



PROGRAMA DE PÓS-GRADUAÇÃO EM ECOLOGIA, EVOLUÇÃO E
BIODIVERSIDADE

EFEITOS DE UMA EXÓTICA INVASORA AMPLAMENTE DISTRIBUÍDA
(*Leucaena leucocephala*) NAS DINÂMICAS LOCAL E REGIONAL DE
COMUNIDADES EM REGENERAÇÃO

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Dissertação apresentada ao Instituto de Biociências do Campus de Rio Claro, Universidade Estadual Paulista, como parte dos requisitos para obtenção do título de Mestre em Ecologia, Evolução e Biodiversidade.

Orientador: Tadeu de Siqueira Barros

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familiares; aos que ainda vivem
e aos que se foram.

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RESUMO

Invasões biológicas são processos complexos cujos efeitos podem se apresentar em mais de uma escala espacial. A ecologia de metacomunidades fornece ferramentas úteis aos estudos com espécies exóticas invasoras (EEI) pois consideram explicitamente escalas locais e regionais, além dos três componentes da biodiversidade: alfa, beta e gama. A leucena (*Leucaena leucocephala* (Lam) de Wit) é uma importante EEI em florestas sob regeneração natural na Mata Atlântica (Brasil). As invasões de leucena fornecem uma estrutura espacial única e interessante: a espécie forma populações densas e homogêneas, em vez de espalhar indivíduos pelas áreas invadidas. Aqui, aproveitamos essa configuração espacial única formada por fragmentos de floresta nativa, áreas em regeneração e manchas de leucena para investigar como as dinâmicas locais e regionais de (meta-)comunidades vegetais sob regeneração natural são afetadas pela leucena. Realizamos levantamentos florísticos em 131 comunidades distribuídas em 29 metacomunidades de florestas em regeneração que tiveram diferentes idades de invasão por leucena. Encontramos (i) respostas espelhadas entre as escalas local e regional: em ambas as escalas, a riqueza de espécies nativas diminuiu com o aumento da idade de invasão, enquanto a riqueza de EEI aumentou; e (ii) a diversidade- β diminuiu com o aumento da idade de invasão, mostrando que o avanço temporal da invasão levou a um efeito homogeneizador. Também encontramos evidências de que o avanço das manchas de leucena em direção às florestas em regeneração aumenta com o tempo e com a proximidade ao fragmento florestal. Nossos resultados podem ser consequências de efeitos bióticos indiretos e sugerem que os efeitos das invasões de leucena em florestas em regeneração podem ser tão vigorosos que se propagam de escala local para regional, através de diminuição da diversidade- β . Reforçamos a importância de considerar abordagens multiescala na avaliação dos efeitos das invasões biológicas por plantas.

Palavras-chave: espécies invasoras; leucena; metacomunidades; regeneração natural; escalas espaciais.

ABSTRACT

Biological invasions are complex scale-dependent processes. Metacommunity ecology frameworks are useful to understand the effects of invasive non-native species (INNS) because they explicitly consider local and regional scales and the three main components of biodiversity: alpha, beta, and gamma. White-popinac (*Leucaena leucocephala* (Lam) de Wit) is an important INNS in regenerating forests in the Atlantic Forest (Brazil). White-popinac invasions form an interesting, unique spatial structure because the species forms dense and homogeneous patches, instead of spreading individuals throughout invaded areas. Here we took advantage of this unique spatial configuration formed by patches of native forest, regenerating areas, and patches of white-popinac to investigate how local and regional dynamics of plant (meta-)communities under natural regeneration are affected by white-popinac. We conducted floristic surveys in 131 communities distributed among 29 metacommunities of regenerating forests that had different ages of white-popinac invasion. We found (i) mirrored responses among local and regional scales: at both scales, native species richness decreased with the increase of invasion age, whereas INNS richness increased; and (ii) β -diversity decreased with the increase of invasion age, showing that time-advance of invasion led to a homogenizing effect. We also found evidence that the advance of white-popinac patches towards regenerating forests increases with time and proximity to the forest fragment. Our results may be consequences of biotic indirect effects, and they suggest that the effects of white-popinac invasions on regenerating forests can be so vigorous that they propagate from local to regional scale, via decrease in β -diversity. We reinforce the importance of considering multiscale approaches on assessing effects of plant invasions.

Key-words: invasive species; white-popinac; metacommunities; natural regeneration; spatial scales.

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1. INTRODUÇÃO

1.1. Apresentação

Este trabalho buscou responder questões acerca dos efeitos da invasão por leucena (*Leucaena leucocephala* (Lam.) de Wit) em comunidades e metacomunidades vegetais sob regeneração natural da vegetação. O trabalho está dividido em partes que seguem a sequência:

- a) **Introdução geral:** esta introdução geral é constituída por uma revisão bibliográfica, em português, em torno de temas que serão abordados no capítulo, incluindo um espaço para uma escrita com caráter levemente opinativo. É dividida em três seções, sendo:
 - i. uma contextualização teórica sobre invasões biológicas, especialmente em comunidades vegetais, com enfoque nas principais hipóteses e modelos teóricos;
 - ii. uma breve descrição da fitofisionomia de Floresta Estacional Semidecidual na Mata Atlântica do estado de São Paulo, sob um contexto de invasão; e uma lista com todas as espécies de plantas exóticas invasoras que foram registradas nos campos deste projeto de mestrado;
 - iii. uma revisão bibliográfica sucinta sobre a espécie-alvo deste trabalho, a leucena (*Leucaena leucocephala* (Lam.) de Wit), nos âmbitos internacional e brasileiro quanto à invasão por esta espécie.
- b) **Capítulo 1:** este capítulo é o conteúdo principal do trabalho. Está escrito em língua inglesa, em formato de artigo científico, que será submetido à revista *Biological Invasions*. A formatação do artigo segue os padrões exigidos pela revista. O artigo conta com sua própria lista de referências e uma seção de Material Suplementar, que é apresentada no Apêndice A dessa dissertação.
- c) **Considerações finais:** esta seção finaliza o texto e estabelece discussões, em português, sobre os resultados gerais do trabalho e traz reflexões sobre perspectivas futuras no âmbito de produção científica acerca de invasões por leucena.

1.2. Introdução geral

1.2.1 Invasões biológicas – um panorama

Espécies exóticas são espécies que se encontram fora de sua área natural de distribuição geográfica (RICHARDSON et al., 2000b). A categorização de uma espécie como exótica depende de conhecimento taxonômico e biogeográfico prévio, visto que sua área de distribuição natural depende de um contexto geográfico de especiação e história natural (LEVIN, 2003). Quando espécies exóticas são introduzidas por ações humanas – intencionalmente ou não -, e são capazes de estabelecer populações sem manejo humano, podendo causar impactos nos ecossistemas naturais e seminaturais, trata-se de um evento de invasão biológica (BLACKBURN et al., 2011; PYŠEK; HULME, 2005). Assim, nem toda espécie exótica é necessariamente invasora, ainda que muitas delas podem, eventualmente e em contextos específicos, tornarem-se invasoras (REJMANEK; RICHARDSON, 1996; STRAUSS; WEBB; SALAMIN, 2006). O processo de invasão é gradativo, passa por estágios com duração de tempo distinta, e não é necessariamente linear (BLACKBURN et al., 2011; THEOHARIDES; DUKES, 2007).

Os eventos de invasão biológica estão intimamente relacionados às movimentações humanas na paisagem ao longo da História. Graças à maior conexão intercontinental, as invasões aumentaram expressivamente nas últimas décadas (HULME, 2009; KANNAN; SHACKLETON; UMA SHAANKER, 2013; WESTPHAL et al., 2008). Atualmente, as invasões biológicas estão listadas entre os principais drivers de perda de diversidade biológica (DÍAZ et al., 2019)

Invasões biológicas podem alterar processos locais e regionais, e às vezes as consequências são extremamente prejudiciais em termos de conservação e áreas protegidas (SHACKLETON et al., 2020). O processo de invasão geralmente causa diferentes respostas de diversidade dependendo da escala (FRIDLEY et al., 2007; SAX; GAINES, 2008), como modificação da sucessão ecológica e processos ecossistêmicos (WALLER et al., 2020), mudanças na diversidade funcional (RENAULT et al., 2022) e alterações em relações tróficas (HOFFMEISTER et al., 2005). Por consequência, invasões biológicas podem culminar em extinção local e regional de espécies

(CHASE et al., 2020) e aumento da suscetibilidade das comunidades a novos eventos de invasão (CAVIERES, 2021).

As invasões biológicas também podem afetar sistemas de produção agropecuária, e trazendo consequências econômicas severas (HANLEY; ROBERTS, 2019; HOFFMANN; BROADHURST, 2016; LOVELL; STONE; FERNANDEZ, 2006; OLSON, 2006; PIMENTEL; ZUNIGA; MORRISON, 2005). Plantas invasoras podem ocupar áreas de agricultura, transmitindo patógenos e competindo por espaço e nutrientes (MANGLA; INDERJIT; CALLAWAY, 2008; MONTESINOS, 2022; PIMENTEL, 2009; SUN et al., 2020). Neste contexto, é comum o emprego de termos adjacentes, não sinônimos a “plantas invasoras”, como “planta daninha” (equivalente a *weed*, em inglês) ou “praga” (*pest*, em inglês, com algumas diferenças no significado); este último se refere a um espectro mais amplo de formas de vida que ameaçam severamente sistemas de produção agropecuária (BRIGHENTI; OLIVEIRA, 2011; CATFORD; JANSSON; NILSSON, 2009).

Animais invasores podem ser vetores de doenças, inclusive de potencial pandêmico, e podem causar danos à produção pecuária (CROWL et al., 2008; KADING et al., 2018; TORCHIN; MITCHELL, 2004). Os sistemas de aquicultura e pesca também são extremamente afetados por espécies invasoras aquáticas, o que levou inclusive ao estabelecimento de regulações internacionais de prevenção, controle e manejo de invasão (BAILEY, 2015; NIIMI, 2004; VAN DER VEER; NENTWIG, 2015; WERSCHKUN et al., 2014).

No contexto de saúde pública, os efeitos de invasões biológicas podem ser severos e custosos. Diferentes patógenos podem aumentar sua distribuição geográfica por meio de introduções de espécies exóticas (DUNN; HATCHER, 2015; HULME, 2014; PYŠEK; RICHARDSON, 2010). Em um contexto brasileiro, a dengue é um marcante exemplo de doença potencialmente fatal que está intimamente relacionada a eventos de invasão biológica (CARRINGTON et al., 2005; LOURENÇO-DE-OLIVEIRA et al., 2004). Modelos teóricos em biologia das invasões são muito úteis para abordar contextos epidemiológicos, especialmente quando consideram vias de dispersão e uma perspectiva espacial multiescala (ANDERSEN et al., 2004). Alguns autores propõem que considerar doenças infecciosas emergentes como espécies invasoras é indispensável para o estabelecimento de políticas internacionais de combate e prevenção efetivos e duradouros (OGDEN et al., 2019). Esta proposta integra o conceito interdisciplinar e unificado de One Biosecurity (HULME, 2021).

As comunidades tradicionais (no Brasil, sendo representadas pelas comunidades indígenas, ribeirinhas, caiçaras, quilombolas, entre outras estabelecidas pelo Decreto nº 8750/2016) constituem um grupo especialmente vulnerável aos efeitos de invasões biológicas (PEJCHAR; MOONEY, 2009; REO et al., 2017), embora haja escassa produção científica neste tema no Brasil. Alterações em serviços ecossistêmicos e disseminação de patógenos são causas principais de consequência socioeconômica que afligem comunidades tradicionais afetadas por espécies invasoras (LINDERS et al., 2020; MCNEELY, 2001). É indiscutível a importância da inclusão destas comunidades como partes interessadas em manejo de espécies invasoras, principalmente em espaços territoriais especialmente protegidos (PEREIRA; SCARDUA, 2008). Apesar dos potenciais problemas concorrentes às invasões biológicas, é importante destacar que, sob pontos de vista etnográficos, algumas comunidades tradicionais consideram espécies invasoras sob outras perspectivas contexto-específicas, disjuntas à dualidade “nativa ou exótica” (HELMREICH, 2005; REO; OGDEN, 2018). No cenário brasileiro, a relação entre visões etnográficas e invasões biológicas é ainda escassamente estudado e há uma importante lacuna de conhecimento neste tema.

1.2.2.1 Terminologia

Em português, o termo mais comum para se referir às espécies invasoras é **espécie exótica invasora** (EEI; LEÃO et al. 2011). O termo **espécie invasora**, sem o adjetivo “exótica” tem significado equivalente, mas abre brechas para o debate de se espécies nativas podem ser consideradas invasoras (VALÉRY et al., 2009). Alguns autores defendem que uma espécie nativa com ampla distribuição, forte dispersão, e que se sobressaia em abundância e/ou densidade em relação às outras da comunidade, causando alterações em diversidade e estrutura dos ecossistemas, deve ser chamada de **superdominante** (PIVELLO et al., 2018). Este debate é intenso e, por esta razão, escolhemos usar o termo espécie exótica invasora (EEI) ao longo do trabalho.

Se em português já temos um termo relativamente consagrado, a terminologia em inglês é mais ampla: *exotic invasive species*, *invasive alien species*, *non-indigenous invasive species* e *invasive non-native species* são exemplos ocorrentes em órgãos de conservação da natureza e em bibliografia científica (EHRENFELD, 2008;

HUGHES et al., 2020; RICHARDSON; REJMÁNEK, 2011; SOUSA et al., 2008). O termo ***invasive non-native species*** (INNS) parece ter se consagrado nos últimos anos. Colautti e MacIsaac (2004) propuseram um *framework* que buscou resolver impasses em terminologia, considerando diferentes estágios de invasão. Os autores discordam que uma abordagem geográfica ao invés de taxonômica resolveria a maior parte das diferenças em escolha de termos para futuros estudos. Ainda assim, todos os termos previamente citados continuam a aparecer em estudos atuais.

Outros termos por vezes intercambiáveis em invasões biológicas são “planta daninha”, “erva daninha” (para plantas não lenhosas); “praga agrícola” e “invasora de culturas” (ALCÂNTARA; NÓBREGA; FERREIRA, 2007; GAZZIERO et al., 2015; POLANCZYK et al., 2006). Vale ressaltar que estes termos se referem a um contexto de produção agrícola, sob um viés econômico e produtivo. Apesar de o senso comum considerar estes termos como equivalentes às espécies exóticas invasoras, principalmente por algumas EEI serem também consideradas plantas daninhas, as terminologias partem de diferentes perspectivas. Especialmente o termo “invasora de culturas”, que parte de um contexto agrícola, pode ser confundido com o conceito de EEI, que parte de um contexto geográfico/taxonômico.

Em inglês, há possibilidade de “planta daninha” ser traduzida como *weed*. Esta tradução pode não ser aplicável em todos contextos. Os termos *pest* ou *noxious* podem ser traduzidos – com ressalvas - como “praga”. Catford e colaboradores (2009) propõem delimitações para os termos “*introduced*”, “*weed*”, “*naturalized*”, “*invasive*” e “*noxious/pest*”, baseadas em estágios de invasão. Apesar de oportuna, essa proposta não foi integralmente adotada pela comunidade científica, e outros autores atribuem distintas nomenclaturas (BLACKBURN et al., 2011; COLAUTTI; MACISAAC, 2004; SAX; GAINES, 2008; SUN et al., 2020).

A denominação **espécie naturalizada** é essencialmente muito problemática. O termo pode se referir a estágios iniciais (RICHARDSON et al., 2000b) ou intermediários (CATFORD; JANSSEN; NILSSON, 2009) de invasão. Talvez o uso mais comum seja para referir a espécies que, apesar de capazes de se dispersarem sem intermédio humano, supostamente não interferem na biodiversidade nativa (ESSL et al., 2019; GALLAGHER; RANDALL; LEISHMAN, 2015; RICHARDSON et al., 2000b). A problemática da terminologia espécie naturalizada é justamente a sua inespecificidade. Espécie naturalizada aproxima-se ao o conceito de nativa (“tão natural que é quase nativa”), criando uma falsa ideia de que a espécie não é invasora ou potencialmente

invasora. Esta terminologia incerta é inclusive adotada pelo Jardim Botânico do Rio de Janeiro (JBRJ, 2023). Este caráter impreciso pode, inclusive, afetar decisões na esfera de políticas públicas.

Uma espécie exótica pode apresentar características ecológicas que a favoreçam num contexto de invasão (REJMANEK; RICHARDSON, 1996). Fatores como plasticidade fenotípica, altas taxas de reprodução, crescimento acelerado e resistência a patógenos podem facilitar o estabelecimento e dispersão da espécie exótica, agora sob a denominação de espécie exótica invasora (LOCKWOOD; CASSEY; BLACKBURN, 2005; RICHARDSON et al., 2000b; WALLER et al., 2020). A este conjunto de características inatas à espécie, damos o nome de **potencial de invasão**. Em inglês, existe o termo *invasiveness* (HUI et al., 2016; REJMANEK; RICHARDSON, 1996), que poderia ser cruamente traduzido para algo como “invasividade”, mas esta palavra não existe em português. O termo potencial de invasão cumpre esta definição e é um conceito amplamente empregado em português (ATTIAS; SIQUEIRA; BERGALLO, 2013; ZIMMERMANN, 2016).

As comunidades ou ecossistemas invadidos apresentam características intrínsecas que os conferem maior ou menor resistência à invasão (LONSDALE, 1999). Uma série de condições abióticas compõem este escopo, juntamente com elementos bióticos como presença de predadores (e.g. DERIVERA et al., 2005), competidores (e.g. CAVIERES, 2021), disponibilidade de recursos e habitat (e.g. HIERRO; MARON; CALLAWAY, 2005), patógenos (e.g. BLUMENTHAL et al., 2009), dispersores (e.g. NORTHFIELD et al., 2018) e mutualismos (e.g. MOLES et al., 2022). Este conjunto de condições das comunidades e ecossistemas passíveis de serem invadidos é definido como **suscetibilidade à invasão**. Assim como o potencial de invasão, este termo também tem um equivalente em inglês, que não existe em português. O termo *invasibility* é frequentemente empregado neste âmbito, e em tradução livre equivaleria a “invasibilidade” (CAVIERES, 2021; HOWETH, 2017; HUI et al., 2016; RENAULT et al., 2022).

Outro conceito essencial no estudo de invasões biológicas é **pressão de propágulos** (*propagule pressure*). Propágulos, no contexto de invasão biológica, são unidades de reprodução de espécies (como sementes, esporos, ou partes vegetativas com potencial reprodutivo) que chegam a um local em cada evento de invasão. A pressão de propágulos compreende o número de propágulos em cada evento de invasão (*propagule size*) e a quantidade de eventos de invasão (*propagule number*; LOCKWOOD;

CASSEY; BLACKBURN, 2005). Desta maneira, alta pressão de propágulos está associada a maior potencial de invasão (KING; HOWETH, 2019; LONSDALE, 1999; PYŠEK; RICHARDSON, 2006). Alguns autores consideram ainda o número de populações-fonte e os padrões espaciais e temporais da chegada de propágulos como partes também componentes do conceito de pressão de propágulos (DE JONG; FOWLER, 2018; SIMBERLOFF, 2009). Ademais, alguns autores também propõem que pressão de propágulos pode funcionar como modelo nulo em invasões biológicas (CASSEY et al., 2018; COLAUTTI; GRIGOROVICH; MACISAAC, 2006).

1.2.2.2 Principais hipóteses

A ecologia de invasões é historicamente marcada por uma abundância de hipóteses e modelos teóricos que, se por um lado representam avanço científico, por outro mostram que há forte redundância em terminologia. Muitos autores propõem termos caso-específicos, com baixo poder de generalização, e com pouca comunicabilidade com estudos prévios. Esta crítica à ecologia de invasões é recorrente, e muitos autores propuseram *frameworks* que visam conciliar hipóteses semelhantes, e padronizar os diferentes termos empregados para explicar os mesmos mecanismos (e.g. ALPERT, 2006; BLACKBURN et al., 2011; BROWN; BARNEY, 2021; CATFORD; JANSSON; NILSSON, 2009; FOXCROFT; PICKETT; CADENASSO, 2011; PERKINS; LEGER; NOWAK, 2011). Nesta seção, abordaremos hipóteses marcantes e mais frequentemente abordadas em estudos de invasão, que estão mais diretamente relacionadas a este trabalho. Todas elas compartilham, em diferentes proporções, componentes bióticos, abióticos e pressão de propágulos (BLACKBURN et al., 2011). Existem muitas outras hipóteses que não serão abordadas aqui (ver CATFORD; JANSSON; NILSSON, 2009).

1.2.2.2.1 Global competition

A hipótese de **global competition** se baseia em pressão de propágulos, mas não é espécie-específica. Propõe que quanto mais espécies exóticas são introduzidas em um local, maiores são as chances de alguma dessas espécies superarem as barreiras impostas pela comunidade nativa (ALPERT, 2006; COLAUTTI; MACISAAC, 2004).

Em essência, este modelo é completamente baseado em uma abordagem de invasão por plantas, proposta por Richardson e colaboradores (2000), onde a invasão é definida pelas barreiras as quais uma espécie invasora supera. Talvez a hipótese de *global competition* seja aplicável a modelos de invasão por animais, baseados em Williamson (1996), mas algumas adaptações conceituais seriam necessárias. Reconhecemos que esta hipótese compartilha, indiretamente, algumas semelhanças com dinâmicas neutras em teoria de metacomunidades (HUBBELL, 2005; THOMPSON et al., 2020) e teoria de biogeografia de ilhas (MACARTHUR; WILSON, 1967).

1.2.2.2.2. Ideal weed

A hipótese de ***ideal weed*** considera história natural e atributos de uma EEI como os principais fatores que facilitam sua invasão (BAKER, 1965; SUTHERLAND, 2004). São exemplos de fatores relacionados à uma *invasive weed*: alta plasticidade fenotípica e genotípica, reprodução precoce e com muitos propágulos, crescimento acelerado, generalismo de habitat e resistência a patógenos (LOCKWOOD; CASSEY; BLACKBURN, 2005; REJMANEK; RICHARDSON, 1996; RICHARDSON et al., 2000b; SULTAN; MATESANZ, 2015; WALLER et al., 2020). O papel dos mutualismos (RICHARDSON et al., 2000a) na hipótese de *ideal weed* é debatido, haja vista evidências recentes têm mostrado que algumas EEI com forte potencial de invasão (*ideal weeds*, per se) são altamente dependentes de relações mutualísticas (MOYANO; RODRIGUEZ-CABAL; NUÑEZ, 2020, 2021).

1.2.2.2.3 Enemy-release

Outra relevante hipótese, certamente entre as mais testadas, é a hipótese de ***enemy release*** (KEANE; CRAWLEY, 2002). Baseia-se na ideia de que, ao invadir uma comunidade, a EEI “é liberada” (*released*) de seus inimigos naturais (predadores, herbívoros, patógenos), os quais ocorrem em sua área de distribuição geográfica original (COLAUTTI et al., 2004; LIU; STILING, 2006; MEIJER et al., 2016). A ausência de inimigos que limitam o crescimento populacional da EEI na área invadida faz com que a população cresça livremente. Alguns autores atribuem a esse tipo de liberação o nome “regulatório” (*regulatory enemy release*; COLAUTTI et al., 2004; KEANE;

CRAWLEY, 2002). A ausência de inimigos também permite com que a população invasora reorganize recursos e energia que outrora seriam usados para sua defesa, e agora estão disponíveis para maior investimento em reprodução e crescimento. Este efeito é tido como “compensatório” (*compensatory enemy release*; LU; DING, 2012; ROGERS; SIEMANN, 2005).

Gruntman et al. (2017) apontam que estudos com esta hipótese devem explicitamente incluir escala temporal, visto que, sob certas circunstâncias, existe uma recuperação de inimigos ao longo do tempo, e os efeitos de *enemy release* diminuem. Existem hipóteses correlatas a esta, com algumas variações: *enemy reduction* (COLAUTTI; MACISAAC, 2004), *evolution of increased competitive ability* (BLOSSEY; NOTZOLD, 1995; JOSHI; VRIELING, 2005) e *resource-enemy release* (BLUMENTHAL et al., 2009; BLUMENTHAL, 2006).

1.2.2.2.4 Biotic resistance e Missed mutualisms

Biotic resistance é uma hipótese que se refere às condições de uma comunidade que a conferem menor suscetibilidade à invasão. Competidores nativos, herbívoros e patógenos limitam colonização e avanço de invasão, uma vez que a EEI não é adaptada às novas interações competitivas que estabelece com a comunidade nativa (ALPERT, 2006; BEAURY et al., 2020; GRUTTERS et al., 2017; LEVINE; ADLER; YELENIK, 2004; PARKER; HAY, 2005). Enquanto *biotic resistance* se refere à menor suscetibilidade de uma comunidade à invasão, a hipótese de **missed mutualisms** aborda a redução do potencial de invasão de uma EEI. Esta hipótese postula que certas espécies perdem relações mutualísticas importantes ao expandirem sua distribuição geográfica para novas áreas, e por isso têm seu potencial de invasão reduzido (ALPERT, 2006; LOPEZ et al., 2021; MITCHELL et al., 2006; MOLES et al., 2022). A ocorrência concomitante de *biotic resistance* e *missed mutualisms* é um tópico ligeiramente discutido (MOYANO et al., 2019).

1.2.2.2.5 Limiting similarity e Empty niche

A hipótese de **limiting similarity** considera que EEI's com sucesso na invasão são funcionalmente distintas das espécies na comunidade invadida (ABRAMS, 1983;

CALLAWAY; RIDENOUR, 2004; MACARTHUR; LEVINS, 1967). De certa maneira, é o inverso de *biotic resistance* (CATFORD; JANSSON; NILSSON, 2009), pois considera que, sendo funcionalmente distintas da espécies nativas, as EEI's têm vantagem na invasão, e não uma redução do potencial competitivo (mecanismo proposto em *biotic resistance*). É uma hipótese antiga e foi inicialmente abordada por Darwin, como um componente de sua hipótese de naturalização (DARWIN, 1859). Também é uma hipótese muito próxima à de ***empty niche***, com a diferença de que *empty niche* explicitamente considera o tamanho do *pool* de espécies nativas como componente da resistência à invasão (ELTON, 1958; HIERRO; MARON; CALLAWAY, 2005; MACARTHUR, 1970). Em *empty niche*, comunidades nativas com baixo *pool* de espécies são possivelmente “insaturadas”, ou seja, há espaço e “nichos vazios” a serem ocupados pela EEI.

Ambas hipóteses são relevantes e explicam processos fundamentais em invasões. Contudo, o papel de disponibilidade de nichos para sucesso de EEI's parece ter sido superestimada historicamente, e evidências recentes sugerem que mesmo EEI funcionalmente semelhantes às nativas, e em comunidades “saturadas”, podem ter sucesso da invasão, por exemplo, ao estabelecer novas relações positivas e de facilitação com outras EEI ou com as próprias espécies nativas. Evidências recentes apontam que a hipótese de *limiting similarity* é (i) mais corroborada em ecossistemas com maior disponibilidade de recursos (EL-BAROUGY et al., 2020); e (ii) não tem relação obrigatória com proximidade filogenética (FRITSCHIE et al., 2014). Apesar de ser uma das primeiras hipóteses em invasões biológicas, há um montante significativo de estudos que encontraram pouca ou nenhuma evidência de sua existência em comunidades vegetais (EMERY, 2007; HESS et al., 2020; PRICE; PÄRTEL, 2013; YANNELLI et al., 2017). A hipótese de *empty niche*, por sua vez, parece ser desafiada em casos de espécies lenhosas invasoras capazes de invadir florestas em estágio avançado de sucessão (FRIDLEY et al., 2023).

1.2.2.2.6 *Biotic indirect effects e Spillover effects*

A hipótese de ***biotic indirect effects*** contempla uma série de mecanismos aos quais uma comunidade invadida é submetida, cujos efeitos se dão por processos indiretos (por exemplo, como “a” afeta “b”, que por sua vez afeta “c” (CATFORD; JANSSON; NILSSON, 2009; MARLER; ZABINSKI; CALLAWAY, 1999; STRAUSS,

1991; WHITE; WILSON; CLARKE, 2006). Exemplos de mecanismos com resultados indiretos são competição aparente (e.g. BONSALL; HASSELL, 1997; HOLT; BONSALL, 2017), mutualismo e comensalismo indiretos (e.g. FATH, 2007; SANDERS; VAN VEEN, 2012), cascatas tróficas (e.g. KIMBRO et al., 2009) e competição indireta (*exploitative competition*; e.g. DAMAS-MOREIRA et al., 2020).

Um outro termo recentemente importado para o contexto de invasões biológicas é o ***spillover effect*** (ALTIERI; IRVING, 2017; FENICHEL; RICHARDS; SHANAFELT, 2014; GOEDKNEGT et al., 2017; WELLS et al., 2015). Os *spillovers* são processos em que um dado processo rompe limitações espaciais/temporais e “transborda” seu efeito a outros elementos, de maneira indireta (JACOBSON, 2014; KUSCHNIG et al., 2021; WANG et al., 2021). Para invasões biológicas, acreditamos que *spillovers* espaciais (*spatial spillovers*) podem ser interpretados como uma escala mais ampla (por exemplo, escala de metacomunidade) de *biotic indirect effects*. Esta interpretação parece já ter sido feita em dinâmicas espaço-temporais de patógenos, mas de maneira indireta, não explicitamente comparando *biotic indirect effects* com *spillover effects* (ALLEN et al., 2018; DILLON; MEENTEMEYER, 2019; DUNN et al., 2012).

1.2.2.2.7 Invasional meltdown

Invasional meltdown é uma hipótese complexa que aborda “efeitos-dominó” em invasões (CATFORD; JANSSON; NILSSON, 2009). A partir de interações simbióticas ou de facilitação, tanto direta quanto indiretamente, EEI’s podem aumentar a suscetibilidade de uma comunidade à invasão (MACK, 2003; RICCIARDI, 2001; SIMBERLOFF; VON HOLLE, 1999). O aumento na suscetibilidade à invasão pode ser ocasionado, por exemplo, por alterações em condições microclimáticas (e.g. LEMBRECHTS et al., 2018), de microbiota do solo (e.g. CHEN; VAN KLEUNEN, 2022; ZHANG et al., 2020), e de regimes de fogo (e.g. D’ANTONIO; HUGHES; TUNISON, 2011; SIMBERLOFF; VON HOLLE, 1999), criando condições adequadas para novos eventos de invasão. Essa interpretação parece ser especialmente apropriada em ecossistemas fragmentados (CACABELOS et al., 2022; GIBSON et al., 2013; WADDELL et al., 2020).

1.2.2 Florestas Estacionais Semidecíduais (FES's) – um cenário de invasão

Florestas Estacionais neotropicais ocorrem da América Central à Argentina, em solos férteis, e são marcadas por uma estação seca bem definida (LIMA et al., 2018). No Brasil, Florestas Estacionais ocorrem, em sua maioria, nas faixas de transição entre florestas ombrófilas e ecossistemas mais abertos, como savanas e estepes. São florestas com pluviosidade anual inferior a 1.100 mm (outros autores consideram valores superiores (e.g. OLIVEIRA-FILHO; FONTES, 2000)), irregularmente distribuída ao longo do ano, visto que, por no mínimo cinco meses, a precipitação mensal é inferior a 100 mm. Nesses períodos de seca, pode ocorrer caducidade foliar (IBGE, 2012; LIMA et al., 2018; OLIVEIRA-FILHO; JARENKOW; RODAL, 2006).

A terminologia em inglês é um desafio: há pouca padronização, e conflitos entre classificações brasileiras e internacionais. Aqui, apresentamos resumidamente os principais termos usados para referir Florestas Estacionais em inglês: *Seasonal forests* (e.g. ZAK; CABIDO; HODGSON, 2004), *Stational forests* (e.g. FIRMINO et al., 2021), *Seasonally dry forests* (e.g. PENNINGTON; PRADO; PENDRY, 2000), *Dry forests* (sensu lato, e.g. RIVAS et al., 2020). Todos estes termos são usados de maneira intercambiável e por vezes considerados sinônimos. Alguns deles, como “*stational forests*”, parecem estar associados a inadequações em tradução de português para inglês. Em português, há divergências entre autores, mas, desde 2012, o Manual Técnico da Vegetação Brasileira, elaborado pelo IBGE, trouxe padronização que passou a ser convenientemente adotada em esferas de políticas públicas e planejamento territorial do meio ambiente (IBGE, 2012). Este manual divide as Florestas Estacionais em três grandes grupos, que são divididos com base em caducidade foliar na estação seca. São eles:

- (i) Floresta Estacional Perenifólia (ou Sempre-Verde): geralmente ocorre em transição com florestas ombrófilas, e apresenta menos de 20% das espécies arbóreas do estrato superior com caducidade foliar na seca. Apesar de haver estação seca definida, os níveis de umidade no solo não diminuem de maneira tão intensa ao ponto de ocasionar expressiva queda de folhas.
- (ii) Floresta Estacional Semidecidual: apresenta de 20 a 50% das árvores do estrato superior com caducidade foliar. Pode ser confundida com savanas florestadas, principalmente em áreas de transição Mata Atlântica-Cerrado.

Este tipo de floresta será abordado ao longo do texto, e é o sistema no qual nossos estudos foram conduzidos.

- (iii) Floresta Estacional Decidual: apresenta mais de 50% de árvores caducifólias, e tendem a se formar em solos mais rasos e com menor capacidade de retenção hídrica. Pode ser confundida com Caatinga.

As Florestas Estacionais Semidecíduais (FES's), focos do nosso trabalho, têm sua maior extensão concentrada nas regiões Sul e Sudeste do Brasil, em áreas de domínio da Mata Atlântica. Tem ocorrência pontual em domínio Amazônico, e no Cerrado ocorre de maneira disjunta em matas ciliares e de galeria (IBGE, 2012). Em campo, FES's podem ser confundidas com savanas florestadas (Cerradão), por similaridades em altura de dossel, sazonalidade e compartilhamento de muitas espécies vegetais. Todavia, savanas florestadas ocorrem em solos muito permeáveis, e apresentam dossel consideravelmente descontínuo, com áreas de abertura de dossel associadas a espécies graminoides (BUISSON et al., 2019; MIATTO; WRIGHT; BATALHA, 2016). Isso não ocorre naturalmente em áreas de FES's, cujo estrato herbáceo é mais sombreado e com maior cobertura por serapilheira (SOUZA et al., 2019).

No domínio de Mata Atlântica, as FES's estão entre as fitofisionomias com maior fragmentação (REZENDE et al., 2018). Uma vez que se encontram em terrenos com menor declive quando comparadas às Florestas Ombrófilas serranas, as FES's sofreram forte desflorestamento por expansão urbana e agropecuária. Atualmente, a cobertura de FES's nas regiões sul e sudeste é essencialmente descontínua, restrita a fragmentos isolados ou com baixa conectividade, e localizados principalmente em áreas de APP nas margens de corpos d'água (IBGE, 2012; REZENDE et al., 2018; RIBEIRO et al., 2011).

Segundo o Mapeamento da Cobertura Vegetal Nativa do Estado de São Paulo de 2020 (IF, 2020), estágios médios e avançados de Floresta Estacional Semidecidual representam em torno de 30% do total de cobertura vegetal atual no estado, e ocupam 7% de seu território. Em sua maioria, são fragmentos de floresta secundária, em processo de regeneração. Este cenário de fragmentação traz consequências negativas severas como aumento dos efeitos de borda (e.g. BANKS-LEITE; EWERS; METZGER, 2010), diminuição da conectividade na paisagem (e.g. PEREIRA et al., 2022), erosão de biodiversidade (e.g. DE LIMA et al., 2020), maior suscetibilidade ao fogo (e.g. SINGH; HUANG, 2022), e repetidos eventos de invasão biológicas (e.g. PIVELLO et al., 2018).

1.2.2.1 *Plantas exóticas invasoras em FES's sob regeneração natural no interior do estado de São Paulo*

As FES's do estado de São Paulo sofrem historicamente com introdução de espécies exóticas e com invasão biológica (CALABONI et al., 2018; ZENNI; ZILLER, 2011). Espécies dos gêneros *Urochloa* (gênero atual da conhecida *Brachiaria* spp.) e *Megathyrsus* (gênero atual de *Panicum* spp.) se destacam como invasoras de áreas em regeneração de FES no estado (SOBANSKI; MARQUES, 2014; ZWIENER et al., 2014). FES's em estado avançado de sucessão também não são imunes às plantas invasoras (ver DECHOUM et al. (2015) para exemplo em floresta estacional em Santa Catarina). O número de espécies de plantas exóticas invasoras que ocorrem nessas áreas de FES's regenerantes é, por vezes subestimado (ZENNI; ZILLER, 2011).

Esta subseção dedica-se a apresentar todas as espécies de plantas exóticas invasoras e potencialmente invasoras que foram registradas ao longo dos trabalhos de campo neste projeto de mestrado. O objetivo é, exclusivamente, fornecer dados de registro de plantas exóticas invasoras em áreas de regeneração natural de Florestas Estacionais Semideciduais. Nem todas as espécies desta lista compuseram o levantamento florístico abordado no capítulo 1. Uma vez que não está diretamente relacionada às perguntas a serem respondidas neste trabalho, trouxemos essa lista como parte da introdução geral. Todas as espécies aqui citadas poderiam ter sido nossa espécie-alvo. A escolha da *Leucaena leucocephala* se deu por motivos específicos, discutidos na seção 1.3. Logo, essa lista reforça que FES's em regeneração natural são áreas submetidas a intensa e contínua invasão por espécies de plantas exóticas, uma realidade que ameaça a rica biodiversidade sustentada por essas florestas.

Todas as espécies apresentadas nesta lista (tabela 1) foram registradas no município de Porto Feliz, interior do estado de São Paulo, em áreas de vegetação secundária dominadas por Floresta Estacional Semidecidual. Classificamos como “potencialmente invasoras” espécies majoritariamente ruderais, cujas populações foram registradas em nossas áreas, mas não é sabido se, de fato, comportam-se como invasoras. As espécies *Lantana camara* L. e *Cardiospermum halicacabum* L. também foram registradas, mas não foram incluídas na lista abaixo pois apresentam distribuição geográfica original incerta, além de discussões taxonômicas importantes (CABI, 2023).

Tabela 1: Espécies de plantas exóticas invasoras ou potencialmente invasoras registradas em áreas de regeneração natural de Floresta Estacional Semidecidual, no município de Porto Feliz – SP.

Família	Espécies
Acanthaceae	<i>Thunbergia alata</i> Bojer ex Sims <i>Thunbergia grandiflora</i> Roxb.
Amaranthaceae	<i>Amaranthus deflexus</i> L. <i>Amaranthus retroflexus</i> L.
Anacardiaceae	<i>Mangifera indica</i> L.
Araliaceae	<i>Heptapleurum actinophyllum</i> (Endl.) Lowry & G.M. Plunkett <i>Heptapleurum arboricola</i> Hayata
Arecaceae	<i>Caryota mitis</i> Lour.
Asparagaceae	<i>Sansevieria trifasciata</i> Prain
Asteraceae	<i>Cosmos sulphureus</i> Cav. <i>Emilia fosbergii</i> Nicolson <i>Youngia japonica</i> (L.) DC.
Balsaminaceae	<i>Impatiens walleriana</i> Hook.f.
Bignoniaceae	<i>Jacaranda mimosifolia</i> D. Don <i>Spathodea campanulata</i> P. Beauv. <i>Tabebuia rosea</i> (Bertol.) Bertero ex A.DC. <i>Tecoma stans</i> (L.) Juss. ex Kunth
Boraginaceae	<i>Cordia africana</i> Lam.
Combretaceae	<i>Terminalia catappa</i> L.
Commelinaceae	<i>Commelina benghalensis</i> L. <i>Tradescantia zebrina</i> Heynh. ex Bosse
Convolvulaceae	<i>Ipomoea cairica</i> (L.) Sweet
Cucurbitaceae	<i>Sicyos edulis</i> Jacq.
Cyperaceae	<i>Cyperus difformis</i> L. <i>Cyperus esculentus</i> L.
Euphorbiaceae	<i>Ricinus communis</i> L.
Fabaceae	<i>Leucaena leucocephala</i> (Lam.) de Wit <i>Mimosa caesalpiniiifolia</i> Benth.

	<i>Neonotonia wightii</i> (Graham ex Wight & Arn.) J.A.Lackey
	<i>Tipuana tipu</i> (Benth.) Kuntze
Lamiaceae	<i>Leonotis nepetifolia</i> (L.) R.Br. <i>Leonurus japonicus</i> Houtt. <i>Mesosphaerum pectinatum</i> (L.) Kuntze
Lauraceae	<i>Persea americana</i> Mill.
Meliaceae	<i>Melia azedarach</i> L.
Moraceae	<i>Artocarpus heterophyllus</i> Lam.
Muntingiaceae	<i>Muntingia calabura</i> L.
Myrtaceae	<i>Psidium guajava</i> L. <i>Syzygium cumini</i> (L.) Skeels
Nyctaginaceae	<i>Mirabilis jalapa</i> L.
Orchidaceae	<i>Oeceoclades maculata</i> (Lindl.) Lindl.
Pinaceae	<i>Pinus elliottii</i> Engelm.
Poaceae	<i>Guadua angustifolia</i> Kunth <i>Megathyrsus maximus</i> (Jacq.) B.K.Simon & S.W.L.Jacobs <i>Melinis minutiflora</i> P.Beauv. <i>Melinis repens</i> (Willd.) Zizka <i>Urochloa decumbens</i> (Stapf) R.D.Webster
Pteridaceae	<i>Pteris vittata</i> L.
Proteaceae	<i>Grevillea robusta</i> A.Cunn. ex R.Br.
Rosaceae	<i>Eriobotrya japonica</i> (Thunb.) Lindl.
Rubiaceae	<i>Coffea arabica</i> L.
Rutaceae	<i>Citrus x limonia</i> Osbeck (pro. sp.) <i>Murraya paniculata</i> (L.) Jack
Thelypteridaceae	<i>Christella dentata</i> (Forssk.) Brownsey & Jermy
Urticaceae	<i>Boehmeria nivea</i> (L.) Gaudich.
Zinziberaceae	<i>Hedychium coronarium</i> J.Koenig

Fonte: elaborado pelo autor.

1.2.3 *Leucaena leucocephala* (Lam.) de Wit

A espécie *Leucaena leucocephala* (Lam. de Wit) é uma árvore de pequeno porte, altamente invasora, nativa da América Central (CABI, 2019). Em português, é popularmente conhecida como “leucena”. Em inglês, recebe nomes como *white-popinac*, *river-tamarind*, *horse-tamarind* e *wild-tamarind*. Essa espécie pertence à família Fabaceae, subfamília Caesalpinioideae (circunscrição atual que integra Mimosoideae; APG - IV (2016)). Tem como sinônimas botânicas o basônimo *Mimosa leucocephala* Lam. e o heterotípico *Leucaena glauca* Benth (JBRJ, 2023). Outras sinônimas são descritas em World Flora Online (2023). Três subespécies são reconhecidas: *L. leucocephala* subsp. *leucocephala*; subsp. *glabrata* e subsp. *ixtahuacana*, esta última tendo pouca importância comercial (CABI, 2019). A subespécie *leucocephala* é conhecida como a variedade “comum” ou “havaiana”, e costuma apresentar porte menor, arbustivo-arborescente. A subespécie *glabrata* é conhecida como a variedade “gigante” ou “Salvador” e apresenta maior porte, explicitamente arbóreo (GISD, 2015).

A plasticidade fenotípica em relação ao porte arbóreo e/ou arbustivo é uma característica marcante. Em regiões insulares do Japão, a leucena é oficialmente descrita como um arbusto, raramente ultrapassando 3 metros de altura (HATA; SUZUKI; KACHI, 2010; OSAWA; HATA; KACHI, 2016). No Brasil, indivíduos isolados podem atingir alturas superiores a 15 metros (MARCHIORI, 2007). A Figura 1 mostra alguns dos maiores indivíduos de leucena registrados neste trabalho.

Figura 1: Grandes indivíduos de *Leucaena leucocephala* registrados em Porto Feliz – SP. O indivíduo à direita teve altura total estimada em aproximadamente 16 metros, com diâmetro à altura do peito (DAP) de 41 centímetros. A título de comparação, a pessoa ao centro da imagem tem 1,70 metros de altura.



Fonte: acervo pessoal.

Leucena é uma espécie de crescimento extremamente rápido, e inicia sua reprodução em idade precoce (1 ano), quando comparada a espécies arbóreas típicas (CABI, 2019). Tem dispersão autocórica, e as sementes costumam cair próximas à árvore-matriz, apenas por ação da gravidade (MARQUES et al., 2014). Por conta dessa dispersão limitada espacialmente, costuma formar grandes maciços populacionais (Figura 2), com alta densidade de indivíduos (GISD, 2015). A formação desses maciços homogêneos também está associada aos efeitos alelopáticos da espécie, que leva a um menor recrutamento de plântulas de outras espécies no interior do maciço.

Há registros históricos de potenciais efeitos alelopáticos de leucena sob outras espécies (AHMED; HOQUE; HOSSAIN, 2008; SINGH; BATISH; KOHLI, 1999), mas um estudo recente confirmou a ação alelopática relacionada à mimosina e outras substâncias presentes em partes da planta (KATO-NOGUCHI; KURNIADIE, 2022) Há registro de dispersão por herbívoros como cabras (HATA; SUZUKI; KACHI, 2010) e

de transporte de sementes por formigas cortadeiras (Figura 3 – registro pessoal), mas pouco se sabe sobre a relevância de episódios de zoocoria para dispersão da espécie.

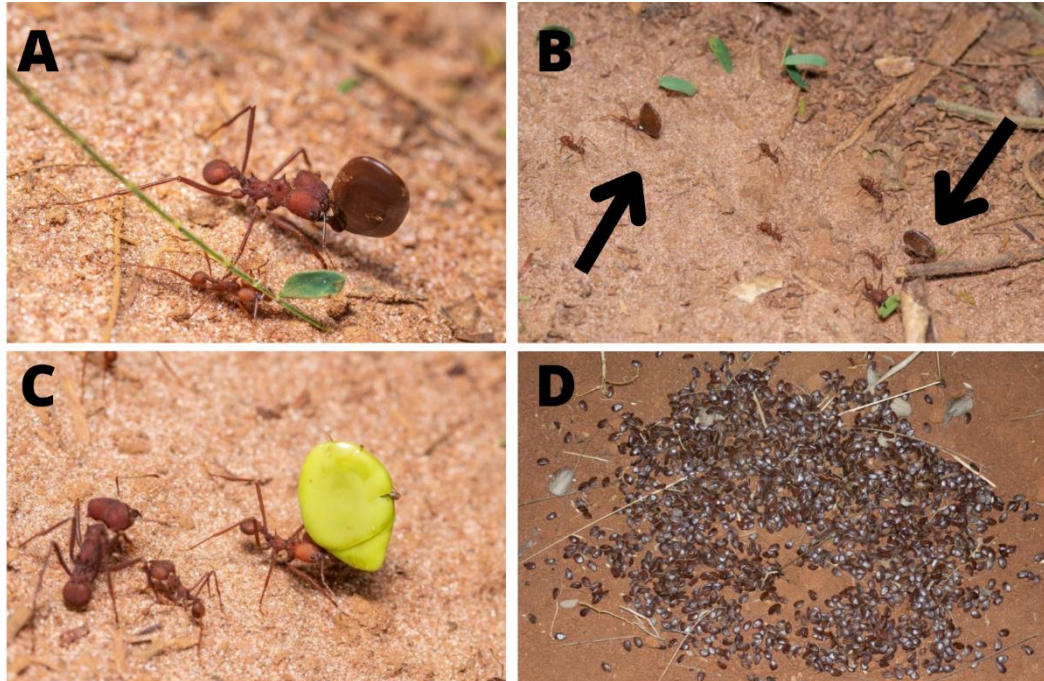
Figura 2: Maciço de leucena em área de regeneração natural de Floresta Estacional Semidecidual, interior do estado de São Paulo.



Fonte: acervo pessoal.

Figura 3: Painel de registros fotográficos de transporte de sementes de leucena (*Leucaena leucocephala*) por formigas cortadeiras (*Atta* sp.), em área de regeneração natural de Floresta Estacional Semidecidual, interior do estado de São Paulo. **A e B:** formigas carregando sementes de leucena; **C:** formiga levando semente imatura de leucena (possível predação); **D:** acúmulo de sementes de leucena em torno de uma das aberturas do formigueiro.

(próxima página)



Fonte: acervo pessoal (registros fotográficos feitos por Wolf J Moeller).

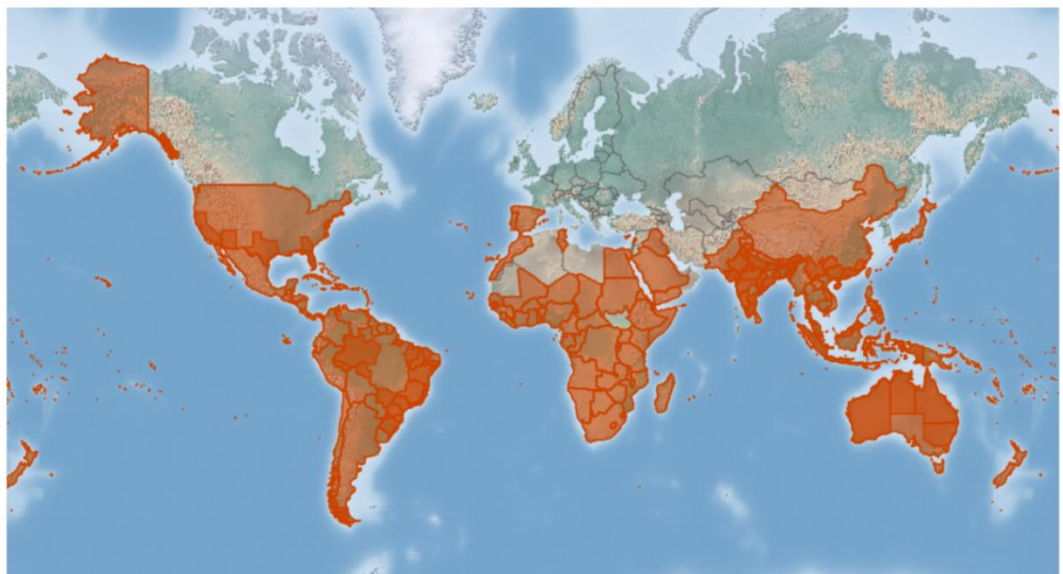
Produz grande quantidade de sementes ao longo de todo o ano, com maior intensidade na estação seca (MARQUES et al., 2014). A chuva de sementes de um indivíduo adulto chega a uma média de 5500 sementes por metro quadrado em um ano (MARQUES et al., 2014). As sementes podem apresentar dormência, permanecendo viáveis no solo por até 20 anos (GISD, 2015). A espécie é majoritariamente polinizada por *Apis mellifera* (Linnaeus, 1758) (DE MELO-SILVA et al., 2014), mas é também viável em autofecundação (BREWBAKER, 1983). É muito resistente a podas drásticas, após as quais costuma apresentar intenso rebrote (COSTA; DURIGAN, 2010).

1.2.3.1 Cenário internacional de invasão

A leucena é nativa da América Central, provavelmente do México. Não há consenso quanto a sua distribuição nativa original, pois esta espécie foi amplamente cultivada ao longo de outros países da América Central em períodos pré e pós Colombianos (CABI, 2019). A subespécie *leucocephala* (comum ou “havaiana”) foi a primeira a ser introduzida em outros países. Esta subespécie tem maior potencial de invasão quando comparada à subespécie *glabrata*. A espécie já estava amplamente distribuída em países da África e Ásia em meados de 1800, sendo considerada, a partir desse período, como uma espécie de ocorrência pantropical (CABI, 2019; GISD, 2015).

Leucena é citada entre as 100 piores espécies invasoras do mundo pela IUCN (ISSG, 2000). Atualmente, é encontrada em todos os continentes exceto Antártica (Figura 4). Apesar de introduzida na Europa, é registrada como invasora apenas nas Ilhas Canárias e Espanha (CABI, 2019). Regiões insulares são regiões drasticamente afetadas com a invasão pela espécie. O país polinésio Reino de Tonga enfrenta grave invasão por leucena, e cita-a como a planta invasora mais prevalente na região (PIER, 2017). As Ilhas Bonin, no Japão, tiveram uma explosão demográfica de leucena após a erradicação de cabras ferais (HATA; SUZUKI; KACHI, 2010; OSAWA; HATA; KACHI, 2016). No Arquipélago de Galápagos, houve implementação de tentativas de manejo, sem sucesso, e a espécie é listada entre as principais plantas invasoras em algumas ilhas (GARDENER; ATKINSON; RENTERÍA, 2010).

Figura 4: Distribuição geográfica atual da leucena (*Leucaena leucocephala*).



Fonte: Invasive Species Compendium (CABI). Disponível em: < <https://www.cabidigitallibrary.org/> >

Apesar do contexto de invasão, a leucena ainda é amplamente cultivada em países da África Subsaariana, Ásia e América do Sul (CABI, 2019). Em regiões com estação seca duradoura, leucena é frequentemente utilizada como planta de forrageamento para gado (e.g. Paquistão; AGANGA; TSHWENYANE, 2003), apesar de sua toxicidade conhecida (MATOS, 2011), principalmente em equinos (PORTO, 2016). Al-

gumas regiões fazem uso de madeira (e.g. Índia; PRASAD et al., 2011), outras implementam plantios de leucena para recuperação de solos degradados (e.g. China; PENG et al., 2013), ou para aumento de fixação de nitrogênio no solo em sistemas agrícolas (e.g. Nigéria; KANG; WILSON; SIPKENS, 1981). Alemán-Raamirez et al. (2022) realizaram uma revisão de usos e potenciais de leucena em produção bioenergética sob uma perspectiva global, destacando sua aplicabilidade para indústria de biocombustíveis.

O desencaixe entre os vieses ecológico e produtivo no que tange espécies invasoras é intrigante. A leucena é, há décadas, descrita como uma espécie altamente invasora, potencialmente causadora de inúmeros problemas em regeneração de áreas naturais de vegetação e ameaça a espécies endêmicas (CABI, 2019; GISD, 2015). Ainda assim, anualmente, diversos estudos insistem em propor a espécie como uma potencial aliada em regeneração de solos e florestas degradados (e.g. COLÓN; LUGO, 2006; COSTA; DURIGAN, 2010; PENG et al., 2013; SANTIAGO-GARCÍA et al., 2008). Campbell et al. (2019) propõem que o fato da leucena ainda ser vista como uma espécie valiosa economicamente seja talvez o principal empecilho às tentativas de prevenção e controle de invasão. É de extrema importância que políticas internacionais de manejo de ecossistemas e espécies invasora passem a considerar, de maneira expressa, o desencorajamento de uso de espécies exóticas invasoras como agentes de restauração ecológica.

1.2.3.2 *Cenário brasileiro de invasão*

Não há consenso sobre a história da introdução de leucena no Brasil. A citação mais antiga de introdução era referida a 1940 (VILELA; PEDREIRA, 1976). Machado (2018) realizou uma extensiva busca em herbários brasileiros e identificou inúmeras introduções anteriores a 1940, sendo o registro mais antigo datado no ano de 1831, no estado da Bahia. Atualmente, a espécie é registrada em todos os estados brasileiros exceto Amapá e Roraima (GBIF, 2022). Os estados de São Paulo, Paraná e Bahia são os que combinam mais registros (MACHADO, 2018).

Zenni e Ziller (2011) apontam que leucena é invasora em áreas de Savana e Savana Estépica, Manguezais, Restingas, Caatinga, Florestas Estacionais Deciduais e Semideciduais, Florestas Ombrófilas e zonas de Ecótonos. Assim como ocorre em outros países tropicais, a espécie no Brasil invade áreas relativamente abertas, não

sendo muito comum em formações florestais antigas ou primárias - com dossel superior bem estruturado (MACHADO, 2018).

Marques e colaboradores (2014) avaliaram potencial de germinação e banco de sementes de leucena em região de ecótono entre Cerrado e Floresta Estacional Semidecidual, e observaram alta produção de sementes, com viabilidade duradoura no solo, formando bancos de sementes viáveis por pelo menos cinco anos. A formação de bancos de sementes duradouros é um dos maiores desafios para o manejo da espécie no Brasil. Frequentemente, prefeituras municipais e outras instituições públicas implementam tentativas de manejo de leucena em Áreas de Preservação Permanente às margens de corpos d'água, parques, terrenos baldios e beira de estradas (e.g. CAMPINAS, 2019; SEMA-VOTORANTIM, 2015). A imensa maioria das tentativas de manejo encontra fracasso em um período curto de tempo, onde ocorre intenso rebrote de indivíduos cortados (COSTA; DURIGAN, 2010), e recrutamento de plântulas no banco de sementes no solo. (MARQUES et al., 2014).

No interior do estado de São Paulo, especialmente na microrregião de Sorocaba, a leucena é uma das espécies exóticas invasoras mais comuns às margens de estradas e corpos d'água urbanos. Essa presença quase ubíqua da espécie fez com que eu, ao longo do meu crescimento e maturação do interesse por ecologia, atentassem-me às áreas invadidas, especialmente florestas em regeneração natural. Como apresentado na seção 1.2.1, dezenas de outras espécies invasoras também estão presentes; contudo, nenhuma delas destacou-se tanto em relação à expansão populacional quanto a leucena. Vi pastos e plantações abandonados tornarem-se unicamente adensamentos de leucena em períodos surpreendentemente curtos. Por esses motivos, e considerando o contexto intenso e problemático de invasão de leucena no Brasil, eu estruturei as perguntas que foram respondidas ao longo deste trabalho.

2. CAPÍTULO 1

Title: Effects of white-popinac (*Leucaena leucocephala*) invasions on regenerating forests propagate from local to regional scale

Introduction

Biological invasions are the result of multiple processes operating at various spatial and temporal scales, from inter-continental (e.g., species introductions) to local (e.g., species establishment), and from diel (e.g., species interactions) to decennial (e.g., homogenization of landscapes) (Cadotte, McMahon, and Fukami 2006). Despite this being widely accepted and the emergence of conceptual frameworks that explicitly incorporate ecological processes at multiple scales (Patrick et al. 2021; Brown and Barney 2021), invasion biology is still predominantly focused on the role of local processes and effects (Powell, Chase, and Knight 2011). Understanding the local effects of biological invasions is still crucial, but this is only part of a more complicated puzzle – we must understand if and how local effects propagate to the regional scale. For example, do invasive non-native species (INNS) reduce the alpha, beta, and gamma facets of biodiversity? Investigating this is particularly important in the context of fragmented landscapes, where local and regional dynamics of invasion are determined by historical land use and species introductions, which comprise broad temporal and spatial scales (Waddell et al. 2020; Gibson et al. 2013; Didham et al. 2007).

Disentangling effects of biological invasions on biodiversity in fragmented landscapes is a challenging task. For example, (i) while in some contexts increasing the spatial connectivity among communities enhances INNS establishment (Chapman et al. 2020), it may also enhance community resistance to invasion (Howeth 2017; Grainger and Gilbert 2016; Cadotte 2006); and (ii) O’Sullivan et al. (2023) proposed that β -diversity patterns explain why some fragmented plant communities go through an increase in species richness after invasion events, whereas others go through opposite dynamics (see also Peng et al. 2019; Jauni and Hyvönen 2012; Fridley et al. 2007). To address these challenges, studies need to be based on hypotheses, sampling designs and variables that explicitly consider multiscale phenomena (Schiesari et al. 2019; Patrick et al. 2021).

The deforestation and fragmentation of the Atlantic Forest has historically been one of the most intense changes in landscape structure and biodiversity in Brazil (Rezende et al. 2018). Recent public policies have defined that private lands in Brazil must have a percentage of their area covered by natural vegetation (Brasil 2012; Rezende et al. 2018). As a consequence, portions of private lands that once were used as pasture and for agriculture have been abandoned so that natural regeneration could occur (César et al. 2018; Crouzeilles et al. 2020). However, regenerating forests in historically fragmented forested landscapes are highly susceptible to biological invasions (Zhang et al. 2021; With 2004; Aguirre-Acosta, Kowaljow, and Aguilar 2014). For example, invasive grasses have invaded the majority of those areas, where they occur abundantly at early regeneration stages (Zenni and Ziller 2011; Sobanski and Marques 2014; Zwiener et al. 2014). Although invasive grasses typically predominate, many other INNS are also often found in regenerating areas, from herbs (Dubbern, Leal, and Pedroso-de-Moraes 2013; Chiba De Castro et al. 2019) to tree species (Londe, de Sousa, and Kozovits 2017; CABI 2023).

Most INNS in regenerating forests can spread themselves across native communities. For example, the INNS *Melia azedarach* L. (China berry) has long-distance dispersal, and therefore is able to spread across invaded communities, with low-density but broad local distribution (Voigt, Farwig, and Johnson 2011; Bhatt et al. 2021). On the other hand, some INNS form dense and homogeneous agglomerations, with high-density of individuals and limited local distribution, usually due to distance-limited dispersal mechanisms (Portela et al. 2009; Chiba De Castro et al. 2019). One interesting example of species that forms dense monospecific patches is white-popinac (*Leucaena leucocephala* Lam. de Wit). White-popinac is a small tree native to Central America, that is broadly distributed in the Brazilian territory, occurring in open ecosystems like savannas and regenerating forest areas (Zenni and Ziller 2011).

In regenerating forests dominated by invasive grasses, the density of white-popinac patches can be so high that grasses and herbaceous species are forced towards the edges of the regenerating area (Hata, Suzuki, and Kachi 2010; Osawa, Hata, and Kachi 2016), due to reduction in sunlight incidence and known allelopathic effects (Kato-Noguchi and Kurniadie 2022). White-popinac can alter soil-nitrogen fixation and has been cited as a potential habitat transformer (CABI 2019; Henderson 2001), especially in regenerating forests (Yoshida and Oka 2000). White-popinac invasions are associated with decrease in diversity of native species (CABI 2019; GISD 2015;

Machado 2018), and enhancement in community susceptibility to invasion by other INNS (Yoshida and Oka 2004). It is reported as an important threat for endemic plant communities (Hughes 1998), especially in insular areas (Mello 2014).

Because of its short-distance seed dispersal, white-popinac usually forms dense and homogeneous patches, with larger individuals at the center (Werema and Wilson 2022; de Melo-Silva et al. 2014). The patches frequently occur in the surrounding of forest fragments, occupying part of the respective regenerating area (Wolfe and Van Bloem 2012; Marod et al. 2012). Here, we took advantage of this unique spatial configuration formed by patches of native forest, regenerating areas, and patches of white-popinac (Figure 1) to investigate how local and regional dynamics of plant (meta-)communities under natural regeneration are affected by white-popinac. More specifically, we analyzed the relationship between various aspects of white-popinac invasions and the local (alpha), regional (gamma) and among-sites (beta) diversity of naturally regenerating areas in the Brazilian Atlantic Forest. To do that, we set up a sampling design that explicitly considered a local (131 communities) and regional scale (29 meta-communities). We expected that the temporal and spatial advance of white-popinac invasion would be associated with (i) a decrease in alpha and gamma diversity; (ii) an increase in the similarity of species composition within metacommunities (reduced beta-diversity); and (iii) an increase of white-popinac abundance in metacommunities and communities, dependent on the community-patch distance.

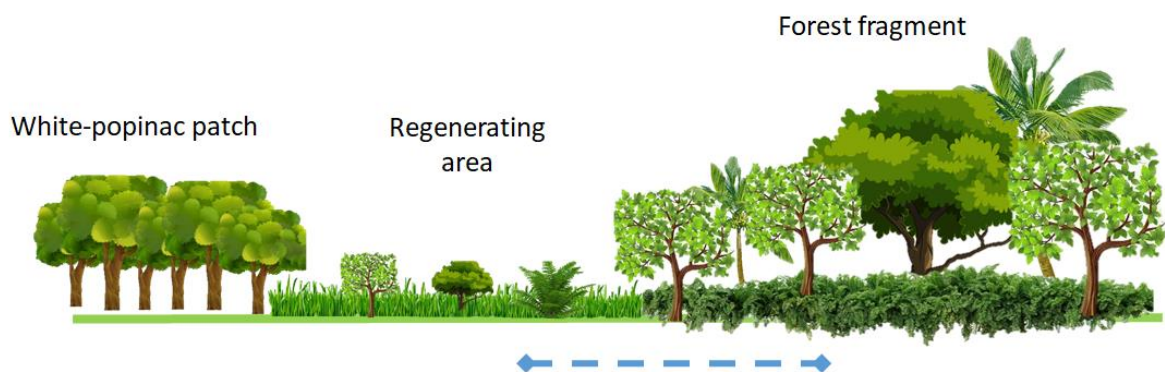


Fig. 1 Horizontal perspective of our regional sampling unit. As time of invasion advances, the white-popinac patch is assumed to expand towards the regenerating area, and therefore affect (meta-)community dynamics.

Methods

Sampling design

We conducted this study in the interior region of the state of São Paulo, south-east Brazil. We visited 38 rural and peri-urban private lands whose owners have destined portions of the property to forest natural regeneration and that were surrounded by at least one major patch of native Seasonal Semideciduous Forest and one patch of white-popinac. We actively chose areas that had represented different ages of invasions, to create a temporal gradient of white-popinac invasion. From 38 areas, we chose 29 that evenly filled the temporal gradient. All fieldwork was done from March 2020 to September 2022. Detailed information about the region can be found in the Supplementary Data section.

Our sampling design explicitly considers two spatial scales – local (131 communities) and regional (29 metacommunities). A metacommunity represented a system composed of a native forest fragment, partially surrounded by a regenerating area, which had at least one patch of white-popinac (Figure 2). Within each metacommunity, communities were defined as transects (14m x 2m) established at the edge between the forest fragment and its respective regenerating area (7m towards the forest fragment and 7m towards the regenerating area). Each metacommunity included four or five communities with varying distances from the white-popinac patch (Figure 2).

At each transect, we estimated the abundance of all species (native and INNS) on the arboreal (trees and shrubs) and herbaceous strata (climbing plants, herbs, ferns, epiphytes and grasses; below 1 m high), as well as of all seedlings. The abundance on the arboreal stratum was estimated via counting individuals, whereas on the herbaceous stratum it was done by visually estimating relative (percentage) ground cover (all methodological procedures and details are presented in Supplementary Data).

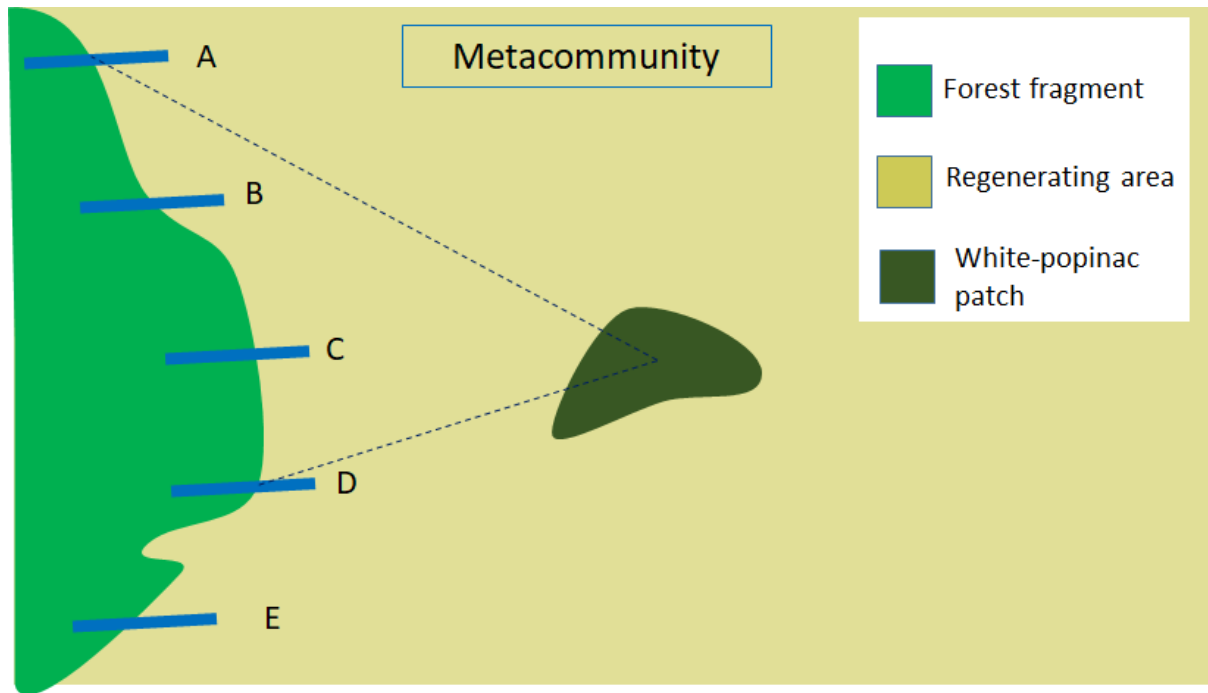


Fig. 2 Schematic representation of our sampling design. A metacommunity ($n = 29$) was defined as a spatial unit composed of a forest fragment, a regenerating area and a white-popinac patch. Within each metacommunity, a community ($n = 131$) was defined as a transect (four or five per metacommunity) placed on the edge between the forest fragment and the regenerating area. Within each metacommunity, local communities were unequally distant from the white-popinac patch.

Response variables

We estimated species richness for each local community ($n = 131$ transects) using sample coverage rarefaction/extrapolation methods in iNext R-package (v3.0.0, Chao et al. 2014; Hsieh, Ma, and Chao 2016). Each region ($n = 29$ metacommunities) had its own sample coverage, considering the maximum sample coverage possible among communities within a metacommunity. This process was made separately for the arboreal and the herbaceous strata. Sample-coverage-based estimation was also applied separately for INNS (excluding white-popinac) and native species. We defined regional (gamma) diversity as the summed richness estimates of all communities within a metacommunity. For the local and regional scales, we also calculated the abundance of white-popinac, with two different metrics: number of white-popinac adult individuals ($\text{dbh} > 2 \text{ cm}$) and relative ground cover by white-popinac seedlings (%).

We estimated variation in species composition among communities within a metacommunity (i.e., β -diversity) using a metric that applies coverage-based rarefaction/extrapolation to infer changes in non-random intraspecific aggregation (β -c; Engel et al. 2021). β -c is not affected by gamma diversity and allows more adequate comparisons of β -diversity among regions with different species pools. We estimated β -diversity separately for the arboreal and herbaceous strata once abundance metrics were different among strata. We used the “betaC” R-package (v0.1.0) to estimate β -c.

Predictor variables

We divide our numeric predictor variables into two groups: (i) time-advance and (ii) space-related variables.

Time-advance variables: we created three different gradients that could work as proxies to describe the temporal progression of invasion in a given metacommunity. We used proxies because it is an observational design; thereby we do not know exactly when the invasion took place initially. Our approach is essentially a time-for-space substitution framework, where the replicates in space comprise different invasion stages, creating a temporal gradient. We detail measuring and standardization methods in Supplementary Data section.

a. Basal area (square meters/plot): basal area was the total area on a given plot that was occupied by tree trunks of white-popinac. We measured diameter at breast height (dbh; breast height = 1.5 m) and calculated trunk area of all white-popinac trees in a standardized plot (25 m²) to estimate the basal area of each patch. We assumed that higher values of basal area should represent older trees and, therefore, older invasion events.

b. Average dbh of the largest trees (centimeters): we calculated the average dbh of trees larger than the superior quantile (75%) for each white-popinac patch. The larger individuals are the most important to provide time-since-invasion estimates. Higher values should also represent older trees.

c. Patch age proxy by satellite imagery (years): white-popinac populations usually occur as large and homogeneous agglomerations that are easily visualized with satellite imagery tools. Because they grow fast, the growth of the patch can be seen through time, using older images provided by Google Earth Software. Because we were able

to detect when the patch firstly appeared on the imagery, we could attribute an estimated age (years) to each patch.

Space-related variable: we measured the distance (meters) between the center of each community to the nearest edge of the white-popinac patch. We expected that longer distances would mean a weaker effect of white-popinac, considering that white-popinac dispersal is distance-limited by its autochory (Hata, Suzuki, and Kachi 2010). This was the only predictor variable applied to each community, instead of one value for the entire metacommunity.

Finally, our models also included a categorical predictor variable to describe strata (herbaceous or arboreal, when the diversity metric was calculated differently between strata, such as β -diversity) and one to describe origin (native or INNS for interaction with time-advance variable).

Model selection

For each of our hypotheses, we built a set of alternative models that represented a potential relationship between one of the response variables and a specific combination of predictor variables (Tables 1 and 2, Supplementary Material). Depending on the spatial scale, alternative models included a combination of predictor variables that had one time-advance variable (basal area, average dbh of the largest trees, or age-proxy by satellite imagery), the space-related variable, and two-level categorical variables (stratum and origin). All numeric predictor variables were standardized before being applied to the models.

For response variables measured at the local community level, we competed Generalized Linear Mixed-Effect Models (GLMM), considering metacommunity identity as a random component. For variables measured at the regional scale, we competed Generalized Linear Models (GLM) for each response variable. In some cases, the same model was fitted (and competed) with different distribution families.

When a model included the categorical variable “origin” (native or INNS), we included it as a potential interaction with the time-advance variable, as we expected that the effects of white-popinac invasion age would be different between native and other INNS. In these cases, we also used interaction-plots to visualize the relationships.

Model selection was based upon an Information-theory approach (Aho, Derryberry, and Peterson 2014), using the corrected Akaike Information Criterion (AICc) ranking, and derived metrics (e.g., Δ_i , AICc weight). The selection and post selection routines followed these steps:

- (i) Model ranking considering AICc values: models with $\Delta_i < 2$ were considered as equally plausible (Δ_i is the subtraction between a model's AICc value and the lowest AICc value among all competitor models [Burnham, Anderson, and Huyvaert 2011]);
- (ii) If the best model had an R^2 (or other equivalent metric of explained variance) lower than 0.05, we discarded that hypothesis and did not interpret the model;
- (iii) We examined the coefficients and their respective confidence intervals (95% CI) for equally plausible models to evaluate their magnitude and direction. Coefficients which CIs included zero were considered to have no effect on the response variable.

For GLMs, we used the “stats” R-package (v.4.2.2), and for GLMMs we used the “glmmTMB” R-package (v1.1.5). For model selection we used the “performance” R-package (v.0.10.0). More details about model structure, distribution families, decisions in selection, software and packages are included in Supplementary Data.

Results

We sampled 328 species: 178 in the arboreal stratum and 150 in herbaceous stratum. We could not identify 21 species due to the absence of taxonomically important structures, especially in case of deciduous species. The 307 identified species were distributed among 218 genus and 73 families, being Fabaceae, Asteraceae and Malvaceae the most representative ones in terms of number of species. The average number of species per metacommunity was 46.5 (minimum [min.] = 25.4; maximum [max.] = 84; standard deviation [s.d.] = 14.2; $n = 29$). For communities, the average number of species was 22.3 (min. = 3.6; max. = 44.1; s.d. = 8.9; $n = 131$). The list of recorded species can be found in Supplementary Data section (Table S5).

Selected models

For the local (community) scale, we found no evidence that either invasion age or community-patch distance were related to species richness. However, when we split the data per stratum, the most plausible model indicated that variation in herbaceous species richness was explained by an interaction between time and origin (Table 1).

Whereas herbaceous native species richness decreased with time, herbaceous INNS richness increased (Fig. 3a). We also found that the number of adults of white-popinac in the regenerating community was negatively related to the distance from the patch of white-popinac (Table 1; Figure 4a).

Table 1 Model selection statistics for equally plausible models at local scale. AICc refers to the corrected Akaike Information Criterion (AIC). Δi refers to the difference between a given AICc value and the lowest among competing models. AICc (W) refers to AICc “weight”, being interpreted as the model’s relative likelihood. R^2c refers to amount of variation explained by the entire model, both fixed and random components, whereas R^2m refers only to the fixed component. ADL = Average-diameter (dbh - cm) of largest white-popinac trees within a metacommunity (region); A = white-popinac patch’s age-proxy (years); BA = white-popinac patch’s basal area (m²). RE = random effects.

Model structure	Distri- bution	AICc	Δi	AICc (W)	R^2c	R^2m
Species richness of the herbaceous stratum						
Time (ADL) * origin + RE	Gamma	1035.79	0	0.221	0.58	0.45
Time (ADL) * origin + distance + RE	Gamma	1035.89	0.091	0.211	0.57	0.47
Time (A) + origin + distance + RE	Gamma	1037.16	1.361	0.112	0.56	0.47
Time (A) + origin + RE	Gamma	1037.67	1.876	0.086	0.56	0.46
White-popinac abundance						
Time (BA) + dis- tance + RE	Poisson	520.32	0	0.392	0.88	0.68
Time (A) + distance + RE	Poisson	520.82	0.503	0.305	0.88	0.68
Time (ADL) + dis- tance + RE	Poisson	520.84	0.52	0.303	0.88	0.68

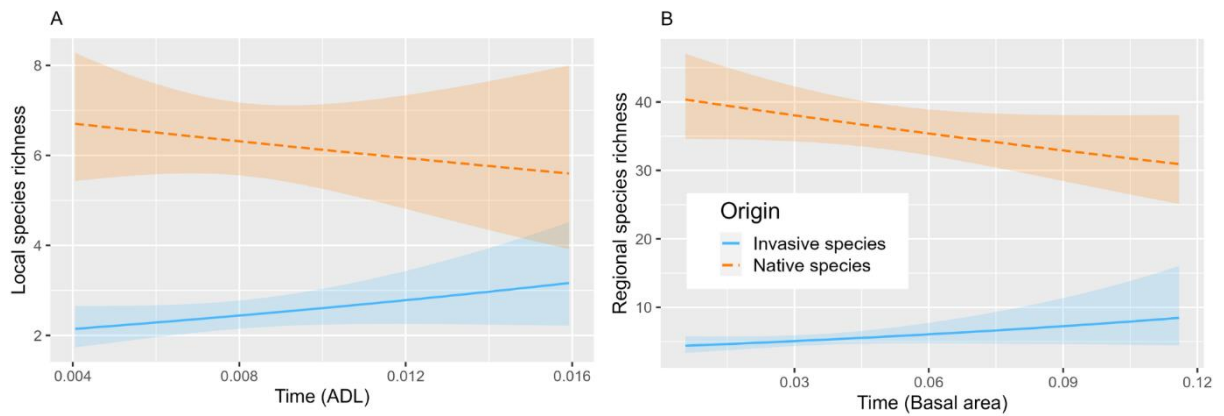


Fig. 3 Interaction plots representing the relationship between species richness and time conditional on origin (native species of INNS). **a.** relationship between local-scale species richness on herbaceous stratum and time conditional on origin. **b.** relationship between regional-scale species richness and time conditional on origin. ADL is a time-advance variable: average diameter (cm) of the largest white-popinac trees within a patch. Basal area (m^2) is a time-advance variable that represents the area occupied by white-popinac trunks within a standardized plot

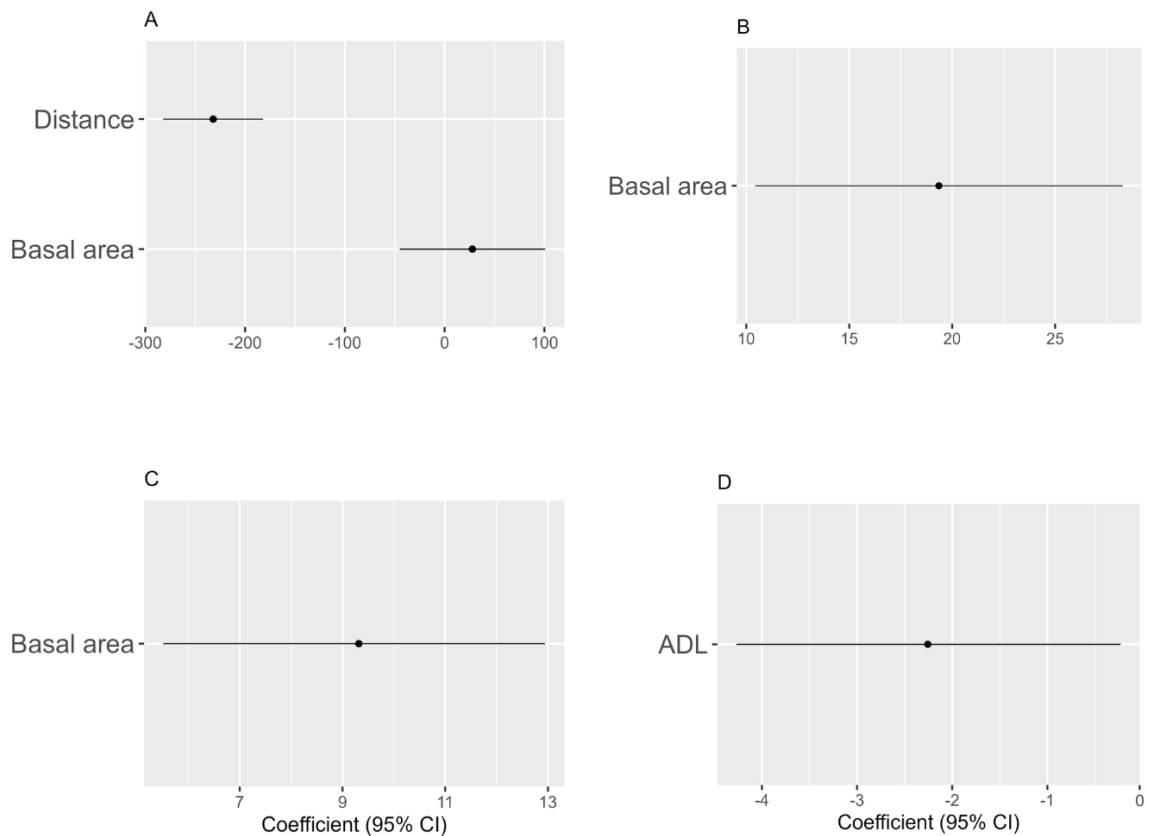


Fig. 4 Confidence interval (95%) plot describing parameter estimates of predictor variables related to: **a.** the abundance of white-popinac trees at regenerating communities (local scale). **b.** relative (%) ground coverage by white-popinac seedlings on metacommunities (regional scale). **c.** number of adult individuals of white-popinac on metacommunities. **d.** for β -diversity modeling. Here, the response variable is β -c, which is a β -diversity metric (see Methods section and Supplementary Data for details). The higher the β -c value, the less homogenous are the communities within a metacommunity in terms of species composition. Basal area (m^2) is the time-advance variable that represents the area occupied by white-popinac trunks within a standardized plot. Distance (m) is a space-related variable that express how far the community is from the main white-popinac patch. ADL is a time-advance variable: average diameter (cm) of the largest white-popinac trees within a patch.

The most plausible models describing variation at the regional scale (metacommunity; Table 2) were in agreement with those at the local scale (Table 1). We found no strong evidence of white-popinac affecting overall metacommunity species richness without discriminating species origin. When we did so, a model including an interaction between time and species origin indicated that native regional species richness decreased with invasion age, whereas the opposite happened with INNS, irrespective of stratum (Table 2; Fig. 3b). Regarding white-popinac abundance, we found that both relative (%) ground cover by white-popinac seedlings and number of adult individuals were positively related to invasion age (Fig. 4b, c). Finally, we found strong evidence that β -diversity decreased with invasion age, irrespective of stratum (Table 2; Fig. 4d).

Table 2 Model selection statistics of equally plausible models describing variation at the metacommunity regional scale and β -diversity. Models fitted with Poisson and Beta distributions are associated with pseudo- R^2 values, whereas models fitted with Gamma distribution are associated with Nagelkerke R^2 , which are mathematically different from a regular R^2 , but may be interpreted in a similar way. Abbreviations as in Table 1.

Model structure	Distribu- tion fam- ily	AICc	Δi	AICc (W)	R ² (Nagelkerke)
Species richness					
Time (A) * origin	Gamma	375.40	0	0.235	0.89
Time (A) + origin	Gamma	375.47	0.063	0.228	0.88
Time (BA) * origin	Gamma	375.61	0.202	0.212	0.88
Time (BA) + origin	Gamma	376.27	0.867	0.152	0.88
Time (ADL) + origin	Gamma	377.04	1.640	0.104	0.88
White-popinac seedlings' relative (%) ground cover					
Time (BA)	Beta	-120.6	0	0.924	0.18
Response variable = Number of white-popinac adult individuals (γ scale)					
Time (BA)	Poisson	472.25	0	0.99	0.52
Response variable = β-c (β-diversity metric)					
Time (ADL) + strata	Gamma	-60.93	0	0.667	0.63
Time (BA) + strata	Gamma	-59.10	1.836	0.266	0.62

Discussion

Our study provides evidence that the effects of white-popinac on regenerating forests are consistently manifested across different spatial scales. Native species richness decreased with time since invasion, whereas the species richness of INNS increased, both at the community and metacommunity levels. This mirrored local-regional negative relationship was due to a clear decrease of β -diversity in function of time since invasion. That is, communities within invaded metacommunities became more homogeneous in terms of species composition as time of invasion advanced.

Together, these results indicate that white-popinac effects on invaded landscapes are so vigorous that they propagate across spatial scales, negatively affecting the three main components of biodiversity.

At the local scale, as time since invasion advanced, the herbaceous stratum became more invaded by other INSS and less species rich of natives. Because the regenerating communities do not contain the main white-popinac patch (only some sparsely distributed individuals), the mechanism that drives this decrease in native species richness is likely to be related to spatial distance. Our results support this view as we found that the farther the regenerating community was from the main white-popinac patch, the lower the abundance of white-popinac adult individuals in the regenerating community (see Hata, Suzuki, and Kachi 2010 for similar results). The advance of white-popinac towards regenerating communities may reduce space available for the regenerating forest, which is already invaded by grass species. This reduction in space availability may enhance competition among native species on the herbaceous stratum and invasive grasses (Martin et al. 2014; Wilsey, Daneshgar, and Polley 2011), and affect trees and shrubs seedlings recruitment (Aronson and Handel 2011; Flory and Clay 2010). Ultimately, an increase in white-popinac abundance in the regenerating community could indirectly lead to the exclusion of less competitive native species by increasing the community's susceptibility to invasion (i.e. invasibility, Lonsdale 1999) by other INNS (similar results were found by Yoshida and Oka (2004)).

The upscaling of the negative relationship between native and INNS richness from local to regional scale is a result rarely seen in invasion studies. A plethora of studies regarding plant invasions has found discrepancy on responses between regional and local scales, which has become a highly debated topic, under the name of "invasion paradox" (Fridley et al. 2007; Peng et al. 2019; Powell, Chase, and Knight 2011; Pyšek et al. 2008; Pyšek and Hulme 2005; Brown and Barney 2021). It is common to find no relationship or even a positive one between native and INNS richness at broader scales (e.g. metacommunities), often due to an increase in β -diversity across communities (e.g. Davies et al. 2005; Chen et al. 2010). As it should be expected, the rare examples of analogous mirrored local-regional effects similarly found a decrease in β -diversity (e.g. Jauni and Hyvönen 2012). Our results show that the effects of white-popinac invasion on regenerating forests can be so strong that the local response dynamics propagate to the regional scale, even though the invasion is considerably recent ($< 20y$).

The β -diversity of both the arboreal and herbaceous strata decreased with the increase of invasion age. This means that communities within invaded metacommunities become more similar to each other as invasion advances in time. Unless demographic stochasticity dominates assembly (Jacobi and Siqueira 2023; Siqueira et al. 2020), competition is likely enhanced in a regenerating forest that is dwindling in size (Cadotte 2007; Amarasekare 2003; Bowker and Maestre 2012). If this process is replicated across various regenerating forest patches, the same set of species may thrive in different communities, which leads to a homogenizing effect (Smart et al. 2006; Solar et al. 2015; McKinney and Lockwood 1999; Arroyo-Rodríguez et al. 2013). For example, all communities within our oldest invaded metacommunity had the same six dominant species, which represented more than 50% of total abundance on each community. Contrastingly, in none of the communities within one of the most recent invaded metacommunities (smallest ADL) there was a clear dominant species – the most abundant species were just slightly more abundant than the others. This decrease in β -diversity along the temporal gradient of invasion explains the propagation of effects from the local to the regional scale (Legendre, Borcard, and Peres-Neto 2005; Anderson et al. 2011; Chao and Jost 2012).

The responses alpha, beta and gamma diversity to white-popinac invasion were essentially temporal. However, we identified indirect spatial effects that likely underlined temporal dynamics. As time of invasion advances, patches of white-popinac tend to grow in area and reduce their distance from regenerating communities (Hata, Suzuki, and Kachi 2010; Osawa, Hata, and Kachi 2016). We found that white-popinac abundance was also affected at regional scale; however, it was not related to distance (as it was for local scale), but to invasion age. We propose that this process describes biotic indirect effects hypothesis (Catford, Jansson, and Nilsson 2009; Marler, Zabinski, and Callaway 1999; White, Wilson, and Clarke 2006; Strauss 1991). The spatiotemporal advance of white-popinac patches may generate indirect effects on regenerating communities by reducing the size of the regenerating area. We suggest that biotic indirect effects provide a likely explanation on how white-popinac invasions disrupt the dynamics of regenerating plant (meta-)communities. In an explicit metacommunity framework, such biotic indirect effects can be seen as analogous to spatial spillover effects (Kuschnig et al. 2021; Wang et al. 2021).

We recognize that other factors than the ones we addressed here can affect how (meta-)communities respond to invasion, such as landscape configuration (Vilà

and Ibáñez 2011; Zhang et al. 2021), historical land use (Didham et al. 2007; Colón and Lugo 2006), fire events (Portela et al. 2009; Flory et al. 2015), and even cattle sporadically sneaking into the regenerating area (Reisner, Doescher, and Pyke 2015; Osawa, Hata, and Kachi 2016). There are also unapproached conditions related to white-popinac invasion, especially propagule and source-populations dynamics. Propagule pressure is as pivotal as time-advance (King and Howeth 2019; Lockwood, Cassey, and Blackburn 2005), but it was not addressed here. Observational field-based studies on biological invasions have major limitations, especially regarding temporal gradients. The use of time-advance proxies and space-for-time substitutions are very common and, despite being useful, these methods may reduce result precision and predictive power. Despite these limitations, our models were associated with reasonably high explanatory power – e.g., R^2 values varied from 0.18 to 0.89. Because the models we compared were all based on sounding hypotheses, we are confident that the relationships and high explanatory of some models we found here are not the result of chance or unaccounted hidden sources of variation.

Investigating how alpha, beta and gamma diversity of regenerating communities are affected by invasive species was only possible because our sampling design explicitly incorporated two aspects that are not frequently considered in other studies: (i) we meticulously surveyed the herbaceous stratum, even though our study region is originally dominated by Seasonal Semideciduous Forests. Had we not included the herbaceous stratum, we would not have found any relationship between species richness and invasion age at the local scale. (ii) We included spatial replication at both the local and regional scales, which allowed us to understand how changes in the local structure of communities upscaled to the metacommunity level. Thus, we reinforce the need to adopting a metacommunity approach that explicitly considers multi-scale consequences of biological invasions (Brown and Barney 2021; Patrick et al. 2021). We suggest that future studies on white-popinac invasions consider both local and regional scales, and explicitly address how β -diversity responds to invasion. Despite white-popinac being considered an arboreal species in Brazil, we recognize that non-arboreal native species should be comprised by floristic surveys on scenarios of forest regeneration (Gilliam 2007; Flory and Clay 2010). Finally, the effects of spatial spillover seem to be crucial for understanding the effects of invasive species that do not spread into communities but establish as isolated patches.

We aimed to understand how various aspects of white-popinac invasions affect communities and metacommunities undergoing through natural regeneration, in a tropical Seasonal Semideciduous Forest system. We found that both communities and metacommunities are negatively affected by white-popinac age of invasion. At both scales, we found that the number of native species decreases as time of invasion advances, concurrently as the regenerating communities become more invaded by other INNS. We also found that communities within invaded metacommunities become more homogeneous regarding species composition with invasion age. The mechanism that underlies these consequences is probably based upon biotic indirect effects. We suggest that future studies on white-popinac invasion address the effects of other facets of biological invasions, especially propagule and source-population dynamics. We also stress that the effects of white-popinac invasions should not be underestimated in terms of management, once the relatively recent invasions that we studied have caused severe diversity declines.

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3. CONSIDERAÇÕES FINAIS

Esta dissertação de mestrado buscou abordar, através de uma perspectiva de ecologia de metacomunidades, os efeitos causados por uma espécie exótica invasora, a árvore leucena (*Leucaena leucocephala*), em áreas de regeneração natural de Floresta Estacional Semidecidual (FES), no interior do estado de São Paulo. Nós observamos que tanto dinâmicas locais (comunidades) quanto regionais (metacomunidades) são negativamente afetadas pelo avanço temporal da invasão de leucena. Nessas duas escalas, a riqueza de espécies nativas diminuiu com o aumento da idade de invasão, enquanto a riqueza de outras espécies exóticas invasoras (EEI) aumentou. Houve também uma forte homogeneização na composição de espécies (diversidade- β) entre comunidades em metacomunidades com invasão mais antiga. A diminuição da diversidade- β é o mecanismo que provoca a propagação dos efeitos encontrados em escala local para a escala regional.

Os resultados sugerem que, em florestas regenerantes, os efeitos da invasão por leucena podem ser tão vigorosos que até mesmo a escala regional é afetada. Este tipo de resultado não é muito comum, sendo frequentes a redução ou até inexistência de efeitos de EEI em escalas regionais (POWELL; CHASE; KNIGHT, 2011). Este estudo é importante ao corpo de evidências que reforçam a importância de considerar leucena como uma espécie invasora que causa, de fato, impactos às (meta-)comunidades nativas. Apesar de o estabelecimento da leucena estar associado a áreas degradadas, seus efeitos como espécie invasora não se restringem, aparentemente, apenas às áreas antropizadas ou extremamente degradadas. No nosso cenário de invasão em FES, os maciços de leucena não invadem o sub-bosque da floresta já estruturada. Logo, ainda que restritos às bordas da floresta e à sua respectiva área

em regeneração, foram capazes de afetar negativamente a riqueza e variação na composição de espécies nativas.

Este trabalho teve dois pontos-fortes que merecem destaque. Nós incluímos amostragem de estrato herbáceo, uma prática não muito frequente em estudos envolvendo sistemas dominados por floresta. Praticamente metade do total de espécies amostradas pertence a este estrato. Foi inclusive nesse estrato onde observamos os efeitos locais da invasão de leucena. A não inclusão teria resultado em ausência de efeitos locais observados. Nós reforçamos a importância de considerar o estrato herbáceo em sistemas de florestas, especialmente sob regeneração natural. A replicação de unidades amostrais em escala regional foi o segundo ponto-forte. O imenso esforço amostral para estabelecer 29 réplicas de metacomunidades foi compensado com uma segura e efetiva capacidade explicativa dos modelos para escala regional. Um esforço amostral reduzido teria prejudicado o potencial oferecido pela escala regional em registrar os efeitos da invasão.

Os desafios para conduzir um estudo observacional como este são diversos. A dificuldade em padronizar unidades amostrais é, talvez, o maior deles. No nosso caso, ainda que a leucena tenha ampla distribuição no interior do estado de São Paulo, encontrar unidades amostrais adequadas não é uma atividade corriqueira. Ao todo, 40 metacomunidades foram visitadas, mas 29 foram escolhidas por compartilharem características mais semelhantes. Tivemos que excluir áreas com evidente passagem de fogo recente, áreas com manejo humano (entrada de tratores, por exemplo), e áreas com entrada de gado. Apesar dessas limitações metodológicas, o estudo teve uma boa condução e foi capaz de apresentar resultados importantes.

Esperamos que mais estudos em invasão biológica contemplem escalas regionais. A ecologia de metacomunidades é promissora à Biologia da Invasão, e fornece

ferramentas importantes para abordar dinâmicas espaciais, como dispersão e conectividade, e possibilita condutas eficazes em análises de risco (BROWN; BARNEY, 2021). Recomendamos que estudos futuros acerca de invasões por leucena considerem também outros componentes da invasão, especialmente dinâmicas de propágulos e populações-fonte, que não foram abordadas neste trabalho. O marcante contraponto entre quantidade de propágulos *versus* distância percorrida na dispersão de leucena é intrigante e certamente oferece respostas importantes à implementação de medidas de manejo.

Finalmente, consideramos que este trabalho atingiu seus objetivos propostos e trouxe resultados importantes para entender o presente cenário de invasão de florestas regenerantes por leucena. A lista de espécies registradas será apresentada a alguma base de dados de ocorrência (por exemplo, GBIF) e também abrirá caminhos a um futuro artigo abordando diversidade funcional em (meta-)comunidades invadidas por leucena.

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APÊNDICE – A

Supplementary Data

Material and methods

Study region

Our study region comprises rural and peri-urban areas in five municipalities within state of São Paulo, southeastern Brazil. Nineteen of the 29 metacommunities were in the city of Porto Feliz. All other ten metacommunities were in neighbor cities, near the division line with Porto Feliz, being: one in Rafard, one in Capivari, two in Itu and six in Boituva (Figure S1). All five cities are relatively small and share similar climatic, vegetation and geological features.

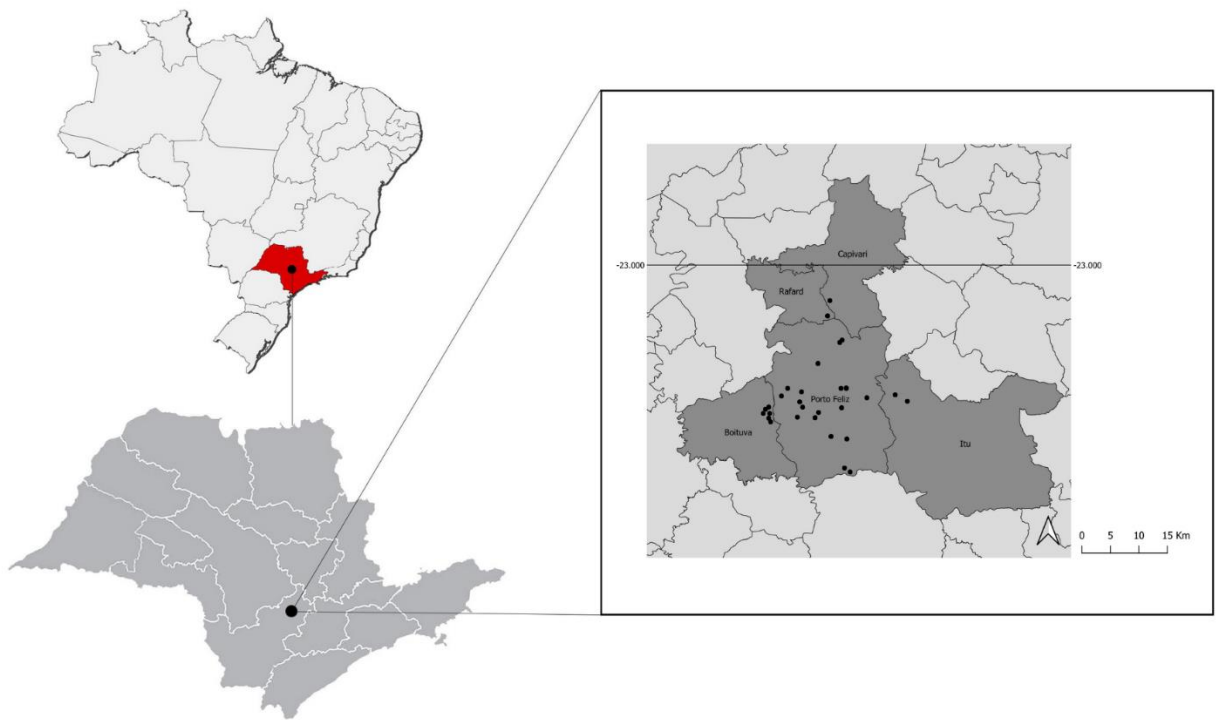


Fig. S1 Location of our study region. State of São Paulo, Brazil. The points on the right side chart are our metacommunities.

The region is located within Paraná's Sedimentary Basin, and the most abundant soil types are Latosols and Ultisols. The climate is classified as CWA by Köppen classification, which is characterized by a warm and rainy summer and a dry winter season. The region is crossed by one of the most important rivers in the state of São Paulo, the Tietê River (Oliver 2016). The following geographical coordinates

delimit the region: North: 23°02'48.64" (S); 47°31'58.92" (W); East: 23°14'59.57" (S); 47°22'23.79" (W); South: 23°21'23.00" (S); 47°29'01.13" (W); West: 23°14'11.83" (S); 47°37'43.70" (W).

Our regional sampling units (metacommunities) were systems composed of a forest fragment and its respective regenerating area, which became invaded by a white-popinac patch (Fig. S2).

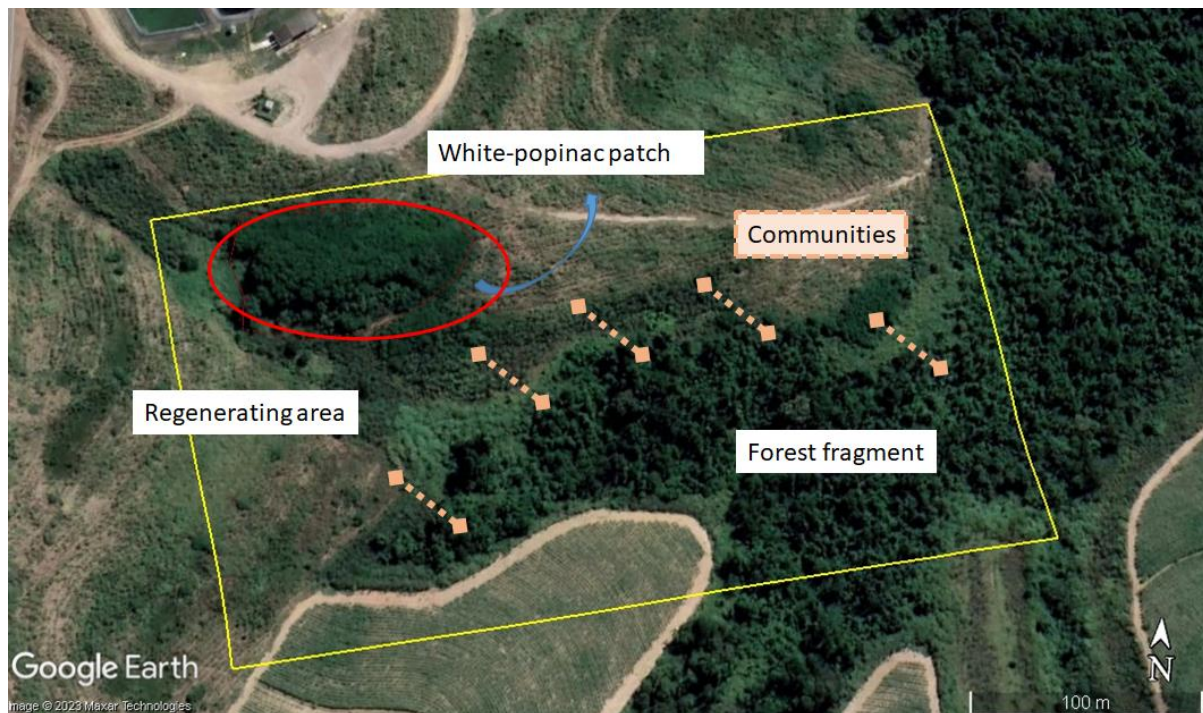


Fig. S2 Example of one of our metacommunities. The yellow rectangle delimits the sampling unit at the regional scale. The patch within the red ellipse is the white-popinac patch. Dotted lines are communities, which comprise both regenerating area and forest fragment.

All fieldwork was done from March 2020 to September 2022. We divided fieldwork into two phases: the first for sampling and measuring white-popinac (predictor variables) and the second for floristic surveying (response variables). Thus, all regional sampling units were visited twice. All sampling units were in private lands, with different history of land management and abandonment. However, the last land uses before abandonment were either pasture or sugar cane cropping. We got this information based on properties' owners and neighbor's reports. We discarded potential areas that had recently gone through fires or cattle (re)introduction.

Details on predictor variables

Bellow, we detail the methods involved in estimating our predictor variables. We created three different gradients to express the potential effects of time-advance of invasion because each one of them captures a different component of time-advance. Despite the age-proxy being more intuitively related to time-advance, the scale of years may not be enough to describe changes in meta(community) dynamics, because white-popinac is an extremely fast-growing tree. Therefore, we understand that other approaches to time-gradients can be useful as well. The use of proxies was necessary because we had an observational design, in which the precise age of a patch is not achievable.

Basal area

Basal area describes the amount of area occupied by tree trunks. The sum of cross-sectional area values of all tree trunks, considering breast height, within a standardized area, gives the basal area value. Usually, it is expressed in square feet per acre, or square meters per hectare (Cancino 2012). In our design, basal area was chosen because it is easy to obtain, and can represent time-advance of invasion. It is expected that as a white-popinac patch grows and develops, the average basal area also increases. To estimate basal area (BA) of a white-popinac patch, we followed this sequence of events:

- 1- Each metacommunity had its own “invader” white-popinac patch. Therefore, the measures that come from that patch were applied for the entire metacommunity and its respective communities.
- 2- For each small patch (total area < 100 m²), we established one 5m x 5m plot, avoiding patch’s edge areas, and placing it on the center of the patch. For larger patches, we established two or even three plots, and the patch’s basal area was the average among them.
- 3- Within each plot, all white-popinac trees higher than 1.70 m were measured. The measuring consists in the trunk’s perimeter on breast height (pbh), which was measured with a measuring tape (precision = 0.1 cm) at breast height (1.5 m).
- 4- We calculated the respective diameter at breast height of each individual applying the perimeter as equal to $2\pi R$, being R the circumference radius to be calculated and multiplied by 2, obtaining the diameter.

- 5- For each patch, the sum of all diameters results in basal area (or average among plots for large patches), which is expressed in square meters (m^2), considering a standardized plot area of 25 m^2 .
- 6- Each basal area value works as a point on the time-gradient proxy. Remind that our approach is based upon a space-for-time substitution method.

Average diameter of the largest white-popinac trees

This variable is also a proxy of time-advance of invasion. We assumed that the largest (largest = greater dbh) white-popinac trees within a patch can be used as models to indirectly express age. The largest trees in a patch are necessarily the oldest ones because (i) the average lifespan of a white-popinac individual is 20-40 years (GISD 2015) and (ii) all of our patches are younger than 18 years (considering our age-proxy approach with Goggle Earth Imagery). Therefore, we can assume that there was not enough time for a complete cycling of individuals in a patch.

We used the same measurements as the ones for basal area. The same data was used to calculate the average of largest trees, but with a second approach to include older trees. Walking into a white-popinac, we actively looked for the largest (greater dbh) individuals, even if they were not included within the plot, to guarantee that we had a representative sampling of older trees. For each path, we filtered the trees that were larger than the superior quantile (75%) in dbh, and estimated the average value, which we call “average diameter of the largest trees” - ADL.

Age proxy

We previously recorded the geographical coordinates of all patches using a Garmin eTrex®10 GPS, and used Google Earth Imagery to visualize the white-popinac patches (which are usually very distinctive on the landscape). We compared older images (from previous years) to current ones using the historical imagery tool, until we could visualize when the patch started to be seen on the imagery, and attribute a value in years. We were aware that we were not considering the exact period where the first propagules arrived or developed. Instead, the “age” variable actually represents how many years ago the white-popinac population structured itself in the sampling unit to the point it was recognizable on the satellite imagery.

Community-patch distance

This was our space-related variable, which described the distance, in meters, between each local community within a metacommunity and the white-popinac patch. All communities (14 m x 2 m transects) had geographical coordinates recorded, so we managed to estimate the distance from them to the patch using line-measurement tool from Google Earth. We considered the coordinates from communities' "center", being the medium point where 7m of transect leaned towards the forest and the other 7m leaned towards the regenerating area. Moreover, the patch's point for reference was not the center, but the edge that was closest to the forest. We decided to consider the edge as reference point because white-popinac has a distance-limited dispersal, and it is plausible to consider that the most "influential" white-popinac individuals are the ones that are closer to the community, and not the central ones (Hata, Suzuki, and Kachi 2010). The overall distance range was 2 – 183 m, highly different throughout metacommunities.

Grouping variables

When arranging the statistical models for selection, we considered both strata separately (arboreal and herbaceous), by creating a categorical variable called "strata", in additive effect to a given time-related predictor variable. We proposed to consider both arboreal and herbaceous strata because we understand that, even though our region is essentially a forest vegetation (Seasonal Semideciduous Forest), herbaceous stratum can bring interesting and important responses that are often disregarded in this type of phytophysiological (Gilliam 2007; Flory and Clay 2010).

We also created a variable called "origin", which groups the species into native and invasive non-native (INNS). In this case, we established models with interaction between the "origin" variable and a time-related variable, once we expected that the effects of time-advance of invasion disrupt different responses between these two groups (native vs. INNS).

We clarify that we obtained the "origin" variable towards species richness estimation (detailed on "Details on response variables" section). We firstly grouped our species into "native" and "INNS" categories and then proceeded to species richness estimation separately between these two categories, for both local and regional

scales. The classification was based upon INNS databases (Invasive Species Compendium – CABI; Global Invasive Species Database – GISD; The Horus Institute for Environmental Conservation and Development). All species were checked in terms of native range and localities where they are reported as INNS. If at least one of these databases reported the species as INNS in our study region, it was considered as INNS for our analysis.

Floristic survey

The floristic surveys conducted in each community were the main methodology to obtain our response variables. Consider that “transect” and “community”, in our framework, are spatially equivalent concepts. All transects were placed in the contact zone between the forest fragment and its respective regenerating area, with 7 meters advancing towards each area. In some cases, especially older-regenerating areas, the spatial delimitation from forest to regenerating area was not explicit. Inherently, the floristic survey provided material for a herbarium, which has been kept by the authors and is available for consultations.

Herbaceous stratum

For the herbaceous stratum, we divided the entire transect into 28 grids (1 m x 1 m). The sampling procedure (Figure S3) consisted in choosing seven grids that were always in the same position. On these seven grids, we recorded the ground cover percentage of each species. Notice that the abundance metric is based into a percentage, instead of counting individuals, once it is often difficult to delimit an individual for some species at this stratum. We considered as part of the grid all species that had vegetative parts comprised by the grid, even if they were not rooted there. We made this decision because climbing species are often included by the grid's area, but are rooted somewhere else. We designed a framework that could fit into our scenario of fragmented Seasonal Semideciduous Forest going through natural regeneration, where we can find extensive ground cover by invasive grasses, many liana species growing like herbs or shrubs (due to the low density of arboreal individuals), other invasive species besides grasses and white-popinac, and scattered young trees and shrubs.

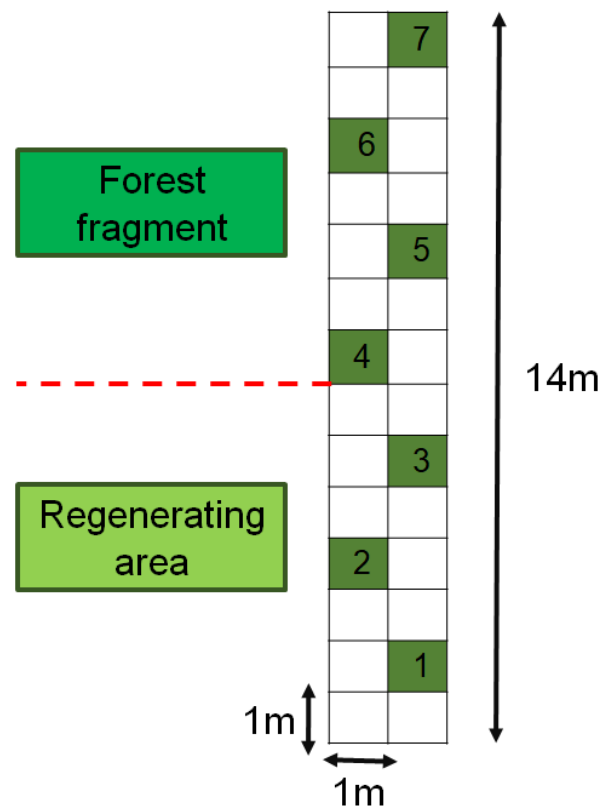


Fig. S3 Diagram of transect division into grids. Among the 28 one-meter-square grids, we chose seven, following the same spatial arrangement as the figure: grid 1 was always the farthest to the forest fragment. From 14 meters in length, 7 entered the forest fragment and 7 entered the regenerating area.

All species except moss and parasite species were identified. In other words, we identified herbs, grasses, climbing plants (lianas and vines), and other types such as ferns and epiphytes. Moreover, we included litter layer, white-popinac seedlings and overall seedlings (not white-popinac) as “species”. Here, it is important to stress that we considered all plant species that were growing below 1 meter height as part of herbaceous strata. That means that climbing species found only on higher ranges, such as upper canopy, were not included.

We visually estimated ground cover percentage of each species we found within a grid, as in *relevé* approach (Braun-Blanquet 1964). Within a grid, the sum of all species ground cover did not necessarily equal to 100, once different plant heights can create “layers” of ground cover that prevent smaller individuals to be recorded if just considering a single-frame vision.

Summarizing, we followed the sequence:

- 1- Delineated a transect;
- 2- Placed the 1 m² grid (made of PVC pipes) within the first sampling spot (grid 1, see figure 3);
- 3- Visually estimated the ground cover percentage of each species found within the grid, and also litter and two classes of seedlings (overall ones and white-popinac ones);
- 4- Collected samples of species with uncertain identification or not yet identified on the survey;
- 5- Followed to grid number 2 (Figure S3) and redoing the previous steps;
- 6- Once all seven grids were sampled, then we moved to arboreal stratum sampling on the same transect.

Arboreal stratum

We considered as components of arboreal stratum all tree and shrub species within the transect area (14 m x 2 m). Distinctively from herbaceous stratum, we did not divide transects into subsamples. All individuals on the arboreal stratum were recorded and counted. In case of saplings - young individuals that are not considered seedlings anymore - they were counted as regular individuals. Abundance in this stratum referred to the number of individuals (absolute abundance) of each species. Individuals that were growing outside of transect but whose brunches were entering the transect space were also included (Figure S4).

The survey of the arboreal stratum was proceed after the herbaceous one, once the walking amid transect area potentially disturbs herbaceous stratum. After accomplishing the first transect, we followed to the second, at least 10 meters of distant from the previous one. This distance among transects could be greater in larger areas, but never lower than that. We tried to place the transects in a way we could achieve a distance-gradient (from transect to white-popinac patch) within the meta-community.

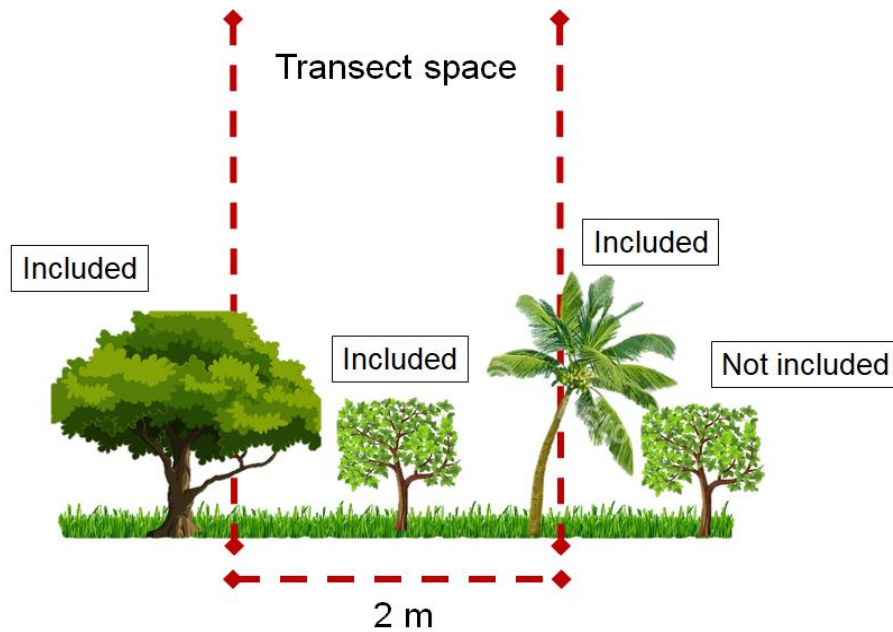


Fig. S4 Diagram representing the inclusion criterion of trees and shrubs regarding the transect area. If an individual was rooted outside but grows branches inside the transect, it was included. If it was totally inside, it was obviously included. In case it was rooted inside, but leans completely towards the outside, it was included as well. Therefore, the condition for not being included is if the individual was fully located outside the transect area.

Details on response variables

In the field, two types of variables were measured: (i) observed species richness, which accounts the total number of species within (meta-)communities; and (ii) abundance, described by two distinct measures: number of individuals (for arboreal stratum) and ground-cover percentage (for herbaceous stratum). We detail estimation methods and R-packages below, considering the three levels of response variables: local scale (alpha – community), regional scale (gamma– metacommunity) and among-sites diversity (β -diversity).

Local scale

This scale refers to the community level. We understand that the observed richness is a variable intrinsically dependent of sample size (total abundance) and number of samples, and may not be the best one to be used for ecological analysis

(Chao et al. 2014). Therefore, we used sample-coverage based methodology to estimate species richness that was developed by Chao and Jost (2012). This approach allows standardizing species richness-values based in community completeness, instead of sample size or sample number. Completeness is described by sample-coverage, a value that ranges from zero to one and refers to the proportion of the total amount of individuals in a community that were comprised by the sample.

We estimated species richness by using iNext package (v3.0.0; Hsieh, Ma, and Chao 2016) on R-Studio software (v4.1.2; R Core Team 2022). We estimated species richness separately between arboreal and herbaceous strata, once their abundance was described with different metrics. The package itself proposes the best sample-coverage that is possible among communities within a particular meta-community, and displays if the estimation for each community was obtained via extrapolation or rarefaction (Figure S5).

For the herbaceous stratum, we initially calculated the average ground cover percentage for each species (average among the 7 grids within a transect). Then, we applied sample-coverage based estimation considering the suggested sample-coverage value for each metacommunity. The average sample-coverage for herbaceous stratum among our 29 metacommunities was 0.98 (sd = 0.03, min = 0.81, max = 1.0). For the arboreal stratum, no averaging was needed once there were not subsamples within transects. The average sample-coverage for this stratum among all metacommunities was 0.74 (sd = 0.13, min = 0.30, max = 0.94).

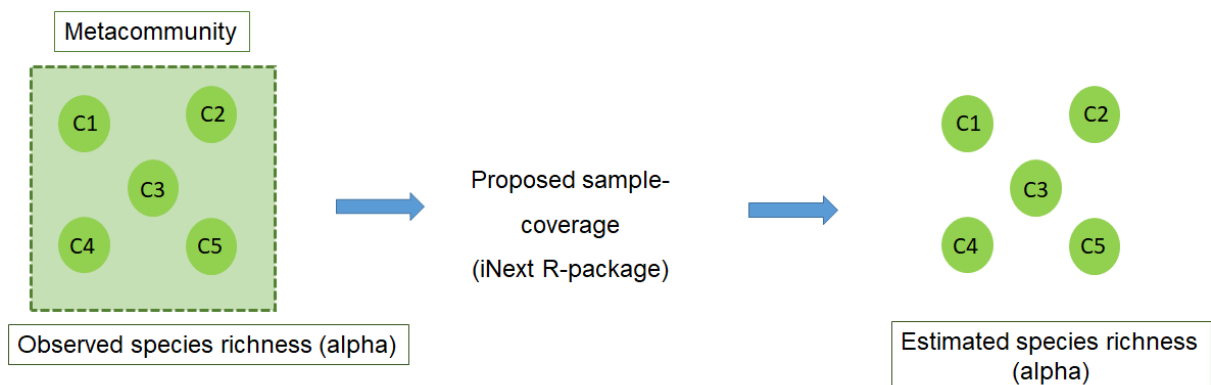


Fig. S5 Conceptual approach to species richness estimation on α -scale (community). C1 to C5 represent communities within a given metacommunity. Each metacommunity has its own proposed sample-coverage, obtained from iNext. This value allows communities to be standardized by their completeness, instead of size.

Regional scale

This scale refers to metacommunity level. We used iNext package (v3.0.0; Hsieh, Ma, and Chao 2016) to estimate species richness considering the same sample coverage among all metacommunities (for the arboreal stratum: 0.88; for the herbaceous stratum: 0.96). Finally, we obtained species richness estimations for this scale, as seen in Figure S6.

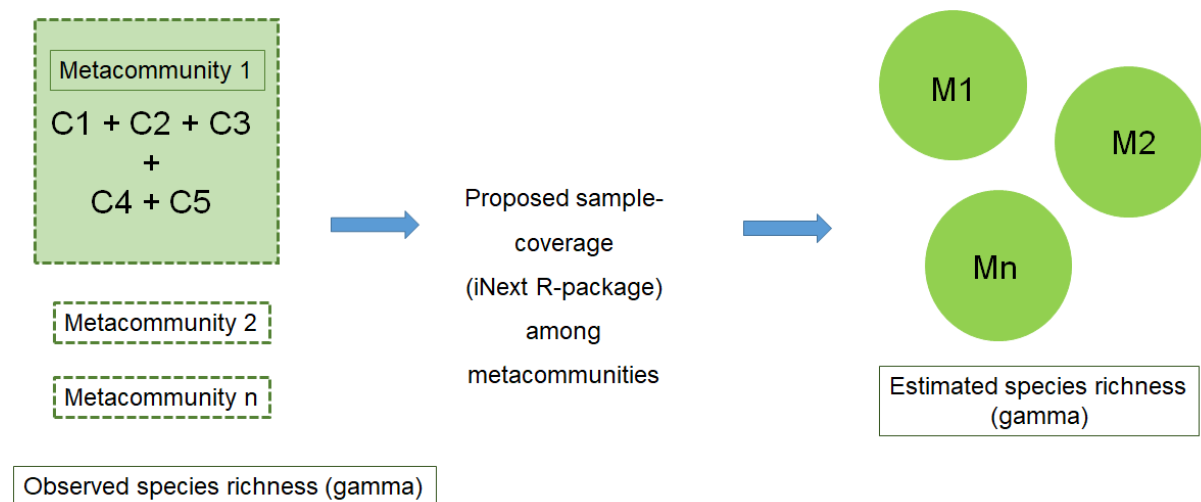


Fig. S6 Conceptual approach to species richness estimation on regional scale (metacommunity). C1 to C5 represent communities within a given metacommunity. Using iNext R-package, we could standardize metacommunities based on their completeness, instead of size.

β -diversity

Comparing differences in species composition among communities among regions is considered challenging as most β -diversity metrics are influenced by γ -diversity. Engel and collaborators (2021) developed a new metric called β -c, which employs the approach of coverage-based estimation to infer changes in non-random intraspecific aggregation, without being affected by γ -diversity (Engel et al. 2021). The new metric considers species composition based in equally complete communities, instead of standardizing them by total abundance. We used the “betaC” R-package (v0.1.0), which provides sample-coverage values for each metacommunity (Figure S7). The standard procedure is to use the smallest provided value (named “C-target”) to estimate β -c metric. The C-target for arboreal stratum was 0.48, which is close to

what is considered the minimum for unbiased estimation (Chao and Jost 2012), whereas for herbaceous it was 0.94.

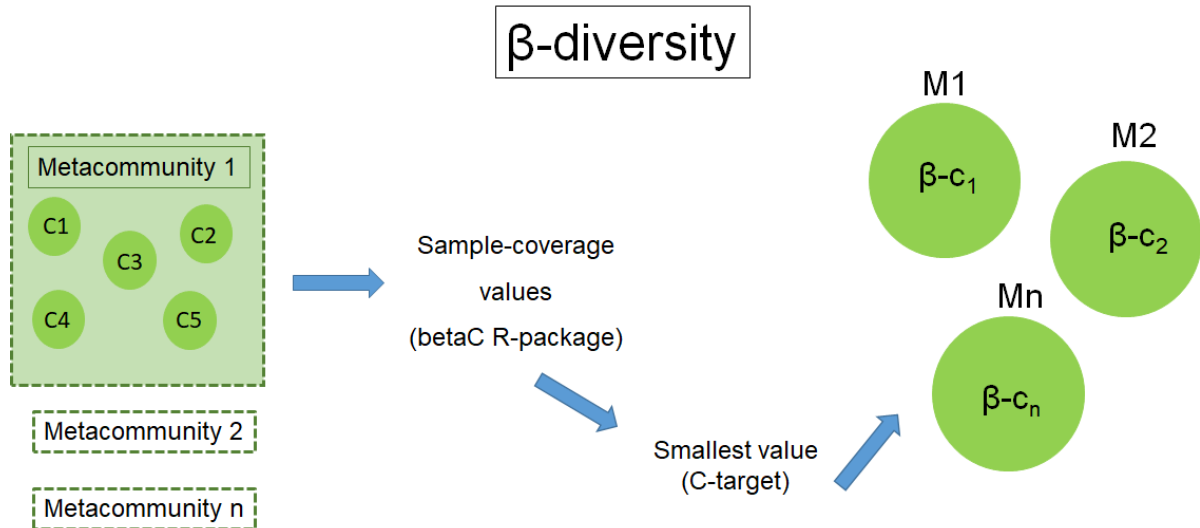


Fig. S7 Conceptual representation of β -diversity estimation (β -c metric). R-package betaC proposes sample-coverage values among metacommunities, and the smallest one (C-target) is chosen to estimate β -c, expressing differences in species composition among communities within a given metacommunity.

Details on model selection

For each response variable Y , we followed the sequence:

- 1- Standardized numeric predictor variables: all numeric predictor variables were standardized by using the "decostand" function in the "vegan" R-package (v2.6-4), with method = "total".
- 2- Chose predictor variables: the number of competing models for a given response variable depended on predictor variables and distribution families:
 - (i) Because we had three different time-related variables, each one was used on an individual model. For the regional scale and β -diversity, the first step stops here and goes to item 2. For the local scale, we followed to sub-items ii and iii before continuing to item 2.
 - (ii) There was a space-related variable at local scale (community-patch distance), so we created models with time and space and with only time.

(iii) All models at local scale had a random component, once our replicas were spatially dependent (communities within a metacommunity are not independent, they are spatially aggregated).

3- Chose the distribution family:

(i) If the response variable was continuous (e.g., estimated species richness), we competed models with two different adjustments in terms of distribution: Gaussian (with identity link function) and Gamma (with log link function).

(ii) If it the response variable was discrete (e.g., number of white-popinac adult individuals), we used the Poisson distribution (with log link function).

(iii) In the case of proportions (e.g., relative groundcover), we used the Beta distribution (logit link function).

4- Chose model type:

(i) For the regional scale and β -diversity, we employed GLMs (Generalized Linear Models), using “glm” function, in the “stats” R-package (v4.2.2).

(ii) For the local scale, we employed GLMMs (Generalized Linear Mixed-Effect Models), using “glmmTMB” function, in the “glmmTMB” R-package (v1.1.5).

5- Competed the models for each response variable:

(i) We used “compare_performance” function in the “performance” R-package (v0.10.1), which provides a series metrics to compare and evaluate models. The ranking was based upon AICc values (corrected Akaike Information Criterion). The lowest the AICc value, the better its predictive power (Aho, Derryberry, and Peterson 2014; Burnham, Anderson, and Huyvaert 2011).

(ii) We considered as equally plausible models those which AICc value was less than two unities greater than the lowest one ($\Delta_i < 2$). This metric is called Δ_i , being the difference between a given AICc value and the lowest AICc value found among the competing models ($\Delta_i = \text{AICc} - [\min]\text{AICc}$).

6- Considered R^2 values:

(i) For the regional scale and β -diversity, we did not interpret models which R^2 values were too small ($< \sim 0.05$), i.e., we discarded the hypothesis that they represented.

(ii) For the local scale, R^2 values were divided into two components: conditional R^2 (R^2_c), which comprises the entire GLMM model, both fixed and random components; and marginal R^2 (R^2_m), which comprises only the fixed component (that were

the predictor variables such as time or distance). If R^2_c was high but R^2_m was too low, it means that most of the variation in the response variable was associated with the random component, i.e., local effects that were not represented by our predictor variables. In cases where R^2_m was lower than approximately 0.05, the models were not interpreted as well (discarded hypothesis).

7- Plotted equally plausible models: this step did not include models with interaction between time-related variables and the categorical variable “origin” (native/INNS).

(i) For each response variable, we plotted the respective coefficient confidence intervals from all equally plausible models. We used the function “modelplot” from “modelsummary” R-package (v1.3.0). Each model was plotted separately.

(ii) After plotting, we checked all coefficient confidence intervals regarding its inclusion of zero. In case of including zero, it means that the relationship was not strong, once “no effect” (zero) is also within the probability range.

(iii) Among all equally plausible models, some of them might have variables whose coefficient confidence intervals included zero, whereas the other ones did not. In those cases, once they are considered as equally plausible, we chose the ones not including zero to be plotted at “Results” section. If none of them included zero, we chose the one with highest R^2 (or correlates) value to be plotted.

8- Final plots:

(i) Plots were made with “modelsummary” R-package (v1.3.0). In case of interaction, we used “interactions” R-package (v1.1.5).

Results

Diversity metrics

Local scale

Table S1 Diversity metrics at local scale. ESR = Estimated Species Richness.

Metacommunity	Community	ESR – Arboreal stratum	ESR – Herbaceous stratum
a	1	5.64	10.77
a	2	4.85	15.51
a	3	9.76	8.04
a	4	9.82	10.02
a	5	7.35	13.73
aa	1	11	8.15
aa	2	15.1	15.14
aa	3	15.41	11.62
aa	4	12.3	10.22
aa	5	14.52	13
bb	1	19.89	5.99
bb	2	2.86	9.42
bb	3	8.61	5.96
bb	4	9.63	5
c	1	6.52	9.96
c	2	14.66	18.89
c	3	13.69	6.92
c	4	12.95	13.97
c	5	7.44	25.06
cc	1	5.7	6.99
cc	2	9.01	10.99
cc	3	3.78	11.62
cc	4	5.67	11
cc	5	4.12	13
dd	1	25.03	8.82
dd	2	13.38	10.79
dd	3	18.05	10.58
dd	4	8.14	11.85

dd	5	10.76	7.34
e	1	9.65	8
e	2	19.84	12.41
e	3	10.47	8
e	4	22.23	16.06
e	5	13.58	9.43
ee	1	24.13	11
ee	2	11.21	15
ee	3	7.49	10
ee	4	4.41	6
f	1	7.1	6.75
f	2	10.57	10.85
f	3	17.11	13.18
f	4	5.91	6.87
f	5	4.33	14.84
g	1	13.09	16.54
g	2	8.27	11.64
g	3	4.87	10.86
g	4	17.87	10.84
g	5	13.95	8.86
gg	1	27.1	14
gg	2	14.46	12
gg	3	14.03	12
gg	4	17.35	13.63
gg	5	3.34	7
h	1	14.2	26.04
h	2	10.77	11.14
h	3	24.17	13.08
h	4	19.19	13.42
ii	1	10.2	4.99
ii	2	4.25	5.43
ii	3	12.35	6
ii	4	19.37	3.99
j	1	6.52	11.18
j	2	9.05	5.81
j	4	21.07	4.86
j	5	7.05	5.96
jj	1	6.31	9.98
jj	2	8.38	13.98
jj	3	32.99	11.18

jj	4	18.06	11.98
jj	5	3.96	9.85
k	1	9.19	9.95
k	2	10.56	11.44
k	3	9.43	12.79
k	4	12.14	14.9
k	5	3.09	8.96
kk	2	9.89	8
kk	3	8.94	13
kk	4	5.06	11.24
kk	5	4.89	6
mm	1	9.89	10.24
mm	2	5.31	6
mm	3	9.89	6
mm	4	3.38	4
mm	5	5.19	6
n	1	21.05	18.86
n	2	18.08	12.99
n	3	16.96	18.7
n	4	21.34	14.46
n	5	12.24	13
nn	1	18.04	7
nn	2	20.08	8
nn	3	23.38	8
nn	4	7.13	9.16
p	1	11.89	8.43
p	2	6.56	19.03
p	3	2.88	2.49
p	4	8.46	1.38
p	5	5.71	3.71
q	1	10.42	6.31
q	2	9.92	3.93
q	3	3.36	11.85
q	4	16.83	11
r	1	1.74	4.98
r	2	15.98	9.85
r	3	5.82	5.95
r	4	4.31	10.84
s	1	4.55	24.63
s	2	2.49	13.88

s	3	4.39	6.96
s	4	14.71	8.97
t	1	11.68	21.86
t	2	17.92	21.04
t	3	28.13	14.99
t	4	18.51	14.08
v	1	16.87	11.69
v	2	15.97	15.76
v	3	23.4	10.16
v	4	21.53	12.9
x	1	12.63	11.64
x	2	12.34	1.53
x	3	15.79	12.11
x	4	32.14	9.51
y	1	14.15	5.84
y	2	9.1	10.98
y	3	23.02	12.73
y	4	8.49	11.07
z	1	6.82	7.99
z	2	19	7.85
z	3	7.73	6.33
z	4	1.62	1.96
z	5	6.71	4.12

Regional scale and β -diversity

Table S1 Diversity metrics at regional scale. ESR = Estimated Species Richness. β -c = among-sites diversity metric (β -diversity).

Metacommunity	ESR – Arboreal stratum	ESR – Herbaceous stratum	β c - Arboreal stratum	β c – Herbaceous stratum
a	18.01	19.56	1.11	1.43
aa	29.45	20.13	1.16	1.49
bb	17	11.57	1.1	1.4
c	21.21	30.9	1.17	1.83
cc	10.33	15.17	1.15	1.29
dd	16.63	13.07	1.01	1.2
e	28.68	17.37	1.24	1.39
ee	35.18	20.37	1.39	1.61

f	19.8	22.49	1.12	1.73
g	37.1	42.17	1.36	1.65
gg	30.83	23.85	1.17	1.78
h	28.48	28.34	1.09	1.6
ii	19.44	6.02	1.17	1.28
j	28.17	13.05	1.13	1.6
jj	60.4	23.64	1.21	1.74
k	27.79	30.85	1.17	1.88
kk	30.47	16.56	1.09	1.52
mm	19.36	13.55	1.08	1.74
n	22.8	22.89	1.09	1.37
nn	26.9	11.04	1.13	1.25
p	27.75	19.18	1.2	1.28
q	22.98	13.88	1.15	1.16
r	18.4	14.97	1.1	1.56
s	25.1	25.38	1.13	1.71
t	35.94	26.99	1.14	1.36
v	39.79	22.47	1.07	1.53
x	30.16	17.27	1.27	1.55
y	30.55	14.75	1.15	1.31
z	26.33	9.26	1.31	1.41

Model selection

Local scale

Table S3 Model selection statistics for all competing models at local scale (community). AICc refers to the corrected Akaike Information Criterion (AIC). Δ_i refers to the difference between a given AICc value and the lowest among competing models. AICc (W) refers to AICc “weight”, being interpreted as the model’s relative likelihood. R^2_c refers to amount of variation explained by the entire model, both fixed and random components, whereas R^2_m refers only to the fixed component. ADL = Average-diameter (dbh - cm) of largest white-popinac trees within a metacommunity (region); A = white-popinac patch’s age-proxy (years); BA = white-popinac patch’s basal area (m^2). S = strata (herbaceous and arboreal); D = community/patch distance (m); O = Origin (native/INNS); RE = random effects. Null models have $y \sim 1$ structure.

Model structure	Distribution	AICc	AICc(wt)	Δi	R ² c	R ² m
Species richness						
Null model 1	Gamma	1544.33	0.33	0	0.18	0
A + S + RE	Gamma	1545.71	0.17	1.37	0.18	0.01
BA + S + RE	Gamma	1545.81	0.16	1.47	0.18	0.01
ADL + S + RE	Gamma	1545.96	0.15	1.62	0.18	0.01
A + D + S + RE	Gamma	1547.73	0.06	3.39	0.19	0.01
BA + D + S + RE	Gamma	1547.77	0.06	3.43	0.19	0.01
ADL + D + S + RE	Gamma	1547.95	0.05	3.61	0.19	0.01
Null model 2	Gaussian	1583.31	0	38.97	0.19	0
ADL + S + RE	Gaussian	1583.70	0	39.36	0.20	0.01
BA + S + RE	Gaussian	1583.74	0	39.40	0.20	0.01
A + S + RE	Gaussian	1583.77	0	39.43	0.20	0.01
ADL + D + S + RE	Gaussian	1585.63	0	41.29	0.22	0.01
BA + D + S + RE	Gaussian	1585.67	0	41.33	0.22	0.01
A + D + S + RE	Gaussian	1585.76	0	41.43	0.22	0.01
Species richness - herbaceous stratum (native/INNS)						
ADL * O + RE	Gamma	1035.79	0.22	0	0.57	0.45
ADL * O + D + RE	Gamma	1035.88	0.21	0.09	0.57	0.46
A + O + D + RE	Gamma	1037.15	0.11	1.36	0.55	0.47
A + O + RE	Gamma	1037.67	0.08	1.87	0.56	0.45
BA + O + RE	Gamma	1038.26	0.06	2.46	0.56	0.45
BA + O + D + RE	Gamma	1038.27	0.06	2.48	0.56	0.46
BA * O + RE	Gamma	1038.99	0.04	3.20	0.56	0.45
ADL * O + D + RE	Gamma	1039.02	0.04	3.23	0.5	0.46
BA * O + D + RE	Gamma	1039.05	0.04	3.26	0.56	0.46
A * O + D + RE	Gamma	1039.22	0.03	3.42	0.55	0.47
ADL + O + RE	Gamma	1039.26	0.03	3.47	0.56	0.44
A * O + RE	Gamma	1039.73	0.03	3.93	0.56	0.45
A + O + RE	Gamma	1199.21	0	163.42	0.48	0.37
A * O + RE	Gaussian	1199.58	0	163.79	0.48	0.38
BA + O + RE	Gaussian	1200.31	0	164.52	0.48	0.37
ADL * O + RE	Gaussian	1200.68	0	164.89	0.49	0.37
ADL + O + RE	Gaussian	1200.69	0	164.90	0.48	0.36
A + O + D + RE	Gaussian	1200.91	0	165.11	0.47	0.37
A * O + D + RE	Gaussian	1201.30	0	165.51	0.48	0.38
BA + O + D + RE	Gaussian	1202.12	0	166.32	0.48	0.37

ADL + O + D + RE	Gaussian	1202.37	0	166.58	0.48	0.37
BA * O + RE	Gaussian	1202.39	0	166.59	0.48	0.37
ADL * O + D + RE	Gaussian	1202.39	0	166.60	0.48	0.37
Null model 1	Gaussian	1202.43	0	166.64	0.08	0
BA * O + D + RE	Gaussian	1204.21	0	168.41	0.48	0.37
Null model 2	Gaussian	1326.72	0	290.93	0.06	0

Relative tree/shrub abundance

Null model	Beta	-70.15	0.34	0	0.64	0
BA + RE	Beta	-69.16	0.21	0.98	0.64	0.05
A + RE	Beta	-68.47	0.15	1.67	0.64	0.02
ADL + RE	Beta	-68.01	0.12	2.13	0.64	0
BA + D + RE	Beta	-67.03	0.07	3.11	0.64	0.05
A + D + RE	Beta	-66.29	0.05	3.85	0.64	0.02
ADL + D + RE	Beta	-65.84	0.04	4.30	0.64	0

Relative ground cover - native/INNS

Null model	Beta	-26.26	0.35	0	0.99	0
ADL + RE	Beta	-25.39	0.23	0.87	0.99	0.05
A + RE	Beta	-24.14	0.12	2.11	0.99	0
BA + RE	Beta	-24.12	0.12	2.13	0.99	0
ADL + D + RE	Beta	-23.32	0.08	2.94	0.99	0.05
A + D + RE	Beta	-22.20	0.04	4.05	0.99	0
BA + D + RE	Beta	-22.18	0.04	4.07	0.99	0

White-popinac abundance (adult)

BA + D + RE	Poisson	520.31	0.39	0	0.87	0.68
A + D + RE	Poisson	520.82	0.30	0.50	0.87	0.67
ADL + D + RE	Poisson	520.83	0.30	0.51	0.87	0.68
BA + RE	Poisson	654.79	0	134.47	0.79	0.08
Null model	Poisson	655.86	0	135.54	0.79	0
ADL + RE	Poisson	657.14	0	136.82	0.79	0.02
A + RE	Poisson	657.95	0	137.63	0.79	0

White-popinac's seedlings relative ground cover

Null model	Beta	-292.32	0.33	0	0.09	0
BA + RE	Beta	-291.23	0.19	1.09	0.11	0.02
A + RE	Beta	-290.35	0.12	1.97	0.09	0
ADL + RE	Beta	-290.12	0.11	2.19	0.10	0
BA + D + RE	Beta	-289.85	0.09	2.47	0.20	0.04

A + D + RE	Beta	-289.03	0.06	3.29	0.19	0.02
ADL + D + RE	Beta	-289.00	0.06	3.32	0.19	0.02

Regional scale and β -diversity

Table S4 Model selection statistics for all competing models describing variation at regional scale (metacommunity) and β -diversity. (*) Models fitted with Poisson and Gamma distributions are with pseudo- R^2 values, whereas models fitted with Gamma distribution are associated with Nagelkerke R^2 , which are mathematically different from a regular R^2 , but may be interpreted in a similar way. Abbreviations as in Table S3.

Model structure	Distribution	AICc	AICc(wt)	Δi	R^2 (*)
Species richness					
BA + S	Gamma	411.02	0.44	0	0.20
ADL + S	Gamma	411.52	0.34	0.49	0.19
A + S	Gamma	412.69	0.19	1.67	0.17
BA + S	Gaussian	419.08	0	8.05	0.18
Null model 1	Gamma	419.12	0	8.10	0
ADL + S	Gaussian	419.77	0	8.74	0.17
A + S	Gaussian	420.73	0	9.71	0.16
Null model 2	Gaussian	426.72	0	15.70	0
Species richness (native/INNS)					
A * O	Gamma	375.40	0.23	0	0.88
A + O	Gamma	375.46	0.22	0.06	0.88
BA * O	Gamma	375.60	0.21	0.20	0.88
BA + O	Gamma	376.27	0.15	0.86	0.88
ADL + O	Gamma	377.04	0.10	1.64	0.88
ADL * O	Gamma	377.87	0.06	2.46	0.88
ADL + O	Gaussian	421.69	0	46.28	0.7
BA + O	Gaussian	421.82	0	46.41	0.78
A + O	Gaussian	422.03	0	46.6	0.78
BA * O	Gaussian	422.39	0	46.99	0.79
ADL * O	Gaussian	422.83	0	47.43	0.78
A * O	Gaussian	424.12	0	48.72	0.78

Null model 1	Gamma	476.33	0	100.93	0
Null model 2	Gaussian	506.18	0	130.78	0
Relative tree/shrub abundance					
Null model	Beta	-34.83	0.42	0	NA
BA	Beta	-34.03	0.28	0.80	0.05
ADL	Beta	-32.82	0.15	2.01	0.01
A	Beta	-32.53	0.13	2.29	0
White-popinac abundance (adult)					
BA	Poisson	472.24	0.99	0	0.52
Null model	Poisson	491.38	0	19.13	0
A	Poisson	493.32	0	21.07	0.01
ADL	Poisson	493.44	0	21.19	0.01
White-popinac's seedlings relative ground cover					
BA	Beta	-120.59	0.92	0	0.18
ADL	Beta	-114.30	0.04	6.29	0.1
Null model	Beta	-113.64	0.03	6.95	NA
A	Beta	-111.01	0.01	9.57	0
β-c					
ADL + S	Gamma	-60.17	0.66	0	0.63
BA + S	Gamma	-58.34	0.26	1.83	0.62
A + S	Gamma	-55.34	0.06	4.83	0.60
ADL + S	Gaussian	-50.26	0	9.91	0.60
BA + S	Gaussian	-48.11	0	12.06	0.58
A + S	Gaussian	-45.21	0	14.95	0.56
Null model 1	Gamma	-6.51	0	53.66	0
Null model 2	Gaussian	-1.83	0	58.34	0

List of recorded species

Table S5 All species (native and INNS) recorded in our floristic survey. Species tagged with (*) are INNS.

Species		Family
<i>Acalypha velamea</i>	Baill.	Euphorbiaceae
<i>Acanthocladus brasiliensis</i>	(A.St.-Hil. & Moq.) Klotzsch ex Hassk.	Polygalaceae
<i>Actinostemon klotzschii</i>	(Didr.) Pax	Euphorbiaceae
<i>Adenocalymma bracteatum</i>	(Cham.) DC.	Bignoniaceae
<i>Albizia polycephala</i>	(Benth.) Killip ex Record	Fabaceae
<i>Alchornea glandulosa</i>	Poepp. & Endl.	Euphorbiaceae
<i>Alchornea sidifolia</i>	Müll.Arg.	Euphorbiaceae
<i>Allophylus edulis</i>	(A.St.-Hil. et al.) Hieron. ex Niederl.	Sapindaceae
<i>Aloysia virgata</i>	(Ruiz & Pav.) Juss.	Verbenaceae
<i>Alternanthera brasiliensis</i>	(L.) Kuntze	Amaranthaceae
<i>Alternanthera philoxeroides</i>	(Mart.) Griseb.	Amaranthaceae
<i>Ambrosia polystachya</i>	DC.	Asteraceae
<i>Anadenanthera colubrina</i>	(Vell.) Brenan	Fabaceae
<i>Anemia phyllitidis</i>	(L.) Sw.	Anemiaceae
<i>Annona dolabripetala</i>	Raddi	Annonaceae
<i>Aristolochia labiata</i>	Willd.	Aristolochiaceae
<i>Aristolochia triangularis</i>	Cham. & Schltld.	Aristolochiaceae
<i>Asclepias curassavica</i>	L.	Apocynaceae
<i>Aspilia pascalioides</i>	Griseb.	Asteraceae
<i>Astronium graveolens</i>	Jacq.	Anacardiaceae
<i>Baccharis dracunculifolia</i>	DC.	Asteraceae
<i>Baccharis trinervis</i>	Pers.	Asteraceae
<i>Banisteriopsis muricata</i>	(Cav.) Cuatrec.	Malpighiaceae
<i>Banisteriopsis nummifera</i>	(A.Juss.) B.Gates	Malpighiaceae
<i>Banisteriopsis</i> sp	-	Malpighiaceae
<i>Banisteriopsis stellaris</i>	(Griseb.) B.Gates	Malpighiaceae
<i>Bastardiopsis densiflora</i>	(Hook. & Arn.) Hassl.	Malvaceae
<i>Bauhinia forficata</i>	Link	Fabaceae
<i>Bauhinia longifolia</i>	(Bong.) Steud.	Fabaceae
<i>Bauhinia unguolata</i>	L.	Fabaceae

<i>Bernardia pulchella</i>	(Baill.) Müll.Arg.	Euphorbiaceae
<i>Bidens pilosa</i>	L.	Asteraceae
<i>Bidens subalternans</i>	DC.	Asteraceae
<i>Blechnum occidentale</i>	L.	Blechnaceae
<i>Boehmeria caudata</i>	Sw.	Urticaceae
<i>Boehmeria nivea</i> *	(L.) Gaudich.	Urticaceae
<i>Buddleja stachyoides</i>	Cham. & Schltl.	Scrophulariaceae
<i>Calliandra foliolosa</i>	Benth.	Fabaceae
<i>Callisia monandra</i>	(Sw.) Schult.f.	Commelinaceae
<i>Callisthene fasciculata</i>	Mart.	Vochysiaceae
<i>Campomanesia guaviroba</i>	(DC.) Kiaersk.	Myrtaceae
<i>Campomanesia sp</i>	-	Myrtaceae
<i>Capsicum baccatum</i>	L.	Solanaceae
<i>Cardiospermum grandiflorum</i>	Sw.	Sapindaceae
<i>Cardiospermum halicacabum</i>	L.	Sapindaceae
<i>Casearia decandra</i>	Jacq.	Salicaceae
<i>Casearia gossypiosperma</i>	Briq.	Salicaceae
<i>Casearia sylvestris</i>	Sw.	Salicaceae
<i>Cecropia pachystachya</i>	Trécul	Urticaceae
<i>Cedrela fissilis</i>	Vell.	Meliaceae
<i>Ceiba speciosa</i>	(A.St.-Hil.) Ravenna	Malvaceae
<i>Celtis iguanaea</i>	(Jacq.) Sarg.	Cannabaceae
<i>Cereus hildmannianus</i>	K.Schum.	Cactaceae
<i>Cestrum mariquitense</i>	Kunth	Solanaceae
<i>Chamaecrista nictitans</i>	(L.) Moench	Fabaceae
<i>Chaptalia integerrima</i>	(Vell.) Burkart	Asteraceae
<i>Chaptalia nutans</i>	(L.) Pol.	Asteraceae
<i>Chionanthus filiformis</i>	(Vell.) P.S.Green	Oleaceae
<i>Christella dentata</i> *	(Forssk.) Brownsey & Jermy	Thelypteridaceae
<i>Chromolaena laevigata</i>	(Lam.) R.M.King & H.Rob.	Asteraceae
<i>Chromolaena maximiliani</i>	(Schrud. ex DC.) R.M.King & H.Rob.	Asteraceae
<i>Chromolaena odorata</i>	(L.) R.M.King & H.Rob.	Asteraceae
<i>Chromolaena squalida</i>	(DC.) R.M.King & H.Rob.	Asteraceae
<i>Chrysophyllum marginatum</i>	(Hook. & Arn.) Radlk.	Sapotaceae
<i>Cissampelos glaberrima</i>	A.St.-Hil.	Menispermaceae

<i>Cissus verticillata</i>	(L.) Nicolson & C.E.Jarvis	Vitaceae
<i>Citharexylum myrianthum</i>	Cham.	Verbenaceae
<i>Citrus x limonia</i> *	Osbeck (pro. sp.)	Rutaceae
<i>Commelina benghalensis</i> *	L.	Commelinaceae
<i>Commelina diffusa</i>	Burm.f.	Commelinaceae
<i>Commelina erecta</i>	L.	Commelinaceae
<i>Condylocarpum isthmicum</i>	(Vell.) A.DC.	Apocynaceae
<i>Copaifera langsdorffii</i>	Desf.	Fabaceae
<i>Cordia africana</i> *	Lam.	Boraginaceae
<i>Cordia americana</i>	(L.) Gottschling & J.S.Mill.	Boraginaceae
<i>Cordia superba</i>	Cham.	Boraginaceae
<i>Cordia trichotoma</i>	(Vell.) Arráb. ex Steud.	Boraginaceae
<i>Cordyline spectabilis</i>	Kunth & Bouché	Asparagaceae
<i>Coutarea hexandra</i>	(Jacq.) K.Schum.	Rubiaceae
<i>Critonia megaphylla</i>	(Baker) R.M.King & H.Rob.	Asteraceae
<i>Crotalaria incana</i>	L.	Fabaceae
<i>Croton floribundus</i>	Spreng.	Euphorbiaceae
<i>Croton urucurana</i>	Baill.	Euphorbiaceae
<i>Ctenodon elegans</i>	(Schltdl. & Cham.) D.B.O.S.Car- doso & A.Delgado	Fabaceae
<i>Cupania vernalis</i>	Cambess.	Sapindaceae
<i>Cuphea carthagenensis</i>	(Jacq.) J.F.Macbr.	Lythraceae
<i>Cyperus aggregatus</i>	(Willd.) Endl.	Cyperaceae
<i>Cyperus difformis</i> *	L.	Cyperaceae
<i>Cyperus esculentus</i> *	L.	Cyperaceae
<i>Cyperus lanceolatus</i>	Poir.	Cyperaceae
<i>Cyperus laxis</i>	Lam.	Cyperaceae
<i>Cyperus surinamensis</i>	Rottb.	Cyperaceae
<i>Cyrtocymura scorpioides</i>	(Lam.) H.Rob.	Asteraceae
<i>Dahlstedtia muehlbergiana</i>	(Hassl.) M.J.Silva & A.M.G.Azevedo	Fabaceae
<i>Dalbergia frutescens</i>	(Vell.) Britton	Fabaceae
<i>Dalechampia triphylla</i>	Lam.	Euphorbiaceae
<i>Dasyphyllum vagans</i>	(Gardner) Cabrera	Asteraceae
<i>Dendropanax cuneatus</i>	(DC.) Decne. & Planch.	Araliaceae
<i>Desmodium incanum</i>	(Sw.) DC.	Fabaceae

<i>Desmodium tortuosum</i>	(Sw.) DC.	Fabaceae
<i>Diatenopteryx sorbifolia</i>	Radlk.	Sapindaceae
<i>Dicella bracteosa</i>	(A.Juss.) Griseb.	Malpighiaceae
<i>Dichondra macrocalyx</i>	Meisn.	Convolvulaceae
<i>Dicksonia sellowiana</i>	Hook.	Dicksoniaceae
<i>Dioscorea multiflora</i>	Mart. ex Griseb.	Dioscoreaceae
<i>Dioscorea piperifolia</i>	Humb. & Bonpl. ex Willd.	Dioscoreaceae
<i>Distimake aegyptius</i>	(L.) A.R. Simões & Staples	Convolvulaceae
<i>Distimake dissectus</i>	(Jacq.) A.R. Simões & Staples	Convolvulaceae
<i>Distimake macrocalyx</i>	(Ruiz & Pav.) A.R. Simões & Staples	Convolvulaceae
<i>Dolichandra unguis-cati</i>	(L.) L.G.Lohmann	Bignoniaceae
<i>Elephantopus mollis</i>	Kunth	Asteraceae
<i>Emilia fosbergii</i> *	Nicolson	Asteraceae
<i>Endlicheria paniculata</i>	(Spreng.) J.F.Macbr.	Lauraceae
<i>Enterolobium contortisiliquum</i>	(Vell.) Morong	Fabaceae
<i>Erythrina speciosa</i>	Andrews	Fabaceae
<i>Erythroxylum deciduum</i>	A.St.-Hil.	Erythroxylaceae
<i>Erythroxylum pelleterianum</i>	A.St.-Hil.	Erythroxylaceae
<i>Esenbeckia febrifuga</i>	(A.St.-Hil.) A. Juss. ex Mart.	Rutaceae
<i>Eugenia uniflora</i>	L.	Myrtaceae
<i>Euphorbia comosa</i>	Vell.	Euphorbiaceae
<i>Ficus guaranitica</i>	Chodat	Moraceae
<i>Fimbristylis autumnalis</i>	(L.) Roem. & Schult.	Cyperaceae
<i>Fridericia chica</i>	(Bonpl.) L.G.Lohmann	Bignoniaceae
<i>Fridericia samyoides</i>	(Cham.) L.G.Lohmann	Bignoniaceae
<i>Garcinia gardneriana</i>	(Planch. & Triana) Zappi	Clusiaceae
<i>Gouania sp</i>	-	Rhamnaceae
<i>Gouania ulmifolia</i>	Hook. & Arn.	Rhamnaceae
<i>Guadua angustifolia</i> *	Kunth	Poaceae
<i>Guarea guidonia</i>	(L.) Sleumer	Meliaceae
<i>Guarea macrophylla</i>	Vahl	Meliaceae
<i>Guazuma ulmifolia</i>	Lam.	Malvaceae
<i>Gymnanthes klotzschiana</i>	Müll.Arg.	Euphorbiaceae
<i>Handroanthus impetiginosus</i>	(Mart. ex DC.) Mattos	Bignoniaceae
<i>Handroanthus umbellatus</i>	(Sond.) Mattos	Bignoniaceae

<i>Heliotropium transalpinum</i>	Vell.	Boraginaceae
<i>Heterocondylus alatus</i>	(Vell.) R.M.King & H.Rob.	Asteraceae
<i>Heteropterys argyrophaea</i>	A.Juss.	Malpighiaceae
<i>Heteropterys sp</i>	-	Malpighiaceae
<i>Hildaea pallens</i>	(Sw.) C.Silva & R.P.Oliveira	Poaceae
<i>Hydrocotyle leucocephala</i>	Cham. & Schltldl.	Araliaceae
<i>Hyptis sp</i>	-	Lamiaceae
<i>Inga edulis</i>	Mart.	Fabaceae
<i>Lochroma arborescens</i>	(L.) J.M.H. Shaw	Solanaceae
<i>Ipomoea bonariensis</i>	Hook.	Convolvulaceae
<i>Ipomoea cairica</i> *	(L.) Sweet	Convolvulaceae
<i>Ipomoea nil</i>	(L.) Roth	Convolvulaceae
<i>Ipomoea saopaulista</i>	O'Donell	Convolvulaceae
<i>Iresine diffusa</i>	Humb. & Bonpl. ex Willd.	Amaranthaceae
<i>Jacaranda mimosifolia</i> *	D. Don	Bignoniaceae
<i>Jacquemontia heterantha</i>	(Nees & Mart.) Hallier f.	Convolvulaceae
<i>Justicia carnea</i>	Lindl.	Acanthaceae
<i>Lafoensia pacari</i>	A.St.-Hil.	Lythraceae
<i>Lantana camara</i>	L.	Verbenaceae
<i>Lantana trifolia</i>	L.	Verbenaceae
<i>Laportea aestuans</i>	(L.) Chew	Urticaceae
<i>Lasiacis ligulata</i>	Hitchc. & Chase	Poaceae
<i>Leandra sp</i>	-	Melastomataceae
<i>Leonotis nepetifolia</i> *	(L.) R.Br.	Lamiaceae
<i>Lepismium cruciforme</i>	(Vell.) Miq.	Cactaceae
<i>Lessingianthus glabratus</i>	(Less.) H.Rob.	Asteraceae
<i>Leucaena leucocephala</i> *	(Lam.) de Wit	Fabaceae
<i>Lippia origanoides</i>	Kunth	Verbenaceae
<i>Lithraea molleoides</i>	(Vell.) Engl.	Anacardiaceae
<i>Luehea divaricata</i>	Mart.	Malvaceae
<i>Machaerium brasiliense</i>	Vogel	Fabaceae
<i>Machaerium hirtum</i>	(Vell.) Stelfeld	Fabaceae
<i>Machaerium nyctitans</i>	(Vell.) Benth.	Fabaceae
<i>Machaerium stiptatum</i>	Vogel	Fabaceae
<i>Machaerium villosum</i>	Vogel	Fabaceae
<i>Matayba elaeagnoides</i>	Radlk.	Sapindaceae

<i>Megathyrsus maximus</i> *	(Jacq.) B.K.Simon & S.W.L.Jacobs	Poaceae
<i>Melia azedarach</i> *	L.	Meliaceae
<i>Melochia oyramidata</i>	L.	Malvaceae
<i>Melochia villosa</i>	(Mill.) Fawc. & Rendle	Malvaceae
<i>Mesosphaerum pectinatum</i> *	(L.) Kuntze	Lamiaceae
<i>Mesosphaerum sidifolium</i>	(L'Hér.) Harley & J.F.B.Pastore	Lamiaceae
<i>Miconia ligustroides</i>	(DC.) Naudin	Melastomataceae
<i>Miconia</i> sp	-	Melastomataceae
<i>Mikania cordifolia</i>	(L.f.) Willd.	Asteraceae
<i>Mikania glomerata</i>	Spreng.	Asteraceae
<i>Mimosa bimucronata</i>	(DC.) Kuntze	Fabaceae
<i>Mimosa caesalpiniiifolia</i> *	Benth.	Fabaceae
<i>Mollinedia widgrenii</i>	A.DC.	Monimiaceae
<i>Momordica charantia</i> *	L.	Cucurbitaceae
<i>Monteverdia aquifolium</i>	(Mart.) Biral	Celastraceae
<i>Monteverdia gonoclada</i>	(Mart.) Biral	Celastraceae
<i>Moquilea tomentosa</i>	Benth.	Chrysobalanaceae
<i>Moquiniastrium polymorphum</i>	(Less.) G. Sancho	Asteraceae
<i>Muelleria campestris</i>	(Mart. ex Benth.) M.J. Silva & A.M.G. Azevedo	Fabaceae
<i>Murraya paniculata</i> *	(L.) Jack	Rutaceae
<i>Myrcia neoclusiifolia</i>	A.R.Lourenço & E.Lucas	Myrtaceae
<i>Myrciaria floribunda</i>	(H.West ex Willd.) O.Berg	Myrtaceae
<i>Myroxylon peruiferum</i>	L.f.	Fabaceae
<i>Myrsine coriacea</i>	(Sw.) R.Br. ex Roem. & Schult.	Primulaceae
<i>Myrsine guianensis</i>	(Aubl.) Kuntze	Primulaceae
<i>Nectandra oppositifolia</i>	Nees & Mart.	Lauraceae
<i>Neonotonia wightii</i> *	(Graham ex Wight & Arn.) J.A.Lackey	Fabaceae
<i>Ocotea puberula</i>	(Rich.) Nees	Lauraceae
<i>Ocotea pulchella</i>	(Nees & Mart.) Mez	Lauraceae
<i>Ocotea velloziana</i>	(Meisn.) Mez	Lauraceae
<i>Oeceoclades maculata</i>	(Lindl.) Lindl.	Orchidaceae
<i>Olyra ciliatifolia</i>	Raddi	Poaceae

<i>Oplismenus hirtellus</i>	(L.) P.Beauv.	Poaceae
<i>Oxalis debilis</i>	Kunth	Oxalidaceae
<i>Oxalis triangularis</i>	A.St.-Hil.	Oxalidaceae
<i>Parapiptadenia rigida</i>	(Benth.) Brenan	Fabaceae
<i>Passiflora edulis</i>	Sims	Passifloraceae
<i>Passiflora suberosa</i>	L.	Passifloraceae
<i>Paullinia elegans</i>	Cambess.	Sapindaceae
<i>Paullinia rhomboidea</i>	Radlk.	Sapindaceae
<i>Pavonia communis</i>	A.St.-Hil.	Malvaceae
<i>Peltophorum dubium</i>	(Spreng.) Taub.	Fabaceae
<i>Pereskia grandifolia</i>	Haw.	Cactaceae
<i>Petrea volubilis</i>	L.	Verbenaceae
<i>Phyllanthus niruri</i>	L.	Phyllanthaceae
<i>Phyllanthus orbiculatus</i>	Rich.	Phyllanthaceae
<i>Piper aduncum</i>	L.	Piperaceae
<i>Piper amalago</i>	L.	Piperaceae
<i>Piper glabratum</i>	Kunth	Piperaceae
<i>Piptadenia gonoacantha</i>	(Mart.) J.F.Macbr.	Fabaceae
<i>Pityrogramma trifoliata</i>	(L.) R.M.Tryon	Pteridaceae
<i>Platypodium elegans</i>	Vogel	Fabaceae
<i>Plinia peruviana</i>	(Poir.) Govaerts	Myrtaceae
<i>Poecilanthus parviflora</i>	Benth.	Fabaceae
<i>Pombalia atropurpurea</i>	(A.St.-Hil.) Paula-Souza	Violaceae
<i>Porophyllum ruderale</i>	(Jacq.) Cass.	Asteraceae
<i>Portulaca oleracea</i>	L.	Portulacaceae
<i>Prunus myrtifolia</i>	(L.) Urb.	Rosaceae
<i>Psidium guajava</i> *	L.	Myrtaceae
<i>Psychotria carthagenensis</i>	Jacq.	Rubiaceae
<i>Pterocaulon lanatum</i>	Kuntze	Asteraceae
<i>Pterocaulon virgatum</i>	(L.) DC.	Asteraceae
<i>Pyrostegia venusta</i>	(Ker Gawl.) Miers	Bignoniaceae
<i>Randia armata</i>	(Sw.) DC.	Rubiaceae
<i>Rhamnidium elaeocarpum</i>	Reissek	Rhamnaceae
<i>Rhipsalis cereuscula</i>	Haw.	Cactaceae
<i>Rhynchosia phaseoloides</i>	(Sw.) DC.	Fabaceae
<i>Richardia brasiliensis</i>	Gomes	Rubiaceae

<i>Ricinus communis</i> *	L.	Euphorbiaceae
<i>Rubus urticifolius</i>	Poir.	Rosaceae
<i>Ruellia jussieuoides</i>	Schltld. & Cham.	Acanthaceae
<i>Ruellia brevifolia</i>	(Pohl) C.Ezcurra	Acanthaceae
<i>Salvia guaranitica</i>	A.St.-Hil. ex Benth.	Lamiaceae
<i>Sapium glandulosum</i>	(L.) Morong	Euphorbiaceae
<i>Schaefferia argentinensis</i>	Speg.	Celastraceae
<i>Schinus terebinthifolia</i>	Raddi	Anacardiaceae
<i>Schizolobium parahyba</i>	(Vell.) Blake	Fabaceae
<i>Scleria gaertneri</i>	Raddi	Cyperaceae
<i>Sicyos edulis</i> *	Jacq.	Cucurbitaceae
<i>Seguiera langsdorffii</i>	Moq.	Phytolaccaceae
<i>Senegalia polyphylla</i>	(DC.) Britton & Rose	Fabaceae
<i>Senna multijuga</i>	(Rich.) H.S.Irwin & Barneby	Fabaceae
<i>Senna pendula</i>	(Humb.& Bonpl.ex Willd.) H.S.Ir- win & Barneby	Fabaceae
<i>Senna pilifera</i>	(Vogel) H.S.Irwin & Barneby	Fabaceae
<i>Senna splendida</i>	(Vogel) H.S.Irwin & Barneby	Fabaceae
<i>Serjania fuscifolia</i>	Radlk.	Sapindaceae
<i>Serjania reticulata</i>	Cambess.	Sapindaceae
<i>Sida planicaulis</i>	Cav.	Malvaceae
<i>Sida rhombifolia</i>	L.	Malvaceae
<i>Sida urens</i>	L.	Malvaceae
<i>Sidastrum micranthum</i>	(A.St.-Hil.) Fryxell	Malvaceae
<i>Sidastrum paniculatum</i>	(L.) Fryxell	Malvaceae
<i>Siparuna guianensis</i>	Aubl.	Siparunaceae
<i>Smilax brasiliensis</i>	Spreng.	Smilacaceae
<i>Smilax elastica</i>	Griseb.	Smilacaceae
<i>Smilax fluminensis</i>	Steud.	Smilacaceae
<i>Solanum americanum</i>	Mill.	Solanaceae
<i>Solanum concinnum</i>	Schott ex Sendtn.	Solanaceae
<i>Solanum granulosoleprosum</i>	Dunal	Solanaceae
<i>Solanum palinacanthum</i>	Dunal	Solanaceae
<i>Solanum paniculatum</i>	L.	Solanaceae
<i>Solanum pseudoquina</i>	A.St.-Hil.	Solanaceae
<i>Solanum robustum</i>	H.L.Wendl	Solanaceae

<i>Solidago chilensis</i>	Meyen	Asteraceae
<i>Spathodea campanulata</i> *	P. Beauv.	Fabaceae
<i>Stizophyllum perforatum</i>	(Cham.) Miers.	Bignoniaceae
<i>Syagrus romanzoffiana</i>	(Cham.) Glassman	Arecaceae
<i>Symplocos pubescens</i>	Klotzsch ex Benth.	Symplocaceae
<i>Syzygium cumini</i> *	(L.) Skeels	Myrtaceae
<i>Tabernaemontana catharinensis</i>	A.DC.	Apocynaceae
<i>Talinum paniculatum</i>	(Jacq.) Gaertn.	Talinaceae
<i>Tapirira guianensis</i>	Aubl.	Anacardiaceae
<i>Tecoma stans</i> *	(L.) Juss. ex Kunth	Bignoniaceae
<i>Teramnus uncinatus</i>	(L.) Sw.	Fabaceae
<i>Terminalia glabrescens</i>	Mart.	Combretaceae
<i>Thalia geniculata</i>	L.	Marantaceae
<i>Thaumatococcus bipinnatifidum</i>	(Schott ex Endl.) Sakur., Calazans & Mayo	Araceae
<i>Thunbergia alata</i> *	Bojer ex Sims	Acanthaceae
<i>Tilesia baccata</i>	(L.) Pruski	Asteraceae
<i>Tradescantia zebrina</i> *	Heynh. ex Bosse	Commelinaceae
<i>Trema micrantha</i>	(L.) Blume	Cannabaceae
<i>Trichilia catigua</i>	A.Juss.	Meliaceae
<i>Trichilia claussoni</i>	C.DC.	Meliaceae
<i>Trichilia elegans</i>	A.Juss.	Meliaceae
<i>Trichilia pallida</i>	Sw.	Meliaceae
<i>Triumfetta bartramia</i>	L.	Malvaceae
<i>Triumfetta semitriloba</i>	Jacq.	Malvaceae
<i>Urera baccifera</i>	(L.) Gaudich. ex Wedd.	Urticaceae
<i>Urochloa decubens</i> *	(Stapf) R.D.Webster	Poaceae
<i>Urvillea laevis</i>	Radlk.	Sapindaceae
<i>Varronia guazumifolia</i>	Desv.	Boraginaceae
<i>Vernonanthura polyanthes</i>	(Sprengel) Vega & Dematteis	Asteraceae
<i>Wissadula hernandioides</i>	(L.Hér.) Garcke	Malvaceae
<i>Xylosma venosa</i>	N.E.Br.	Salicaceae
<i>Zanthoxylum caribaeum</i>	Lam.	Rutaceae
<i>Zanthoxylum rhoifolium</i>	Lam.	Rutaceae
<i>Zanthoxylum riedelianum</i>	Engl.	Rutaceae

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