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**POSSÍVEL AÇÃO SINÉRGICA DE COMPONENTES DA
PRÓPOLIS SOBRE CÉLULAS DE CARCINOMA DE
LARINGE HUMANA (HEp-2): MECANISMOS DE
RESISTÊNCIA E MORTE CELULAR**

LÍVIA MATSUMOTO DA SILVA

Dissertação apresentada ao Instituto de Biociências, Campus de Botucatu, UNESP, para obtenção do título de Mestre no Programa de Pós-Graduação em Biologia Geral e Aplicada, Área de concentração *Biomoléculas: estrutura e função*.

Prof. Dr. José Maurício Sforcin

**BOTUCATU – SP
2016**



UNIVERSIDADE ESTADUAL PAULISTA

“Júlio de Mesquita Filho”

INSTITUTO DE BIOCIÊNCIAS DE BOTUCATU

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Palavras-chave: Células HEp-2; Citotoxicidade; Compostos fenólicos; Glicoproteina-P; Própolis.

“Cada pessoa tem seu próprio deserto a atravessar. E a cada vez será necessário desmascarar as miragens e também contemplar os milagres: o instante, a aliança, a douta ignorância e a fecunda vacuidade.”

Jean-Yves Leloup

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Revisão de Literatura

Resumo

Própolis e seus compostos fenólicos são conhecidos por apresentarem propriedades antioxidantes e anticâncer. Recentemente, os mecanismos de ação da propolis têm sido objeto de investigação. Este estudo teve como objetivo elucidar os efeitos da própolis e três compostos fenólicos (ácido cafeico - Caf, ácido dihidrocinâmico - Cin; ácido *p*-cumárico - Cou) na mesma proporção que são encontrados em nossa amostra de própolis, isoladamente ou em combinação, sobre células de carcinoma epidermóide de laringe humano (HEp-2). A viabilidade celular, tipo de morte e parada do ciclo celular, geração de espécies reativas de oxigênio (ROS) e a possível capacidade da própolis em induzir o efluxo de doxorrubicina (DOX) via inibidor de glicoproteína-P (P-gp) foram avaliadas. A própolis exerceu um efeito citotóxico em células HEp-2 e apresentaram um valor de IC₅₀ igual a 80 µg/mL, enquanto que os compostos isolados (isoladamente ou em combinação) não mostraram efeito sobre a viabilidade celular após 72 h. Assim, concentrações mais elevadas destes compostos foram testadas e Caf (IC₅₀: 1.332 µM) induziu necrose em células HEp-2, enquanto que a própolis induziu apoptose em células HEp-2, ambos, provavelmente devido à geração de ROS. A amostra de própolis induziu parada do ciclo celular na fase G2/M e Caf na fase S. Própolis ou seus componentes, com exceção de Caf, pode agir como um potencial inibidor de P-gp por modulação da atividade da P-gp, inibindo o efluxo de DOX. Sendo assim, os dados sugeriram que a própolis exerceu efeitos citotóxicos contra células HEp-2 e alguns mecanismos são discutidos. O seu potencial como um fármaco anti-tumoral deve ser investigado em ensaios futuros.

Palavras-chave: Própolis; Compostos fenólicos; Efeito citotóxico; células HEp-2.

Abstract

Propolis and its phenolic compounds are known for their antioxidant and anticancer properties. Propolis mechanisms of action have been the subject of research recently. This study aimed to elucidate the effects of propolis and three phenolic compounds (caffeic acid – Caf; dihydrocinnamic acid – Cin; *p*-coumaric acid – Cou) in the same proportion they are found in our propolis sample, alone or in combination, towards human larynx epidermoid carcinoma (HEp-2) cell. Cell viability, apoptosis/necrosis and cell cycle arrest, generation of reactive oxygen species (ROS) and the ability of propolis to induce doxorubicin (DOX) efflux using a P-glycoprotein (P-gp) inhibitor (verapamil) were assayed. Propolis exerted a cytotoxic effect in HEp-2 cells and exhibited an IC₅₀ value of 80 µg/mL, whereas the isolated compounds (alone or in combination) had no effect on cell viability after 72 h. Hence, higher concentrations of these compounds were tested and Caf (IC₅₀: 1.332 µM) induced necrosis in HEp-2 cells, while propolis induced apoptosis, both, probably due to ROS generation. Propolis induced cell cycle arrest at G2/M phase and, Caf at S phase. Propolis or its components, except Caf, can act as a P-gp inhibitor by modulating P-gp activity and inhibiting the efflux of DOX. Altogether, data suggested that propolis exerted cytotoxic effects against HEp-2 cells and some mechanisms are discussed. Its potential as an antitumor drug should be investigated in further assays.

Key words: Propolis; Phenolic compounds; Cytotoxic effect; Hep-2 cells.

1. Câncer: características gerais

Câncer é o termo genérico dado a um conjunto de doenças que apresentam o crescimento desordenado de células que podem vir a invadir tecidos e órgãos, podendo espalhar-se para outras regiões do corpo (1). Um acúmulo de alterações genéticas permite que as células rompam uma série de barreiras proliferativas, conferindo algumas características comuns a todas as células tumorais como: autossuficiência em sinais de crescimento, ausência de resposta a sinais anti-crescimento, evasão da apoptose, potencial replicativo ilimitado, angiogênese, invasão tecidual e metástase (2). As condições para o desenvolvimento tumoral são multifatoriais e podem agir em conjunto ou sequencialmente para promover sua iniciação, quando o organismo está suscetível à instabilidade genética e metabólica (3, 4).

O câncer é uma das principais causas de morte em todo o mundo, apresentando uma grande complexidade etiológica e complexas interações entre o tumor e seu microambiente. Além disso, a incidência cresce conforme a longevidade e a adoção de hábitos considerados fatores de risco como consumo de álcool e tabaco, hábitos alimentares, exposição à radiações, etc. (5, 6). Apesar de tantos anos dedicados a estudos para a sua compreensão, o câncer ainda continua sendo uma doença que carece de maiores investigações e terapias eficazes (7).

Linhagens celulares imortalizadas são uma ferramenta valiosa na investigação molecular, bioquímica, genética e imunológica de células derivadas de tecidos neoplásicos, usadas como modelo experimental *in vitro*. O uso destas linhagens mostra-se crível, pois permite avaliar as contribuições funcionais das anormalidades genômicas, visto que a transcrição desregulada presente nos tumores primários humanos é conservada nas linhagens celulares (8).

Diversas células têm sido utilizadas nos estudos de câncer de cabeça e pescoço, como HEp-2, HEp-3 e KB. Em trabalhos realizados por nosso grupo de pesquisa, temos utilizado as células da linhagem HEp-2, frequentemente utilizada como modelo de estudos sobre carcinogênese e mutagênese (9-11). Essa linhagem tem uma alta taxa de proliferação e ciclo celular de 23 horas (12). Elas foram descritas inicialmente como sendo derivadas de carcinoma epidermóide da laringe, mas subsequentemente foi encontrado um perfil genético que a identifica como a linhagem celular HeLa, com base em análise de isoenzimas, provavelmente por contaminação com estas células (ATCC® CCL-23™).

Sendo assim, é interessante que haja estudos que promovam uma melhor compreensão das características moleculares que podem predizer ou indicar a via de ação de agentes terapêuticos de diferentes tipos de carcinomas, e que possam fornecer subsídios para tratamentos alternativos aos tradicionais quimioterápicos.

2. Produtos naturais e câncer

O uso de plantas medicinais e metabólitos microbianos tem ampliado a expectativa de vida, seja empiricamente ou a partir da síntese de medicamentos de origem natural (13). Nos últimos 25 anos, mais de 75% dos produtos antitumorais foram obtidos de fontes naturais ou sintetizados industrialmente a partir de princípios ativos purificados das mesmas. Dos 140 agentes antitumorais disponíveis desde a década de 40, mais de 60% são oriundos de compostos naturais (14). Esses agentes atuam através de mecanismos que incluem indução de apoptose, clivagem do DNA mediada pela inibição pela topoisomerase I ou II, permeabilização mitocondrial, inibição de enzimas envolvidas na transdução de sinal ou no metabolismo celular, e por inibição da angiogênese (13).

Estudos também têm investigado o efeito da administração de produtos naturais concomitantemente com fármacos (15, 16), a fim de obter uma interação entre ambos por efeito aditivo, sinérgico, antagônico ou potencializador, com o intuito de compreender os efeitos causados pelos agentes antitumorais *in vitro*, para que, futuramente, esta combinação possa ser útil em novos tratamentos, sem interferência na eficiência, diminuindo os efeitos colaterais dos quimioterápicos.

Além de plantas medicinais e metabólitos microbianos, existem também os opoterápicos obtidos a partir de glândulas, órgãos, tecidos ou secreções animais (17). Um exemplo de opoterápico é a própolis produzida por abelhas africanizadas (*Apis mellifera*). Vários autores têm relatado que a própolis e seus componentes representam candidatos promissores para a elaboração de novos fármacos, com efeitos imunomodulador, radioprotetor e principalmente como agentes citotóxicos, antitumoral e quimiopreventivo contra vários tipos de tumores, contribuindo com a expansão do arsenal existente de medicamentos contra o câncer (18-22).

Conclusão

Esta amostra de própolis inibiu o crescimento de células HEp-2 e induziu apoptose mediada pela produção de ROS. Os compostos fenólicos isolados (Caf, Cin e Cou) em concentrações encontradas na amostra de própolis não afetou a viabilidade celular, no entanto, maiores concentrações exercearam efeitos citotóxicos significativos. Particularmente, Caf exibiu uma atividade citotóxica maior do que Cin e Cou, induzindo necrose das células HEp-2. Além disso, a própolis foi capaz de modular a sensibilidade de células HEp-2 à DOX por inibição da atividade de P-gp. São necessárias maiores investigações para elucidar o envolvimento de própolis contra células tumorais.

ANEXO 1:

TABLE 1: Relative percentages of compounds, determined by GC-MS, from ethanolic extract of Brazilian propolis.

Component	Retention time (min)	% of total
Benzoic acid	9.3	0.193
Dihydrocinnamic acid	14	2.180
Dihydrocinnamic acid	22	0.860
Coumaric acid	26.5	0.382
Cafeic acid	28.6	0.297
Prenyl- <i>p</i> -coumaric acid	32.5	6.560
Flavonoids	35.8	1.142
Artepillin C	37.7	16.750
Trihydroxymethoxy flavonon	40.5	0.666
Tetrahydroxy flavonon	40.8	0.228
Triterpene	47.6	0.777
Triterpene	51	0.309

Retirado de: Conti BJ, Bufalo MC, Golim Mde A, Bankova V, Sforcin JM. Cinnamic Acid is partially involved in propolis immunomodulatory action on human monocytes. Evidence-based complementary and alternative medicine: eCAM. 2013;2013:109864. Epub 2013/06/14.

Referências:

1. INCA INDCJDAGDS. O que é câncer. 2016 [cited 2016]; Available from: http://www1.inca.gov.br/conteudo_view.asp?id=322.
2. Hanahan D, Weinberg Robert A. Hallmarks of Cancer: The Next Generation. *Cell*. 2011;144(5):646-74.
3. de Martel C, Ferlay J, Franceschi S, Vignat J, Bray F, Forman D, et al. Global burden of cancers attributable to infections in 2008: a review and synthetic analysis. *The Lancet Oncology*. 2012;13(6):607-15. Epub 2012/05/12.
4. Global action plan for the prevention and control of NCDs 2013-2020. [database on the Internet]. 2014.
5. Siegel R, Naishadham D, Jemal A. Cancer statistics, 2013. *CA: a cancer journal for clinicians*. 2013;63(1):11-30. Epub 2013/01/22.
6. Torre LA, Bray F, Siegel RL, Ferlay J, Lortet-Tieulent J, Jemal A. Global cancer statistics, 2012. *CA: a cancer journal for clinicians*. 2015;65(2):87-108. Epub 2015/02/06.
7. Sonnenschein C, Soto AM, Rangarajan A, Kulkarni P. Competing views on cancer. *Journal of biosciences*. 2014;39(2):281-302. Epub 2014/04/17.
8. Neve RM, Chin K, Fridlyand J, Yeh J, Baehner FL, Fevr T, et al. A collection of breast cancer cell lines for the study of functionally distinct cancer subtypes. *Cancer cell*. 2006;10(6):515-27. Epub 2006/12/13.
9. Grem JL, Fischer PH. Enhancement of 5-fluorouracil's anticancer activity by dipyridamole. *Pharmacol Ther*. 1989;40(3):349-71. Epub 1989/01/01.
10. Sinico RA, Bollini B, Sabadini E, Di Toma L, Radice A. The use of laboratory tests in diagnosis and monitoring of systemic lupus erythematosus. *Journal of nephrology*. 2002;15 Suppl 6:S20-7. Epub 2003/01/08.
11. Valdovinos MR, Gomez B. Establishment of respiratory syncytial virus persistence in cell lines: association with defective interfering particles. *Intervirology*. 2003;46(3):190-8. Epub 2003/07/18.
12. Den Beste HE, Fjelde A, Jackson JL, Andresen WF, Kerr HA, Evans VJ. Adaptation, growth, and chromosomal analysis of HEp-2 cells in chemically defined medium. *Journal of the National Cancer Institute*. 1966;36(6):1075-88. Epub 1966/06/01.
13. Demain AL, Vaishnav P. Natural products for cancer chemotherapy. *Microb Biotechnol*. 2011;4:687.
14. Cragg GM, Newman DJ. Antineoplastic agents from natural sources: achievements and future directions. *Expert opinion on investigational drugs*. 2000;9(12):2783-97. Epub 2000/11/28.
15. Ambroz M, Hanusova V, Skarka A, Bousova I, Kralova V, Langhasova L, et al. Essential Oil from Myrica rubra Leaves Potentiated Antiproliferative and Prooxidative Effect of Doxorubicin and its Accumulation in Intestinal Cancer Cells. *Planta medica*. 2016;82(1-02):89-96. Epub 2015/10/21.
16. Surmeli Z, Gursoy P, Erdogan AP, Bozkurt E, Atmaca H, Uzunoglu S, et al. Combination of zoledronic acid and serine/threonine phosphatase inhibitors induces synergistic cytotoxicity and apoptosis in human breast cancer cells via inhibition of PI3K/Akt pathway. *Tumour biology : the journal of the International Society for Oncodevelopmental Biology and Medicine*. 2015. Epub 2015/10/16.
17. [Internet] ANdVS. Câmara Técnica de Medicamentos Fitoterápicos - CATEF. <http://www.anvisa.gov.br/medicamentos/catef/propolis.htm.2016> [cited 2016]; Available from: Available from: <http://www.anvisa.gov.br/medicamentos/catef/propolis.htm>.
18. Banskota AH, Tezuka Y, Kadota S. Recent progress in pharmacological research of propolis. *Phytotherapy Research*. 2001;15(7):561-71.
19. Frión-Herrera Y, Díaz-García A, Ruiz-Fuentes J, Rodríguez-Sánchez H, Sforcin JM. Brazilian green propolis induced apoptosis in human lung cancer A549 cells through mitochondrial-mediated pathway. *Journal of Pharmacy and Pharmacology*. 2015;67(10):1448-56.

20. Oršolić N. A review of propolis antitumour action in vivo and in vitro. *Journal of Api Product and ApiMedical Science* 2010;2 (1):1 - 20
21. Sforcin JM, Bankova V. Propolis: is there a potential for the development of new drugs? *Journal of ethnopharmacology*. 2011;133(2):253-60. Epub 2010/10/26.
22. Watanabe MA, Amarante MK, Conti BJ, Sforcin JM. Cytotoxic constituents of propolis inducing anticancer effects: a review. *The Journal of pharmacy and pharmacology*. 2011;63(11):1378-86. Epub 2011/10/13.
23. Burdock GA. Review of the biological properties and toxicity of bee propolis (propolis). *Food and chemical toxicology : an international journal published for the British Industrial Biological Research Association*. 1998;36(4):347-63. Epub 1998/07/03.
24. Castaldo S, Capasso F. Propolis, an old remedy used in modern medicine. *Fitoterapia*. 2002;73 Suppl 1:S1-6. Epub 2002/12/24.
25. Lustosa SR, Galindo AB, Nunes LCC, Randau KP, Rolim Neto PJ. Própolis: atualizações sobre a química e a farmacologia. *Revista Brasileira de Farmacognosia*. 2008;18:447-54.
26. Sforcin JM. Propolis and the immune system: a review. *Journal of ethnopharmacology*. 2007;113(1):1-14. Epub 2007/06/21.
27. Salatino A, Teixeira EW, Negri G, Message D. Origin and Chemical Variation of Brazilian Propolis. *Evidence-based complementary and alternative medicine : eCAM*. 2005;2(1):33-8. Epub 2005/04/21.
28. Ahn MR, Kunimasa K, Ohta T, Kumazawa S, Kamihira M, Kaji K, et al. Suppression of tumor-induced angiogenesis by Brazilian propolis: major component artepillin C inhibits in vitro tube formation and endothelial cell proliferation. *Cancer letters*. 2007;252(2):235-43. Epub 2007/03/09.
29. Aso K, Kanno S, Tadano T, Satoh S, Ishikawa M. Inhibitory effect of propolis on the growth of human leukemia U937. *Biological & pharmaceutical bulletin*. 2004;27(5):727-30. Epub 2004/05/11.
30. Orsolic N, Basic I. Antitumor, hematostimulative and radioprotective action of water-soluble derivative of propolis (WSDP). *Biomedicine & pharmacotherapy = Biomedecine & pharmacotherapie*. 2005;59(10):561-70. Epub 2005/10/06.
31. Oršolić N, Bašić I. Water-soluble derivative of propolis and its polyphenolic compounds enhance tumoricidal activity of macrophages. *Journal of ethnopharmacology*. 2005;102(1):37-45.
32. Sawicka D, Car H, Borawska MH, Niklinski J. The anticancer activity of propolis. *Folia histochemica et cytobiologica / Polish Academy of Sciences, Polish Histochemical and Cytochemical Society*. 2012;50(1):25-37. Epub 2012/04/26.
33. Kartal M, Kaya S, Kurucu S. GC-MS analysis of propolis samples from two different regions of Turkey. *Zeitschrift fur Naturforschung C, Journal of biosciences*. 2002;57(9-10):905-9. Epub 2002/11/21.
34. Krell R. VALUE-ADDED PRODUCTS FROM BEEKEEPING. *Food and Agriculture Organization of the United Nations Rome*

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35. Bankova Vea, Boudourova-Krasteva G, Popov S, Sforcin JM, Funari SRC. . Seasonal variations of the chemical composition of Brazilian propolis. *Apidologie*. 1998b;29:361-6.
36. Conti BJ, Bufalo MC, Golim Mde A, Bankova V, Sforcin JM. Cinnamic Acid is partially involved in propolis immunomodulatory action on human monocytes. *Evidence-based complementary and alternative medicine : eCAM*. 2013;2013:109864. Epub 2013/06/14.
37. G. Boudourova-Krasteva, Vassya Bankova, J. M. Sforcin, Nikolova N, Popov S. Phenolics from Brazilian Propolis. *Zeitschrift für Naturforschung*. 1997;52c: 676-9.
38. Bankova V. Chemical diversity of propolis and the problem of standardization. *Journal of ethnopharmacology*. 2005;100(1-2):114-7. Epub 2005/07/05.
39. Bankova V, Boudourova-Krasteva G, Sforcin JM, Frete X, Kujumgiev A, Maimoni-Rodella R, et al. Phytochemical evidence for the plant origin of Brazilian propolis from Sao

- Paulo state. Zeitschrift fur Naturforschung C, Journal of biosciences. 1999;54(5-6):401-5. Epub 1999/08/04.
40. Conti BJ, Bankova V, Sforcin JM. Chemical Composition of the Same Brazilian Propolis Sample Analyzed in 1997 and in 2012: No Freezing Effect. Natural product communications. 2015;10(7):1279-80. Epub 2015/09/29.
41. Quideau S, Deffieux D, Douat-Casassus C, Pouysegu L. Plant polyphenols: chemical properties, biological activities, and synthesis. Angew Chem Int Ed Engl. 2011;50(3):586-621. Epub 2011/01/13.
42. Boudet AM. Evolution and current status of research in phenolic compounds. Phytochemistry. 2007;68(22-24):2722-35. Epub 2007/07/24.
43. Cheynier V, Comte G, Davies KM, Lattanzio V, Martens S. Plant phenolics: recent advances on their biosynthesis, genetics, and ecophysiology. Plant physiology and biochemistry : PPB / Societe francaise de physiologie vegetale. 2013;72:1-20. Epub 2013/06/19.
44. de Sousa JP, da Silva Filho AA, Bueno PC, Gregorio LE, Furtado NA, Jorge RF, et al. A validated reverse-phase HPLC analytical method for the quantification of phenolic compounds in Baccharis dracunculifolia. Phytochemical analysis : PCA. 2009;20(1):24-32. Epub 2008/08/30.
45. Arts IC, Hollman PC. Polyphenols and disease risk in epidemiologic studies. The American journal of clinical nutrition. 2005;81(1 Suppl):317S-25S. Epub 2005/01/11.
46. Gonzalez S, Fernandez M, Cuervo A, Lasheras C. Dietary intake of polyphenols and major food sources in an institutionalised elderly population. Journal of human nutrition and dietetics : the official journal of the British Dietetic Association. 2014;27(2):176-83. Epub 2013/03/26.
47. Yang CS, Landau JM, Huang MT, Newmark HL. Inhibition of carcinogenesis by dietary polyphenolic compounds. Annual review of nutrition. 2001;21:381-406. Epub 2001/05/26.
48. SOARES SE. Ácidos fenólicos como antioxidantes. Revista de Nutrição. 2002;15(1):71-81.
49. Croft KD. The chemistry and biological effects of flavonoids and phenolic acids. Annals of the New York Academy of Sciences. 1998;854:435-42. Epub 1999/02/03.
50. Licandro-Seraut H, Roussel C, Perpetuini G, Gervais P, Cavin JF. Sensitivity to vinyl phenol derivatives produced by phenolic acid decarboxylase activity in Escherichia coli and several food-borne Gram-negative species. Applied microbiology and biotechnology. 2013;97(17):7853-64. Epub 2013/07/13.
51. THIMANN KV. Physiology of Plant Growth and Development. McGraw-Hill, editor. The auxins, in: M.B. Wilkins (Ed.)1969. 2-45 p.
52. Costa G, Francisco V, Lopes MC, Cruz MT, Batista MT. Intracellular signaling pathways modulated by phenolic compounds: application for new anti-inflammatory drugs discovery. Current medicinal chemistry. 2012;19(18):2876-900. Epub 2012/04/24.
53. Campos FM, Couto JA, Figueiredo AR, Toth IV, Rangel AO, Hogg TA. Cell membrane damage induced by phenolic acids on wine lactic acid bacteria. International journal of food microbiology. 2009;135(2):144-51. Epub 2009/09/08.
54. Manach C, Scalbert A, Morand C, Remesy C, Jimenez L. Polyphenols: food sources and bioavailability. The American journal of clinical nutrition. 2004;79(5):727-47. Epub 2004/04/29.
55. Castelli F, Uccella N, Trombetta D, Saija A. Differences between Coumaric and Cinnamic Acids in Membrane Permeation As Evidenced by Time-Dependent Calorimetry. Journal of agricultural and food chemistry. 1999;47(3):991-5.
56. Jaganathan SK, Supriyanto E, Mandal M. Events associated with apoptotic effect of p - Coumaric acid in HCT-15 colon cancer cells. World Journal of Gastroenterology. 2013;19(43):7726-34.
57. Akao Y, Maruyama H, Matsumoto K, Ohguchi K, Nishizawa K, Sakamoto T, et al. Cell growth inhibitory effect of cinnamic acid derivatives from propolis on human tumor cell lines. Biological & pharmaceutical bulletin. 2003;26(7):1057-9. Epub 2003/07/05.

58. Nieva Moreno MI, Zampini IC, Ordóñez RM, Jaime GS, Vattuone MA, Isla MI. Evaluation of the cytotoxicity, genotoxicity, mutagenicity, and antimutagenicity of propolis from Tucuman, Argentina. *Journal of agricultural and food chemistry*. 2005;53(23):8957-62. Epub 2005/11/10.
59. Kumazaki M, Shinohara H, Taniguchi K, Yamada N, Ohta S, Ichihara K, et al. Propolis cinnamic acid derivatives induce apoptosis through both extrinsic and intrinsic apoptosis signaling pathways and modulate of miRNA expression. *Phytomedicine : international journal of phytotherapy and phytopharmacology*. 2014;21(8-9):1070-7. Epub 2014/05/24.
60. Khosravi AR, Shokri H, Darvishi S, Taghavi M. Immunomodulatory efficacy of ethanol extract of propolis on tumor-bearing mice with disseminated candidiasis. *Journal de Mycologie Médicale / Journal of Medical Mycology*. 2014;24(4):e143-e8.
61. Missima F, Pagliarone AC, Orsatti CL, Araujo JP, Jr., Sforzin JM. The Effect of propolis on Th1/Th2 cytokine expression and production by melanoma-bearing mice submitted to stress. *Phytotherapy research : PTR*. 2010;24(10):1501-7. Epub 2010/09/30.
62. Orsolic N, Sver L, Terzic S, Basic I. Peroral application of water-soluble derivative of propolis (WSDP) and its related polyphenolic compounds and their influence on immunological and antitumour activity. *Veterinary research communications*. 2005;29(7):575-93. Epub 2005/09/06.
63. Archer EJ, Simpson MA, Watts NJ, O'Kane R, Wang B, Erie DA, et al. Long-range architecture in a viral RNA genome. *Biochemistry*. 2013;52(18):3182-90. Epub 2013/04/26.
64. Fuchs Y, Steller H. Live to die another way: modes of programmed cell death and the signals emanating from dying cells. *Nature reviews Molecular cell biology*. 2015;16(6):329-44. Epub 2015/05/21.
65. Vangestel C, Van de Wiele C, Mees G, Peeters M. Forcing cancer cells to commit suicide. *Cancer biotherapy & radiopharmaceuticals*. 2009;24(4):395-407. Epub 2009/08/22.
66. Leist M, Jaattela M. Four deaths and a funeral: from caspases to alternative mechanisms. *Nature reviews Molecular cell biology*. 2001;2(8):589-98. Epub 2001/08/03.
67. Kerr JF, Wyllie AH, Currie AR. Apoptosis: a basic biological phenomenon with wide-ranging implications in tissue kinetics. *British journal of cancer*. 1972;26(4):239-57. Epub 1972/08/01.
68. Seino K, Setoguchi Y, Ogino T, Kayagaki N, Akiba H, Nakano H, et al. Protection against Fas-mediated and tumor necrosis factor receptor 1-mediated liver injury by blockade of FADD without loss of nuclear factor- κ B activation. *Annals of surgery*. 2001;234(5):681-8. Epub 2001/10/31.
69. Mohan S, Abdelwahab SI, Kamalidehghan B, Syam S, May KS, Harmal NSM, et al. Involvement of NF- κ B and Bcl2/Bax signaling pathways in the apoptosis of MCF7 cells induced by a xanthone compound Pyranocycloartobiloxanthone A. *Phytomedicine : international journal of phytotherapy and phytopharmacology*. 2012;19(11):1007-15.
70. Jada SR, Matthews C, Saad MS, Hamzah AS, Lajis NH, Stevens MFG, et al. Benzylidene derivatives of andrographolide inhibit growth of breast and colon cancer cells in vitro by inducing G1 arrest and apoptosis. *Br J Pharmacol*. 2008;155.
71. Verhoven B, Schlegel RA, Williamson P. Mechanisms of phosphatidylserine exposure, a phagocyte recognition signal, on apoptotic T lymphocytes. *J Exp Med*. 1995;182.
72. Chen JH, Shao Y, Huang MT, Chin CK, Ho CT. Inhibitory effect of caffeic acid phenethyl ester on human leukemia HL-60 cells. *Cancer letters*. 1996;108(2):211-4. Epub 1996/11/29.
73. Amaravadi RK, Thompson CB. The Roles of Therapy-Induced Autophagy and Necrosis in Cancer Treatment. *Clinical Cancer Research*. 2007;13(24):7271-9.
74. Zeh HJ, Lotze MT. Addicted to death - Invasive cancer and the immune response to unscheduled cell death. *Journal of Immunotherapy*. 2005;28(1):1-9.
75. Festjens N, Vanden Berghe T, Vandenebeeck P. Necrosis, a well-orchestrated form of cell demise: Signalling cascades, important mediators and concomitant immune response. *Biochimica et Biophysica Acta (BBA) - Bioenergetics*. 2006;1757(9-10):1371-87.
76. Goldstein JA, Gallagher MJ, O'Neill WW, Ross MA, O'Neil BJ, Raff GL. A randomized controlled trial of multi-slice coronary computed tomography for evaluation of

- acute chest pain. *Journal of the American College of Cardiology*. 2007;49(8):863-71. Epub 2007/02/27.
77. Proskuryakov SY, Gabai VL. Mechanisms of tumor cell necrosis. *Current pharmaceutical design*. 2010;16(1):56-68. Epub 2010/03/11.
 78. Gajski G, Čimbora-Zovko T, Rak S, Rožman M, Osmak M, Garaj-Vrhovac V. Combined antitumor effects of bee venom and cisplatin on human cervical and laryngeal carcinoma cells and their drug resistant sublines. *Journal of Applied Toxicology*. 2014;34(12):1332-41.
 79. Malumbres M, Barbacid M. Cell cycle, CDKs and cancer: a changing paradigm. *Nature reviews Cancer*. 2009;9(3):153-66. Epub 2009/02/25.
 80. Paulovich AG, Toczyski DP, Hartwell LH. When checkpoints fail. *Cell*. 1997;88(3):315-21. Epub 1997/02/07.
 81. Ogino A, Yoshino A, Katayama Y, Watanabe T, Ota T, Komine C, et al. The p15(INK4b)/p16(INK4a)/RB1 pathway is frequently deregulated in human pituitary adenomas. *Journal of neuropathology and experimental neurology*. 2005;64(5):398-403. Epub 2005/05/17.
 82. Ekholm SV, Zickert P, Reed SI, Zetterberg A. Accumulation of cyclin E is not a prerequisite for passage through the restriction point. *Molecular and cellular biology*. 2001;21(9):3256-65. Epub 2001/04/05.
 83. Pardee AB. A restriction point for control of normal animal cell proliferation. *Proceedings of the National Academy of Sciences of the United States of America*. 1974;71(4):1286-90. Epub 1974/04/01.
 84. Sun D, Urrabaz R, Buzello C, Nguyen M. Effects of cisplatin on expression of DNA ligases in MiaPaCa human pancreatic cancer cells. *Biochem Biophys Res Commun*. 2002;298:537.
 85. Yoshikawa R, Kusunoki M, Yanagi H, Noda M, Furuyama JI, Yamamura T, et al. Dual antitumor effects of 5-fluorouracil on the cell cycle in colorectal carcinoma cells: a novel target mechanism concept for pharmacokinetic modulating chemotherapy. *Cancer Res*. 2001;61:1029.
 86. Gallagher BM, Jr. Microtubule-stabilizing natural products as promising cancer therapeutics. *Current medicinal chemistry*. 2007;14(28):2959-67. Epub 2008/01/29.
 87. Singh RP, Agarwal R. Natural flavonoids targeting deregulated cell cycle progression in cancer cells. *Current drug targets*. 2006;7(3):345-54. Epub 2006/03/07.
 88. Rasul A, Khan M, Yu B, Ma T, Yang H. Xanthoxyletin, a coumarin induces S phase arrest and apoptosis in human gastric adenocarcinoma SGC-7901 cells. *Asian Pacific journal of cancer prevention : APJCP*. 2011;12(5):1219-23. Epub 2011/08/31.
 89. Yang X, Li X, An L, Bai B, Chen J. Silibinin induced the apoptosis of Hep-2 cells via oxidative stress and down-regulating survivin expression. *Eur Arch Otorhinolaryngol*. 2013;270(8):2289-97. Epub 2013/04/13.
 90. Sforzin JM, Bankova V. Propolis: Is there a potential for the development of new drugs? *Journal of ethnopharmacology*. 2011;133(2):253-60.
 91. Havsteen BH. The biochemistry and medical significance of the flavonoids. *Pharmacology & Therapeutics*. 2002;96(2-3):67-202.
 92. Akbas SH, Timur M, Ozben T. The effect of quercetin on topotecan cytotoxicity in MCF-7 and MDA-MB 231 human breast cancer cells. *J Surg Res*. 2005;125.
 93. Chua PJ, Yip GWC, Bay BH. Cell cycle arrest induced by hydrogen peroxide is associated with modulation of oxidative stress related genes in breast cancer cells. *Exp Biol Med*. 2009;234.
 94. Pelicano H, Carney D, Huang P. ROS stress in cancer cells and therapeutic implications. *Drug resistance updates : reviews and commentaries in antimicrobial and anticancer chemotherapy*. 2004;7(2):97-110. Epub 2004/05/26.
 95. Trachootham D, Zhou Y, Zhang H, Demizu Y, Chen Z, Pelicano H, et al. Selective killing of oncogenically transformed cells through a ROS-mediated mechanism by beta-phenylethyl isothiocyanate. *Cancer cell*. 2006;10(3):241-52. Epub 2006/09/09.
 96. Loo G. Redox-sensitive mechanisms of phytochemical-mediated inhibition of cancer cell proliferation (review). *The Journal of nutritional biochemistry*. 2003;14(2):64-73. Epub 2003/04/02.

97. Holohan C, Van Schaeybroeck S, Longley DB, Johnston PG. Cancer drug resistance: an evolving paradigm. *Nature reviews Cancer*. 2013;13(10):714-26. Epub 2013/09/26.
98. Juliano RL, Ling V. A surface glycoprotein modulating drug permeability in Chinese hamster ovary cell mutants. *Biochimica et biophysica acta*. 1976;455(1):152-62. Epub 1976/11/11.
99. Muller M, Meijer C, Zaman GJ, Borst P, Schepers RJ, Mulder NH, et al. Overexpression of the gene encoding the multidrug resistance-associated protein results in increased ATP-dependent glutathione S-conjugate transport. *Proceedings of the National Academy of Sciences of the United States of America*. 1994;91(26):13033-7. Epub 1994/12/20.
100. Szakacs G, Paterson JK, Ludwig JA, Booth-Genthe C, Gottesman MM. Targeting multidrug resistance in cancer. *Nat Rev Drug Discov*. 2006;5(3):219-34. Epub 2006/03/07.
101. Gottesman MM, Fojo T, Bates SE. Multidrug resistance in cancer: role of ATP-dependent transporters. *Nature reviews Cancer*. 2002;2(1):48-58. Epub 2002/03/21.
102. Choi CH. ABC transporters as multidrug resistance mechanisms and the development of chemosensitizers for their reversal. *Cancer cell international*. 2005;5:30. Epub 2005/10/06.
103. Ambudkar SV, Dey S, Hrycyna CA, Ramachandra M, Pastan I, Gottesman MM. Biochemical, cellular, and pharmacological aspects of the multidrug transporter. *Annual review of pharmacology and toxicology*. 1999;39:361-98. Epub 1999/05/20.
104. Triller N, Korosec P, Kern I, Kosnik M, Debeljak A. Multidrug resistance in small cell lung cancer: expression of P-glycoprotein, multidrug resistance protein 1 and lung resistance protein in chemo-naive patients and in relapsed disease. *Lung Cancer*. 2006;54(2):235-40. Epub 2006/08/29.
105. Thomas H, Coley HM. Overcoming multidrug resistance in cancer: an update on the clinical strategy of inhibiting p-glycoprotein. *Cancer control : journal of the Moffitt Cancer Center*. 2003;10(2):159-65. Epub 2003/04/25.
106. Nooter K, Brutel de la Riviere G, Look MP, van Wingerden KE, Henzen-Logmans SC, Schepers RJ, et al. The prognostic significance of expression of the multidrug resistance-associated protein (MRP) in primary breast cancer. *British journal of cancer*. 1997;76(4):486-93. Epub 1997/01/01.
107. Azeredo F J., Uchôa F. T., Costa TD. P-glycoprotein role on drug pharmacokinetics and interactions. *Revista Brasileira de Farmacognosia*. 2009;90(4).
108. Schuurhuis GJ, van Heijningen TH, Cervantes A, Pinedo HM, de Lange JH, Keizer HG, et al. Changes in subcellular doxorubicin distribution and cellular accumulation alone can largely account for doxorubicin resistance in SW-1573 lung cancer and MCF-7 breast cancer multidrug resistant tumour cells. *British journal of cancer*. 1993;68(5):898-908. Epub 1993/11/01.
109. Milroy R. A randomised clinical study of verapamil in addition to combination chemotherapy in small cell lung cancer. West of Scotland Lung Cancer Research Group, and the Aberdeen Oncology Group. *British journal of cancer*. 1993;68(4):813-8. Epub 1993/10/01.
110. Passerini N, Perissutti B, Albertini B, Voinovich D, Moneghini M, Rodriguez L. Controlled release of verapamil hydrochloride from waxy microparticles prepared by spray congealing. *Journal of controlled release : official journal of the Controlled Release Society*. 2003;88(2):263-75. Epub 2003/03/12.
111. Abdallah HM, Al-Abd AM, El-Dine RS, El-Halawany AM. P-glycoprotein inhibitors of natural origin as potential tumor chemo-sensitizers: A review. *Journal of advanced research*. 2015;6(1):45-62. Epub 2015/02/17.
112. Zhao D, Liu N, Shi K, Wang X, Wu G. Preparation of a multifunctional verapamil-loaded nano-carrier based on a self-assembling PEGylated prodrug. *Colloids and surfaces B, Biointerfaces*. 2015;135:682-8. Epub 2015/09/05.
113. Dong X, Mumford RJ. Nanomedicinal strategies to treat multidrug-resistant tumors: current progress. *Nanomedicine (Lond)*. 2010;5(4):597-615. Epub 2010/06/10.
114. Eid SY, El-Readi MZ, Wink M. Synergism of three-drug combinations of sanguinarine and other plant secondary metabolites with digitonin and doxorubicin in multi-drug resistant cancer cells. *Phytomedicine : international journal of phytotherapy and phytopharmacology*. 2012;19(14):1288-97. Epub 2012/11/14.

115. Middleton E, Jr., Kandaswami C, Theoharides TC. The effects of plant flavonoids on mammalian cells: implications for inflammation, heart disease, and cancer. *Pharmacological reviews*. 2000;52(4):673-751. Epub 2000/12/21.
116. Tewey KM, Rowe TC, Yang L, Halligan BD, Liu LF. Adriamycin-induced DNA damage mediated by mammalian DNA topoisomerase II. *Science*. 1984;226(4673):466-8. Epub 1984/10/26.
117. Triton TR, Yee G. The anticancer agent adriamycin can be actively cytotoxic without entering cells. *Science*. 1982;217(4556):248-50. Epub 1982/07/16.
118. Cole SP, Bhardwaj G, Gerlach JH, Mackie JE, Grant CE, Almquist KC, et al. Overexpression of a transporter gene in a multidrug-resistant human lung cancer cell line. *Science*. 1992;258(5088):1650-4. Epub 1992/12/04.
119. Fojo A, Akiyama S, Gottesman MM, Pastan I. Reduced drug accumulation in multiply drug-resistant human KB carcinoma cell lines. *Cancer research*. 1985;45(7):3002-7. Epub 1985/07/01.
120. Takara K, Fujita M, Matsubara M, Minegaki T, Kitada N, Ohnishi N, et al. Effects of propolis extract on sensitivity to chemotherapeutic agents in HeLa and resistant sublines. *Phytotherapy research : PTR*. 2007;21(9):841-6. Epub 2007/05/09.
121. Markham KR, Mitchell KA, Wilkins AL, Daldy JA, Lu Y. HPLC and GC-MS identification of the major organic constituents in New Zealand propolis. *Phytochemistry*. 1996;42(1):205-11.
122. Grunberger D, Banerjee R, Eisinger K, Oltz EM, Efros L, Caldwell M, et al. Preferential cytotoxicity on tumor cells by caffeic acid phenethyl ester isolated from propolis. *Experientia*. 1988;44(3):230-2. Epub 1988/03/15.
123. Liao HF, Chen YY, Liu JJ, Hsu ML, Shieh HJ, Liao HJ, et al. Inhibitory effect of caffeic acid phenethyl ester on angiogenesis, tumor invasion, and metastasis. *Journal of agricultural and food chemistry*. 2003;51(27):7907-12. Epub 2003/12/24.
124. Fiúza SM, Gomes C, Teixeira LJ, Girao da Cruz MT, Cordeiro MN, Milhazes N, et al. Phenolic acid derivatives with potential anticancer properties--a structure-activity relationship study. Part 1: methyl, propyl and octyl esters of caffeic and gallic acids. *Bioorganic & medicinal chemistry*. 2004;12(13):3581-9. Epub 2004/06/10.
125. Cai H, Huang X, Xu S, Shen H, Zhang P, Huang Y, et al. Discovery of novel hybrids of diaryl-1,2,4-triazoles and caffeic acid as dual inhibitors of cyclooxygenase-2 and 5-lipoxygenase for cancer therapy. *European journal of medicinal chemistry*. 2016;108:89-103.
126. Peng W, Wu JG, Jiang YB, Liu YJ, Sun T, Wu N, et al. Antitumor activity of 4-O-(2"-O-acetyl-6"-O-p-coumaroyl-beta-D-glucopyranosyl)-p-coumaric acid against lung cancers via mitochondrial-mediated apoptosis. *Chemico-biological interactions*. 2015;233:8-13. Epub 2015/04/01.
127. Kimoto T, Arai S, Kohguchi M, Aga M, Nomura Y, Micallef MJ, et al. Apoptosis and suppression of tumor growth by artepillin C extracted from Brazilian propolis. *Cancer detection and prevention*. 1998;22(6):506-15. Epub 1998/11/21.
128. He N, Shi X, Zhao Y, Tian L, Wang D, Yang X. Inhibitory effects and molecular mechanisms of selenium-containing tea polysaccharides on human breast cancer MCF-7 cells. *Journal of agricultural and food chemistry*. 2013;61(3):579-88. Epub 2012/12/29.
129. Lu CC, Yang JS, Huang AC, Hsia TC, Chou ST, Kuo CL, et al. Chrysophanol induces necrosis through the production of ROS and alteration of ATP levels in J5 human liver cancer cells. *Molecular nutrition & food research*. 2010;54(7):967-76. Epub 2010/02/20.
130. Kuo YY, Jim WT, Su LC, Chung CJ, Lin CY, Huo C, et al. Caffeic Acid phenethyl ester is a potential therapeutic agent for oral cancer. *International journal of molecular sciences*. 2015;16(5):10748-66. Epub 2015/05/20.
131. Rzepecka-Stojko A, Kabala-Dzik A, Mozdzierz A, Kubina R, Wojtyczka RD, Stojko R, et al. Caffeic Acid phenethyl ester and ethanol extract of propolis induce the complementary cytotoxic effect on triple-negative breast cancer cell lines. *Molecules*. 2015;20(5):9242-62. Epub 2015/05/27.

132. Zilius M, Ramanauskiene K, Briedis V. Release of propolis phenolic acids from semisolid formulations and their penetration into the human skin in vitro. Evidence-based complementary and alternative medicine : eCAM. 2013;2013:958717. Epub 2013/06/14.
133. Chuu CP, Lin HP, Ciaccio MF, Kokontis JM, Hause RJ, Jr., Hiipakka RA, et al. Caffeic acid phenethyl ester suppresses the proliferation of human prostate cancer cells through inhibition of p70S6K and Akt signaling networks. Cancer Prev Res (Phila). 2012;5(5):788-97. Epub 2012/05/09.
134. Dorai T, Aggarwal BB. Role of chemopreventive agents in cancer therapy. Cancer letters. 2004;215(2):129-40. Epub 2004/10/19.
135. Ho HC, Chang HC, Ting CT, Kuo CY, Yang VC. Caffeic acid phenethyl ester inhibits proliferation and migration, and induces apoptosis in platelet-derived growth factor-BB-stimulated human coronary smooth muscle cells. Journal of vascular research. 2012;49(1):24-32. Epub 2011/10/12.
136. Kuo HC, Kuo WH, Lee YJ, Lin WL, Chou FP, Tseng TH. Inhibitory effect of caffeic acid phenethyl ester on the growth of C6 glioma cells in vitro and in vivo. Cancer letters. 2006;234(2):199-208. Epub 2005/05/12.
137. Orban Z, Mitsiades N, Burke TR, Jr., Tsokos M, Chrousos GP. Caffeic acid phenethyl ester induces leukocyte apoptosis, modulates nuclear factor-kappa B and suppresses acute inflammation. Neuroimmunomodulation. 2000;7(2):99-105. Epub 2000/02/25.
138. Su ZZ, Lin J, Prewett M, Goldstein NI, Fisher PB. Apoptosis mediates the selective toxicity of caffeic acid phenethyl ester (CAPE) toward oncogene-transformed rat embryo fibroblast cells. Anticancer research. 1995;15(5B):1841-8. Epub 1995/09/01.
139. Storz P. Reactive oxygen species in tumor progression. Frontiers in bioscience : a journal and virtual library. 2005;10:1881-96. Epub 2005/03/17.
140. Wang Y, Tang Q, Jiang S, Li M, Wang X. Anti-colorectal cancer activity of macrostemonoside A mediated by reactive oxygen species. Biochemical and biophysical research communications. 2013;441(4):825-30.
141. Xuan H, Zhao J, Miao J, Li Y, Chu Y, Hu F. Effect of Brazilian propolis on human umbilical vein endothelial cell apoptosis. Food and chemical toxicology : an international journal published for the British Industrial Biological Research Association. 2011;49(1):78-85. Epub 2010/10/12.
142. Tsai Y-C, Wang Y-H, Liou C-C, Lin Y-C, Huang H, Liu Y-C. Induction of Oxidative DNA Damage by Flavonoids of Propolis: Its Mechanism and Implication about Antioxidant Capacity. Chemical Research in Toxicology. 2012;25(1):191-6.
143. Orsi RO, Funari SRC, Soares AMVC, Calvi SA, Oliveira SL, Sforcin JM, et al. Immunomodulatory action of propolis on macrophage activation. Journal of Venomous Animals and Toxins. 2000;6:205-19.
144. Aroui S, Brahim S, Waard MD, Kenani A. Cytotoxicity, intracellular distribution and uptake of doxorubicin and doxorubicin coupled to cell-penetrating peptides in different cell lines: a comparative study. Biochemical and biophysical research communications. 2010;391(1):419-25. Epub 2009/11/17.
145. Minotti G, Menna P, Salvatorelli E, Cairo G, Gianni L. Anthracyclines: molecular advances and pharmacologic developments in antitumor activity and cardiotoxicity. Pharmacological reviews. 2004;56(2):185-229. Epub 2004/06/01.
146. Silva R, Carmo H, Dinis-Oliveira R, Cordeiro-da-Silva A, Lima SC, Carvalho F, et al. In vitro study of P-glycoprotein induction as an antidotal pathway to prevent cytotoxicity in Caco-2 cells. Archives of toxicology. 2011;85(4):315-26. Epub 2010/09/22.
147. Ullah MF. Cancer multidrug resistance (MDR): a major impediment to effective chemotherapy. Asian Pacific journal of cancer prevention : APJCP. 2008;9(1):1-6. Epub 2008/04/29.
148. Harbottle A, Daly AK, Atherton K, Campbell FC. Role of glutathione S-transferase P1, P-glycoprotein and multidrug resistance-associated protein 1 in acquired doxorubicin resistance. International Journal of Cancer. 2001;92(6):777-83.