification programs and safe doses of selenium to combat nutritional deficiencies through agronomic strategies.

Keywords: *Vigna Unguiculata*, Agronomic Biofortification, Hidden Hunger, Absorption March, Selenium.

CHAPTER 1 - SELENIUM BIOFORTIFICATION: IMPROVE HUMAN NUTRITION AND RESILIENCE TO ECONOMIC CRISES

Maria Gabriela Dantas Bereta Lanza¹, André Rodrigues dos Reis^{1,2*}.

¹ São Paulo State Univeristy – UNESP, Rua Domingos da Costa Lopes 780, Tupã-SP, Postal Code 17602-496, Brazil.

² São Paulo State Univeristy – UNESP, Via de Acesso Prof. Paulo Donato Castellane s/n – Jaboticabal-SP, Postal Code 14884-900, Brazil.

*Author for correspondence:

André Rodrigues dos Reis

Tel: +55 14 3704 4265

E-mail: andre.reis@unesp.br

ABSTRACT

Malnutrition can be caused by hidden hunger, resulting in deficiencies in essential micronutrients such as selenium, iron, zinc, copper, calcium, and magnesium. This study developed a meta-analysis of documents from the Web of Science using the keywords "Biofortification," "Selenium," and "Cowpea" from 1990 to 2024. It analyzed the number of documents, keywords, citations, institutions, and H-index to understand the research landscape in this field. Biofortification is a quick, efficient, and sustainable method to reduce micronutrient deficiencies. Selenium (Se) is an essential micronutrient for human and animal health due to its ability to support antioxidant defense systems. Agronomic selenium biofortification has demonstrated significant success in Finland, where selenium-enriched fertilizers have substantially improved public health by boosting selenium levels in food crops. Its significance became particularly evident during the COVID-19 pandemic, as adequate selenium levels were correlated with reduced infection severity, emphasizing its role in global health crises. Economically, selenium biofortification enhances agricultural sustainability by increasing crop resilience to abiotic stresses such as drought and salinity and reducing phytic acid content, thus improving mineral bioavailability. In plants, Se can regulate the activity of antioxidant enzymes, such as superoxide dismutase (SOD), peroxidase (POX), ascorbate peroxidase (APX), and catalase (CAT), which help neutralize reactive oxygen species (ROS) generated under stress conditions. Furthermore, leveraging genotypic variability ensures that different crops optimally uptake selenium, thereby enhancing their nutritional value. By promoting better human nutrition and more robust agricultural systems, selenium biofortification is vital for achieving the UN Sustainable Development Goals, especially Goal 2 (Zero Hunger), which seeks to end hunger, achieve food security and improved nutrition, and promote sustainable agriculture by 2030.

Keywords: micronutrients, abiotic stress, phytic acid, food safety, hidden hunger, agronomic biofortification, zero hunger, human health.

1. INTRODUCTION

Micronutrient malnutrition, also referred to as hidden hunger, pertains to insufficient essential vitamins and minerals, leading to significant adverse impacts on human health (Bouis and Saltzman 2017a). By enhancing the nutritional quality of food crops, agronomic biofortification contributes to increased food security, which is always defined as the consistent access of all individuals to safe and nutritious food (Simkin 2019).

Hidden hunger affects over two billion individuals globally, with higher prevalence in Africa, South Asia, and Northeast Brazil (Siwela et al. 2020). Approximately 811 million people suffer from chronic malnutrition. (Langyan et al. 2022). Thus, One-third of women of childbearing age are anemic, 22% of children under the age of five suffer from stunted growth, and 50% of the global population of pregnant women experience micronutrient deficiencies. (Alae-Carew et al. 2019; PNUD 2022).

Biofortification can contribute to the achievement of various United Nations Sustainable Development Goals (SDGs), including eradicating hunger, achieving food security, improving nutrition, and promoting sustainable agriculture (Rehman et al. 2021). Additionally, biofortification is regarded as a highly cost-effective strategy to combat micronutrient malnutrition on a global scale, particularly when compared to other interventions such as dietary diversification or micronutrient supplementation. (Wakeel et al. 2018)

Among the critical minerals involved are iron, iodine, zinc, copper, calcium, magnesium, and selenium (Shankar 2020; White et al. 2009). The global deficiency rates for Fe, Zn, I, and Se are approximately 60%, 30%, 30%, and 15%, respectively. Additionally, deficiencies in Zn, Fe, and I, along with vitamin A deficiency, can result in a mortality rate of approximately 20% among children under the age of five. (Imdad et al. 2022; White et al. 2009). Over the past twenty years, global governments have invested more than US\$ 15 billion in vitamin A supplementation capsules as a medical strategy (Imdad et al. 2017a).

Biofortification involves the enhancement of micronutrient levels in food crops through agricultural technologies (Garg et al. 2018). This technique is essential in agriculture and food systems, as it has the potential to enhance the nutritional quality of crops and subsequently improve public health, particularly for populations with limited access to diverse diets or other micronutrient interventions (Langyan et al. 2022). This study aims to provide a comprehensive overview of the scientific evolution of agronomic biofortification and its integration into food systems. It highlights various success cases that have reduced regional food insecurities and contributed to the development and health of the global population.

5. CONCLUSION

The agronomic biofortification of cowpeas with Se proves to be an efficient way to increase the grain yield and nutritional quality of grains. Doses up to 50 g ha⁻¹ positively increase the grain yield. Foliar Se application is revealed to be the most efficient strategy to increase Se concentration in leaves and grains. Low doses of Se (10 and 25 g ha⁻¹) applied via foliar are sufficient to meet the nutritional needs of a population that consumes cowpeas and meet the requirements established by the *Codex Alimentarius* (60 µg day⁻¹). The application of Se also plays synergistic/antagonistic roles with other nutrients that contribute to plant development, such as photosynthesis. This study presents important information for Se management recommendations in field-grown cowpea plants aiming to increase Se concentration in grains at a safety level to combat malnutrition. And reveals that the agronomic biofortification of cowpeas with Se is a cheap and accessible strategy to increase the quality of staple foods, like cowpea, aiming to combat malnutrition in developing countries.

7. REFERENCES

- Abebe BK, Alemayehu MT (2022) A review of the nutritional use of cowpea (*Vigna unguiculata* L. Walp) for human and animal diets. *J Agric Food Res* 10:100383. <https://doi.org/10.1016/J.JAFR.2022.100383>
- Adebayo A, Yakubu O, Bakare-Akpata O (2020) Uptake, Metabolism and Toxicity of Selenium in Tropical Plants. In: Importance of Selenium in the Environment and Human Health. IntechOpen
- Affrifah NS, Phillips RD, Saalia FK (2022) Cowpeas: Nutritional profile, processing methods and products—A review. *Legume Science* 4:e131. <https://doi.org/10.1002/LEG3.131>
- Alae-Carew C, Bird FA, Choudhury S, Harris F, Aleksandrowicz L, Milner J, Joy EJ, Agrawal S, Dangour AD, Green R (2019) Future diets in India: A systematic review of food consumption projection studies. *Glob Food Sec* 23:182–190. <https://doi.org/10.1016/J.GFS.2019.05.006>
- Alexieva V, Sergiev I, Mapelli S, Karanov E (2001) The effect of drought and ultraviolet radiation on growth and stress markers in pea and wheat. *Plant Cell Environ* 24:1337–1344. <https://doi.org/10.1046/j.1365-3040.2001.00778.x>
- Alfthan G, Aspila P, Ekholm P, Eurola M, Hartikainen H, Hero H, Hietaniemi V, Root T, Salminen P, Venäläinen ER, Aro A (2010) Nationwide supplementation of sodium selenate to commercial fertilizers: History and 25-year results from the Finnish selenium monitoring programme. *Combating Micronutrient Deficiencies: Food-based Approaches* 312–337. <https://doi.org/10.1079/9781845937140.0312>
- Alves LR, Rossatto DR, Rossi ML, Martinelli AP, Gratão PL (2020) Selenium improves photosynthesis and induces ultrastructural changes but does not alleviate cadmium-stress damages in tomato plants. *Protoplasma* 257:597–605. <https://doi.org/10.1007/S00709-019-01469-W/FIGURES/5>
- Andrade FR, da Silva GN, Guimarães KC, Barreto HBF, de Souza KRD, Guilherme LRG, Faquin V, Reis AR dos (2018) Selenium protects rice plants from water deficit stress. *Ecotoxicol Environ Saf* 164:562–570. <https://doi.org/10.1016/j.ecoenv.2018.08.022>
- Antonio C, Santos F (2012) Melhoramento do feijão-caupi para temperaturas moderadas e elevadas no Vale do São Francisco.
- Asri-Rezaei S, Nourian A, Shalazar-Jalali A, Najafi G, Nazarizadeh A, Koohestani M, Karimi A (2018) Selenium supplementation in the form of selenium nanoparticles and selenite sodium improves mature male mice reproductive performances. *Iran J Basic Med Sci* 21:577. <https://doi.org/10.22038/IJBMS.2018.26023.6397>
- Bailey RL, West KP, Black RE (2015) The Epidemiology of Global Micronutrient Deficiencies. *Ann Nutr Metab* 66:22–33. <https://doi.org/10.1159/000371618>
- Bairu M, Laurie S, Mavengahama S, Mekonnen TW, Shegro Gerrano A, Mbuma NW, Labuschagne MT (2022) Breeding of Vegetable Cowpea for Nutrition and Climate

- Resilience in Sub-Saharan Africa: Progress, Opportunities, and Challenges. *Plants* 2022, Vol 11, Page 1583 11:1583. <https://doi.org/10.3390/PLANTS11121583>
- Bajaj M, Eiche E, Neumann T, Winter J, Gallert C (2011) Hazardous concentrations of selenium in soil and groundwater in North-West India. *J Hazard Mater* 189:640–646. <https://doi.org/10.1016/J.JHAZMAT.2011.01.086>
- Bao P, Huang H, Hu ZY, Häggblom MM, Zhu YG (2013) Impact of temperature, CO₂ fixation and nitrate reduction on selenium reduction, by a paddy soil *Clostridium* strain. *J Appl Microbiol* 114:703–712. <https://doi.org/10.1111/JAM.12084>
- Barker A V., Pilbeam DJ (2016) *Handbook of plant nutrition*
- Barros JRA, Guimarães MJM, Silva RM e., Rêgo MTC, de Melo NF, de Melo Chaves AR, Angelotti F (2021) Selection of cowpea cultivars for high temperature tolerance: physiological, biochemical and yield aspects. *Physiology and Molecular Biology of Plants* 27:29–38. <https://doi.org/10.1007/S12298-020-00919-7/METRICS>
- Bhattacharjee S (2010) Sites of generation and physicochemical basis of formation of reactive oxygen species in plant cell. In: *Reactive Oxygen Species and Antioxidants in Higher Plants*. CRC Press New York, NY, USA, pp 1–30
- Bielecki RL, Turner NA (1966a) Separation and estimation of amino acids in crude plant extracts by thin-layer electrophoresis and chromatography. *Anal Biochem* 17:278–293. [https://doi.org/10.1016/0003-2697\(66\)90206-5](https://doi.org/10.1016/0003-2697(66)90206-5)
- Bielecki RL, Turner NA (1966b) Separation and estimation of amino acids in crude plant extracts by thin-layer electrophoresis and chromatography. *Anal Biochem* 17:278–293. [https://doi.org/10.1016/0003-2697\(66\)90206-5](https://doi.org/10.1016/0003-2697(66)90206-5)
- Blamey FPC, Hernandez-Soriano MC, Cheng M, Tang C, Paterson DJ, Lombi E, Wang WH, Scheckel KG, Kopittke PM (2015) Synchrotron-based techniques shed light on mechanisms of plant sensitivity and tolerance to high manganese in the root environment. *Plant Physiol* 169:2006–2020. <https://doi.org/10.1104/pp.15.00726>
- Bloot APM, Kalschne DL, Amaral JAS, Baraldi IJ, Canan C (2023) A Review of Phytic Acid Sources, Obtention, and Applications. *Food Reviews International* 39:73–92. <https://doi.org/10.1080/87559129.2021.1906697>
- Bohn L, Meyer AS, Rasmussen SK (2008) Phytate: Impact on environment and human nutrition. A challenge for molecular breeding. *J Zhejiang Univ Sci B* 9:165–191. <https://doi.org/10.1631/jzus.B0710640>
- Bohrer AS, Kopriva S, Takahashi H (2015) Plastid-cytosol partitioning and integration of metabolic pathways for APS/PAPS biosynthesis in *Arabidopsis thaliana*. *Front Plant Sci* 5:1–4. <https://doi.org/10.3389/FPLS.2014.00751/BIBTEX>
- Bouis H, Saltzman A (2017a) Melhorando a nutrição por meio da biofortificação: uma revisão das evidências do HarvestPlus, 2003 a 2016. *Segurança alimentar global*

- Bouis H, Saltzman A, Birol E (2019) Improving Nutrition Through Biofortification. In: Agriculture for improved nutrition: seizing the momentum. pp 47–57
- Bouis HE, Hotz C, McClafferty B, Meenakshi J V., Pfeiffer WH (2011) Biofortification: a new tool to reduce micronutrient malnutrition. Food Nutr Bull 32:. <https://doi.org/10.1177/15648265110321S105>
- Bouis HE, Saltzman A (2017b) Improving nutrition through biofortification: A review of evidence from HarvestPlus, 2003 through 2016. Glob Food Sec 12:49–58
- Boukar O, Fatokun CA, Roberts PA, Abberton M, Boukar O, Fatokun CA, Roberts PA, Abberton M, Huynh BL, Close TJ, Kyei-Boahen S, Higgins TJ V, Ehlers JD (2015) Caupi. SpringerO Boukar, CA Fatokun, PA Roberts, M Abberton, BL Huynh, TJ Close, S Kyei-Boahen. Leguminosas para grãos, 2015•Springer 219–250. https://doi.org/10.1007/978-1-4939-2797-5_7
- Bradford MM (1976) A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. Anal Biochem 72:248–254. [https://doi.org/10.1016/0003-2697\(76\)90527-3](https://doi.org/10.1016/0003-2697(76)90527-3)
- Broadley MR, White PJ, Hammond JP, Zelko I, Lux A (2007) Zinc in plants. New Phytologist 173:677–702. <https://doi.org/10.1111/J.1469-8137.2007.01996.X>
- Cabral Gouveia GC, Galindo FS, Dantas Bereta Lanza MG, Caroline da Rocha Silva A, Pereira de Brito Mateus M, Souza da Silva M, Rimoldi Tavanti RF, Tavanti TR, Lavres J, Reis AR dos (2020) Selenium toxicity stress-induced phenotypical, biochemical and physiological responses in rice plants: Characterization of symptoms and plant metabolic adjustment. Ecotoxicol Environ Saf 202:110916. <https://doi.org/10.1016/j.ecoenv.2020.110916>
- Camaschella C (2019) Iron deficiency. Blood 133:30–39
- Campos F, Rodrigues Freire Filho F, Lopes Â, Ribeiro V, Querino R, Rocha M (2000a) Ciclo fenológico em caupi (*Vigna unguiculata* L.Walp.): uma proposta de escala de desenvolvimento. Rev Cient Rural 5:110–116
- Campos FL, Ribeiro VQ, Rocha M (2000b) Ciclo fenológico em caupi (*Vigna unguiculata* L.Walp.): uma proposta de escala de desenvolvimento Universidade Federal do Delta do Parnaíba
- Cantarella H, Van Raij B, Quaggio JA (1998) Soil and plant analyses for lime and fertilizer recommendations in Brazil. In: Communications in Soil Science and Plant Analysis. Marcel Dekker Inc., pp 1691–1706
- Cappa JJ, Cappa PJ, El Mehdawi AF, McAleer JM, Simmons MP, Pilon-Smits EAH (2014) Characterization of selenium and sulfur accumulation across the genus *Stanleya* (Brassicaceae): A field survey and common-garden experiment. Am J Bot 101:830–839. <https://doi.org/10.3732/AJB.1400041>
- Carlquist S (1975) Ecological strategies of xylem evolution

- Carvalho AFU, de Sousa NM, Farias DF, da Rocha-Bezerra LCB, da Silva RMP, Viana MP, Gouveia ST, Sampaio SS, de Sousa MB, de Lima GPG, de Moraes SM, Barros CC, Filho FRF (2012) Nutritional ranking of 30 Brazilian genotypes of cowpeas including determination of antioxidant capacity and vitamins. *Journal of Food Composition and Analysis* 26:81–88. <https://doi.org/10.1016/J.JFCA.2012.01.005>
- Carvalho M, Castro I, Moutinho-Pereira J, Correia C, Egea-Cortines M, Matos M, Rosa E, Carnide V, Lino-Neto T (2019) Evaluating stress responses in cowpea under drought stress. *J Plant Physiol* 241:153001. <https://doi.org/10.1016/J.JPLPH.2019.153001>
- Carvalho M, Lino-Neto T, Rosa E, Carnide V (2017a) Cowpea: a legume crop for a challenging environment. *J Sci Food Agric* 97:4273–4284. <https://doi.org/10.1002/JSFA.8250>
- Carvalho M, Lino-Neto T, Rosa E, Carnide V (2017b) Cowpea: a legume crop for a challenging environment. *J Sci Food Agric* 97:4273–4284
- Cataldo DA, Haroon MH, Schrader LE, Youngs VL (1975) Rapid colorimetric determination of nitrate in plant tissue by nitration of salicylic acid. *Commun Soil Sci Plant Anal* 6:71–80. <https://doi.org/10.1080/00103627509366547>
- Chauhan R, Awasthi S, Srivastava S, Dwivedi S, Pilon-Smits EAH, Dhankher OP, Tripathi RD (2019) Understanding selenium metabolism in plants and its role as a beneficial element. *Crit Rev Environ Sci Technol* 49:1937–1958. <https://doi.org/10.1080/10643389.2019.1598240>
- Chauhan R, Awasthi S, Tripathi P, Mishra S, Dwivedi S, Niranjana A, Mallick S, Tripathi P, Tripathi RD, Chauhan R, Pande V (2017) Selenite modulates the level of phenolics and nutrient element to alleviate the toxicity of arsenite in rice (*Oryza sativa* L.). *Ecotoxicol Environ Saf* 138:47–55. <https://doi.org/10.1016/j.ecoenv.2016.11.015>
- Chilimba ADC, Young SD, Black CR, Rogerson KB, Louise Ander E, Watts MJ, Lammel J, Broadley MR (2011a) Maize grain and soil surveys reveal suboptimal dietary selenium intake is widespread in Malawi. *Scientific Reports* 1:1–9. <https://doi.org/10.1038/srep00072>
- Chilimba ADC, Young SD, Black CR, Rogerson KB, Louise Ander E, Watts MJ, Lammel J, Broadley MR (2011b) Maize grain and soil surveys reveal suboptimal dietary selenium intake is widespread in Malawi. *Sci Rep* 1:1–9. <https://doi.org/10.1038/srep00072>
- Combs GF (2001a) Selenium in global food systems. *British Journal of Nutrition* 85:517–547. <https://doi.org/10.1079/bjn2000280>
- Combs J (2001b) Impact of selenium and cancer-prevention findings on the nutrition-health paradigm. *Nutr Cancer* 40:6–11
- CONAB (2023) Boletim da Safra de Grãos 2023/2024. <https://www.conab.gov.br/info-agro/safras/graos/boletim-da-safra-de-graos>. Accessed 9 Aug 2024
- Crizel RL, Deitos ÊC, Ferreira CD, Ziegler V, Hoffmann JF, Pinto VZ (2023) Role of Proteins in Chickpea and Cowpea Considering Global Food Security, Especially Protein-Based

- Diet Security. Chickpea and Cowpea: Nutritional Profile, Processing, Health Prospects and Commercial Uses 195–226. <https://doi.org/10.1201/9781003382027-8/ROLE-PROTEINS-CHICKPEA-COWPEA-CONSIDERING-GLOBAL-FOOD-SECURITY-ESPECIALLY-PROTEIN-BASED-DIET-SECURITY-ROSANE-LOPES-CRIZEL->
- Cui EJ, Song NY, Shrestha S, Chung IS, Kim JY, Jeong TS, Baek NI (2012) Flavonoid glycosides from cowpea seeds (*Vigna sinensis* K.) inhibit LDL oxidation. *Food Sci Biotechnol* 21:619–624. <https://doi.org/10.1007/S10068-012-0080-7/METRICS>
- Dahlen CR, Reynolds LP, Caton JS (2022) Selenium supplementation and pregnancy outcomes. *Front Nutr* 9:1011850. <https://doi.org/10.3389/FNUT.2022.1011850/BIBTEX>
- Dai H, Wei S, Twardowska I (2020) Biofortification of soybean (*Glycine max* L.) with Se and Zn, and enhancing its physiological functions by spiking these elements to soil during flowering phase. *Science of the Total Environment* 740:139648. <https://doi.org/10.1016/j.scitotenv.2020.139648>
- Dakora FD, Belane AK (2019) Evaluation of Protein and Micronutrient Levels in Edible Cowpea (*Vigna Unguiculata* L. Walp.) Leaves and Seeds. *Front Sustain Food Syst* 3:70. <https://doi.org/10.3389/fsufs.2019.00070>
- Dall'Acqua S, Ertani A, Pilon-Smits EAH, Fabrega-Prats M, Schiavon M (2019) Selenium Biofortification Differentially Affects Sulfur Metabolism and Accumulation of Phytochemicals in Two Rocket Species (*Eruca Sativa* Mill. and *Diplotaxis Tenuifolia*) Grown in Hydroponics. *Plants* 2019, Vol 8, Page 68 8:68. <https://doi.org/10.3390/PLANTS8030068>
- Dareus R, Acharya JP, Paudel DR, Lopes De Souza CH, Tome Gouveia B, Chase CA, DiGennaro P, Mulvaney MJ, Koenig R, Rios EF (2021) Phenotypic diversity for phenological and agronomic traits in the UC-Riverside cowpea (*Vigna unguiculata* L. Walp) mini-core collection. *Crop Sci* 61:3551–3563. <https://doi.org/10.1002/CSC2.20544>
- de Cassia Felix Alvarez R, Taveira AC, de Lima SF, Teodoro LPR, de Oliveira JT, dos Santos A, Rodrigues EV, Ceccon G, Teodoro PE (2021) Genetic Diversity of Cowpea Genotypes Grown in the Brazilian Cerrado. *HortScience* 56:28–29. <https://doi.org/10.21273/HORTSCI15514-20>
- de Souza Marinho JT, Lessa LS, da Costa CR (2021) Agronomic performance of cowpea genotypes in southwestern Brazilian Amazon. *Pesqui Agropecu Bras* 56:e02046. <https://doi.org/10.1590/S1678-3921.PAB2021.V56.02046>
- de Valença AW, Bake A, Brouwer ID, Giller KE (2017) Agronomic biofortification of crops to fight hidden hunger in sub-Saharan Africa. *Glob Food Sec* 12:8–14. <https://doi.org/10.1016/J.GFS.2016.12.001>
- Demidchik V, Cuin TA, Svistunenko D, Smith SJ, Miller AJ, Shabala S, Sokolik A, Yurin V (2010) Arabidopsis root K⁺-efflux conductance activated by hydroxyl radicals: Single-channel properties, genetic basis and involvement in stress-induced cell death. *J Cell Sci* 123:1468–1479. <https://doi.org/10.1242/jcs.064352>

- Deshpande SK, Mani BR, Desai SA, Nagarathna TK, Hanchinal RR (2018) Review on Characterization of Cowpea Germplasm in Terms of Distinctness, Uniformity, Stability and Novelty for Morphological, Quality and Yield Attributing Parameters. *IntJCurrMicrobiolAppSci* 7:1124–1139. <https://doi.org/10.20546/ijcmas.2018.706.134>
- Diao M, Ma L, Wang J, Cui J, Fu A, Liu H ying (2014) Selenium Promotes the Growth and Photosynthesis of Tomato Seedlings Under Salt Stress by Enhancing Chloroplast Antioxidant Defense System. *J Plant Growth Regul* 33:671–682. <https://doi.org/10.1007/s00344-014-9416-2>
- Dixon NE, Gazzola C, Blakeley RL, Zerner B (1975) Jack Bean Urease (EC 3.5.1.5). A Metalloenzyme. A Simple Biological Role for Nickel? *J Am Chem Soc* 97:4131–4133. <https://doi.org/10.1021/JA00847A045>
- dos Reis AR, El-Ramady H, Santos EF, Grato PL, Schomburg L (2017) Overview of Selenium Deficiency and Toxicity Worldwide: Affected Areas, Selenium-Related Health Issues, and Case Studies. 209–230. https://doi.org/10.1007/978-3-319-56249-0_13
- Dubois M, Gilles KA, Hamilton JK, Rebers PA, Smith F (1956) Colorimetric Method for Determination of Sugars and Related Substances. *Anal Chem* 28:350–356. <https://doi.org/10.1021/ac60111a017>
- DuBois Michel, Gilles KA, Hamilton JK, Rebers PA, Smith Fred (1956) Colorimetric Method for Determination of Sugars and Related Substances. *Anal Chem* 28:350–356. <https://doi.org/10.1021/ac60111a017>
- Duntas LH, Benvenga S (2015) Selenium: an element for life. *Endocrine* 48:756–775. <https://doi.org/10.1007/S12020-014-0477-6/FIGURES/2>
- Echer FR, Dominato JC, Creste JE (2009) Absorção de nutrientes e distribuição da massa fresca e seca entre órgãos de batata-doce. *Hortic Bras* 27:176–182. <https://doi.org/10.1590/s0102-05362009000200010>
- Eggersdorfer M, Akobundu U, Bailey RL, et al (2018) Hidden hunger: Solutions for America's aging populations. *Nutrients* 10:1210
- El Mehdawi AF, Jiang Y, Guignardi ZS, Esmat A, Pilon M, Pilon-Smits EAH, Schiavon M (2018) Influence of sulfate supply on selenium uptake dynamics and expression of sulfate/selenate transporters in selenium hyperaccumulator and nonhyperaccumulator Brassicaceae. *New Phytologist* 217:194–205. <https://doi.org/10.1111/NPH.14838>
- El Mehdawi AF, Pilon-Smits EAH (2012) Ecological aspects of plant selenium hyperaccumulation. *Plant Biol* 14:1–10
- Elkelish AA, Soliman MH, Alhaithloul HA, El-Esawi MA (2019) Selenium protects wheat seedlings against salt stress-mediated oxidative damage by up-regulating antioxidants and osmolytes metabolism. *Plant Physiology and Biochemistry* 137:144–153. <https://doi.org/10.1016/J.PLAPHY.2019.02.004>

- Eurola M, Alainen T, Berlin T, Ekholm P, Erlund I, Hieta-Niemi V, Mannio J, Mykkänen S, Pulkkinen M, Root T, Seppänen M, Siimes K, Venäläinen E-R, Ylivainio K (2022) Report of the Selenium Working Group 2022
- Fairweather-Tait S, Reviews RH-NR, 1996 undefined (1996) Bioavailability of minerals and trace elements: Members of EC flair concerted action no. 10: Measurements of micronutrient absorption and status. *cambridge.org* S Fairweather-Tait, RF Hurrell *Nutrition Research Reviews*, 1996•*cambridge.org* 9:295–324. <https://doi.org/10.1079/NRR19960016>
- Fairweather-Tait SJ, Bao Y, Broadley MR, Collings R, Ford D, Hesketh JE, Hurst R (2011a) Selenium in human health and disease. *Antioxid Redox Signal* 14:1337–1383
- Fairweather-Tait SJ, Bao Y, Broadley MR, Collings R, Ford D, Hesketh JE, Hurst R (2011b) Selenium in human health and disease. *Antioxid Redox Signal* 14:1337–1383
- Fanzo J, Hawkes C, Udomkesmalee E, Afshin A (2018) 2018 Global Nutrition Report: Shining a light to spur action on nutrition
- FAO (2023) Crops and Livestock Products. <https://www.fao.org/faostat/en/#data/QCL>. Accessed 9 Aug 2024
- FAO (2017) The State of Food Insecurity in the World: Building Resilience for Peace and Food Security. Rome
- FAO F and agriculture organization of the UNations (2020a) The State of Food Security and Nutrition in the World 2020
- FAO F and agriculture organization of the UNations (2020b) The Global Economy of Pulses
- Farias VD da S, Costa DLPC, Souza PJ de OP de, Takaki AY, Lima MJA de (2015) Temperaturas basais e necessidade térmica para o ciclo de desenvolvimento do feijão-caupi. *Enciclopédia Biosfera* 11:1781
- Faye M, Jooste A, ... JL-D-SAJ of, 2006 undefined Impact of sucrose contents and cooking time on cowpea prices in Senegal: economics. *journals.co.za* MD Faye, A Jooste, J Lowenberg-Deboer, J Fulton *South African Journal of Economic and Management Sciences*, 2006•*journals.co.za*
- Feizollahi E, Mirmahdi RS, Zoghi A, Zijlstra RT, Roopesh MS, Vasanthan T (2021) Review of the beneficial and anti-nutritional qualities of phytic acid, and procedures for removing it from food products. *Food Research International* 143:110284. <https://doi.org/10.1016/J.FOODRES.2021.110284>
- Feng R, Wei C, Tu S (2013) The roles of selenium in protecting plants against abiotic stresses. *Environ Exp Bot* 87:58–68
- Ferreira RLU, Sena-Evangelista KCM, de Azevedo EP, Pinheiro FI, Cobucci RN, Pedrosa LFC (2021) Selenium in Human Health and Gut Microflora: Bioavailability of Selenocompounds and Relationship With Diseases. *Front Nutr* 8:685317. <https://doi.org/10.3389/FNUT.2021.685317/BIBTEX>

- FILHO FF, FILHO FF (2011) Feijão-caupi no Brasil: produção, melhoramento genético, avanços e desafios.
- Fredlund K, Isaksson M, Rossander-Hulthén L, Almgren A, Sandberg AS (2006) Absorption of zinc and retention of calcium: Dose-dependent inhibition by phytate. *Journal of Trace Elements in Medicine and Biology* 20:49–57. <https://doi.org/10.1016/J.JTEMB.2006.01.003>
- Galindo FS, Teixeira Filho MCM, da Silva EC, Buzetti S, Fernandes GC, Rodrigues WL (2020) Technical and economic viability of cowpea co-inoculated with *Azospirillum brasilense* and *Bradyrhizobium* spp. and nitrogen doses. *Revista Brasileira de Engenharia Agrícola e Ambiental* 24:304–311. <https://doi.org/10.1590/1807-1929/AGRIAMBI.V24N5P304-311>
- Gammelgaard B, Jackson MI, Gabel-Jensen C (2010a) Surveying selenium speciation from soil to cell—forms and transformations. *Analytical and Bioanalytical Chemistry* 2010 399:5 399:1743–1763. <https://doi.org/10.1007/S00216-010-4212-8>
- Gammelgaard B, Jackson MI, Gabel-Jensen C (2010b) Surveying selenium speciation from soil to cell—forms and transformations. *Analytical and Bioanalytical Chemistry* 2010 399:5 399:1743–1763. <https://doi.org/10.1007/S00216-010-4212-8>
- Garg M, Sharma N, Sharma S, Kapoor P, Kumar A, Chunduri V, Arora P (2018) Biofortified Crops Generated by Breeding, Agronomy, and Transgenic Approaches Are Improving Lives of Millions of People around the World. *Front Nutr* 5:. <https://doi.org/10.3389/FNUT.2018.00012/FULL>
- Gigolashvili T, Kopriva S (2014a) Transporters in plant sulfur metabolism. *Front Plant Sci* 5:107212. <https://doi.org/10.3389/FPLS.2014.00442/BIBTEX>
- Gigolashvili T, Kopriva S (2014b) Transporters in plant sulfur metabolism. *Front Plant Sci* 5:442
- Gill SS, Tuteja N (2010) Reactive oxygen species and antioxidant machinery in abiotic stress tolerance in crop plants. *Plant Physiology and Biochemistry* 48:909–930
- Gondwe TM, Alamu EO, Mdziniso P, Maziya-Dixon B (2019) Cowpea (*Vigna unguiculata* (L.) Walp) for food security: an evaluation of end-user traits of improved varieties in Swaziland. *Sci Rep* 9:1–6. <https://doi.org/10.1038/s41598-019-52360-w>
- Gouveia GCC, Galindo FS, Dantas Bereta Lanza MG, Caroline da Rocha Silva A, Pereira de Brito Mateus M, Souza da Silva M, Rimoldi Tavanti RF, Tavanti TR, Lavres J, Reis AR (2020a) Selenium toxicity stress-induced phenotypical, biochemical and physiological responses in rice plants: Characterization of symptoms and plant metabolic adjustment. *Ecotoxicol Environ Saf* 202:110916. <https://doi.org/10.1016/j.ecoenv.2020.110916>
- Gouveia GCC, Galindo FS, Lanza MGDB, Silva AC da R, Mateus MP de B, Silva MS da, Tavanti RFR, Tavanti TR, Lavres J, Reis AR dos (2020b) Selenium toxicity stress-induced phenotypical, biochemical and physiological responses in rice plants: Characterization of

- symptoms and plant metabolic adjustment. *Ecotoxicol Environ Saf* 202:.. <https://doi.org/10.1016/j.ecoenv.2020.110916>
- Guignardi Z, Schiavon M (2017) Biochemistry of Plant Selenium Uptake and Metabolism. 21–34. https://doi.org/10.1007/978-3-319-56249-0_2
- Gupta M, Gupta S (2017a) An overview of selenium uptake, metabolism, and toxicity in plants. *Front Plant Sci* 7:234638. <https://doi.org/10.3389/FPLS.2016.02074/BIBTEX>
- Gupta M, Gupta S (2017b) An overview of selenium uptake, metabolism, and toxicity in plants. *Front Plant Sci* 7:2074. <https://doi.org/10.3389/fpls.2016.02074>
- Gupta M, Gupta S (2017c) An overview of selenium uptake, metabolism, and toxicity in plants. *Front Plant Sci* 7:2074. <https://doi.org/10.3389/fpls.2016.02074>
- Gupta PK, Balyan HS, Sharma S, Kumar R (2021) Biofortification and bioavailability of Zn, Fe and Se in wheat: present status and future prospects. *Theoretical and Applied Genetics* 134:1–35. <https://doi.org/10.1007/S00122-020-03709-7/FIGURES/8>
- Ha HY, Alfulaij N, Berry MJ, Seale LA (2019) From Selenium Absorption to Selenoprotein Degradation. *Biological Trace Element Research* 2019 192:1 192:26–37. <https://doi.org/10.1007/S12011-019-01771-X>
- Hall AE (2012a) Phenotyping cowpeas for adaptation to drought. *Front Physiol* 3 MAY: <https://doi.org/10.3389/FPHYS.2012.00155/FULL>
- Hall AE (2012b) Phenotyping cowpeas for adaptation to drought. *Front Physiol* 3 MAY:155. <https://doi.org/10.3389/fphys.2012.00155>
- Handa N, Kohli SK, Sharma A, Thukral AK, Bhardwaj R, Abd_Allah EF, Alqarawi AA, Ahmad P (2019) Selenium modulates dynamics of antioxidative defence expression, photosynthetic attributes and secondary metabolites to mitigate chromium toxicity in *Brassica juncea* L. plants. *Environ Exp Bot* 161:180–192. <https://doi.org/10.1016/J.ENVEXPBOT.2018.11.009>
- Harvestplus (2021) Biofortification: The Nutrition Revolution is Now. In: [harvestplus.org](https://www.harvestplus.org/biofortification-nutrition-revolution-now). <https://www.harvestplus.org/biofortification-nutrition-revolution-now>. Accessed 10 Feb 2021
- Hawkesford M, Horst W, Kichey T, Lambers H, Schjoerring J, Møller IS, White P (2012) Functions of Macronutrients. In: Petra Marschner (ed) *Marschner's Mineral Nutrition of Higher Plants: Third Edition, 3rd edn*. Elsevier Inc., pp 135–189
- He R, Gao M, Shi R, Song S, Zhang Y, Su W, Liu H (2020) The combination of selenium and LED light quality affects growth and nutritional properties of broccoli sprouts. *Molecules* 25:4788. <https://doi.org/10.3390/molecules25204788>
- Heath RL, Packer L (1968) Photoperoxidation in isolated chloroplasts. I. Kinetics and stoichiometry of fatty acid peroxidation. *Arch Biochem Biophys* 125:189–198. [https://doi.org/10.1016/0003-9861\(68\)90654-1](https://doi.org/10.1016/0003-9861(68)90654-1)

- Hossain A, Skalicky M, Brestic M, et al (2021) Selenium Biofortification: Roles, Mechanisms, Responses and Prospects. *Molecules* 2021, Vol 26, Page 881 26:881. <https://doi.org/10.3390/MOLECULES26040881>
- Huang QQ, Wang Q, Wan YN, Yu Y, Jiang RF, Li HF (2017) Application of X-ray absorption near edge spectroscopy to the study of the effect of sulphur on selenium uptake and assimilation in wheat seedlings. *Biol Plant* 61:726–732. <https://doi.org/10.1007/s10535-016-0698-z>
- Hummel M, Talsma EF, Taleon V, Londoño L, Brychkova G, Gallego S, Raatz B, Spillane C (2020) Iron, Zinc and Phytic Acid Retention of Biofortified, Low Phytic Acid, and Conventional Bean Varieties When Preparing Common Household Recipes. *Nutrients* 2020, Vol 12, Page 658 12:658. <https://doi.org/10.3390/NU12030658>
- Imdad A, Mayo-Wilson E, Haykal MR, Regan A, Sidhu J, Smith A, Bhutta ZA (2022) Vitamin A supplementation for preventing morbidity and mortality in children from six months to five years of age. *Cochrane Database of Systematic Reviews* 2022:. <https://doi.org/10.1002/14651858.CD008524.PUB4/MEDIA/CDSR/CD008524/URN:X-WILEY:14651858:MEDIA:CD008524:CD008524-CMP-001.21>
- Imdad A, Mayo-Wilson E, Herzer K, Bhutta ZA (2017a) Vitamin A supplementation for preventing morbidity and mortality in children from six months to five years of age. *Cochrane Database Syst Rev* 3:. <https://doi.org/10.1002/14651858.CD008524.PUB3>
- Imdad A, Mayo-Wilson E, Herzer K, Bhutta ZA (2017b) Vitamin A supplementation for preventing morbidity and mortality in children from six months to five years of age. *Cochrane Database of Systematic Reviews* 2017:. <https://doi.org/10.1002/14651858.CD008524.pub3>
- Irani S, Todd CD (2016) Ureide metabolism under abiotic stress in *Arabidopsis thaliana*. *J Plant Physiol* 199:87–95. <https://doi.org/10.1016/j.jplph.2016.05.011>
- Ismael MA, Elyamine AM, Moussa MG, Cai M, Zhao X, Hu C (2019) Cadmium in plants: uptake, toxicity, and its interactions with selenium fertilizers. *Metallomics* 11:255–277. <https://doi.org/10.1039/C8MT00247A>
- Jafari S, McClements D (2017) Abordagens nanotecnológicas para aumentar a biodisponibilidade de nutrientes. *Avanços na pesquisa em alimentos e nutrição*
- Jang HS, Lee J, Lee HJ, Park EY (2020) Phytate-mediated phosphorylation of maize, rice, and potato starches at different pH conditions. *Int J Biol Macromol* 165:857–864. <https://doi.org/10.1016/J.IJBIOMAC.2020.09.245>
- Jiang H, Lin W, Jiao H, Liu J, Chan L, Liu X, Wang R, Chen T (2021) Uptake, transport, and metabolism of selenium and its protective effects against toxic metals in plants: a review. *Metallomics* 13:40. <https://doi.org/10.1093/MTOMCS/MFAB040>
- Jiang Y, Schiavon M, Lima LW, Tripti, Jones RR, El Mehdawi AF, Royer S, Zeng Z, Hu Y, Pilon-Smits EAH, Pilon M (2018) Comparison of ATP sulfurylase 2 from selenium hyperaccumulator *Stanleya pinnata* and non-accumulator *Stanleya elata* reveals

- differential intracellular localization and enzyme activity levels. *Biochimica et Biophysica Acta* (BBA) - General Subjects 1862:2363–2371. <https://doi.org/10.1016/J.BBAGEN.2018.03.014>
- Jiang Y, Tang X, Zhou B, Sun T, Chen H, Zhao X, Wang Y (2017) The ROS-mediated pathway coupled with the MAPK-p38 signalling pathway and antioxidant system plays roles in the responses of *Mytilus edulis* haemocytes induced by BDE-47. *Aquatic Toxicology* 187:55–63. <https://doi.org/10.1016/j.aquatox.2017.03.011>
- Jones GD, Droz B, Greve P, Gottschalk P, Poffet D, McGrath SP, Seneviratne SI, Smith P, Winkel LHE (2017a) Selenium deficiency risk predicted to increase under future climate change. *Proc Natl Acad Sci U S A* 114:2848–2853. https://doi.org/10.1073/PNAS.1611576114/SUPPL_FILE/PNAS.201611576SI.PDF
- Jones GD, Droz B, Greve P, Gottschalk P, Poffet D, McGrath SP, Seneviratne SI, Smith P, Winkel LHE (2017b) Selenium deficiency risk predicted to increase under future climate change. *Proc Natl Acad Sci U S A* 114:2848–2853. <https://doi.org/10.1073/pnas.1611576114>
- Joy EJM, Kalimbira AA, Sturgess J, et al (2022) Biofortified Maize Improves Selenium Status of Women and Children in a Rural Community in Malawi: Results of the Addressing Hidden Hunger With Agronomy Randomized Controlled Trial. *Front Nutr* 8:788096. <https://doi.org/10.3389/FNUT.2021.788096/BIBTEX>
- Kato T, Read R, Rozga J, Burk RF (1992) Evidence for intestinal release of absorbed selenium in a form with high hepatic extraction. <https://doi.org/10.1152/ajpgi.1992.262.5.G854> <https://doi.org/10.1152/AJPGI.1992.262.5.G854>
- Kieliszek M (2019) Selenium–Fascinating Microelement, Properties and Sources in Food. *Molecules* 2019, Vol 24, Page 1298 24:1298. <https://doi.org/10.3390/MOLECULES24071298>
- Kieliszek M, Błaż S, Santi C, Bagnoli L (2016) Current Knowledge on the Importance of Selenium in Food for Living Organisms: A Review. *Molecules* 2016, Vol 21, Page 609 21:609. <https://doi.org/10.3390/MOLECULES21050609>
- Kieliszek M, Błażej S, Gientka I, Bzducha-Wróbel A (2015) Accumulation and metabolism of selenium by yeast cells. *Appl Microbiol Biotechnol* 99:5373–5382. <https://doi.org/10.1007/S00253-015-6650-X/FIGURES/5>
- Kolbert Z, Lehotai N, Molnár Á, Feigl G (2016) “The roots” of selenium toxicity: A new concept. *Plant Signal Behav* 11:e1241935. <https://doi.org/10.1080/15592324.2016.1241935>
- Kolbert Z, Molnár, Feigl G, Van Hoewyk D (2019) Plant selenium toxicity: Proteome in the crosshairs. *J Plant Physiol* 232:291–300. <https://doi.org/10.1016/J.JPLPH.2018.11.003>
- König P, Tautenhahn S, Cornelissen JHC, Kattge J, Bönisch G, Römermann C (2018) Advances in flowering phenology across the Northern Hemisphere are explained by functional traits. *Global Ecology and Biogeography* 27:310–321. <https://doi.org/10.1111/GEB.12696>

- Kovalchuk I (2010) Multiple roles of radicals in plants. In: Reactive oxygen species and antioxidants in higher plants
- Kumar P, Kumar S, Tripathi M, ... NM-V, 2013 undefined Flavonoids in the development of functional meat products: A review. cyberleninka.org P Kumar, S Kumar, MK Tripathi, N Mehta, R Ranjan, ZF Bhat, PK Singh Veterinary World, 2013•cyberleninka.org
- Kumar S, Singh A, Dikshit HK, Mishra GP, Aski M, Bansal R, Gupta P (2024) Global Scenario of Biofortification. Biofortification for Nutrient-Rich Crops 26–42. <https://doi.org/10.1201/9781032690636-3>
- Kumar V, Sinha AK, Makkar HPS, Becker K (2010) Dietary roles of phytate and phytase in human nutrition: A review. Food Chem 120:945–959. <https://doi.org/10.1016/J.FOODCHEM.2009.11.052>
- Lacour M, Zunder T, Restle A, Schwarzer G (2004) No evidence for an impact of selenium supplementation on environment associated health disorders – a systematic review. Int J Hyg Environ Health 207:1–13. <https://doi.org/10.1078/1438-4639-00254>
- Langyan S, Yadava P, Khan FN, Bhardwaj R, Tripathi K, Bhardwaj V, Bhardwaj R, Gautam RK, Kumar A (2022) Nutritional and Food Composition Survey of Major Pulses Toward Healthy, Sustainable, and Biofortified Diets. Front Sustain Food Syst 6:878269. <https://doi.org/10.3389/FSUFS.2022.878269/BIBTEX>
- Lanza MGDB, Reis AR dos (2021) Roles of selenium in mineral plant nutrition: ROS scavenging responses against abiotic stresses. Plant Physiology and Biochemistry 164:27–43. <https://doi.org/10.1016/j.plaphy.2021.04.026>
- Lanza MGDB, Silva VM, Montanha GS, Lavres J, Pereira de Carvalho HW, Reis AR dos (2020a) Assessment of selenium spatial distribution using μ -XFR in cowpea (*Vigna unguiculata* (L.) Walp.) plants: Integration of physiological and biochemical responses. Ecotoxicol Environ Saf 207:111216. <https://doi.org/10.1016/j.ecoenv.2020.111216>
- Lanza MGDB, Silva VM, Montanha GS, Lavres J, Pereira de Carvalho HW, Reis AR dos (2020b) Assessment of selenium spatial distribution using μ -XFR in cowpea (*Vigna unguiculata* (L.) Walp.) plants: Integration of physiological and biochemical responses. Ecotoxicol Environ Saf 207:111216. <https://doi.org/10.1016/j.ecoenv.2020.111216>
- Lanza MGDB, Silva VM, Montanha GS, Lavres J, Pereira de Carvalho HW, Reis AR dos (2021) Assessment of selenium spatial distribution using μ -XFR in cowpea (*Vigna unguiculata* (L.) Walp.) plants: Integration of physiological and biochemical responses. Ecotoxicol Environ Saf 207:111216. <https://doi.org/10.1016/j.ecoenv.2020.111216>
- Lara TS, Lessa JH de L, de Souza KRD, Corguinha APB, Martins FAD, Lopes G, Guilherme LRG (2019) Selenium biofortification of wheat grain via foliar application and its effect on plant metabolism. Journal of Food Composition and Analysis 81:10–18. <https://doi.org/10.1016/j.jfca.2019.05.002>

- Lavres J, Reis AR, Rossi ML, Cabral CP, Nogueira N de L, Malavolta E (2010) Changes in the ultrastructure of soybean cultivars in response to manganese supply in solution culture. *Sci Agric* 67:287–294. <https://doi.org/10.1590/S0103-90162010000300006>
- Lazard M, Dauplais M, Blanquet S, Plateau P (2017) Recent advances in the mechanism of selenoamino acids toxicity in eukaryotic cells. *Biomol Concepts* 8:93–104. https://doi.org/10.1515/BMC-2017-0007/ASSET/GRAPHIC/J_BMC-2017-0007_FIG_002.JPG
- Lemire M, Philibert A, Fillion M, Passos CJS, Guimarães JRD, Barbosa F, Mergler D (2012) No evidence of selenosis from a selenium-rich diet in the Brazilian Amazon. *Environ Int* 40:128–136. <https://doi.org/10.1016/J.ENVINT.2011.07.005>
- Lewis J (2019) Codex nutrient reference values. Rome
- Li X, Wu Y, Li B, Yang Y, Yang Y (2018) Selenium accumulation characteristics and biofortification potentiality in turnip (*Brassica rapa* var. *rapa*) supplied with selenite or selenate. *Front Plant Sci* 8:2207. <https://doi.org/10.3389/fpls.2017.02207>
- Liang Y, Li D, Chen Y, Cheng J, Zhao G, Fahima T, Yan J (2020) Selenium mitigates salt-induced oxidative stress in durum wheat (*Triticum durum* Desf.) seedlings by modulating chlorophyll fluorescence, osmolyte accumulation, and antioxidant system. *3 Biotech* 10:1–14. <https://doi.org/10.1007/S13205-020-02358-3/FIGURES/5>
- Lichtenthaler HK, Wellburn AR (1983) Determinations of total carotenoids and chlorophylls a and b of leaf extracts in different solvents. *Biochem Soc Trans* 11:591–592. <https://doi.org/10.1042/bst0110591>
- Liliana García-Bañuelos M, Pedro Sida-Arreola J, Sánchez E, Mónica García-Bañuelos ML, Juan Pedro Sida-Arreola M (2014) Biofortification - promising approach to increasing the content of iron and zinc in staple food crops. *J Elem* 19:. <https://doi.org/10.5601/JELEM.2014.19.3.708>
- Lima LW, Pilon-Smits EAH, Schiavon M (2018a) Mechanisms of selenium hyperaccumulation in plants: A survey of molecular, biochemical and ecological cues. *Biochimica et Biophysica Acta (BBA) - General Subjects* 1862:2343–2353. <https://doi.org/10.1016/J.BBAGEN.2018.03.028>
- Lima LW, Pilon-Smits EAH, Schiavon M (2018b) Mechanisms of selenium hyperaccumulation in plants: A survey of molecular, biochemical and ecological cues. *Biochim Biophys Acta Gen Subj* 1862:2343–2353
- Lindblom SD, Wangeline AL, Valdez Barillas JR, Devilbiss B, Fakra SC, Pilon-Smits EAH (2018) Fungal endophyte *alternaria tenuissima* can affect growth and selenium accumulation in its hyperaccumulator host *astragalus bisulcatus*. *Front Plant Sci* 9:1213. <https://doi.org/10.3389/fpls.2018.01213>
- Liu H, Xiao C, Qiu T, Deng J, Cheng H, Cong X, Cheng S, Rao S, Zhang Y (2022) Selenium Regulates Antioxidant, Photosynthesis, and Cell Permeability in Plants under Various

- Abiotic Stresses: A Review. *Plants* 2023, Vol 12, Page 44 12:44. <https://doi.org/10.3390/PLANTS12010044>
- Liu Q, Zhao X, Ma J, Mu Y, Wang Y, Yang S, Wu Y, Wu F, Zhou Y (2021) Selenium (Se) plays a key role in the biological effects of some viruses: Implications for COVID-19. *Environ Res* 196:110984. <https://doi.org/10.1016/J.ENVRES.2021.110984>
- Loechl CU, Datta-Mitra A, Fenlason L, Green R, Hackl L, Itzkowitz L, Koso-Thomas M, Moorthy D, Owino VO, Pachón H, Stoffel N, Zimmerman MB, Raiten DJ (2023) Approaches to Address the Anemia Challenge. *J Nutr* 153:S42–S59. <https://doi.org/10.1016/J.TJNUT.2023.07.017>
- Luciano De Souza Ferraz R, Soares De Melo A, Fernandes Suassuna J, Barbosa De Brito ME, Dantas Fernandes P, Da Silva E, Júnior N (2012) Trocas gasosas e eficiência fotossintética em ecótipos de feijoeiro cultivados no semiárido. *Pesqui Agropecu Trop* 42:181–188. <https://doi.org/10.1590/S1983-40632012000200010>
- Mafakheri K, Bihamta MR, Abbasi AR (2017) Assessment of genetic diversity in cowpea (*Vigna unguiculata* L.) germplasm using morphological and molecular characterisation. *Cogent Food Agric* 3:1327092. <https://doi.org/10.1080/23311932.2017.1327092>
- Malavolta E, Haag HP, Gargantini H, Garcia Blanco H (1967) Absorção de nutrientes pela cultura do feijoeiro. *Bragantia* 26:381–391. <https://doi.org/10.1590/s0006-87051967000100030>
- Malhotra H, Vandana, Sharma S, Pandey R (2018) Phosphorus nutrition: Plant growth in response to deficiency and excess. In: *Plant Nutrients and Abiotic Stress Tolerance*. Springer Singapore, pp 171–190
- Manaf HH (2016) Beneficial effects of exogenous selenium, glycine betaine and seaweed extract on salt stressed cowpea plant. *Annals of Agricultural Sciences* 61:41–48. <https://doi.org/10.1016/j.aoas.2016.04.003>
- Mangels AR, Moser-Veillon PB, Patterson KY, Veillon C (1990) Selenium utilization during human lactation by use of stable-isotope tracers. *Am J Clin Nutr* 52:621–627. <https://doi.org/10.1093/AJCN/52.4.621>
- Manzeke MG, Mtambanengwe F, Nezomba H, Watts MJ, Broadley MR, Mapfumo P (2017) Zinc fertilization increases productivity and grain nutritional quality of cowpea (*Vigna unguiculata* [L.] Walp.) under integrated soil fertility management. *Field Crops Res* 213:231–244. <https://doi.org/10.1016/j.fcr.2017.08.010>
- Maria Machado Maia F, Tadeu Abreu Oliveira J, Moreira R, Maia FM, Oliveira JT, Matos MR, Moreira RA, Vasconcelos IM (2000) Proximate composition, amino acid content and haemagglutinating and trypsin-inhibiting activities of some Brazilian *Vigna unguiculata* (L) Walp cultivars. Wiley Online Library FMM Maia, JTA Oliveira, MRT Matos, RA Moreira, IM Vasconcelos *Journal of the Science of Food and Agriculture*, 2000•Wiley Online Library. [https://doi.org/10.1002/\(SICI\)1097-0010\(200003\)80:4<453::AID-JSFA548>3.0.CO;2-X](https://doi.org/10.1002/(SICI)1097-0010(200003)80:4<453::AID-JSFA548>3.0.CO;2-X)

- Marinho R de CN, Ferreira L de VM, da Silva AF, Martins LMV, Nóbrega RSA, Fernandes-Júnior PI (2017) Symbiotic and agronomic efficiency of new cowpea rhizobia from Brazilian Semi-Arid. *Bragantia* 76:273–281. <https://doi.org/10.1590/1678-4499.003>
- Marschner P (2011) *Marschner's Mineral Nutrition of Higher Plants: Third Edition*. Elsevier Inc.
- Martineau E, Domec JC, Bosc A, Dannoura M, Gibon Y, Bénard C, Jordan-Meille L (2017) The role of potassium on maize leaf carbon exportation under drought condition. *Acta Physiol Plant* 39:1–13. <https://doi.org/10.1007/S11738-017-2515-5/TABLES/4>
- Martins MDPSC, Lopes AF de S, Jean A, Damasceno-Silva KJ, E Martins M do C de C, Rocha M de M (2023) Characterization of cowpea cultivars for grain size, color, and biofortification. *Revista Caatinga* 36:207–214. <https://doi.org/10.1590/1983-21252023V36N122RC>
- Mateus MP de B, Tavanti RFR, Galindo FS, Silva AC da R, Gouveia GCC, Aparecido CFF, Carr NF, Feitosa YB, Santos EF, Lavres J, Reis AR dos (2020a) *Coffea arabica* seedlings genotypes are tolerant to high induced selenium stress: Evidence from physiological plant responses and antioxidative performance. *Ecotoxicol Environ Saf* 203:111016. <https://doi.org/10.1016/j.ecoenv.2020.111016>
- Mateus MP de B, Tavanti RFR, Galindo FS, Silva AC da R, Gouveia GCC, Aparecido CFF, Carr NF, Feitosa YB, Santos EF, Lavres J, Reis AR dos (2020b) *Coffea arabica* seedlings genotypes are tolerant to high induced selenium stress: Evidence from physiological plant responses and antioxidative performance. *Ecotoxicol Environ Saf* 203:111016. <https://doi.org/10.1016/j.ecoenv.2020.111016>
- Mateus MP de B, Tavanti RFR, Tavanti TR, Santos EF, Jalal A, Reis AR dos (2021a) Selenium biofortification enhances ROS scavenge system increasing yield of coffee plants. *Ecotoxicol Environ Saf* 209:111772. <https://doi.org/10.1016/j.ecoenv.2020.111772>
- Mateus MP de B, Tavanti RFR, Tavanti TR, Santos EF, Jalal A, Reis AR dos (2021b) Selenium biofortification enhances ROS scavenge system increasing yield of coffee plants. *Ecotoxicol Environ Saf* 209:111772. <https://doi.org/10.1016/j.ecoenv.2020.111772>
- Mathur S, Sharma MP, Jajoo A (2018) Improved photosynthetic efficacy of maize (*Zea mays*) plants with arbuscular mycorrhizal fungi (AMF) under high temperature stress. *J Photochem Photobiol B* 180:149–154. <https://doi.org/10.1016/J.JPHOTOBIO.2018.02.002>
- Matos RP, Lima VMP, Windmüller CC, Nascentes CC (2017) Correlation between the natural levels of selenium and soil physicochemical characteristics from the Jequitinhonha Valley (MG), Brazil. *J Geochem Explor* 172:195–202. <https://doi.org/10.1016/j.gexplo.2016.11.001>
- Melo AS de, Costa RR da, Sá FV da S, et al (2024) Modulation of Drought-Induced Stress in Cowpea Genotypes Using Exogenous Salicylic Acid. *Plants* 2024, Vol 13, Page 634 13:634. <https://doi.org/10.3390/PLANTS13050634>

- Melo F de B, Cardoso MJ (2017) Cultivo de Feijão-Caupi: Dados Sistema de Produção Embrapa Meio-Norte, Embrapa Amazônia Ocidental e Embrapa Agrobiologia
- Mendes NAC, Cunha MLO, Bosse MA, Silva VM, Moro AL, Agathokleous E, Vicente EF, Reis AR dos (2023) Physiological and biochemical role of nickel in nodulation and biological nitrogen fixation in *Vigna unguiculata* L. Walp. *Plant Physiology and Biochemistry* 201:107869. <https://doi.org/10.1016/J.PLAPHY.2023.107869>
- Miller G, Suzuki N, Ciftci-Yilmaz S, Mittler R (2010) Reactive oxygen species homeostasis and signalling during drought and salinity stresses. *Plant Cell Environ* 33:453–467. <https://doi.org/10.1111/j.1365-3040.2009.02041.x>
- Mimmo T, Tiziani R, Valentinuzzi F, Lucini L, Nicoletto C, Sambo P, Scampicchio M, Pii Y, Cesco S (2017) Selenium biofortification in *fragaria* × *ananassa*: Implications on strawberry fruits quality, content of bioactive health beneficial compounds and metabolomic profile. *Front Plant Sci* 8:1887. <https://doi.org/10.3389/fpls.2017.01887>
- Mishra GP, Dikshit HK, Priti, Kukreja B, Aski M, Yadava DK, Sarker A, Kumar S (2022) Historical Overview of Biofortification in Crop Plants and Its Implications. *Biofortification of Staple Crops* 31–61. https://doi.org/10.1007/978-981-16-3280-8_2
- Moghaddam A, Heller RA, Sun Q, Seelig J, Cherkezov A, Seibert L, Hackler J, Seemann P, Diegmann J, Pilz M, Bachmann M, Minich WB, Schomburg L (2020) Selenium Deficiency Is Associated with Mortality Risk from COVID-19. *Nutrients* 12:2098. <https://doi.org/10.3390/nu12072098>
- Montanha GS, Rodrigues ES, Marques JPR, De Almeida E, Dos Reis AR, Pereira De Carvalho HW (2020) X-ray fluorescence spectroscopy (XRF) applied to plant science: Challenges towards: In vivo analysis of plants. *Metallomics* 12:183–192. <https://doi.org/10.1039/c9mt00237e>
- Moraes MF, Welch RM, Nutti MR, Carvalho JL V, Watanabe E (2009) Evidences of selenium deficiency in Brazil: from soil to human nutrition. *First International Conference on Selenium in the Environment and Human Health* 73–74
- Moreno-Martin G, Sanz-Landaluze J, León-Gonzalez ME, Madrid Y (2019) In-vivo solid phase microextraction for quantitative analysis of volatile organoselenium compounds in plants. *Anal Chim Acta* 1081:72–80. <https://doi.org/10.1016/J.ACA.2019.06.061>
- Mostofa MG, Ghosh A, Li ZG, Siddiqui MN, Fujita M, Tran LSP (2018) Methylglyoxal – a signaling molecule in plant abiotic stress responses. *Free Radic Biol Med* 122:96–109. <https://doi.org/10.1016/J.FREERADBIOMED.2018.03.009>
- Mostofa MG, Hossain MA, Siddiqui MN, Fujita M, Tran LSP (2017) Phenotypical, physiological and biochemical analyses provide insight into selenium-induced phytotoxicity in rice plants. *Chemosphere* 178:212–223. <https://doi.org/10.1016/j.chemosphere.2017.03.046>

- Muñoz-Amatriaín M, Lo S, Herniter IA, Boukar O, Fatokun C, Carvalho M, Castro I, Guo YN, Huynh BL, Roberts PA, Carnide V, Close TJ (2021) The UCR Minicore: a resource for cowpea research and breeding. *Legume Science* 3:e95. <https://doi.org/10.1002/LEG3.95>
- Muratore C, Espen L, Prinsi B (2021) Nitrogen Uptake in Plants: The Plasma Membrane Root Transport Systems from a Physiological and Proteomic Perspective. *Plants* 2021, Vol 10, Page 681 10:681. <https://doi.org/10.3390/PLANTS10040681>
- Nascimento SP, Bastos EA, Araújo ECE, Freire Filho FR, da Silva EM (2011) Tolerance to water deficit of cowpea genotypes. *Revista Brasileira de Engenharia Agrícola e Ambiental* 15:853–860. <https://doi.org/10.1590/S1415-43662011000800013>
- Nassar M, Nassar R, Maki H, Al-Yagoob A, Hachim M, Senok A, Williams D, Hiraishi N (2021) Phytic Acid: Properties and Potential Applications in Dentistry. *Front Mater* 8:638909. <https://doi.org/10.3389/FMATS.2021.638909/BIBTEX>
- Nassourou MA, Noubissié TJB, Njintang YN, Bell JM (2017) Diallel analyses of soluble sugar content in cowpea (*Vigna unguiculata* L. Walp.). *Crop J* 5:553–559. <https://doi.org/10.1016/J.CJ.2017.05.005>
- Natasha, Shahid M, Niazi NK, Khalid S, Murtaza B, Bibi I, Rashid MI (2018) A critical review of selenium biogeochemical behavior in soil-plant system with an inference to human health. *Environmental Pollution* 234:915–934. <https://doi.org/10.1016/j.envpol.2017.12.019>
- Naz FS, Yusuf M, Khan TA, Fariduddin Q, Ahmad A (2015) Low level of selenium increases the efficacy of 24-epibrassinolide through altered physiological and biochemical traits of *Brassica juncea* plants. *Food Chem* 185:441–448. <https://doi.org/10.1016/j.foodchem.2015.04.016>
- Ngigi PB, Lachat C, Masinde PW, Du Laing G (2019) Agronomic biofortification of maize and beans in Kenya through selenium fertilization. *Environ Geochem Health* 41:2577–2591. <https://doi.org/10.1007/s10653-019-00309-3>
- Nickel A, Kottra G, Schmidt G, Danier J, Hofmann T, Daniel H (2009) Characteristics of transport of selenoamino acids by epithelial amino acid transporters. *Chem Biol Interact* 177:234–241. <https://doi.org/10.1016/J.CBI.2008.09.008>
- Nissar J, Ahad T, Naik HR, Hussain SZ (2017) A review phytic acid: As antinutrient or nutraceutical. *J Pharmacogn Phytochem* 6:1554–1560
- Nkhoma N, Shimelis H, Laing MD, Shayanowako A, Mathew I (2020) Assessing the genetic diversity of cowpea [*Vigna unguiculata* (L.) Walp.] germplasm collections using phenotypic traits and SNP markers. *BMC Genet* 21:1–16. <https://doi.org/10.1186/S12863-020-00914-7/TABLES/9>
- Nutti M, Rocha M, Watanabe E, Carvalho J, Freire Filho F, Silva K (2009) Biofortificação de feijão-caupi no Brasil. In: Embrapa Meio-Norte-Artigo em anais de congresso (ALICE). In: CONGRESSO NACIONAL DE FEIJÃO-CAUPI, 2., 2009, Belém, PA. Da agricultura

- de subsistência ao agronegócio: anais. Belém, PA: Embrapa Amazônia Oriental, 2009. 26–38
- Oliveira A, Fernandes E, Agrícola TR-E, 2005 undefined Condutância estomática como indicador de estresse hídrico em feijão. SciELO BrasilAD Oliveira, EJ Fernandes, TJD RodriguesEngenharia Agrícola, 2005•SciELO Brasil
- Paudel D, Dareus R, Rosenwald J, Muñoz-Amatriaín M, Rios EF (2021) Genome-Wide Association Study Reveals Candidate Genes for Flowering Time in Cowpea (*Vigna unguiculata* [L.] Walp.). *Front Genet* 12:. <https://doi.org/10.3389/FGENE.2021.667038/FULL>
- Pereira EG, Amaral MB, Bucher CA, Santos LA, Fernandes MS, Vieira Rossetto CA (2021) Proline osmopriming improves the root architecture, nitrogen content and growth of rice seedlings. *Biocatal Agric Biotechnol* 33:101998. <https://doi.org/10.1016/J.BCAB.2021.101998>
- Peters KM, Galinn SE, Tsuji PA (2016) Selenium: Dietary Sources, Human Nutritional Requirements and Intake Across Populations. *Selenium: Its Molecular Biology and Role in Human Health, Fourth Edition* 295–305. https://doi.org/10.1007/978-3-319-41283-2_25
- Pilon-Smits EA, Quinn CF, Tapken W, Malagoli M, Schiavon M (2009) Physiological functions of beneficial elements. *Curr Opin Plant Biol* 12:267–274
- Pilon-Smits EAH (2015) Selenium in Plants. 93–107. https://doi.org/10.1007/978-3-319-08807-5_4
- Pilon-Smits EAH (2019a) On the Ecology of Selenium Accumulation in Plants. *Plants* 2019, Vol 8, Page 197 8:197. <https://doi.org/10.3390/PLANTS8070197>
- Pilon-Smits EAH (2019b) On the ecology of selenium accumulation in plants. *Plants* 8:197
- PNUD (2022) Towards 2021/2022 Human Development Report. *Human Development Reports*
- Prasad R, Shivay YS (2020) Agronomic biofortification of plant foods with minerals, vitamins and metabolites with chemical fertilizers and liming. *J Plant Nutr* 43:1534–1554
- Raboy V (2020) Low phytic acid Crops: Observations Based on Four Decades of Research. *Plants* 2020, Vol 9, Page 140 9:140. <https://doi.org/10.3390/PLANTS9020140>
- Raboy V (2003) myo-Inositol-1,2,3,4,5,6-hexakisphosphate. *Phytochemistry* 64:1033–1043. [https://doi.org/10.1016/S0031-9422\(03\)00446-1](https://doi.org/10.1016/S0031-9422(03)00446-1)
- Raggi L, Caproni L, Carboni A, Negri V (2019) Genome-Wide Association Study Reveals Candidate Genes for Flowering Time Variation in Common Bean (*Phaseolus vulgaris* L.). *Front Plant Sci* 10:. <https://doi.org/10.3389/FPLS.2019.00962/FULL>
- Rahman M, Rahman K, Sathi KS, Alam MM, Nahar K, Fujita M, Hasanuzzaman M (2021) Supplemental Selenium and Boron Mitigate Salt-Induced Oxidative Damages in Glycine

- max L. *Plants* 2021, Vol 10, Page 2224 10:2224. <https://doi.org/10.3390/PLANTS10102224>
- Raij B van, Andrade JC, Cantarella H, Quaggio JA (2001a) Análise química para avaliação da fertilidade de solos tropicais
- Raij B van, Andrade JC, Cantarella H, Quaggio JA (2001b) Análise química para avaliação da fertilidade de solos tropicais
- Ramos HMM, Bastos EA, Cardoso MJ, Ribeiro VQ, Do Nascimento FN (2014) Produtividade de grãos verdes do feijão-caupi sob diferentes regimes hídricos. *Engenharia Agrícola* 34:683–694. <https://doi.org/10.1590/S0100-69162014000400008>
- Rayman MP (2012a) Selenium and human health. *The Lancet* 379:1256–1268
- Rayman MP (2012b) Selenium and human health. *The Lancet* 379:1256–1268
- Rayman MP, Duntas LH (2019) Selenium Deficiency and Thyroid Disease. In: *The Thyroid and Its Diseases*. Springer International Publishing, pp 109–126
- Rehman A ur, Masood S, Khan NU, Abbasi ME, Hussain Z, Ali I (2021) Molecular basis of Iron Biofortification in crop plants; A step towards sustainability. *Plant Breeding* 140:12–22. <https://doi.org/10.1111/PBR.12886>
- Reis AR, El-Ramady H, Santos EF, Gratão PL, Schomburg L (2017a) Overview of Selenium Deficiency and Toxicity Worldwide: Affected Areas, Selenium-Related Health Issues, and Case Studies. In: Springer. pp 209–230
- Reis AR, El-Ramady H, Santos EF, Gratão PL, Schomburg L (2017b) Overview of Selenium Deficiency and Toxicity Worldwide: Affected Areas, Selenium-Related Health Issues, and Case Studies. Springer, Cham, pp 209–230
- Reis AR dos, Boleta EHM, Alves CZ, Cotrim MF, Barbosa JZ, Silva VM, Porto RL, Lanza MGDB, Lavres J, Gomes MHF, Carvalho HWP de (2020a) Selenium toxicity in upland field-grown rice: Seed physiology responses and nutrient distribution using the μ -XRF technique. *Ecotoxicol Environ Saf* 190:. <https://doi.org/10.1016/j.ecoenv.2019.110147>
- Reis HPG, Barcelos JP de Q, Junior EF, Santos EF, Silva VM, Moraes MF, Putti FF, Reis AR dos (2018a) Agronomic biofortification of upland rice with selenium and nitrogen and its relation to grain quality. *J Cereal Sci* 79:508–515. <https://doi.org/10.1016/j.jcs.2018.01.004>
- Reis HPG, Barcelos JP de Q, Junior EF, Santos EF, Silva VM, Moraes MF, Putti FF, Reis AR dos (2018b) Agronomic biofortification of upland rice with selenium and nitrogen and its relation to grain quality. *J Cereal Sci* 79:508–515. <https://doi.org/10.1016/j.jcs.2018.01.004>
- Reis HPG, Barcelos JP de Q, Junior EF, Santos EF, Silva VM, Moraes MF, Putti FF, Reis AR dos (2018c) Agronomic biofortification of upland rice with selenium and nitrogen and its relation to grain quality. *J Cereal Sci* 79:508–515. <https://doi.org/10.1016/j.jcs.2018.01.004>

- Reis HPG, de Queiroz Barcelos JP, Silva VM, Santos EF, Tavanti RFR, Putti FF, Young SD, Broadley MR, White PJ, dos Reis AR (2020b) Agronomic biofortification with selenium impacts storage proteins in grains of upland rice. *J Sci Food Agric* 100:1990–1997. <https://doi.org/10.1002/jsfa.10212>
- Research HB-FC, 1999 undefined Economics of enhanced micronutrient density in food staples. ElsevierHE BouisField Crops Research, 1999•Elsevier
- Ribeiro DM, Silva Júnior DD, Cardoso FB, Martins AO, Silva WA, Nascimento VL, Araújo WL (2016) Growth inhibition by selenium is associated with changes in primary metabolism and nutrient levels in *Arabidopsis thaliana*. *Plant Cell Environ* 39:2235–2246. <https://doi.org/10.1111/pce.12783>
- Rivas R, Falcão HM, Ribeiro R V., Machado EC, Pimentel C, Santos MG (2016) Drought tolerance in cowpea species is driven by less sensitivity of leaf gas exchange to water deficit and rapid recovery of photosynthesis after rehydration. *South African Journal of Botany* 103:101–107. <https://doi.org/10.1016/J.SAJB.2015.08.008>
- Ros GH, van Rotterdam AMD, Bussink DW, Bindraban PS (2016) Selenium fertilization strategies for bio-fortification of food: an agro-ecosystem approach. *Plant Soil* 404:99–112. <https://doi.org/10.1007/S11104-016-2830-4/FIGURES/5>
- Rua RM, Nogales F, Carreras O, Ojeda ML (2023) Selenium, selenoproteins and cancer of the thyroid. *Journal of Trace Elements in Medicine and Biology* 76:127115. <https://doi.org/10.1016/J.JTEMB.2022.127115>
- Saltzman A, Birol E, Oparinde A, Andersson MS, Asare-Marfo D, Diressie MT, Gonzalez C, Lividini K, Moursi M, Zeller M (2017) Availability, production, and consumption of crops biofortified by plant breeding: current evidence and future potential. *Ann N Y Acad Sci* 1390:104–114. <https://doi.org/10.1111/NYAS.13314>
- Santos EF, Figueiredo CO, Rocha MAP, Lanza MGDB, Silva VM, Reis AR (2023) Phosphorus and Selenium Interaction Effects on Agronomic Biofortification of Cowpea Plants. *J Soil Sci Plant Nutr* 23:4385–4395. <https://doi.org/10.1007/S42729-023-01357-6/FIGURES/5>
- Santos EF, Kondo Santini JM, Paixão AP, Júnior EF, Lavres J, Campos M, Reis AR dos (2017) Physiological highlights of manganese toxicity symptoms in soybean plants: Mn toxicity responses. *Plant Physiology and Biochemistry* 113:6–19. <https://doi.org/10.1016/j.plaphy.2017.01.022>
- Santos JA da. S, Soares CMG, Correa AM, Teodoro PE, Ribeiro LP, de Abreu HKA (2014) Agronomic performance and genetic dissimilarity among cowpea (*Vigna unguiculata* (L.) Walp.) genotypes. *GLOBAL ADVANCED RESEARCH JOURNAL OF AGRICULTURAL SCIENCE*
- Sarkhel S, Roy A (2022) Phytic acid and its reduction in pulse matrix: Structure–function relationship owing to bioavailability enhancement of micronutrients. *J Food Process Eng* 45:e14030. <https://doi.org/10.1111/JFPE.14030>

- Schiavon M, Lima LW, Jiang Y, Hawkesford MJ (2017) Effects of Selenium on Plant Metabolism and Implications for Crops and Consumers. In: Selenium in plants. Springer, Cham, pp 257–275
- Schiavon M, Nardi S, dalla Vecchia F, Ertani A (2020) Selenium biofortification in the 21st century: status and challenges for healthy human nutrition. *Plant Soil* 453:245–270. <https://doi.org/10.1007/s11104-020-04635-9>
- Schiavon M, Pilon-Smits EAH (2017) The fascinating facets of plant selenium accumulation – biochemistry, physiology, evolution and ecology. *New Phytologist* 213:1582–1596
- Sebetha E, Modi A, Agricultural LO-J of, 2015 undefined (2015) Proteína bruta do feijão-caupi em função do sistema de cultivo, local e adubação nitrogenada. *researchgate.net* ET Sebetha, AT Modi, LG Owoeye *Jornal de Ciências Agrícolas*, 2015•*researchgate.net* 7:. <https://doi.org/10.5539/jas.v7n1p224>
- Seliem MK, Hafez Y, El-Ramady H (2020) Using Nano - Selenium in Reducing the Negative Effects of High Temperature Stress on *Chrysanthemum morifolium* Ramat. *Journal of Sustainable Agricultural Sciences* 46:47–60. <https://doi.org/10.21608/JSAS.2020.23905.1203>
- Selle PH, Cowieson AJ, Cowieson NP, Ravindran V (2012) Protein–phytate interactions in pig and poultry nutrition: a reappraisal. *Nutr Res Rev* 25:1–17. <https://doi.org/10.1017/S0954422411000151>
- Shafiq M, Qadir A, Ahmad SR (2019) Biofortification: A sustainable agronomic strategy to increase selenium content and antioxidant activity in Garlic. *Appl Ecol Environ Res* 17:1685–1704. https://doi.org/10.15666/aeer/1702_16851704
- Shankar AH (2020) Mineral Deficiencies. *Hunter’s Tropical Medicine and Emerging Infectious Diseases* 1048–1054. <https://doi.org/10.1016/B978-0-323-55512-8.00145-9>
- Sharma S, Sharma A, Singh D (2018) Effect of sodium selenate on photosynthetic efficiency, antioxidative defence system and micronutrients in maize (*Zea mays*). *Biologia (Poland)* 73:137–144. <https://doi.org/10.2478/s11756-018-0017-6>
- Shastun S, Chauhan AK, Singh RB, Singh M, Singh RP, Itharat A, Chibisov S, De Meester F, Wilson DW, Halabi G (2016) Can functional food security decrease the epidemic of obesity and metabolic syndrome? A viewpoint. *World Heart J* 8:273–280
- Shini S, Sultan A, Bryden WL (2015) Selenium Biochemistry and Bioavailability: Implications for Animal Agriculture. *Agriculture* 2015, Vol 5, Pages 1277–1288 5:1277–1288. <https://doi.org/10.3390/AGRICULTURE5041277>
- Siddique KHM, Li X, Gruber K (2021) Rediscovering Asia’s forgotten crops to fight chronic and hidden hunger. *Nature Plants* 2021 7:2 7:116–122. <https://doi.org/10.1038/s41477-021-00850-z>
- Silva AC da, Santos D da C, Junior DLT, Silva PB da, Siviero RC dos S and A, Silva AC da, Santos D da C, Junior DLT, Silva PB da, Siviero RC dos S and A (2018a) Cowpea: A

Strategic Legume Species for Food Security and Health. Legume Seed Nutraceutical Research. <https://doi.org/10.5772/INTECHOPEN.79006>

- Silva VM, Boleta EH, Martins JT, dos Santos FLM, da Rocha Silva AC, Alcock TD, Wilson L, de Sá ME, Young SD, Broadley MR, White PJ, dos Reis AR (2019a) Agronomic biofortification of cowpea with selenium: effects of selenate and selenite applications on selenium and phytate concentrations in seeds. *J Sci Food Agric* 99:5969–5983. <https://doi.org/10.1002/jsfa.9872>
- Silva VM, Boleta EH, Martins JT, dos Santos FLM, da Rocha Silva AC, Alcock TD, Wilson L, de Sá ME, Young SD, Broadley MR, White PJ, dos Reis AR (2019b) Agronomic biofortification of cowpea with selenium: effects of selenate and selenite applications on selenium and phytate concentrations in seeds. *J Sci Food Agric* 99:5969–5983. <https://doi.org/10.1002/jsfa.9872>
- Silva VM, Boleta EH, Martins JT, Santos FL, Rocha Silva AC, Alcock TD, Wilson L, Sá ME, Young SD, Broadley MR, White PJ, Reis AR (2019c) Agronomic biofortification of cowpea with selenium: effects of selenate and selenite applications on selenium and phytate concentrations in seeds. *J Sci Food Agric* 99:5969–5983. <https://doi.org/10.1002/jsfa.9872>
- Silva VM, Boleta EHM, Lanza MGDB, Lavres J, Martins JT, Santos EF, dos Santos FLM, Putti FF, Junior EF, White PJ, Broadley MR, Carvalho HWP de, Reis AR dos (2018b) Physiological, biochemical, and ultrastructural characterization of selenium toxicity in cowpea plants. *Environ Exp Bot* 150:172–182. <https://doi.org/10.1016/j.envexpbot.2018.03.020>
- Silva VM, Boleta EHM, Lanza MGDB, Lavres J, Martins JT, Santos EF, dos Santos FLM, Putti FF, Junior EF, White PJ, Broadley MR, Carvalho HWP de, Reis AR dos (2018c) Physiological, biochemical, and ultrastructural characterization of selenium toxicity in cowpea plants. *Environ Exp Bot* 150:172–182. <https://doi.org/10.1016/j.envexpbot.2018.03.020>
- Silva VM, Boleta EHM, Lanza MGDB, Lavres J, Martins JT, Santos EF, dos Santos FLM, Putti FF, Junior EF, White PJ, Broadley MR, Carvalho HWP de, Reis AR dos (2018d) Physiological, biochemical, and ultrastructural characterization of selenium toxicity in cowpea plants. *Environ Exp Bot* 150:172–182. <https://doi.org/10.1016/j.envexpbot.2018.03.020>
- Silva VM, Boleta EHM, Martins JT, dos Santos FLM, da Rocha Silva AC, Alcock TD, Wilson L, de Sá ME, Young SD, Broadley MR, White PJ, dos Reis AR (2019d) Agronomic biofortification of cowpea with selenium: effects of selenate and selenite applications on selenium and phytate concentrations in seeds. *J Sci Food Agric* 99:5969–5983. <https://doi.org/10.1002/JSFA.9872>
- Silva VM, Nardeli AJ, Mendes NAC, Gabriel Filho LRA, Gabriel CPC, Reis AR dos (2023) Does Selenium Application Increase the Yield of Cowpea Plants? Evidence from 29

- Genotypes on Ureides and Sugar Index Affecting the Yield. *J Soil Sci Plant Nutr* 23:5899–5908. <https://doi.org/10.1007/S42729-023-01448-4/METRICS>
- Silva VM, Rimoldi Tavanti RF, Gratão PL, Alcock TD, Reis AR dos (2020a) Selenate and selenite affect photosynthetic pigments and ROS scavenging through distinct mechanisms in cowpea (*Vigna unguiculata* (L.) walp) plants. *Ecotoxicol Environ Saf* 201:110777. <https://doi.org/10.1016/j.ecoenv.2020.110777>
- Silva VM, Rimoldi Tavanti RF, Gratão PL, Alcock TD, Reis AR dos (2020b) Selenate and selenite affect photosynthetic pigments and ROS scavenging through distinct mechanisms in cowpea (*Vigna unguiculata* (L.) walp) plants. *Ecotoxicol Environ Saf* 201:110777. <https://doi.org/10.1016/j.ecoenv.2020.110777>
- Silva VM, Rimoldi Tavanti RF, Gratão PL, Alcock TD, Reis AR dos (2020c) Selenate and selenite affect photosynthetic pigments and ROS scavenging through distinct mechanisms in cowpea (*Vigna unguiculata* (L.) walp) plants. *Ecotoxicol Environ Saf* 201:110777. <https://doi.org/10.1016/j.ecoenv.2020.110777>
- Sim MS, Ogata H, Lubitz W, Adkins JF, Sessions AL, Orphan VJ, McGlynn SE (2019) Role of APS reductase in biogeochemical sulfur isotope fractionation. *Nature Communications* 2019 10:1 10:1–9. <https://doi.org/10.1038/s41467-018-07878-4>
- Simkin AJ (2019) Genetic Engineering for Global Food Security: Photosynthesis and Biofortification. *Plants* 2019, Vol 8, Page 586 8:586. <https://doi.org/10.3390/PLANTS8120586>
- Simpson JL, Bailey LB, Pietrzik K, Shane B, Holzgreve W (2010) Micronutrients and women of reproductive potential: required dietary intake and consequences of dietary deficiency or excess. Part I – Folate, Vitamin B12, Vitamin B6. *The Journal of Maternal-Fetal & Neonatal Medicine* 23:1323–1343. <https://doi.org/10.3109/14767051003678234>
- Sindhu M, Kumar A, Yadav H, Chaudhary D, Jaiwal R, Jaiwal PK (2019) Current advances and future directions in genetic enhancement of a climate resilient food legume crop, cowpea (*Vigna unguiculata* L. Walp.). *Plant Cell Tissue Organ Cult* 139:429–453. <https://doi.org/10.1007/S11240-019-01695-3/TABLES/1>
- Singh B (2020a) Feijão-caupi: a leguminosa alimentar do século 21
- Singh B (2020b) Cowpea: the food legume of the 21st century. John Wiley & Sons
- Singh SK, Raja Reddy K (2011) Regulation of photosynthesis, fluorescence, stomatal conductance and water-use efficiency of cowpea (*Vigna unguiculata* [L.] Walp.) under drought. *J Photochem Photobiol B* 105:40–50. <https://doi.org/10.1016/J.JPHOTOBIO.2011.07.001>
- Sinha R, Kawatra A (2003) Effect of processing on phytic acid and polyphenol contents of cowpeas [*Vigna unguiculata* (L) Walp]. *Plant Foods for Human Nutrition* 58:1–8. <https://doi.org/10.1023/B:QUAL.0000040322.01063.D4/METRICS>

- Sitko K, Opała-Owczarek M, Jemioła G, Gieroń Ż, Szopiński M, Owczarek P, Rudnicka M, Małkowski E (2022) Effect of drought and heavy metal contamination on growth and photosynthesis of silver birch trees growing on post-industrial heaps. *Cells* 11:53. <https://doi.org/10.3390/CELLS11010053/S1>
- Siwela M, Pillay K, Govender L, Lottering S, Mudau FN, Modi AT, Mabhaudhi T (2020) Biofortified Crops for Combating Hidden Hunger in South Africa: Availability, Acceptability, Micronutrient Retention and Bioavailability. *Foods* 2020, Vol 9, Page 815 9:815. <https://doi.org/10.3390/FOODS9060815>
- Sun W, Zhu J, Li S, Tang C, Zhao Q, Zhang J (2020) Selenium supplementation protects against oxidative stress-induced cardiomyocyte cell cycle arrest through activation of PI3K/AKT. *Metallomics* 12:1965–1978. <https://doi.org/10.1039/D0MT00225A>
- Tani LSK, Dennouni-Medjati N, Toubhans B, Charlet L (2020) Selenium deficiency-from soil to thyroid cancer. *Applied Sciences (Switzerland)* 10:. <https://doi.org/10.3390/APP10155368>
- Tavanti TR, Melo AAR de, Moreira LDK, Sanchez DEJ, Silva R dos S, Silva RM da, Reis AR dos (2021a) Micronutrient fertilization enhances ROS scavenging system for alleviation of abiotic stresses in plants. *Plant Physiology and Biochemistry* 160:386–396
- Tavanti TR, Melo AAR de, Moreira LDK, Sanchez DEJ, Silva R dos S, Silva RM da, Reis AR dos (2021b) Micronutrient fertilization enhances ROS scavenging system for alleviation of abiotic stresses in plants. *Plant Physiology and Biochemistry* 160:386–396. <https://doi.org/10.1016/J.PLAPHY.2021.01.040>
- Teka TA, Retta N, Bultosa G, Admassu H, Astatkie T (2020) Protein fractions, in vitro protein digestibility and amino acid composition of select cowpea varieties grown in Ethiopia. *Food Biosci* 36:100634. <https://doi.org/10.1016/j.fbio.2020.100634>
- Thomas CL, Alcock TD, Graham NS, et al (2016a) Root morphology and seed and leaf ionic traits in a *Brassica napus* L. diversity panel show wide phenotypic variation and are characteristic of crop habit. *BMC Plant Biol* 16:1–18. <https://doi.org/10.1186/s12870-016-0902-5>
- Thomas CL, Alcock TD, Graham NS, et al (2016b) Root morphology and seed and leaf ionic traits in a *Brassica napus* L. diversity panel show wide phenotypic variation and are characteristic of crop habit. *BMC Plant Biol* 16:1–18. <https://doi.org/10.1186/s12870-016-0902-5>
- Timpanaro G, Bellia C, Foti VT, Scuderi A (2020) Consumer Behaviour of Purchasing Biofortified Food Products. *Sustainability* 2020, Vol 12, Page 6297 12:6297. <https://doi.org/10.3390/SU12166297>
- Toyota M, Spencer D, Sawai-Toyota S, Jiaqi W, Zhang T, Koo AJ, Howe GA, Gilroy S (2018) Glutamate triggers long-distance, calcium-based plant defense signaling. *Science* (1979) 361:1112–1115. <https://doi.org/10.1126/science.aat7744>

- Tran TAT, Dinh QT, Zhou F, Zhai H, Xue M, Du Z, Bañuelos GS, Liang D (2021) Mechanisms underlying mercury detoxification in soil–plant systems after selenium application: a review. *Environmental Science and Pollution Research* 28:34 28:46852–46876. <https://doi.org/10.1007/S11356-021-15048-1>
- Tripathi AD, Mishra R, Maurya KK, Singh RB, Wilson DW (2018) Estimates for world population and global food availability for global health. In: *The Role of Functional Food Security in Global Health*. Elsevier, pp 3–24
- Tripathi K, Gore PG, Ahlawat SP, Tyagi V, Semwal DP, Gautam NK, Rana JC, Kumar A (2019) Cowpea genetic resources and its utilization: Indian perspective – A review. *Legume Research* 42:437–446
- Turck D, Bohn T, Castenmiller J, et al (2023) Scientific opinion on the tolerable upper intake level for selenium. *EFSA Journal* 21:e07704. <https://doi.org/10.2903/J.EFSA.2023.7704>
- Turrubiates-Hernández FJ, Márquez-Sandoval YF, González-Estevez G, Reyes-Castillo Z, Muñoz-Valle JF (2020) The Relevance of Selenium Status in Rheumatoid Arthritis. *Nutrients* 2020, Vol 12, Page 3007 12:3007. <https://doi.org/10.3390/NU12103007>
- Ulhassan Z, Gill RA, Huang H, Ali S, Mwamba TM, Ali B, Huang Q, Hamid Y, Khan AR, Wang J, Zhou W (2019) Selenium mitigates the chromium toxicity in *Brassica napus* L. by ameliorating nutrients uptake, amino acids metabolism and antioxidant defense system. *Plant Physiology and Biochemistry* 145:142–152. <https://doi.org/10.1016/j.plaphy.2019.10.035>
- US (2000) Dietary Reference Intakes for Vitamin C, Vitamin E, Selenium, and Carotenoids. National Academies Press, Washington, D.C.
- Van Der Straeten D, Bhullar NK, De Steur H, et al (2020) Multiplying the efficiency and impact of biofortification through metabolic engineering. *Nat Commun* 11:1–10. <https://doi.org/10.1038/s41467-020-19020-4>
- Van Handel E (1968) Direct microdetermination of sucrose. *Anal Biochem* 22:280–283. [https://doi.org/10.1016/0003-2697\(68\)90317-5](https://doi.org/10.1016/0003-2697(68)90317-5)
- Vandemark GJ, Grusak MA, McGee RJ (2018) Mineral concentrations of chickpea and lentil cultivars and breeding lines grown in the U.S. Pacific Northwest. *Crop Journal* 6:253–262. <https://doi.org/10.1016/J.CJ.2017.12.003>
- Vasconcelos IM, Maia FMM, Farias DF, Campello CC, Carvalho AFU, de Azevedo Moreira R, de Oliveira JTA (2010) Protein fractions, amino acid composition and antinutritional constituents of high-yielding cowpea cultivars. *Journal of Food Composition and Analysis* 23:54–60. <https://doi.org/10.1016/j.jfca.2009.05.008>
- Veiga N, Macho I, Gómez K, González G, Kremer C, Torres J (2015) Potentiometric and spectroscopic study of the interaction of 3d transition metal ions with inositol hexakisphosphate. *J Mol Struct* 1098:55–65. <https://doi.org/10.1016/J.MOLSTRUC.2015.05.034>

- Victor N. E, Justina Y. T, Sunday A. M, Aderonke I. O (2012) DPPH Radical Scavenging Capacity of Phenolic Extracts from African Yam Bean (*Sphenostylis stenocarpa*). Food Nutr Sci 2012:7–13. <https://doi.org/10.4236/FNS.2012.31002>
- Vinceti M, Filippini T, Wise LA (2018) Environmental Selenium and Human Health: an Update. Curr Environ Health Rep 5:464–485. <https://doi.org/10.1007/s40572-018-0213-0>
- Vogels G, biochemistry CV der D-A, 1970 undefined Análises diferenciais de derivados de glioxilato. ElsevierGD Vogels, C Van der DriftBioquímica analítica, 1970•Elsevier
- Vogels GD, Van Der Drift C (1970) Differential analyses of glyoxylate derivatives. Anal Biochem 33:143–157. [https://doi.org/10.1016/0003-2697\(70\)90448-3](https://doi.org/10.1016/0003-2697(70)90448-3)
- Wakeel A, Farooq M, Bashir K, Ozturk L (2018) Micronutrient Malnutrition and Biofortification: Recent Advances and Future Perspectives. Plant Micronutrient Use Efficiency: Molecular and Genomic Perspectives in Crop Plants 225–243. <https://doi.org/10.1016/B978-0-12-812104-7.00017-4>
- Wang P, Lombi E, Zhao FJ, Kopittke PM (2016) Nanotechnology: A New Opportunity in Plant Sciences. Trends Plant Sci 21:699–712
- Wang R, Guo S (2021) Phytic acid and its interactions: Contributions to protein functionality, food processing, and safety. Compr Rev Food Sci Food Saf 20:2081–2105. <https://doi.org/10.1111/1541-4337.12714>
- Wang Y, Yang XD, Ali A, Lv GH, Long YX, Wang YY, Ma YG, Xu CC (2020) Flowering Phenology Shifts in Response to Functional Traits, Growth Form, and Phylogeny of Woody Species in a Desert Area. Front Plant Sci 11:531690. <https://doi.org/10.3389/FPLS.2020.00536/BIBTEX>
- Wang Y-D, Wang X, Wong Y (2012) Proteomics analysis reveals multiple regulatory mechanisms in response to selenium in rice. J Proteomics 75:1849–1866. <https://doi.org/10.1016/j.jprot.2011.12.030>
- Welch RM (2002) Breeding strategies for biofortified staple plant foods to reduce micronutrient malnutrition globally. Journal of Nutrition 132:. <https://doi.org/10.1093/JN/132.3.495S>
- Wessells KR, Brown KH (2012) Estimating the Global Prevalence of Zinc Deficiency: Results Based on Zinc Availability in National Food Supplies and the Prevalence of Stunting. PLoS One 7:. <https://doi.org/10.1371/journal.pone.0050568>
- White P, phytologist MB-N, 2009 undefined (2009) Biofortificação de culturas com sete elementos minerais frequentemente ausentes na dieta humana - ferro, zinco, cobre, cálcio, magnésio, selênio e iodo. Wiley Online LibraryPJ Branco, MR BroadleyNovo fitologista, 2009•Biblioteca Online Wiley 182:49–84. <https://doi.org/10.1111/j.1469-8137.2008.02738.x>
- White PJ (2016a) Selenium accumulation by plants. Ann Bot 117:217–235
- White PJ (2018a) Selenium metabolism in plants. Biochim Biophys Acta Gen Subj 1862:2333–2342

- White PJ (2016b) Selenium accumulation by plants. *Ann Bot* 117:217–235
- White PJ (2018b) Selenium metabolism in plants. *Biochim Biophys Acta Gen Subj* 1862:2333–2342
- White PJ, Broadley MR (2009) Biofortification of crops with seven mineral elements often lacking in human diets – iron, zinc, copper, calcium, magnesium, selenium and iodine. *New Phytologist* 182:49–84. <https://doi.org/10.1111/J.1469-8137.2008.02738.X>
- Wu H, Zhang X, Giraldo JP, Shabala S (2018) It is not all about sodium: revealing tissue specificity and signalling roles of potassium in plant responses to salt stress. *Plant Soil* 431:1–17
- Xu W, Chai Y-R (2017) Differences in Selenium Uptake, Distribution and Expression of Selenium Metabolism Genes in Tomatoes Article in. *Int J Agric Biol.* <https://doi.org/10.17957/IJAB/15.0330>
- Yadav DN, Bansal S, Tushir S, Kaur J, Sharma K (2020) Advantage of biofortification over fortification technologies. *Wheat and Barley Grain Biofortification* 257–273. <https://doi.org/10.1016/B978-0-12-818444-8.00010-9>
- Yang J, Yu F, Fu Z, Fu Y, Liu S, Chen M, Li Y, Sun Q, Chang H, Zhou W, Wang X, Zhang L (2019) Pathway and driving forces of selenite absorption in wheat leaf blades. *Plant Soil Environ* 65:609–614. <https://doi.org/10.17221/542/2019-PSE>
- Yemm EW, Cocking EC, Ricketts RE (1955) The determination of amino-acids with ninhydrin. *Analyst* 80:209–214. <https://doi.org/10.1039/AN9558000209>
- Yildiztugay E, Ozfidan-Konakci C, Kucukoduk M, Tekis SA (2017) The impact of selenium application on enzymatic and non-enzymatic antioxidant systems in *Zea mays* roots treated with combined osmotic and heat stress. *Arch Agron Soil Sci* 63:261–275. <https://doi.org/10.1080/03650340.2016.1201810>
- Yu Y, Liu Z, Luo LY, Fu PN, Wang Q, Li HF (2019) Selenium Uptake and Biotransformation in Brassica rapa Supplied with Selenite and Selenate: A Hydroponic Work with HPLC Speciation and RNA-Sequencing. *J Agric Food Chem.* https://doi.org/10.1021/ACS.JAFC.9B05359/ASSET/IMAGES/LARGE/JF9B05359_0005.JPEG
- Zahedi SM, Hosseini MS, Daneshvar Hakimi Meybodi N, Teixeira da Silva JA (2019) Foliar application of selenium and nano-selenium affects pomegranate (*Punica granatum* cv. Malase Saveh) fruit yield and quality. *South African Journal of Botany* 124:350–358. <https://doi.org/10.1016/j.sajb.2019.05.019>
- Zhang YY, Stockmann R, Ng K, Ajlouni S (2022) Revisiting phytate-element interactions: implications for iron, zinc and calcium bioavailability, with emphasis on legumes. *Crit Rev Food Sci Nutr* 62:1696–1712. <https://doi.org/10.1080/10408398.2020.1846014>

Zhou B, Cao H, Wu Q, Mao K, Yang X, Su J, Zhang H (2023) Agronomic and Genetic Strategies to Enhance Selenium Accumulation in Crops and Their Influence on Quality. *Foods* 2023, Vol 12, Page 4442 12:4442. <https://doi.org/10.3390/FOODS12244442>