

# RESSALVA

Atendendo solicitação do(a) autor(a), o texto completo desta dissertação será disponibilizado somente a partir de 26/04/2024.

UNIVERSIDADE ESTADUAL PAULISTA  
FACULDADE DE MEDICINA VETERINÁRIA E ZOOTECNIA

BIOMARCADORES EM CORDEIRAS (*Ovis aries*) NATURALMENTE  
INFECTADAS POR PARASITOS GASTRINTESTINAIS SUPLEMENTADAS  
COM CAROÇO DE ALGODÃO EM SISTEMA INTEGRADO DE  
PRODUÇÃO AGROPECUÁRIA

VITOLDO ANTONIO KOZLOWSKI NETO

Botucatu – SP

2022

UNIVERSIDADE ESTADUAL PAULISTA  
FACULDADE DE MEDICINA VETERINÁRIA E ZOOTECNIA

BIOMARCADORES EM CORDEIRAS (*Ovis aries*) NATURALMENTE  
INFECTADAS POR PARASITOS GASTRINTESTINAIS SUPLEMENTADAS  
COM CAROÇO DE ALGODÃO EM SISTEMA INTEGRADO DE  
PRODUÇÃO AGROPECUÁRIA

VITOLDO ANTONIO KOZLOWSKI NETO

Dissertação apresentada junto ao  
Programa de Pós-Graduação em  
Medicina Veterinária para a obtenção do  
título de Mestre.

Orientador: Prof. Dr. Alessandro  
Francisco Talamini do Amarante

FICHA CATALOGRÁFICA ELABORADA PELA SEÇÃO TÉC. AQUIS. TRATAMENTO DA INFORM.  
DIVISÃO TÉCNICA DE BIBLIOTECA E DOCUMENTAÇÃO - CÂMPUS DE BOTUCATU - UNESP

BIBLIOTECÁRIA RESPONSÁVEL: ROSEMEIRE APARECIDA VICENTE-CRB 8/5651

Kozlowski Neto, Vitoldo Antonio.

Biomarcadores em cordeiras (*Ovis aries*) naturalmente infectadas por parasitos gastrintestinais suplementadas com caroço de algodão em sistema integrado de produção agropecuária / Vitoldo Antonio Kozlowski Neto. - Botucatu, 2022

Dissertação (mestrado) - Universidade Estadual Paulista "Júlio de Mesquita Filho", Faculdade de Medicina Veterinária e Zootecnia

Orientador: Alessandro Francisco Talamini do Amarante  
Capes: 50502042

1. Ovinos. 2. Cordeiros. 3. Caroço de algodão como ração.  
4. Intestinos - Parasitos. 5. Estresse oxidativo.

Palavras-chave: Caroço de algodão; Estresse oxidativo;  
Ovinos; Parasitos; Proteínas de fase aguda.

Nome do autor: Vitoldo Antonio Kozlowski Neto

Título: BIOMARCADORES EM CORDEIRAS (*Ovis aries*) NATURALMENTE INFECTADAS POR PARASITOS GASTRINTESTINAIS SUPLEMENTADAS COM CAROÇO DE ALGODÃO EM SISTEMA INTEGRADO DE PRODUÇÃO AGROPECUÁRIA

### COMISSÃO EXAMINADORA

Prof. Dr. Alessandro Francisco Talamini do Amarante

Presidente e Orientador

Departamento de Bioestatística, Biologia Vegetal, Parasitologia e Zoologia,  
Instituto de Biociências, Universidade Estadual Paulista Júlio de Mesquita Filho

Prof. Dr. Livio Martins Costa Junior

Membro Titular

Departamento de Patologia, Centro de Ciências Biológicas e da Saúde,  
Universidade Federal do Maranhão

Dr. César Cristiano Bassetto

Membro Titular

Embrapa Pecuária Sudeste – São Carlos

Data da Defesa: 26 de abril de 2022.

## DEDICATÓRIA

A Deus, pela saúde,  
inspiração e força.

## AGRADECIMENTOS

Aos meus pais, Aparecida de Cássia Queiroz Kozlowski e Vitoldo Antonio Kozlowski Junior, pelos ensinamentos, apoio durante todo o tempo de estudo em Botucatu, conforto, orientações, incentivo, exemplos de pessoas na minha construção pessoal e profissional, e principalmente pela paciência em todos os momentos. Sou eternamente grato e tenho muito orgulho por tê-los como pais.

Ao Prof. Dr. Alessandro Francisco Talamini do Amarante pela orientação e por ter me aceito desde o primeiro momento, pela transmissão de conhecimento, exemplo de ética profissional, paciência, por ser acessível e estar sempre disponível.

A Profa. Dra. Elizabeth Moreira dos Santos Schmidt, pelas orientações desde a residência, pela oportunidade de crescimento profissional e pessoal, ensinamentos, disponibilidade, atenção, por estar sempre presente, pelas reuniões, pelo exemplo de ética profissional e por ter aceitado e participado da minha qualificação.

A Dra. Camila Peres Rubio, pela orientação, por ser acessível, pelas reuniões mesmo com a diferença do fuso horário, por ter trabalhado em conjunto e principalmente pelas análises laboratoriais deste estudo, assim como toda a equipe do Interlab-UMU.

Ao Prof. Dr. Ciniro Costa por ter aceitado e participado da minha banca de qualificação, com sugestões e orientações, além da oportunidade de ter trabalhado em conjunto na parte experimental deste estudo.

Ao Prof. Dr. Livio Martins Costa Junior e Dr. César Cristiano Bassetto por terem aceitado o convite como membros titulares na banca de defesa.

Aos meus familiares que sempre me incentivaram e em especial a minha irmã Ana Paula Kozlowski, pelas risadas, brincadeiras, conversas e carinho.

A minha namorada Juliana Pedroso Mendes por ser uma pessoa iluminada, atenciosa, companheira, carinhosa, por sempre estar presente e de me incentivar em todos os momentos.

Aos Professores, Funcionários, Amigos e Colegas da Faculdade de Medicina Veterinária e Zootecnia/UNESP, Botucatu, que fizeram parte do meu crescimento profissional ao longo da residência e mestrado.

A República Zona Azul que me acolheu, forneceu grandes amizades, trocas de experiências e culturas.

A equipe do Laboratório de Helminologia Veterinária que forneceu experiência e ensinamentos, em especial ao José Gabriel Gonçalves Lins e a Naiara Mirelly Marinho da Silva que me auxiliou nas análises laboratoriais.

A Renata Tardivo pelo trabalho em conjunto com os animais, parte experimental e informações adicionais.

O presente trabalho foi realizado com apoio da Coordenação de Aperfeiçoamento de Pessoal de Nível Superior - Brasil (CAPES) - Código de Financiamento 001.

A Fundação de Amparo à Pesquisa do Estado de São Paulo – FAPESP, processo: 17/05044-7, linha de fomento: auxílio à pesquisa – regular, pesquisador responsável e beneficiário: Ciniro Costa.

## LISTA DE TABELAS

Tabela 1. Proteínas de fase aguda em ovinos e caprinos (Fonte: Adaptado de Iliev; Georgieva, 2018). .....	12
Table 1. Formulation and chemical composition of the experimental trial diets	41
Supplementary File 1. Mean ( $\pm$ standard deviation) of eggs per gram of faeces (EPG) and oocysts per gram of faeces (OPG) of crossbred lambs naturally infected with gastrointestinal nematodes and <i>Eimeria</i> spp., receiving two different treatment diets (whole cottonseed and control) at seven different time points (TP1 – TP7). Between each time point there was an interval of 14 days.....	77
Supplementary File 2. Mean ( $\pm$ standard deviation) of body weight (kg) values of crossbred lambs naturally infected with gastrointestinal nematodes and <i>Eimeria</i> spp., receiving two different treatment diets (whole cottonseed and control) at seven different time points (TP1 – TP7). Between each time point there was an interval of 14 days. ....	78
Supplementary File 3. Mean ( $\pm$ standard deviation) of average daily weight gain values of crossbred lambs naturally infected with gastrointestinal nematodes and <i>Eimeria</i> spp., receiving two different treatment diets (whole cottonseed and control) at seven different time points (TP1 – TP7). Between each time point there was an interval of 14 days.....	79
Supplementary File 4. Mean ( $\pm$ standard deviation) concentrations of the biochemical analytes of crossbred lambs naturally infected with gastrointestinal nematodes and <i>Eimeria</i> spp., receiving two different treatment diets (whole cottonseed and control) at seven different time points (TP1 – TP7). Between each time point there was an interval of 14 days. ....	80
Supplementary File 5. Mean ( $\pm$ standard deviation) concentrations of the biomarkers of oxidative stress of crossbred lambs naturally infected with gastrointestinal nematodes and <i>Eimeria</i> spp., receiving two different treatment diets (whole cottonseed and control) at seven different time points (TP1 – TP7). Between each time point there was an interval of 14 days. ....	82

## LISTA DE FIGURAS

Figura 1. Resposta da fase aguda. Início da produção de proteínas de fase aguda após dano tecidual, envolvendo a ativação de macrófagos/monócitos, liberação de citocinas pró-inflamatórias como interleucina 6, 1 e fator de necrose tumoral alfa. Seguido da ativação do fígado e outros tecidos, induzindo a síntese de proteínas de fase aguda. Fonte: Adaptado de Cray, 2012.....	10
Figura 2. Fontes endógenas e exógenas de espécies reativas (ER). Espécies reativas de oxigênio (ERO), espécies reativas de nitrogênio (ERN), espécies reativas derivados do enxofre (ERS), metais de transição [M <sup>(n+)</sup> ], espécies reativas de cloro (ERCl) e espécies reativas de carbono (ERC). Sistema de defesa antioxidante, endógeno e exógeno, atuando para controlar a produção de espécies reativas, a fim de promover a homeostase. Resultado de estresse oxidativo e conseqüentemente danos celulares a partir da produção descontrolada de espécies reativas. Fonte: Adaptado de Finkel; Holbrook, (2000); Vasconcelos et al. (2007). .....	16
Figure 1. Timeline of the experimental trial. Management (M) of crossbred lambs that were tagged, vaccinated, adapted to the diets (whole cottonseed and control), and evaluated through faecal examination. Two treatments were performed, first treatment (1°TN) with 1% moxidectin against gastrointestinal nematodes infection and second treatment (2°TE) with 5% toltrazuril against <i>Eimeria</i> spp. infection. The experimental trial samples were collected at seven different time points (TP1 – TP7). Between each time point and management there was an interval of 14 days.....	39
Figure 2. Distribution of crossbred lambs according to treatment. ....	40
Figure 3. Median (interquartile range) of eggs per gram of faeces (EPG) of crossbred lambs naturally infected with gastrointestinal nematodes, receiving two different treatment diets (whole cottonseed and control) at seven different time points (TP1 – TP7). Between each time point there was an interval of 14 days. ....	44
Figure 4. Median (interquartile range) of <i>Eimeria</i> spp. oocysts per gram of faeces (OPG) of crossbred lambs naturally infected, receiving two different treatment diets (whole cottonseed and control) at seven different time points (TP1 – TP7).	

Between each time point there was an interval of 14 days. The arrow indicates the treatment with 5% toltrazuril. ....	45
Figure 5. Mean ( $\pm$ standard error of the mean) of body weight (kg) values of crossbred lambs naturally infected with gastrointestinal nematodes and <i>Eimeria</i> spp., receiving two different treatment diets (whole cottonseed and control) at seven different time points (TP1 – TP7). Between each time point there was an interval of 14 days. ....	46
Figure 6. Mean ( $\pm$ standard error of the mean) of average daily weight gain (ADG) values of crossbred lambs naturally infected with gastrointestinal nematodes and <i>Eimeria</i> spp., receiving two different treatment diets (whole cottonseed and control) at seven different time points (TP1 – TP7). Between each time point there was an interval of 14 days. ....	47
Figure 7. Mean ( $\pm$ standard error of the mean) of albumin concentrations of crossbred lambs naturally infected with gastrointestinal nematodes and <i>Eimeria</i> spp., receiving two different treatment diets (whole cottonseed and control) at seven different time points (TP1 – TP7). Between each time point there was an interval of 14 days. ....	48
Figure 8. Mean ( $\pm$ standard error of the mean) of total protein concentrations of crossbred lambs naturally infected with gastrointestinal nematodes and <i>Eimeria</i> spp., receiving two different treatment diets (whole cottonseed and control) at seven different time points (TP1 – TP7). Between each time point there was an interval of 14 days. ....	49
Figure 9. Mean ( $\pm$ standard error of the mean) of globulins concentrations of crossbred lambs naturally infected with gastrointestinal nematodes and <i>Eimeria</i> spp., receiving two different treatment diets (whole cottonseed and control) at seven different time points (TP1 – TP7). Between each time point there was an interval of 14 days. ....	50
Figure 10. Mean ( $\pm$ standard error of the mean) of cholesterol concentrations of crossbred lambs naturally infected with gastrointestinal nematodes and <i>Eimeria</i> spp., receiving two different treatment diets (whole cottonseed and control) at seven different time points (TP1 – TP7). Between each time point there was an interval of 14 days. ....	51
Figure 11. Mean ( $\pm$ standard error of the mean) of trolox equivalent antioxidant capacity (TEAC) concentrations of crossbred lambs naturally infected with	

gastrointestinal nematodes and <i>Eimeria</i> spp., receiving two different treatment diets (whole cottonseed and control) at seven different time points (TP1 – TP7). Between each time point there was an interval of 14 days. ....	52
Figure 12. Mean ( $\pm$ standard error of the mean) of cupric reducing antioxidant capacity (CUPRAC) concentrations of crossbred lambs naturally infected with gastrointestinal nematodes and <i>Eimeria</i> spp., receiving two different treatment diets (whole cottonseed and control) at seven different time points (TP1 – TP7). Between each time point there was an interval of 14 days. ....	53
Figure 13. Mean ( $\pm$ standard error of the mean) of ferric reducing ability of plasma (FRAP) concentrations of crossbred lambs naturally infected with gastrointestinal nematodes and <i>Eimeria</i> spp., receiving two different treatment diets (whole cottonseed and control) at seven different time points (TP1 – TP7). Between each time point there was an interval of 14 days. ....	53
Figure 14. Mean ( $\pm$ standard error of the mean) of thiol concentrations of crossbred lambs naturally infected with gastrointestinal nematodes and <i>Eimeria</i> spp., receiving two different treatment diets (whole cottonseed and control) at seven different time points (TP1 – TP7). Between each time point there was an interval of 14 days. ....	54
Figure 15. Mean ( $\pm$ standard error of the mean) of uric acid concentrations of crossbred lambs naturally infected with gastrointestinal nematodes and <i>Eimeria</i> spp., receiving two different treatment diets (whole cottonseed and control) at seven different time points (TP1 – TP7). Between each time point there was an interval of 14 days. ....	55
Figure 16. Mean ( $\pm$ standard error of the mean) of paraoxonase-1 (PON-1) concentrations of crossbred lambs naturally infected with gastrointestinal nematodes and <i>Eimeria</i> spp., receiving two different treatment diets (whole cottonseed and control) at seven different time points (TP1 – TP7). Between each time point there was an interval of 14 days. ....	56
Figure 17. Mean ( $\pm$ standard error of the mean) of advanced oxidation protein products (AOPP) concentrations of crossbred lambs naturally infected with gastrointestinal nematodes and <i>Eimeria</i> spp., receiving two different treatment diets (whole cottonseed and control) at seven different time points (TP1 – TP7). Between each time point there was an interval of 14 days. ....	57

- Figure 18. Mean ( $\pm$  standard error of the mean) of total oxidant status (TOS) concentrations of crossbred lambs naturally infected with gastrointestinal nematodes and *Eimeria* spp., receiving two different treatment diets (whole cottonseed and control) at seven different time points (TP1 – TP7). Between each time point there was an interval of 14 days. .... 58
- Figure 19. Mean ( $\pm$  standard error of the mean) of ferric-xylenol orange (FOX) concentrations of crossbred lambs naturally infected with gastrointestinal nematodes and *Eimeria* spp., receiving two different treatment diets (whole cottonseed and control) at seven different time points (TP1 – TP7). Between each time point there was an interval of 14 days. .... 59
- Figure 20. Mean ( $\pm$  standard error of the mean) of reactive oxygen metabolites derived compounds (d-ROMs) concentrations of crossbred lambs naturally infected with gastrointestinal nematodes and *Eimeria* spp., receiving two different treatment diets (whole cottonseed and control) at seven different time points (TP1 – TP7). Between each time point there was an interval of 14 days. .... 60

## SUMÁRIO

CAPÍTULO I .....	1
1 INTRODUÇÃO .....	2
2 REVISÃO DE LITERATURA .....	3
2.1 Sistema Integrado de Produção Agropecuária.....	3
2.2 Carozo de algodão.....	4
2.3 Parasitos gastrintestinais de ovinos .....	5
2.3.1 Nematódeos.....	5
2.3.2 <i>Eimeria</i> spp. ....	8
2.4 Proteínas de fase aguda .....	9
2.4.1 Haptoglobina.....	11
2.4.2 Albumina .....	12
2.5 Estresse oxidativo e biomarcadores .....	13
2.5.1 Capacidade antioxidante total.....	18
2.5.2 Tiol .....	19
2.5.3 Ácido Úrico.....	20
2.5.4 Paraoxonase-1.....	20
2.5.5 Estado oxidante total .....	20
2.5.6 Método férrico-xilenol.....	21
2.5.7 Produtos da oxidação avançada de proteínas .....	21
2.5.8 Derivados de metabólicos reativos de oxigênio .....	21
3 OBJETIVOS .....	22
3.1 Objetivo geral .....	22
3.2 Objetivos específicos .....	22
REFERÊNCIAS.....	23
CAPÍTULO II .....	33
Abstract.....	34
1. Introduction .....	35
2. Material and Methods.....	37
2.1. Experimental area description.....	37
2.2. Animals and management.....	38

2.3. Body weight, faecal and blood samples .....	41
2.3.1 Eggs and oocysts counting .....	42
2.3.2 Biochemical analytes .....	42
2.3.3. Biomarkers of oxidative stress .....	43
2.4. Statistical analysis .....	43
3. Results .....	44
4. Discussion.....	60
5. Conclusion .....	67
References.....	68
SUPPLEMENTARY FILE .....	77

KOZLOWSKI NETO, V. A. **Biomarcadores em cordeiras (*Ovis aries*) naturalmente infectadas por parasitos gastrintestinais suplementadas com caroço de algodão em sistema integrado de produção agropecuária.** Botucatu, 2022. 100p. Dissertação (Mestrado) – Faculdade de Medicina Veterinária e Zootecnia, Campus de Botucatu, Universidade Estadual Paulista “Júlio de Mesquita Filho”.

## RESUMO

Em animais de fazenda, as perdas econômicas são amplamente descritas devido aos parasitos. Para auxiliar no monitoramento dessas infecções e na suplementação alimentar, as análises bioquímicas e biomarcadores do estresse oxidativo estão se tornando ferramentas importantes na Medicina Veterinária. A presente investigação teve como objetivo avaliar os níveis séricos de biomarcadores do estresse oxidativo e análises bioquímicas em cordeiras cruzadas, naturalmente infectadas por nematódeos gastrintestinais e *Eimeria* spp., com e sem a inclusão do caroço de algodão nas dietas, em sistema integrado de produção agropecuária (SIPA). O experimento foi realizado na Fazenda Experimental do Lageado, pertencente à FMVZ/UNESP, com 36 cordeiras cruzadas (cruzamento: Ile de France x White Dorper x Texel), separadas em 18 animais para cada tratamento (com inclusão de caroço de algodão e controle, sem inclusão de caroço de algodão) na fase de recria. O peso corporal, coleta de sangue e análise fecal de contagens de ovos e oocistos por grama de fezes (OPG e OoPG, respectivamente) foram realizados para cada animal a cada 14 dias (total de sete tempos). O soro obtido foi avaliado quanto às concentrações das análises bioquímicas: haptoglobina, albumina, globulina, proteína total, colesterol e de biomarcadores do estresse oxidativo: capacidade antioxidante cúprica redutora (CUPRAC: *cupric reducing antioxidant capacity*), capacidade de redução férrica do plasma (FRAP: *ferric reducing ability of plasma*), capacidade antioxidante equivalente ao Trolox (TEAC: *Trolox equivalent antioxidant capacity*), tiol, ácido úrico, paraoxonase-1 (PON-1), estado oxidante total (TOS: *total oxidant status*), método férrico-xilenol (FOX: *ferric-xyleneol orange*), produtos da oxidação avançada de proteínas (AOPP: *advanced oxidation protein products*) e derivados de metabólitos reativos de oxigênio (d-ROMs: *reactive oxygen metabolites derived compounds*). A haptoglobina estava abaixo do limite de detecção do ensaio para todos os animais em todos os tempos. O tratamento com inclusão de caroço de algodão apresentou maiores ( $p < 0,05$ ) concentrações de TEAC, AOPP e d-ROMs. Enquanto que a contagem de OoPG, albumina, tiol e FOX foram maiores ( $p < 0,05$ ) para o tratamento controle. Não foram observadas diferenças significativas entre os tratamentos ( $p > 0,05$ ) para a contagem de OPG, peso corporal, ganho de peso médio diário e outras variáveis. A inclusão de caroço de algodão sugeriu o benefício no controle de *Eimeria* spp. além de induzir aumento de oxidantes e antioxidantes em cordeiras naturalmente infectadas por parasitos gastrintestinais. A combinação de caroço de algodão e SIPA pode ser útil no controle de infecções por endoparasitos sem afetar o desempenho da produção.

**Palavras-chave:** Nematódeos gastrintestinais; *Eimeria* spp.; Estresse oxidativo; Inflamação.

KOZLOWSKI NETO, V. A. **Biomarkers in lambs (*Ovis aries*) naturally infected by gastrointestinal parasites supplemented with whole cottonseed in an integrated crop-livestock system.** Botucatu, 2022. 100p. Dissertação (Mestrado) – Faculdade de Medicina Veterinária e Zootecnia, Campus de Botucatu, Universidade Estadual Paulista “Júlio de Mesquita Filho”.

## **ABSTRACT**

In farm animals, economic losses are widely described due to parasites. To assist in monitoring these infections and the feed supplementation, the biochemical analytes and the oxidative stress biomarkers are becoming important tools in Veterinary Medicine. The present investigation aimed to evaluate serum levels of oxidative stress biomarkers and biochemical analytes in crossbred lambs naturally infected by gastrointestinal nematodes and *Eimeria* spp., with and without the dietary inclusion of whole cottonseed (WCS) in an integrated crop-livestock system (ICLS). The experiment was carried out at the Experimental Farm of Lageado, belonging to the FMVZ/UNESP, with 36 crossbred lambs (cross: Ile de France x White Dorper x Texel), separated into 18 animals for each treatment (with WCS inclusion and control, without WCS inclusion) in the rearing phase. Body weight, blood collection and faecal analysis of egg and oocyst counting per gram of faeces (EPG and OPG, respectively) were performed for each animal every 14 days (total of seven time points). The serum obtained were evaluated for concentrations of biochemical analytes: haptoglobin, albumin, globulin, total protein, cholesterol and biomarkers of oxidative stress: cupric reducing antioxidant capacity (CUPRAC), ferric reducing ability of plasma (FRAP), trolox equivalent antioxidant capacity (TEAC), thiol, uric acid, paraoxonase-1 (PON-1), total oxidant status (TOS), ferric-xylenol orange (FOX), advanced oxidation protein products (AOPP) and reactive oxygen metabolites derived compounds (d-ROMs). The haptoglobin were below the detection limit of the assay for all animals at all time points. The treatment with the WCS inclusion had higher ( $p < 0.05$ ) TEAC, AOPP, and d-ROMs concentrations. Whereas the OPG counting, albumin, thiol and FOX were higher ( $p < 0.05$ ) for the control treatment. No significant differences were observed between treatments ( $p > 0.05$ ) for the EPG counting, body weight, average daily weight gain and other variables. The inclusion of WCS suggested the benefit in controlling *Eimeria* spp. infection as well as inducing increase in oxidants and antioxidants in lambs naturally infected by gastrointestinal parasites. The combination of WCS and ICLS could be useful in controlling endoparasite infection without affecting the production performance.

**Keywords:** Gastrointestinal nematodes; *Eimeria* spp.; Oxidative stress; Inflammation.

## REFERÊNCIAS

ALMEIDA, F. A.; PIZA, M. L. S. T.; BASSETTO, C. C.; STARLING, R. Z. C.; ALBUQUERQUE, A. C. A.; PROTES, V. M.; PARIZ, C. M.; CASTILHOS, A. M.; COSTA, C.; AMARANTE, A. F. T. Infection with gastrointestinal nematodes in lambs in different integrated crop-livestock systems (ICL). **Small Ruminant Research**, v. 166, October 2017, p. 66–72, 2018. Disponível em: <<https://doi.org/10.1016/j.smallrumres.2018.07.009>>

AMARANTE, A. F. T. **Os parasitas de ovinos**. São Paulo: Editora UNESP, 2014. a. Disponível em: <<http://books.scielo.org>>

AMARANTE, A. F. T. Sustainable worm control practices in South America. **Small Ruminant Research**, v. 118, n. 1–3, p. 56–62, 2014. b. Disponível em: <<http://dx.doi.org/10.1016/j.smallrumres.2013.12.016>>

AMARANTE, A. F. T.; BARBOSA, M. A. Species of coccidia occurring in lambs in São Paulo State, Brazil. **Veterinary Parasitology**, v. 41, p. 189–193, 1992.

ANDERSON, R. C. **Nematode parasites of vertebrates : their development and transmission**. CABI Publishing, 2000.

ARAB, K.; STEGHENS, J. P. Plasma lipid hydroperoxides measurement by an automated xylenol orange method. **Analytical Biochemistry**, v. 325, n. 1, p. 158–163, 2004.

ARNAO, M. B.; CANO, A.; HERNÁNDEZ-RUIZ, J.; GARCÍA-CÁNOVAS, F.; ACOSTA, M. Inhibition by L-Ascorbic Acid and Other Antioxidants of the 2,2'-Azino-bis(3-ethylbenzthiazoline-6-sulfonic Acid) Oxidation Catalyzed by Peroxidase: A New Approach for Determining Total Antioxidant Status of Foods. **Analytical Biochemistry**, v. 236, n. 2, p. 255–261, 1996.

BALBINO, L. C.; CORDEIRO, L. A. M.; PORFÍRIO-DA-SILVA, V.; MORAES, A. De; MARTÍNEZ, G. B.; ALVARENGA, R. C.; KICHEL, A. N.; FONTANELI, R. S.; SANTOS, H. P. Dos; FRANCHINI, J. C.; GALERANI, P. R. Evolução tecnológica e arranjos produtivos de sistemas de integração lavoura-pecuária-floresta no Brasil. **Pesq. agropec. bras.**, v. 46, n. 10, p. i–xii, 2011.

BAPTISTIOLLI, L.; NARCISO, L. G.; DE ALMEIDA, B. F. M.; BOSCO, A. M.; DE SOUZA, J. C.; TORRECILHA, R. B. P.; PEREIRA, P. P.; FIGUEIREDO, R. N.; GARCIA, J. F.; KANETO, C. N.; CIARLINI, P. C. Systemic oxidative stress in Suffolk and Santa Ines sheep experimentally infected with *Haemonchus contortus*. **Acta Parasitologica**, v. 63, n. 3, p. 504–514, 2018.

BENZIE, I. F. F.; STRAIN, J. J. The Ferric Reducing Ability of Plasma (FRAP) as a Measure of “Antioxidant Power”: The FRAP Assay. **Analytical Biochemistry**, v. 239, p. 70–76, 1996.

BOTTGER, G. T.; SHEEHAN, E. T.; LUKEFAHR, M. J. Relation of gossypol content of cotton plants to insect resistance. **Journal of Economic Entomology**, v. 57, n. 2, p. 283–285, 1964. Disponível em: <<https://academic.oup.com/jee/article/57/2/283/876790>>

BOWMAN, D. D. **Georgis Parasitologia Veterinária**. 9. ed. Rio de Janeiro: Elsevier, 2010.

CAMA, V. A.; MATHISON, B. A. Infections by Intestinal Coccidia and *Giardia duodenalis*. **Clinics in Laboratory Medicine**, v. 35, n. 2, p. 423–444, 2015. Disponível em: <<http://dx.doi.org/10.1016/j.cll.2015.02.010>>

CAMPOS, C.; GUZMÁN, R.; LÓPEZ-FERNÁNDEZ, E.; CASADO, Á. Evaluation of the copper(II) reduction assay using bathocuproinedisulfonic acid disodium salt for the total antioxidant capacity assessment: The CUPRAC-BCS assay. **Analytical Biochemistry**, v. 392, n. 1, p. 37–44, 2009. Disponível em: <<http://dx.doi.org/10.1016/j.ab.2009.05.024>>

CARVALHO, P. C. de F.; MORAES, A. De; PONTES, L. da S.; ANGHINONI, I.; SULC, R. M.; BATELLO, C. Definições e terminologias para Sistema Integrado de Produção Agropecuária. **Revista Ciência Agronômica**, v. 45, n. 5, p. 1040–1046, 2014. Disponível em: <[www.ccarevista.ufc.br](http://www.ccarevista.ufc.br)>

CECILIANI, F.; CERON, J. J.; ECKERSALL, P. D.; SAUERWEIN, H. Acute phase proteins in ruminants. **Journal of Proteomics**, v. 75, n. 14, p. 4207–4231, 2012. Disponível em: <<http://dx.doi.org/10.1016/j.jprot.2012.04.004>>

CESARONE, M. R.; BELCARO, G.; CARRATELLI, M.; CORNELLI, U. A simple test to monitor oxidative stress. **International angiology**, v. 18, n. 2, p. 127–30,

1999.

CHAICHISEMSARI, M.; ESHRATKHAH, B.; MAHERISIS, N.; SADAGHIAN, M.; HASSANPOUR, S. Evaluation of Total Protein, Albumin, Globulin and Blood Urea Nitrogen Concentrations in Gastrointestinal Nematodes Infected Sheep. **Global Veterinaria**, v. 6, n. 5, p. 433–437, 2011.

CHARTIER, C.; PARAUD, C. Coccidiosis due to *Eimeria* in sheep and goats, a review. **Small Ruminant Research**, v. 103, n. 1, p. 84–92, 2012.

COSTA, C. M. Da; SANTOS, R. C. C. Dos; LIMA, E. S. A simple automated procedure for thiol measurement in human serum samples. **J Bras Patol Med Lab**, v. 42, n. 5, p. 345–350, 2006.

COSTA, J. A. A. Da; PARIZ, C. M.; FROTA, M. N. L. Da; REIS, F. A.; COSTA, C.; ARAÚJO NETO, R. B. De; TEIXEIRA NETO, M. L.; MEIRELLES, P. R. de L.; FEIJÓ, G. L. D.; CASTILHOS, A. M. De; CATTO, J. B.; CRUSCIOL, C. A. C.; CARVALHO, G. M. C. Produção de ovinos de corte em sistemas integrados. In: BUNGENSTAB, D. J.; ALMEIDA, R. G. De; LAURA, V. A.; BALBINO, L. C.; FERREIRA, A. D. (Eds.). **ILPF: inovação com integração de lavoura, pecuária e floresta**. 1. ed. Brasília, DF: Embrapa, 2019. p. 241–261.

CRAY, C. **Acute phase proteins in animals**. 1. ed.: Elsevier Inc., 2012. v. 105  
Disponível em: <http://dx.doi.org/10.1016/B978-0-12-394596-9.00005-6>

CUNHA, M. G. G.; CARVALHO, F. F. R.; NETO, S. G.; CEZAR, M. F. Características quantitativas de carcaça de ovinos Santa Inês confinados alimentados com rações contendo diferentes níveis de caroço de algodão integral. **Revista Brasileira de Zootecnia**, v. 37, n. 6, p. 1112–1120, 2008.  
Disponível em: <[www.sbz.org.br](http://www.sbz.org.br)>

DI MASI, A.; DE SIMONE, G.; CIACCIO, C.; D'ORSO, S.; COLETTA, M.; ASCENZI, P. Haptoglobin: From hemoglobin scavenging to human health. **Molecular Aspects of Medicine**, v. 73, p. 100851, 2020. Disponível em: <<https://doi.org/10.1016/j.mam.2020.100851>>

DIAS-SILVA, T. P.; FILHO, A. L. A.; KATIKI, L. M.; AMARANTE, A. F. T.; ABDALLA, A. L.; LOUVANDINI, H. *Trichostrongylus Colubriformis* infection in santa inês lambs: Impact on feed digestibility, blood markers, and nitrogen

balance. **Revista Brasileira de Parasitologia Veterinaria**, v. 29, n. 2, p. 1–7, 2020.

DINLER, C.; ULUTAS, B.; VOYVODA, H.; ULUTAS, P. A.; URAL, K.; KARAGENC, T. Haptoglobin and serum amyloid-A concentrations and their relationship with oocyst count in neonatal lambs experimentally infected with *Cryptosporidium parvum*. **Veterinary Parasitology**, v. 247, n. September, p. 49–56, 2017. Disponível em: <<http://dx.doi.org/10.1016/j.vetpar.2017.09.023>>

EREL, O. A new automated colorimetric method for measuring total oxidant status. **Clinical Biochemistry**, v. 38, p. 1103–1111, 2005.

FAO. An international consultation on integrated crop-livestock systems for development The Way Forward for Sustainable Production. **Integrated Crop Management**, v. 13, n. 1, p. 79, 2010.

FARID, A. S.; NAKAHARA, K.; MURAKAMI, N.; HAYASHI, T.; HORII, Y. Decreased serum paraoxonase-1 activity during intestinal nematode (*Nippostrongylus brasiliensis*) infection in rats. **American Journal of Tropical Medicine and Hygiene**, v. 78, n. 5, p. 770–776, 2008.

FARID, A. S.; SHIMOHIRA, T.; KOBAYASHI, I.; SAWADA, J.; HORII, Y. Intestinally implanted *Nippostrongylus brasiliensis* adult worms decrease serum paraoxonase-1 activity in rats. **Parasitology International**, v. 58, n. 2, p. 178–183, 2009. Disponível em: <<http://dx.doi.org/10.1016/j.parint.2009.03.001>>

FERREIRA, A. L. A.; MATSUBARA, L. S. Radicais livres: conceitos, doenças relacionadas, sistema de defesa e estresse oxidativo. **Rev Ass Med Brasil**, v. 43, n. 1, p. 61–8, 1997.

FINKEL, T.; HOLBROOK, N. J. Oxidants, oxidative stress and the biology of ageing. **Nature**, v. 408, p. 239–247, 2000.

GERON, L. J. V.; MEXIA, A. A.; GARCIA, J.; ZEOULA, L. M.; GARCIA, R. R. F.; MOURA, D. C. De. DESEMPENHO DE CORDEIROS EM TERMINAÇÃO SUPLEMENTADOS COM CAROÇO DE ALGODÃO (*GOSSYPIUM HIRSUTUM* L.) E GRÃO DE MILHO MOÍDO (*ZEA MAYS* L.). **Archives of Veterinary Science**, v. 17, n. 4, p. 34–42, 2012. Disponível em: <[www.ser.ufpr.br/veterinary](http://www.ser.ufpr.br/veterinary)>

GONZÁLEZ, F. H. D.; TECLES, F.; MARTÍNEZ-SUBIELA, S.; TVARIJONAVICIUTE, A.; VASCO, L. S.; CERÓN, J. J. Acute phase protein response in goats. **Journal of Veterinary Diagnostic Investigation**, v. 20, n. 5, p. 580–584, 2008.

GYURÁSZOVÁ, M.; KOVALČÍKOVÁ, A.; JANŠÁKOVÁ, K.; ŠEBEKOVÁ, K.; CELEC, P.; TÓTHOVÁ, L. Markers of oxidative stress and antioxidant status in the plasma, urine and saliva of healthy mice. **Physiological Research**, v. 67, p. 921–934, 2018.

HUANG, D.; BOXIN, O. U.; PRIOR, R. L. The chemistry behind antioxidant capacity assays. **Journal of Agricultural and Food Chemistry**, v. 53, n. 6, p. 1841–1856, 2005.

ILIEV, P. T.; GEORGIEVA, T. M. Acute phase proteins in sheep and goats – function, reference ranges and assessment methods: an overview. **Bulgarian Journal of Veterinary Medicine**, v. 21, n. 1, p. 1–16, 2018.

KANDYLIS, K.; NIKOKYRIS, P. N.; DELIGIANNIS, K. Performance of Growing-Fattening Lambs Fed Whole Cotton Seed. **Journal of the Science of Food and Agriculture**, v. 78, n. 2, p. 281–289, 1998.

KEETON, S. T. N.; NAVARRE, C. B. Coccidiosis in Large and Small Ruminants. **Veterinary Clinics of North America - Food Animal Practice**, v. 34, n. 1, p. 201–208, 2018. Disponível em: <<https://doi.org/10.1016/j.cvfa.2017.10.009>>

KNUTSEN, H. K.; BARREGÅRD, L.; BIGNAMI, M.; BRÜSCHWEILER, B.; CECCATELLI, S.; DINOVI, M.; EDLER, L.; GRASL-KRAUPP, B.; HOGSTRAND, C.; HOOGENBOOM, L.; NEBBIA, C. S.; OSWALD, I. P.; PETERSEN, A.; ROSE, M.; ROUDOT, A. C.; SCHWERDTLE, T.; VLEMINCKX, C.; VOLLMER, G.; WALLACE, H.; ALEXANDER, J.; COTTRILL, B.; MACKAY, K. Presence of free gossypol in whole cottonseed. **EFSA Journal**, v. 15, n. 7, 2017.

LYKKESFELDT, J.; SVENDSEN, O. Oxidants and antioxidants in disease: Oxidative stress in farm animals. **The Veterinary Journal**, v. 173, n. 3, p. 502–511, 2007.

MACHADO, V.; DA SILVA, A. S.; SCHAFFER, A. S.; AIRES, A. R.; TONIN, A. A.; OLIVEIRA, C. B.; HERMES, C. L.; ALMEIDA, T. C.; MORESCO, R. N.; STEFANI,

L. M.; LOPES, S. T. A.; MENDES, R. E.; SCHWERTZ, C. I.; LEAL, M. L. R. Relationship between oxidative stress and pathological findings in abomasum of infected lambs by *Haemonchus contortus*. **Pathology Research and Practice**, v. 210, n. 12, p. 812–817, 2014. Disponível em: <<http://dx.doi.org/10.1016/j.prp.2014.09.006>>

MACKNESS, B.; DURRINGTON, P. N.; MACKNESS, M. I. Human Serum Paraoxonase. **General Pharmacology**, v. 31, n. 3, p. 329–336, 1998.

MACKNESS, M. I.; ARROL, S.; DURRINGTON, P. N. Paraoxonase prevents accumulation of lipoperoxides in low-density lipoprotein. **FEBS Letters**, v. 286, n. 1–2, p. 152–154, 1991.

MADRUGA, M. S.; VIEIRA, T. R. D. L.; CUNHA, M. das G. G.; FILHO, J. M. P.; QUEIROGA, R. de C. R. do E.; SOUSA, W. H. De. Efeito de dietas com níveis crescentes de caroço de algodão integral sobre a composição química e o perfil de ácidos graxos da carne de cordeiros Santa Inês. **Revista Brasileira de Zootecnia**, v. 37, n. 8, p. 1496–1502, 2008. Disponível em: <[www.sbz.org.br](http://www.sbz.org.br)>

MATHISON, B. A.; BRADBURY, R. S.; PRITT, B. S. Medical Parasitology Taxonomy Update, January 2018 to May 2020. **Journal of Clinical Microbiology**, v. 59, n. 2, 2021.

MILLER, N. J.; RICE-EVANS, C.; DAVIES, M. J.; GOPINATHAN, V.; MILNER, A. A novel method for measuring antioxidant capacity and its application to monitoring the antioxidant status in premature neonates. **Clinical Science**, v. 84, n. 4, p. 407–412, 1993.

MONTEIRO, S. G. **Parasitologia na medicina veterinária**. 2. ed. Rio de Janeiro: Roca, 2017.

MORAES, A. De; CARVALHO, P. C. de F.; CRUSCIOL, C. A. C.; LANG, C. R.; PARIZ, C. M.; DEISS, L.; SULC, R. M. Integrated crop-livestock systems as a solution facing the destruction of pampa and cerrado biomes in south america by intensive monoculture systems. **Agroecosystem Diversity: Reconciling Contemporary Agriculture and Environmental Quality**, p. 257–273, 2019.

NAGY, O.; TÓTHOVÁ, C.; KLEIN, R.; CHOVANOVÁ, F. The electrophoretic pattern of serum proteins in sheep with naturally acquired gastrointestinal

nematode infections. **Acta Veterinaria-Beograd**, v. 70, n. 3, p. 316–328, 2020.

NRC. National Research Council. **Nutrient Requirements of Small Ruminants: Sheep, Goats, Cervids and New Camelids**. Washington, DC: The National Academies Press, 2007.

ODDEN, A.; DENWOOD, M. J.; STUEN, S.; ROBERTSON, L. J.; RUIZ, A.; HAMNES, I. S.; HEKTOEN, L.; ENEMARK, H. L. Field evaluation of anticoccidial efficacy: A novel approach demonstrates reduced efficacy of toltrazuril against ovine *Eimeria* spp. in Norway. **International Journal for Parasitology: Drugs and Drug Resistance**, v. 8, n. 2, p. 304–311, 2018.

OLIVEIRA, P. V. S.; LAURINDO, F. R. M. Implications of plasma thiol redox in disease. **Clinical Science**, v. 132, p. 1257–1280, 2018.

ÖZYÜREK, M.; GÜÇLÜ, K.; TÜTEM, E.; BAKAN, K. S.; ERÇAĞ, E.; ESİN ÇELİK, S.; BAKI, S.; YILDIZ, L.; KARAMAN, Ş.; APAK, R. A comprehensive review of CUPRAC methodology. **Analytical Methods**, v. 3, p. 2439–2453, 2011.

PATHAK, A. K.; DUTTA, N.; PATTANAIK, A. K.; SHARMA, K.; BANERJEE, P. S.; GOSWAMI, T. K. The effect of condensed tannins supplementation through *Ficus infectoria* and *Psidium guajava* leaf meal mixture on erythrocytic antioxidant status, immune response and gastrointestinal nematodes in lambs (*Ovis aries*). **Veterinarski Arhiv**, v. 87, n. 2, p. 139–156, 2017.

PIECHOTA-POLANCZYK, A.; FICHNA, J. Review article: The role of oxidative stress in pathogenesis and treatment of inflammatory bowel diseases. **Naunyn-Schmiedeberg's Archives of Pharmacology**, v. 387, n. 7, p. 605–620, 2014.

PIVOTO, F. L.; TORBITZ, V. D.; AIRES, A. R.; DA ROCHA, J. F. X.; SEVERO, M. M.; GRANDO, T. H.; PEITER, M.; MORESCO, R. N.; DA ROCHA, J. B. T.; LEAL, M. L. do R. Oxidative stress by *Haemonchus contortus* in lambs: Influence of treatment with zinc edetate. **Research in Veterinary Science**, v. 102, p. 22–24, 2015. Disponível em: <<http://dx.doi.org/10.1016/j.rvsc.2015.07.001>>

RALPH, A.; O'SULLIVAN, M. V. N.; SANGSTER, N. C.; WALKER, J. C. Abdominal pain and eosinophilia in suburban goat keepers. **Medical Journal of Australia**, v. 184, n. 9, p. 467–469, 2006.

RASHID, S.; IRSHADULLAH, M. Evaluation of antioxidant and oxidant status of goats (*Capra aegagrus hircus*) naturally infected with *Haemonchus contortus*. **Journal of Helminthology**, v. 94, n. May, p. 1–6, 2020.

RIVERA-GOMIS, J.; RUBIO, C. P.; CONESA, C. M.; SALAVERRI, J. O.; CERÓN, J. J.; TORTOSA, D. E.; PABLO, M. J. C. Effects of dietary supplementation of garlic and oregano essential oil on biomarkers of oxidative status, stress and inflammation in postweaning piglets. **Animals**, v. 10, p. 1–17, 2020.

RODRÍGUEZ, C. C.; MENGE, F. W.; CERÓN, J. J. Oxidative Stress in Veterinary Medicine. **Veterinary Medicine International**, v. 2011, p. 1–1, 2011. Disponível em: <http://www.hindawi.com/journals/vmi/2011/812086/>

ROGÉRIO, M. C. P.; BORGES, I.; SANTIAGO, G. S.; TEIXEIRA, D. A. B. Uso do caroço de algodão na alimentação de ruminantes. **Arq. ciên. vet. zool. UNIPAR**, v. 6, n. 1, p. 85–90, 2003.

ROGERS, G. M.; POORE, M. H.; PASCHAL, J. C. Feeding cotton products to cattle. **Veterinary Clinics: Food Animal Practice**, v. 18, n. 2, p. 267–294, 2002.

ROSA, V. D. Da; BORDINHÃO, T.; DIAS, J. B.; OKINO, A. M.; VENTURINI, D. Nível de ácido úrico como biomarcador diagnóstico e prognóstico de doenças cardiovasculares. **Ciências Biológicas e da Saúde**, v. 36, n. 1, p. 159–168, 2015.

RUBIO, C. P.; CERÓN, J. J. Spectrophotometric assays for evaluation of Reactive Oxygen Species (ROS) in serum: general concepts and applications in dogs and humans. **BMC veterinary research**, v. 17, n. 1, p. 226, 2021.

RUBIO, C. P.; CONTRERAS-AGUILAR, M. D.; QUILES, A.; LÓPEZ-ARJONA, M.; CERÓN, J. J.; MARTÍNEZ-SUBIELA, S.; HEVIA, M. L.; ESCRIBANO, D.; TECLES, F. Biomarkers of oxidative stress in saliva of sheep: Analytical performance and changes after an experimentally induced stress. **Research in Veterinary Science**, v. 123, p. 71–76, 2019. Disponível em: <<https://doi.org/10.1016/j.rvsc.2018.12.015>>

RUBIO, C. P.; HERNÁNDEZ-RUIZ, J.; MARTINEZ-SUBIELA, S.; TVARIJONAVICIUTE, A.; CERON, J. J. Spectrophotometric assays for total

antioxidant capacity (TAC) in dog serum: An update. **BMC Veterinary Research**, v. 12, n. 1, p. 1–7, 2016. Disponível em: <<http://dx.doi.org/10.1186/s12917-016-0792-7>>

SHUNMOOGAM, N.; NAIDOO, P.; CHILTON, R. Paraoxonase (PON)-1: A brief overview on genetics, structure, polymorphisms and clinical relevance. **Vascular Health and Risk Management**, v. 14, p. 137–143, 2018.

SIHAG, M. K.; PATEL, A.; KUMAR, V. Cottonseed (*Gossypium hirsutum*). In: **Oilseeds: Health Attributes and Food Applications**. Singapore: Springer Singapore, 2021. p. 73–92.

SMITH, N. C.; BRYANT, C. Free radical generation during primary infections with *Nippostrongylus brasiliensis*. **Parasite Immunology**, v. 11, n. 2, p. 147–160, 1989.

TAYLOR, M. A.; COOP, R. L.; WALL, R. L. **Veterinary Parasitology**. 3. ed. Oxford: Blackwell Publishing Ltd, 2007. Disponível em: <[www.BlackwellVet.com](http://www.BlackwellVet.com)>

TAYLOR, M. A.; COOP, R. L.; WALL, R. L. **Parasitologia Veterinária**. 4. ed. Rio de Janeiro: Editora Guanabara Koogan Ltda., 2017.

TOTHOVA, C.; NAGY, O.; KOVAC, G. Acute phase proteins and their use in the diagnosis of diseases in ruminants: A review. **Veterinarni Medicina**, v. 59, n. 4, p. 163–180, 2014.

TVARIJONAVICIUTE, A.; TECLES, F.; CALDIN, M.; TASCA, S.; CERÓN, J. Validation of spectrophotometric assays for serum paraoxonase type-1 measurement in dogs. **American Journal of Veterinary Research**, v. 73, n. 1, p. 34–41, 2012.

VASCONCELOS, S. M. L.; GOULART, M. O. F.; MOURA, J. B. de F.; BENFATO, V. M. e M. da S.; KUBOTA, L. T. Espécies reativas de oxigênio e de nitrogênio, antioxidantes e marcadores de dano oxidativo em sangue humano: principais métodos analíticos para sua determinação. **Quim. Nova**, v. 30, n. 5, p. 1323–1338, 2007.

VILLANUEVA, C.; KROSS, R. D. Antioxidant-induced stress. **International**

**Journal of Molecular Sciences**, v. 13, n. 2, p. 2091–2109, 2012.

WITKO-SARSAT, V.; FRIEDLANDER, M.; CAPELLÈRE-BLANDIN, C.; NGUYEN-KHOA, T.; NGUYEN, A. T.; ZINGRAFF, J.; JUNGERS, P.; DESCAMPS-LATSCHA, B. Advanced oxidation protein products as a novel marker of oxidative stress in uremia. **Kidney International**, v. 49, n. 5, p. 1304–1313, 1996

WŁODEK, L. Beneficial and harmful effects of thiols. **Polish Journal of Pharmacology**, v. 54, p. 215–223, 2002.

ZAJAC, A. M. Gastrointestinal Nematodes of Small Ruminants: Life Cycle, Anthelmintics, and Diagnosis. **Veterinary Clinics of North America - Food Animal Practice**, v. 22, n. 3, p. 529–541, 2006.

## Oxidative stress biomarkers in lambs naturally infected by gastrointestinal nematodes and *Eimeria* spp. supplemented with whole cottonseed

### Abstract

In farm animals, economic losses are widely described due to gastrointestinal parasites infections. To assist in monitoring these infections and the feed supplementation, the oxidative stress biomarkers and biochemical analytes are relevant tools in Veterinary Medicine. The purpose of this trial was to evaluate serum levels of oxidative stress biomarkers and biochemical analytes in crossbred lambs naturally infected by gastrointestinal nematodes and *Eimeria* spp., with and without the dietary inclusion of whole cottonseed (WCS) in an integrated crop-livestock system (ICLS). The experiment was carried out with 36 crossbred lambs (cross: Ile de France x White Dorper x Texel), in the rearing phase. Body weight, blood collection and faecal analysis of eggs and oocysts counting per gram of faeces (EPG and OPG, respectively) were performed for each animal every 14 days, with a total of seven time points within 85 days of experiment. Serum biochemical analytes were determined: total protein, albumin, globulin, cholesterol, haptoglobin, and oxidative stress biomarkers: cupric reducing antioxidant capacity (CUPRAC), ferric reducing ability of plasma (FRAP), trolox equivalent antioxidant capacity (TEAC), thiol, uric acid, paraoxonase-1 (PON-1), total oxidant status (TOS), ferric-xylenol orange (FOX), advanced oxidation protein products (AOPP), and reactive oxygen metabolites derived compounds (d-ROMs). The treatment with the WCS inclusion had higher ( $p < 0.05$ ) TEAC, AOPP, and d-ROMs concentrations. Whereas the OPG counting, albumin, thiol and FOX were higher ( $p < 0.05$ ) for the control treatment (without the WCS). No significant differences were observed between treatments ( $p > 0.05$ ) for EPG counting, body weight, average daily weight gain, and other biomarkers of oxidative stress and biochemical analytes. The inclusion of WCS suggested the benefit in controlling *Eimeria* spp. infection as well as inducing an increase in oxidants and antioxidants in lambs naturally infected by gastrointestinal parasites. The combination of WCS and ICLS could be useful in controlling gastrointestinal parasites infection without affecting the production performance.

**Keywords:** *Haemonchus*; *Trichostrongylus*; antioxidant; oxidant; biochemical analytes; integrated crop-livestock system

## References

- Alam, R.T.M., Hassanen, E.A.A., El-Mandrawy, S.A.M., 2020. *Heamonchus contortus* infection in Sheep and Goats: alterations in haematological, biochemical, immunological, trace element and oxidative stress markers. *J. Appl. Anim. Res.* 48, 357–364. <https://doi.org/10.1080/09712119.2020.1802281>
- Almeida, F.A., Piza, M.L.S.T., Bassetto, C.C., Starling, R.Z.C., Albuquerque, A.C.A., Protes, V.M., Pariz, C.M., Castilhos, A.M., Costa, C., Amarante, A.F.T., 2018. Infection with gastrointestinal nematodes in lambs in different integrated crop-livestock systems (ICL). *Small Rumin. Res.* 166, 66–72. <https://doi.org/10.1016/j.smallrumres.2018.07.009>
- Alvares, C.A., Stape, J.L., Sentelhas, P.C., Gonçalves, J.L. de M., Sparovek, G., 2013. Köppen's climate classification map for Brazil. *Meteorol. Z.* 22, 711–728. <https://doi.org/10.1127/0941-2948/2013/0507>
- Amarante, A.F.T., Barbosa, M.A., 1992. Species of coccidia occurring in lambs in São Paulo State, Brazil. *Vet. Parasitol.* 41, 189–193. [https://doi.org/10.1016/0304-4017\(92\)90078-N](https://doi.org/10.1016/0304-4017(92)90078-N)
- Amarante, A.F.T., Barbosa, M.A., Sequeira, J.L., 1993. Coccidiose em cordeiros em Botucatu - SP, relato de dois casos. *Brazil J. Vet. Parasitol.* 2, 73–74.
- Arab, K., Steghens, J.-P., 2004. Plasma lipid hydroperoxides measurement by an automated xylenol orange method. *Anal. Biochem.* 325, 158–163. <https://doi.org/10.1016/j.ab.2003.10.022>
- Arnao, M.B., Cano, A., Hernández-Ruiz, J., García-Cánovas, F., Acosta, M., 1996. Inhibition byl-Ascorbic Acid and Other Antioxidants of the 2,2'-Azino-bis(3-ethylbenzthiazoline-6-sulfonic Acid) Oxidation Catalyzed by Peroxidase: A New Approach for Determining Total Antioxidant Status of Foods. *Anal. Biochem.* 236, 255–261. <https://doi.org/10.1006/abio.1996.0164>

- Baptistiolli, L., Narciso, L.G., Almeida, B.F.M. de, Bosco, A.M., Souza, J.C. de, Torrecilha, R.B.P., Pereira, P.P., Figueiredo, R.N., Garcia, J.F., Kaneto, C.N., Ciarlini, P.C., 2018. Systemic oxidative stress in Suffolk and Santa Ines sheep experimentally infected with *Haemonchus contortus*. *Acta Parasitol.* 63, 504–514. <https://doi.org/10.1515/ap-2018-0060>
- Benzie, I.F.F., Strain, J.J., 1996. The Ferric Reducing Ability of Plasma (FRAP) as a Measure of “Antioxidant Power”: The FRAP Assay. *Anal. Biochem.* 239, 70–76. <https://doi.org/10.1006/abio.1996.0292>
- Bottger, G.T., Sheehan, E.T., Lukefahr, M.J., 1964. Relation of Gossypol Content of Cotton Plants to Insect Resistance. *J. Econ. Entomol.* 57, 283–285. <https://doi.org/10.1093/jee/57.2.283>
- Câmara, A.C.L., do Vale, A.M., Mattoso, C.R.S., Melo, M.M., Soto-Blanco, B., 2016. Effects of gossypol from cottonseed cake on the blood profile in sheep. *Trop. Anim. Health. Prod.* 48, 1037–1042. <https://doi.org/10.1007/s11250-016-1039-0>
- Campos, C., Guzmán, R., López-Fernández, E., Casado, Á., 2009. Evaluation of the copper(II) reduction assay using bathocuproinedisulfonic acid disodium salt for the total antioxidant capacity assessment: The CUPRAC–BCS assay. *Anal. Biochem.* 392, 37–44. <https://doi.org/10.1016/j.ab.2009.05.024>
- Cardia, D.F.F., Rocha-Oliveira, R.A., Tsunemi, M.H., Amarante, A.F.T., 2011. Immune response and performance of growing Santa Ines lambs to artificial *Trichostrongylus colubriformis* infections. *Vet. Parasitol.* 182, 248–258. <https://doi.org/10.1016/j.vetpar.2011.05.017>
- Carneiro, P.G., Sasse, J.P., Silva, A.C. dos S., Seixas, M. de, Paschoal, A.T.P., Minutti, A.F., Martins, T.A., Cardim, S.T., Rodrigues, F. de S., Barros, L.D. de, Garcia, J.L., 2022. Prevalence and risk factors of *Eimeria* spp. natural infection in sheep from northern Paraná, Brazil. *Braz. J. Vet. Parasitol.* 31. <https://doi.org/10.1590/s1984-29612022004>

- Carvalho, N., Neves, J.H. das, Pennacchi, C.S., Castilhos, A.M. de, Amarante, A.F.T. do, 2021. Performance of lambs under four levels of dietary supplementation and artificially mix-infected with *Haemonchus contortus* and *Trichostrongylus colubriformis*. *Braz. J. Vet. Parasitol.* 30. <https://doi.org/10.1590/s1984-29612021010>
- Catchpole, J., Gregory, M.W., 1985. Pathogenicity of the coccidium *Eimeria crandallis* in laboratory lambs. *Parasitology* 91, 45–52. <https://doi.org/10.1017/S003118200005650X>
- Cesarone, M.R., Belcaro, G., Carratelli, M., Cornelli, U., de Sanctis, M.T., Incandela, L., Barsotti, A., Terranova, R., Nicolaidis, A., 1999. A simple test to monitor oxidative stress. *Int. Angiol.* 18, 127–30.
- Chartier, C., Paraud, C., 2012. Coccidiosis due to *Eimeria* in sheep and goats, a review. *Small Rumin. Res.* 103, 84–92. <https://doi.org/10.1016/j.smallrumres.2011.10.022>
- Cornell Net Carbohydrate and Protein System, 2000. The Net Carbohydrate and Protein System for Evaluating Herd Nutrition and Nutrients Excretion. Version 5.0. CNCPS, Ithaca, 237 pp.
- Costa, C.M. da, Santos, R.C.C. dos, Lima, E.S., 2006. A simple automated procedure for thiol measurement in human serum samples. *J. Bras. Patol. Med. Lab.* 42, 345–350. <https://doi.org/10.1590/S1676-24442006000500006>
- Costa, J.A.A. da, Pariz, C.M., Frota, M.N.L. da, Reis, F.A., Costa, C., Araújo Neto, R.B. de, Teixeira Neto, M.L., Meirelles, P.R. de L., Feijó, G.L.D., Castilhos, A.M. de, Catto, J.B., Crusciol, C.A.C., Carvalho, G.M.C., 2019. Produção de ovinos de corte em sistemas integrados, in: Bungenstab, D.J., Almeida, R.G. de, Laura, V.A., Balbino, L.C., Ferreira, A.D. (Eds.), *ILPF: Inovação Com Integração de Lavoura, Pecuária e Floresta*. Embrapa, Brasília, DF, pp. 241–261.

- Cunha, A.R. da, Martins, D., 2009. Climatic classification for the Districts of Botucatu and São Manuel, SP. *Irriga* 14, 1–11. <https://doi.org/10.15809/irriga.2009v14n1p1-11>
- Demir, H., Can, A., 2019. Effect of various levels of dietary whole cottonseed on blood parameters and performance of Awassi lambs under heat stress. *S. Afr. J. Anim. Sci.* 49, 50–55. <https://doi.org/10.4314/sajas.v49i1.6>
- Erel, O., 2005. A new automated colorimetric method for measuring total oxidant status. *Clin. Biochem.* 38, 1103–1111. <https://doi.org/10.1016/j.clinbiochem.2005.08.008>
- Escobedo, J.F., Gomes, E.N., Oliveira, A.P., Soares, J., 2011. Ratios of UV, PAR and NIR components to global solar radiation measured at Botucatu site in Brazil. *Renew. Energy.* 36, 169–178. <https://doi.org/10.1016/j.renene.2010.06.018>
- FAO, 2006. World reference base for soil resources 2006: A framework for international classification, correlation and communication. Food and Agriculture Organization of the United Nations, Rome, 128 pp.
- Fossati, P., Prencipe, L., Berti, G., 1980. Use of 3,5-Dichloro-2-hydroxybenzenesulfonic Acid/4-Aminophenazone Chromogenic System in Direct Enzymic Assay of Uric Acid in Serum and Urine. *Clin. Chem.* 26, 227–231. <https://doi.org/10.1093/clinchem/26.2.227>
- Frazzoli, C., Bocca, B., Mantovani, A., 2015. The One Health Perspective in Trace Elements Biomonitoring. *J. Toxicol. Environ. Health B Crit. Rev.* 18, 344–370. <https://doi.org/10.1080/10937404.2015.1085473>
- González-Garza, M.T., Matlin, S.A., Mata-Cárdenas, B.D., Said-Fernández, S., 1993. Differential effects of the (+)- and (–)-gossypol enantiomers upon *Entamoeba histolytica* axenic cultures. *J. Pharm. Pharmacol.* 45, 144–145. <https://doi.org/10.1111/j.2042-7158.1993.tb03701.x>

- Gordon, H.M., Whitlock, H. v., 1939. A New Technique for Counting Nematode Eggs in Sheep Faeces. *J. Counc. Sci. Ind Res.* 12, 50–52.
- Kandyliis, K., Nikokyris, P.N., Deligiannis, K., 1998. Performance of Growing-Fattening Lambs Fed Whole Cotton Seed. *J. Sci. Food Agric.* 78, 281–289. [https://doi.org/10.1002/\(SICI\)1097-0010\(199810\)78:2<281::AID-JSFA116>3.0.CO;2-W](https://doi.org/10.1002/(SICI)1097-0010(199810)78:2<281::AID-JSFA116>3.0.CO;2-W)
- Kaneko, J.J., Harvey, J.W., Bruss, M.L., 2008. *Clinical Biochemistry of Domestic Animals*, 6th ed. Academic Press. <https://doi.org/10.1016/B978-0-12-370491-7.X0001-3>
- Kluthcouski, João, Yokoyama, L.P., 2003. Opções de integração lavoura-pecuária, in: Kluthcouski, J., Stone, L.F., Aidar, H. (Eds.), *Integração Lavoura-Pecuária*. Embrapa Arroz e Feijão, Santo Antônio de Goiás, pp. 129–141.
- Knutsen, H.K., Barregård, L., Bignami, M., Brüscheweiler, B., Ceccatelli, S., Dinovi, M., Edler, L., Grasl-Kraupp, B., Hogstrand, C., Hoogenboom, L. (Ron), Nebbia, C.S., Oswald, I.P., Petersen, A., Rose, M., Roudot, A., Schwerdtle, T., Vleminckx, C., Vollmer, G., Wallace, H., Alexander, J., Cottrill, B., Mackay, K., 2017. Presence of free gossypol in whole cottonseed. *EFSA J.* 15, 1–15. <https://doi.org/10.2903/j.efsa.2017.4850>
- Lykkesfeldt, J., Svendsen, O., 2007. Oxidants and antioxidants in disease: Oxidative stress in farm animals. *Vet. J.* 173, 502–511. <https://doi.org/10.1016/j.tvjl.2006.06.005>
- Machado, V., da Silva, A.S., Schafer, A.S., Aires, A.R., Tonin, A.A., Oliveira, C.B., Hermes, C.L., Almeida, T.C., Moresco, R.N., Stefani, L.M., Lopes, S.T.A., Mendes, R.E., Schwertz, C.I., Leal, M.L.R., 2014. Relationship between oxidative stress and pathological findings in abomasum of infected lambs by *Haemonchus contortus*. *Pathol. Res. Pract.* 210, 812–817. <https://doi.org/10.1016/j.prp.2014.09.006>

- Mena, H., Santos, J.E.P., Huber, J.T., Tarazon, M., Calhoun, M.C., 2004. The Effects of Varying Gossypol Intake from Whole Cottonseed and Cottonseed Meal on Lactation and Blood Parameters in Lactating Dairy Cows. *J. Dairy Sci.* 87, 2506–2518. [https://doi.org/10.3168/jds.S0022-0302\(04\)73375-5](https://doi.org/10.3168/jds.S0022-0302(04)73375-5)
- Mohamaden, W.I., Sallam, N.H., Abouelhassan, E.M., 2018. Prevalence of *Eimeria* species among sheep and goats in Suez Governorate, Egypt. *Int. J. Vet. Sci. Med.* 6, 65–72. <https://doi.org/10.1016/j.ijvsm.2018.02.004>
- Montamat, E.E., Burgos, C., Gerez de Burgos, N.M., Rovai, L.E., Blanco, A., Segura, E.L., 1982. Inhibitory Action of Gossypol on Enzymes and Growth of *Trypanosoma cruzi*. *Science* 218, 288–289. <https://doi.org/10.1126/science.6750791>
- Moretti, D.B., Jimenez, C.R., Trinca, H.M., Machado-Neto, R., Louvandini, H., 2019. Cottonseed feeding changes oxidative stress markers in ewes during the peripartum period and increases the quality of colostrum. *Vet. J.* 247, 32–37. <https://doi.org/10.1016/j.tvjl.2019.02.009>
- NRC, 2007. *Nutrient Requirements of Small Ruminants: Sheep, Goats, Cervids, and New World Camelids*. The National Academies Press, Washington, D.C., 384 pp. <https://doi.org/10.17226/11654>
- Pariz, C.M., Costa, N.R., Costa, C., Crusciol, C.A.C., de Castilhos, A.M., Meirelles, P.R. de L., Calonego, J.C., Andreotti, M., Souza, D.M. de, Cruz, I.V., Longhini, V.Z., Protes, V.M., Sarto, J.R.W., Piza, M.L.S. de T., Melo, V.F. de P., Sereia, R.C., Fachiolli, D.F., Almeida, F.A. de, Souza, L.G.M. de, Franzluebbbers, A.J., 2020. An Innovative Corn to Silage-Grass-Legume Intercropping System With Oversown Black Oat and Soybean to Silage in Succession for the Improvement of Nutrient Cycling. *Front. Sustain. Food Syst.* 4, 1–20. <https://doi.org/10.3389/fsufs.2020.544996>
- Pathak, A.K., Dutta, N., Pattanaik, A.K., Sharma, K., Banerjee, P.S., Goswami, T.K., 2017. The effect of condensed tannins supplementation through *Ficus infectoria* and *Psidium guajava* leaf meal mixture on erythrocytic antioxidant

- status, immune response and gastrointestinal nematodes in lambs (*Ovis aries*). *Vet. Arh.* 87, 139–156.
- Risco, C.A., Adams, A.L., Seebohm, S., Thatcher, M.-J., Staples, C.R., van Horn, H.H., McDowell, L.R., Calhoun, M.C., Thatcher, W.W., 2002. Effects of Gossypol from Cottonseed on Hematological Responses and Plasma  $\alpha$ -Tocopherol Concentration of Dairy Cows. *J. Dairy Sci.* 85, 3395–3402. [https://doi.org/10.3168/jds.S0022-0302\(02\)74427-5](https://doi.org/10.3168/jds.S0022-0302(02)74427-5)
- Risco, C.A., Holmberg, C.A., Kutches, A., 1992. Effect of Graded Concentrations of Gossypol on Calf Performance: Toxicological and Pathological Considerations. *J. Dairy Sci.* 75, 2787–2798. [https://doi.org/10.3168/jds.S0022-0302\(92\)78042-4](https://doi.org/10.3168/jds.S0022-0302(92)78042-4)
- Rogers, G.M., Poore, M.H., Paschal, J.C., 2002. Feeding cotton products to cattle. *Vet. Clin. North Am. Food Anim. Pract.* 18, 267–294. [https://doi.org/10.1016/S0749-0720\(02\)00020-8](https://doi.org/10.1016/S0749-0720(02)00020-8)
- Rubio, C.P., Cerón, J.J., 2021. Spectrophotometric assays for evaluation of Reactive Oxygen Species (ROS) in serum: general concepts and applications in dogs and humans. *BMC Vet. Res.* 17, 226. <https://doi.org/10.1186/s12917-021-02924-8>
- Rubio, C.P., Contreras-Aguilar, M.D., Quiles, A., López-Arjona, M., Cerón, J.J., Martínez-Subiela, S., Hevia, M.L., Escribano, D., Tecles, F., 2019. Biomarkers of oxidative stress in saliva of sheep: Analytical performance and changes after an experimentally induced stress. *Res. Vet. Sci.* 123, 71–76. <https://doi.org/10.1016/j.rvsc.2018.12.015>
- Rubio, C.P., Hernández-Ruiz, J., Martínez-Subiela, S., Tvarijonaviciute, A., Ceron, J.J., 2016. Spectrophotometric assays for total antioxidant capacity (TAC) in dog serum: an update. *BMC Vet. Res.* 12, 166. <https://doi.org/10.1186/s12917-016-0792-7>
- Schmidt, E.M. dos S., Fachiolli, D.F., de Oliveira, R.M., Almeida, F.A., Pariz, C.M., de Lima Meirelles, P.R., Costa, C., Tvarijonaviciute, A., Erel, O., Neselioglu,

- S., Ceron, J.J., Rubio, C.P., 2021. Changes in Serum Thiol-Disulphide Homeostasis in Sheep with Gastrointestinal Nematodes. *Animals* 11, 2856. <https://doi.org/10.3390/ani11102856>
- Sihag, M.K., Patel, A., Kumar, V., 2021. Cottonseed (*Gossypium hirsutum*), in: Tanwar, B., Goyal, A. (Eds.), *Oilseeds: Health Attributes and Food Applications*. Springer, Singapore, pp. 73–92. [https://doi.org/10.1007/978-981-15-4194-0\\_3](https://doi.org/10.1007/978-981-15-4194-0_3)
- Starling, R.Z.C., Almeida, F.A. de, Viana, M.V.G., Castilhos, A.M. de, Amarante, A.F.T. do, 2019. Losses caused by gastrointestinal nematode infections in Dorper lambs under two nutritional status. *Braz. J. Vet. Parasitol.* 28, 652–660. <https://doi.org/10.1590/s1984-29612019084>
- Tardivo, R., 2021. Performance and beginning of semi-confined rubber puberty supplemented with cotton core in integrated agricultural production systems. Dissertation (Master's degree). São Paulo State University (UNESP), School of Veterinary Medicine and Animal Science (FMVZ), Botucatu, 62 pp.
- Teye, G.A., Ansah, T., Adjei, B.G.K., 2010. Effect of whole cottonseed supplementation on worm load in Djallonke sheep in the Tamale Metropolis; A case study in the Saatingli and Zaagyuli communities. *Development Spectrum* 3, 83–90.
- Tvarijonaviciute, A., Tecles, F., Caldin, M., Tasca, S., Cerón, J., 2012. Validation of spectrophotometric assays for serum paraoxonase type-1 measurement in dogs. *Am. J. Vet. Res.* 73, 34–41. <https://doi.org/10.2460/ajvr.73.1.34>
- Ueno, H., Gonçalves, P.C., 1998. *Manual para diagnóstico das helmintoses de ruminantes*, 4th ed. Japan International Cooperation Agency, Tokyo, Japan.
- Wilmsen, M.O., Silva, B.F., Bassetto, C.C., Amarante, A.F.T. do, 2014. Gastrointestinal nematode infections in sheep raised in Botucatu, state of São Paulo, Brazil. *Braz. J. Vet. Parasitol.* 23, 348–354. <https://doi.org/10.1590/S1984-29612014058>

- Witko-Sarsat, V., Friedlander, M., Capeillère-Blandin, C., Nguyen-Khoa, T., Nguyen, A.T., Zingraff, J., Jungers, P., Descamps-Latscha, B., 1996. Advanced oxidation protein products as a novel marker of oxidative stress in uremia. *Kidney Int.* 49, 1304–1313. <https://doi.org/10.1038/ki.1996.186>
- Yılmaz, K., Çakırca, G., Erel, Ö., 2021. Impaired thiol/disulphide homoeostasis in children with steroid-sensitive nephrotic syndrome. *Int. J. Clin. Pract.* 75. <https://doi.org/10.1111/ijcp.13794>
- Zajac, A.M., 2006. Gastrointestinal Nematodes of Small Ruminants: Life Cycle, Anthelmintics, and Diagnosis. *Vet. Clin. North Am. Food Anim. Pract.* 22, 529–541. <https://doi.org/10.1016/j.cvfa.2006.07.006>