

RESSALVA

Atendendo solicitação do(a) autor(a), o texto completo desta dissertação será disponibilizado somente a partir de 02/06/2023.

UNIVERSIDADE ESTADUAL PAULISTA “JULIO DE MESQUITA FILHO”

FACULDADE DE MEDICINA VETERINÁRIA E ZOOTECNIA

CAMPUS DE BOTUCATU

POTENCIAL DE INIBIÇÃO DA PROLIFERAÇÃO DE LINFÓCITOS IN VITRO PELAS
CELULAS TRONCO MESENQUIMAIS CANINAS DERIVADAS DO TECIDO ADIPOSEO

DIEGO PETROCINO CAETANO

Botucatu – SP

2022

UNIVERSIDADE ESTADUAL PAULISTA “JULIO DE MESQUITA FILHO”

FACULDADE DE MEDICINA VETERINÁRIA E ZOOTECNIA

CAMPUS DE BOTUCATU

POTENCIAL DE INIBIÇÃO DA PROLIFERAÇÃO DE LINFÓCITOS IN VITRO
PELAS CELULAS TRONCO MESENQUIMAIS CANINAS DERIVADAS DO
TECIDO ADIPOSE

DIEGO PETROCINO CAETANO

Dissertação apresentada junto ao Programa de Pós-Graduação em Medicina
Veterinária para obtenção do título de Mestre

Orientador: Prof. Dr. Rogério Martins Amorim

Botucatu – SP

2022

FICHA CATALOGRÁFICA ELABORADA PELA SEÇÃO TÊC. AQUIS. TRATAMENTO DA INFORM.
DIVISÃO TÉCNICA DE BIBLIOTECA E DOCUMENTAÇÃO - CAMPUS DE BOTUCATU - UNESP
BIBLIOTECÁRIA RESPONSÁVEL: ROSEMEIRE APARECIDA VICENTE-CRB 8/5651

Caetano, Diego Petrocino.

Potencial de inibição da proliferação de linfócitos in vitro pelas células tronco mesenquimais caninas derivadas do tecido adiposo / Diego Petrocino Caetano. - Botucatu, 2022

Dissertação (mestrado) - Universidade Estadual Paulista "Júlio de Mesquita Filho", Faculdade de Medicina Veterinária e Zootecnia

Orientador: Rogério Martins Amorim

Capes: 50501003

1. Cães. 2. Células-tronco mesenquimais. 3. Doenças autoimunes. 4. Regeneração. 5. linfócitos. 6. Medicina regenerativa.

Palavras-chave: Autoimune; Pbmnc; Reação mista de leucócitos; Regeneração.

Título: POTENCIAL DE INIBIÇÃO DA PROLIFERAÇÃO DE LINFÓCITOS IN VITRO PELAS CELULAS TRONCO MESENQUIMAIS CANINAS DE TECIDO ADIPOSEO

COMISSÃO EXAMINADORA

Prof. Dr. Rogério Martins Amorim
Presidente e Orientador
Departamento de Clínica Veterinária
FMVZ – UNESP – Botucatu, São Paulo.

Prof. Dra. Fernanda da Cruz Landim
Membro da Banca
Departamento de Cirurgia Veterinária e Reprodução Animal
FMVZ – UNESP – Botucatu, São Paulo.

Prof. Dra Márjorie de Assim Golim
Membro da Banca
Núcleo de Pesquisa do Hospital das Clínicas
FMB – UNESP – Botucatu, São Paulo.

Data da defesa: 04 de dezembro de 2022

AGRADECIMENTOS

Agradeço primeiramente à minha família: minha esposa Tatiane que abraçou minha ideia maluca de largar emprego e mudar de cidade, em busca de um sonho meu; minha mãe Eliana, minha irmã Fernanda, que mesmo de longe, sempre estiveram apoiando, perguntando, e incentivando sobre todos os processos em que me envolvi.

Ao professor e orientador Rogério, que sequer me conhecia antes de começarmos a conversar sobre o processo seletivo do mestrado, por toda paciência e orientação tanto técnica quanto burocrática.

A todos os amigos da UNESP que acompanharam essa jornada; o Vinícius, que me acompanhou em vários contratempos no laboratório; professora Márjorie e Aline do laboratório de citometria, por todo o conhecimento e acompanhamento de leitura das amostras.

E a todos os familiares e amigos que acompanharam e incentivaram o andamento do mestrado, desde o processo seletivo.

O presente trabalho foi realizado com apoio da Coordenação de Aperfeiçoamento de Pessoal de Nível Superior - Brasil (CAPES) - Código de Financiamento 001.

LISTA DE FIGURAS

Figura 1: Esquema da ação das MSCs sobre linfócitos. Quando há contato direto, a inibição da proliferação das células T se dá por ligações de proteínas de membrana, além de secreção de citocinas e fatores solúveis; quando não há contato direto, por exemplo por separação física por membranas transwell, a inibição se torna mais limitada, ocorrendo somente pelas citocinas e fatores solúveis.....	11
Figura 2: Teste de MLR. Linfócitos de cães saudáveis foram plaqueados com AT-MSCs e PMSCs provenientes de banco de células já caracterizadas, em situações com e sem contato direto.....	18
Figura 4: Citometria de fluxo PBMCs em co-cultura com uma das linhagens de MSCs, demonstrando fases do ciclo celular de linfócitos T mantidos em cultura. Nota-se uma redução da Fase S (proliferativa) de linfócitos quando em co-cultura com MSCs. Todas as culturas continham o mitogênico Concanavalina A. Análise feita em citômetro de fluxo.....	21
Figura 5: Fases do ciclo celular comparando as co-culturas com PBMCs e MSCs em contato direto e indireto. Os gráficos apresentam proporções normalizadas em relação à cultura sem MSCs, ou seja, somente PBMCs e ConA. A: Debris + apoptose celular; B: Fase S (proliferativa) do ciclo celular; C: Fase G0/G1; D: Fase G2/M. * indica diferença estatística ($p < 0.05$).....	21
Figura 6: Médias e desvio padrão de contagem de linfócitos por citometria de fluxo, normalizada com base na contagem de cultura de PBMCs. Todas as contagens são referentes a culturas com ConA.....	22
Figura 3: Co-culturas de PBMCs. Nas figuras c e d é possível observar as MSCs aderidas. a: PBMCs; b: PBMCs + ConA; c: PBMCs: ConA + MSCs; d: PBMCs + ConA + MSCs + transwell.....	22

LISTA DE ABREVIACÕES

APC = Células Apresentadoras De Antígenos
AT-MSC = Células Tronco Mesenquimais de Tecido Adiposo
BDNF = Fator Neurotrófico Derivado do Cérebro
Breg = Célula B Reguladora
BTLA = Atenuador de Linfócitos B e T
CCL2 = Quimiocina De Motivo C-C Ligante 2
ConA = Concanavalina A
CTLA-4 = Receptores Antígeno 4 Associado ao Linfócito T Citotóxico
DC = Célula Dendrítica
EAE = Encefalomielite Autoimune Experimental
EM = Esclerose Múltipla
EPHB2 = Produtor De Eritropoietina B 2
EPHB4 = Produtor De Eritropoietina B 4
Fas = Proteína Transmembrânica Indutora de Apoptose
FasL = Ligante de Proteína Transmembrânica Indutora de Apoptose
HGF = Fator de Crescimento de Hepatócitos
HLA-G = Antígeno De Locus De Histocompatibilidade-g
HO-1 = Heme Oxigenase 1 (HO-1)
IDO = Indoleamina 2,3-dioxigenase
IFN- γ = Interferon-Gama
IL-1 = Interleucina 1
IL-10 = Interleucina 10
IL-17 = Interleucina 17
IL-1 β = Interleucina-1 Beta
IL-2 = Interleucina 2
IL-33 = Interleucina 33
IL-6 = Interleucina 6
IL-7 = Interleucina 7
IL1RA = Antagonista Do Receptor De Interleucina-1
ILC = Célula Linfoide Inata
LAG3 = Gene de Ativação de Linfócitos 3
MDSC = Célula Mieloide Supressora Derivadas
MEN = Meningoencefalite Granulomatosa

MHC = Complexo Principal de Histocompatibilidade
MLR = Reação Mista de Leucócitos
MSC = Mesenchymal Stem Cells - Células Tronco Mesenquimais
MUO = Meningoencefalites de Origem Desconhecida
NGF = Fator De Crescimento Nervoso
NK = Natural Killer
NKT = Célula T Natural Killer
NO = Óxido Nítrico
NT-3 = Neurotrofina 3
PBMC = Células Mononucleares de Sangue Periférico
PD-1 = Proteína de Morte Celular Programada 1
PD-L1 = Ligante 1 de Proteína de Morte Celular Programada 1
PD-L2 = Ligante 2 de Proteína de Morte Celular Programada 1
PGE2 = Prostaglandina E2
TCR = Receptores de Células T
TGF- β = Fator de Crescimento Transformador β
Th = Célula T Helper
Th1 = Célula T Auxiliar Tipo 1
Th17 = Célula T Auxiliar Tipo 17
Th2 = Célula T Auxiliar Tipo 2
TIM-1 = Domínios de Mucina e Imunoglobulina de Célula T 1
TIM-2 = Domínios de Mucina e Imunoglobulina de Célula T 2
TIM-3 = Domínios de Mucina e Imunoglobulina de Célula T 3
TIM-4 = Domínios de Mucina e Imunoglobulina de Célula T 4
TNF- α = Fator De Necrose Tumoral Alfa
Treg = Células T Reguladoras
TSG6 = Gene 6 Estimulado Pelo Fator De Necrose Tumoral
VEGF = Fator De Crescimento Endotelial Vascular
 μg = Microgramas
 μL = Microlitros

RESUMO

A terapia com células-tronco mesenquimais (MSCs) tem se demonstrado uma biotecnologia promissora em medicina regenerativa, principalmente pelo seu potencial imunomodulador. Sua ação ocorre por diversos mecanismos, como secreção de fatores solúveis e interação direta entre células. Dentre as diversas interações, a redução da proliferação de linfócitos T é fundamental para controle de doenças autoimunes em geral. Dentre as formas de avaliar o potencial imunomodulador das MSCs *in vitro*, destaca-se o teste de reação mista de leucócitos (MLR), que avalia a inibição da proliferação de leucócitos, particularmente linfócitos. O objetivo deste estudo foi avaliar *in vitro* o ciclo celular e o potencial de inibição da proliferação de linfócitos estimulados com concanavalina A, quando em co-cultura (em contato direto e indireto) com MSCs derivadas do tecido adiposo (AT-MSCs) canino e células monocleares de sangue periférico (PBMC). Para tal, foram realizados testes MLR utilizando-se PBMCs provenientes de cães doadores clinicamente saudáveis obtidas por meio de gradiente de separação com Histopaque 1077, em co-cultura com AT-MSCs caninas (n=4). Foram realizadas culturas em duplicatas de PBMCs (10^6 células por poço), PBMCs estimuladas com concanavalina A (10^6 células + 5ug/mL de ConA), PBMCs estimuladas com concanavalina A em co-cultura direta com AT-MSCs (10^6 PBMCs + 5ug/mL de ConA + 2×10^5 AT-MSCs), e PBMCs estimuladas com concanavalina A em co-cultura indireta (membranas transwell) com AT-MSCs (10^6 PBMCs + 5ug/mL de ConA + 2×10^5 AT-MSCs). As placas foram mantidas em cultivo por 96h, sendo adicionado BrdU nas últimas 24h. Em seguida, as amostras dos cultivos foram incubadas com anticorpos anti-CD3 canino, anti-BrdU e 7-AAD para avaliação por citometria de fluxo. As AT-MSCs caninas em co-cultura com as PBMCs induziram a redução da fase proliferativa dos linfócitos CD3+, possivelmente deslocando-os para a fase G0/G1 do ciclo celular. A redução da fase proliferativa dos linfócitos CD3+ foi evidenciada tanto nas co-culturas de AT-MSCs e PBMCs com contato direto quanto indireto, indicando que os mecanismos de ação também dependem de

mediadores químicos secretados no meio. Apesar de não observamos diferença estatística, houve tendência de inibição da proliferação de linfócitos CD3+ estimulados com concanavalina A quando em contato direto e indireto com as AT-MSCs. Nossos resultados indicam que as AT-MSCs caninas apresentam potencial imunomodulador. Novos estudos devem ser realizados para elucidar os mecanismos de ação imunomoduladores das AT-MSCs caninas sobre linfócitos.

Palavras-chave: PBMC, reação mista de leucócitos, autoimune, linfócito CD3+

ABSTRACT

Mesenchymal stem cells (MSCs) therapy has been shown to be a promising biotechnology in regenerative medicine, mainly due to its immunomodulatory potential. Its action occurs through several mechanisms, such as secretion of soluble factors and direct interaction between cells. Among the various interactions, the reduction of T lymphocyte proliferation is fundamental for the control of autoimmune diseases in general. Among the ways to evaluate the immunomodulatory potential of MSCs in vitro, the mixed leukocyte reaction test (MLR) is one of the most important, which evaluates the inhibition of leukocyte proliferation, particularly lymphocytes. The aim of this study was to evaluate in vitro the cell cycle and proliferation inhibition potential of lymphocytes stimulated with concanavalin A, when in co-culture (in direct and indirect contact) with canine adipose tissue-derived MSCs (AT-MSCs) and peripheral blood mononuclear cells (PBMC). For this purpose, MLR tests were performed using PBMCs from clinically healthy donor dogs obtained through separation gradient with Histopaque 1077, in co-culture with canine AT-MSCs (n=4). Cultures were performed in duplicates of PBMCs (10^6 cells per well), PBMCs stimulated with concanavalin A (10^6 cells + 5ug/ml ConA), PBMCs stimulated with concanavalin A in direct co-culture with AT-MSCs (10^6 PBMCs + 5ug/ mL of ConA + 2×10^5 AT-MSCs), and PBMCs stimulated with concanavalin A in indirect co-culture (transwell membranes) with AT-MSCs (10^6 PBMCs + 5ug/mL of ConA + 2×10^5 AT-MSCs). The plates were kept in culture for 96h, with BrdU being added in the last 24h. Then, culture samples were incubated with anti-CD3 canine, anti-BrdU and 7-AAD antibodies for evaluation in flow cytometry. Canine AT-MSCs in co-culture with PBMCs induced a reduction in the proliferative phase of CD3+ lymphocytes, possibly shifting them to the G0/G1 phase of the cell cycle. The reduction in the proliferative phase of CD3+ lymphocytes was evidenced both in co-cultures of AT-MSCs and PBMCs with direct and indirect contact, indicating that the mechanisms of action also depend on chemical mediators secreted in the medium. Although we did not observe any statistical difference, there was a tendency to inhibit the proliferation of CD3+ lymphocytes stimulated with

concanavalin A when in direct and indirect contact with AT-MSCs. Our results indicate that canine AT-MSCs have immunomodulatory potential. New studies must be carried out to elucidate the immunomodulatory mechanisms of action of canine AT-MSCs on lymphocytes.

Keywords: PBMC, mixed lymphocyte reaction, autoimmune, CD3+ lymphocyte

SUMÁRIO

1	INTRODUÇÃO E JUSTIFICATIVA.....	1
2	REVISÃO DE LITERATURA.....	2
2.1	Papel de Linfócitos T em Doenças Autoimunes.....	3
2.2	Citocinas e Fatores Solúveis Relevantes.....	6
2.3	Potencial Imunomodulador de Células Tronco Mesenquimais (MSCs).....	10
2.4	Potencial Terapêutico das MSCs.....	12
2.5	Relevância Clínica.....	13
2.6	Teste de Reação Mista de Leucócitos – Mixed Leukocyte Reaction (MLR).....	15
3	OBJETIVOS.....	16
4	MATERIAIS E MÉTODOS.....	16
4.1	Amostras de AT-MSCs.....	17
4.2	Amostras de PBMCs.....	17
4.3	Teste de Reação Mista de Leucócitos (MLR).....	18
4.4	Análise do Ciclo Celular.....	19
4.5	Análise Estatística.....	19
5	RESULTADOS.....	21
5.1	AT-MSCs promovem alteração do ciclo celular dos linfócitos T CD3+.....	22
5.2	A ação das AT-MSCs não dependeu do contato direto com os linfócitos T CD3.....	22
5.3	Contagem de linfócitos T CD3 ativados.....	22
6	DISCUSSÃO.....	24
7	CONCLUSÃO.....	26
8	REFERÊNCIAS.....	27
9	ANEXOS.....	40
9.1	TABELA DE LEITURAS DE BRDU-APC OBTIDAS EM CITOMETRIA DE FLUXO.....	40
9.2	ANEXO II – ATESTADO DE APROVAÇÃO NA CEUA – COMITÊ DE ÉTICA NO USO DE ANIMAIS.....	42

8 REFERÊNCIAS

ABBAS, A. K.; LICHTMAN, L. H.; PILLAI, S. Immunologic tolerance and autoimmunity. Em: **Cellular and Molecular Immunology**. Philadelphia: Elsevier Saunders, 2012. p. 326.

AL JUMAH, M. A.; ABUMAREE, M. H. The Immunomodulatory and Neuroprotective Effects of Mesenchymal Stem Cells (MSCs) in Experimental Autoimmune Encephalomyelitis (EAE): A Model of Multiple Sclerosis (MS). **International Journal of Molecular Sciences**, v. 13, n. 7, p. 9298–9331, jul. 2012.

AMORIM, R. M. et al. Placenta-derived multipotent mesenchymal stromal cells: a promising potential cell-based therapy for canine inflammatory brain disease. **Stem Cell Research & Therapy**, v. 11, n. 1, p. 304, 22 jul. 2020.

ANAYA, J.-M. Common mechanisms of autoimmune diseases (the autoimmune tautology). **Autoimmunity Reviews**, v. 11, n. 11, p. 781–784, 1 set. 2012.

ANDERSON, P. et al. Allogeneic Adipose-Derived Mesenchymal Stromal Cells Ameliorate Experimental Autoimmune Encephalomyelitis by Regulating Self-Reactive T Cell Responses and Dendritic Cell Function. **Stem Cells International**, v. 2017, p. e2389753, 30 jan. 2017.

BACH, J. F. Regulatory T cells under scrutiny. **Nature Reviews Immunology**, v. 3, n. 3, p. 189–198, 2003.

BERNARDO, M. E.; FIBBE, W. E. Mesenchymal stromal cells: sensors and switchers of inflammation. **Cell Stem Cell**, v. 13, n. 4, p. 392–402, 3 out. 2013.

BIKKER, A. et al. Interleukin-7: A key mediator in T cell-driven autoimmunity, inflammation, and tissue destruction. **Current Pharmaceutical Design**, v. 18, n. 16, p. 2347–2356, 2012.

BOYMAN, O.; SPRENT, J. The role of interleukin-2 during homeostasis and activation of the immune system. **Nature Reviews Immunology**, v. 12, n. 3, p. 180–190, 2012.

BULATI, M. et al. The Immunomodulatory Properties of the Human Amnion-Derived Mesenchymal Stromal/Stem Cells Are Induced by INF- γ Produced by Activated Lymphomonocytes and Are Mediated by Cell-To-Cell Contact and Soluble Factors. **Frontiers in Immunology**, v. 11, 2020.

CARRADE, D. D.; BORJESSON, D. L. Immunomodulation by Mesenchymal Stem Cells in Veterinary Species. **Comparative Medicine**, v. 63, n. 3, p. 207–217, jun. 2013.

CHOI, S.; SCHWARTZ, R. H. Molecular mechanisms for adaptive tolerance and other T cell anergy models. **Seminars in Immunology**, Molecular Mechanisms Supporting Peripheral T cell Tolerance: Potential Therapeutic Approaches to Autoimmunity and Allograft Rejection. v. 19, n. 3, p. 140–152, 1 jun. 2007.

CIPRIANI, P. et al. Stem cells in autoimmune diseases: Implications for pathogenesis and future trends in therapy. **Autoimmunity Reviews**, v. 12, n. 7, p. 709–716, maio 2013.

CLARK, K. C.; AMADOR, A.; WANG, A. Convergence of human and veterinary medicine: leveraging canine naturally occurring neurological disorders to develop regenerative treatments. **Neural Regeneration Research**, v. 18, n. 3, p. 541–542, mar. 2023.

CROP, M. J. et al. Human Adipose Tissue-Derived Mesenchymal Stem Cells Induce Explosive T-Cell Proliferation. **Stem Cells and Development**, v. 19, n. 12, p. 1843–1853, dez. 2010.

DAI, C. et al. Hepatocyte growth factor preserves beta cell mass and mitigates hyperglycemia in streptozotocin-induced diabetic mice. **The Journal of Biological Chemistry**, v. 278, n. 29, p. 27080–27087, 18 jul. 2003.

DE WOLF, C.; VAN DE BOVENKAMP, M.; HOEFNAGEL, M. Regulatory perspective on in vitro potency assays for human mesenchymal stromal cells used in immunotherapy. **Cytotherapy**, v. 19, n. 7, p. 784–797, jul. 2017.

DELAROSA, O.; DALEMANS, W.; LOMBARDO, E. Toll-like receptors as modulators of mesenchymal stem cells. **Frontiers in Immunology**, v. 3, p. 182, 2012.

DENDROU, C. A.; FUGGER, L.; FRIESE, M. A. Immunopathology of multiple sclerosis. **Nature Reviews. Immunology**, v. 15, n. 9, p. 545–558, 15 set. 2015.

DI IANNI, M. et al. Mesenchymal cells recruit and regulate T regulatory cells. **Experimental Hematology**, v. 36, n. 3, p. 309–318, mar. 2008.

DI NICOLA, M. et al. Human bone marrow stromal cells suppress T-lymphocyte proliferation induced by cellular or nonspecific mitogenic stimuli. **Blood**, v. 99, n. 10, p. 3838–3843, 15 maio 2002.

DOMBROWSKI, Y. et al. Regulatory T cells promote myelin regeneration in the central nervous system. **Nature Neuroscience**, v. 20, n. 5, p. 674–680, maio 2017.

DOMINICI, M. et al. Minimal criteria for defining multipotent mesenchymal stromal cells. The International Society for Cellular Therapy position statement. **Cytotherapy**, v. 8, n. 4, p. 315–317, 2006.

DU, W. et al. Tim-3 as a target for cancer immunotherapy and mechanisms of action. **International Journal of Molecular Sciences**, v. 18, n. 3, 2017.

ERKERS, T. et al. Decidual stromal cells promote regulatory T cells and suppress alloreactivity in a cell contact-dependent manner. **Stem Cells and Development**, v. 22, n. 19, p. 2596–2605, 1 out. 2013.

FRIESE, M. A.; SCHATTLING, B.; FUGGER, L. Mechanisms of neurodegeneration and axonal dysfunction in multiple sclerosis. **Nature Reviews Neurology**, v. 10, n. 4, p. 225–238, abr. 2014.

FURTADO, G. C. et al. Swift Entry of Myelin-Specific T Lymphocytes into the Central Nervous System in Spontaneous Autoimmune Encephalomyelitis. **The Journal of Immunology**, v. 181, n. 7, p. 4648–4655, 1 out. 2008.

GARCÍA-GONZÁLEZ, P. et al. Tolerogenic dendritic cells for reprogramming of lymphocyte responses in autoimmune diseases. **Autoimmunity Reviews**, v. 15, n. 11, p. 1071–1080, nov. 2016.

GLENNIE, S. et al. Bone marrow mesenchymal stem cells induce division arrest anergy of activated T cells. **Blood**, v. 105, n. 7, p. 2821–2827, 1 abr. 2005.

GOLDBERG, M. V.; DRAKE, C. G. LAG-3 in Cancer Immunotherapy. Em: DRANOFF, G. (Ed.). **Cancer Immunology and Immunotherapy**. Current Topics in Microbiology and Immunology. Berlin, Heidelberg: Springer, 2011. p. 269–278.

GONZALEZ-PUJANA, A. et al. Multifunctional biomimetic hydrogel systems to boost the immunomodulatory potential of mesenchymal stromal cells. **Biomaterials**, v. 257, p. 120266, out. 2020.

GREER, K. A. et al. Necrotizing meningoencephalitis of Pug dogs associates with dog leukocyte antigen class II and resembles acute variant forms of multiple sclerosis. **Tissue Antigens**, v. 76, n. 2, p. 110–118, ago. 2010.

GU, Y. et al. Different roles of PD-L1 and FasL in immunomodulation mediated by human placenta-derived mesenchymal stem cells. **Human Immunology**, v. 74, n. 3, p. 267–276, mar. 2013.

HOFFMAN, A. M.; DOW, S. W. Concise Review: Stem Cell Trials Using Companion Animal Disease Models. **Stem Cells (Dayton, Ohio)**, v. 34, n. 7, p. 1709–1729, jul. 2016.

HSU, W.-T. et al. Prostaglandin E2 potentiates mesenchymal stem cell-induced IL-10+IFN- γ +CD4⁺ regulatory T cells to control transplant arteriosclerosis. **Journal of Immunology (Baltimore, Md.: 1950)**, v. 190, n. 5, p. 2372–2380, 1 mar. 2013.

JIANG, W.; XU, J. Immune modulation by mesenchymal stem cells. **Cell Proliferation**, v. 53, n. 1, p. e12712, 2020.

KAMRADT, T.; MITCHISON, N. A. Tolerance and Autoimmunity. **New England Journal of Medicine**, v. 344, n. 9, p. 655–664, 1 mar. 2001.

KANG, J. W. et al. Soluble factors-mediated immunomodulatory effects of canine adipose tissue-derived mesenchymal stem cells. **Stem Cells and Development**, v. 17, n. 4, p. 681–693, ago. 2008.

KEATING, A. Mesenchymal stromal cells: new directions. **Cell Stem Cell**, v. 10, n. 6, p. 709–716, 14 jun. 2012.

KHAN, U. et al. Immunotherapy-associated autoimmune hemolytic anemia. **Journal for ImmunoTherapy of Cancer**, v. 5, n. 1, 2017.

KHAN, U.; GHAZANFAR, H. Chapter Three - T Lymphocytes and Autoimmunity. Em: GALLUZZI, L.; RUDQVIST, N.-P. (Eds.). **International Review of Cell and Molecular Biology**. Biology of T Cells - Part A. [s.l.] Academic Press, 2018. v. 341p. 125–168.

KODA, M. et al. Brain-Derived Neurotrophic Factor Suppresses Delayed Apoptosis of Oligodendrocytes after Spinal Cord Injury in Rats. **Journal of Neurotrauma**, v. 19, n. 6, p. 777–785, jun. 2002.

KOL, A. et al. Companion animals: Translational scientist's new best friends. **Science Translational Medicine**, v. 7, n. 308, p. 308ps21-308ps21, 7 out. 2015.

KOLACZKOWSKA, E.; KUBES, P. Neutrophil recruitment and function in health and inflammation. **Nature Reviews. Immunology**, v. 13, n. 3, p. 159–175, mar. 2013.

KORN, T. et al. IL-17 and Th17 cells. **Annual Review of Immunology**, v. 27, p. 485–517, 2009.

KRAMPERA, M. et al. Bone marrow mesenchymal stem cells inhibit the response of naive and memory antigen-specific T cells to their cognate peptide. **Blood**, v. 101, n. 9, p. 3722–3729, 1 maio 2003.

KRAMPERA, M. et al. Regenerative and immunomodulatory potential of mesenchymal stem cells. **Current Opinion in Pharmacology**, v. 6, n. 4, p. 435–441, ago. 2006.

KUEHN, H. S. et al. FAS Haploinsufficiency Is a Common Disease Mechanism in the Human Autoimmune Lymphoproliferative Syndrome. **The Journal of Immunology**, v. 186, n. 10, p. 6035–6043, 15 maio 2011.

KYURKCHIEV, D. et al. Secretion of immunoregulatory cytokines by mesenchymal stem cells. **World Journal of Stem Cells**, v. 6, n. 5, p. 552–570, 26 nov. 2014.

LAFFERTY, K.; CUNNINGHAM, A. A New Analysis of Allogeneic Interactions. **Australian Journal of Experimental Biology and Medical Science**, v. 53, n. 1, p. 27–42, 1975.

LE BLANC, K. et al. Mesenchymal stem cells inhibit and stimulate mixed lymphocyte cultures and mitogenic responses independently of the major histocompatibility complex. **Scandinavian Journal of Immunology**, v. 57, n. 1, p. 11–20, jan. 2003.

LEACH, D. R.; KRUMMEL, M. F.; ALLISON, J. P. Enhancement of antitumor immunity by CTLA-4 blockade. **Science**, v. 271, n. 5256, p. 1734–1736, 1996.

LEJKOWSKA, R. et al. Preclinical Evaluation of Long-Term Neuroprotective Effects of BDNF-Engineered Mesenchymal Stromal Cells as Intravitreal Therapy for Chronic Retinal Degeneration in Rd6 Mutant Mice. **International Journal of Molecular Sciences**, v. 20, n. 3, p. 777, jan. 2019.

LI, S. et al. A naturally occurring CD8(+)CD122(+) T-cell subset as a memory-like Treg family. **Cellular & Molecular Immunology**, v. 11, n. 4, p. 326–331, jul. 2014.

LIAO, W.; LIN, J.-X.; LEONARD, W. J. Interleukin-2 at the Crossroads of Effector Responses, Tolerance, and Immunotherapy. **Immunity**, v. 38, n. 1, p. 13–25, 24 jan. 2013.

LIN, R. et al. Bone marrow-derived mesenchymal stem cells favor the immunosuppressive T cells skewing in a *Helicobacter pylori* model of gastric cancer. **Stem Cells and Development**, v. 22, n. 21, p. 2836–2848, 1 nov. 2013.

LINKER, R. A. et al. Functional role of brain-derived neurotrophic factor in neuroprotective autoimmunity: therapeutic implications in a model of multiple sclerosis. **Brain**, v. 133, n. 8, p. 2248–2263, 1 ago. 2010.

MAETA, N. et al. Comparative analysis of canine mesenchymal stem cells and bone marrow-derived mononuclear cells. **Veterinary World**, v. 14, n. 4, p. 1028–1037, abr. 2021.

MAHMOOD, A.; LU, D.; CHOPP, M. Intravenous Administration of Marrow Stromal Cells (MSCs) Increases the Expression of Growth Factors in Rat Brain after Traumatic Brain Injury. **Journal of Neurotrauma**, v. 21, n. 1, p. 33–39, jan. 2004.

MAKI, C. B. et al. Intra-articular Administration of Allogeneic Adipose Derived MSCs Reduces Pain and Lameness in Dogs With Hip Osteoarthritis: A Double Blinded, Randomized, Placebo Controlled Pilot Study. **Frontiers in Veterinary Science**, v. 7, 2020.

MALTMAN, D. J.; HARDY, S. A.; PRZYBORSKI, S. A. Role of mesenchymal stem cells in neurogenesis and nervous system repair. **Neurochemistry International**, v. 59, n. 3, p. 347–356, set. 2011.

MANCHES, O. et al. HIV-activated human plasmacytoid DCs induce Tregs through an indoleamine 2,3-dioxygenase–dependent mechanism. **The Journal of Clinical Investigation**, v. 118, n. 10, p. 3431–3439, 1 out. 2008.

MANSOOR, S. R.; ZABIHI, E.; GHASEMI-KASMAN, M. The potential use of mesenchymal stem cells for the treatment of multiple sclerosis. **Life Sciences**, v. 235, p. 116830, 15 out. 2019.

MARKUS, A.; PATEL, T. D.; SNIDER, W. D. Neurotrophic factors and axonal growth. **Current Opinion in Neurobiology**, v. 12, n. 5, p. 523–531, out. 2002.

MAURI, C.; BOSMA, A. Immune regulatory function of B cells. **Annual Review of Immunology**, v. 30, p. 221–241, 2012.

MEISEL, R. et al. Human bone marrow stromal cells inhibit allogeneic T-cell responses by indoleamine 2,3-dioxygenase-mediated tryptophan degradation. **Blood**, v. 103, n. 12, p. 4619–4621, 15 jun. 2004.

MELIEF, S. M. et al. Adipose tissue-derived multipotent stromal cells have a higher immunomodulatory capacity than their bone marrow-derived counterparts. **Stem Cells Translational Medicine**, v. 2, n. 6, p. 455–463, jun. 2013.

MELLOR, A. L.; MUNN, D. H. IDO expression by dendritic cells: tolerance and tryptophan catabolism. **Nature Reviews. Immunology**, v. 4, n. 10, p. 762–774, out. 2004.

MIYARA, M.; ITO, Y.; SAKAGUCHI, S. TREG-cell therapies for autoimmune rheumatic diseases. **Nature Reviews. Rheumatology**, v. 10, n. 9, p. 543–551, set. 2014.

MIZUNO, K. et al. Hepatocyte growth factor stimulates growth of hematopoietic progenitor cells. **Biochemical and Biophysical Research Communications**, v. 194, n. 1, p. 178–186, 15 jul. 1993.

MIZUNO, S.; MATSUMOTO, K.; NAKAMURA, T. HGF as a renoprotective and anti-fibrotic regulator in chronic renal disease. **Frontiers in Bioscience: A Journal and Virtual Library**, v. 13, p. 7072–7086, 1 maio 2008.

MIZUNO, S.; NAKAMURA, T. Prevention of neutrophil extravasation by hepatocyte growth factor leads to attenuations of tubular apoptosis and renal dysfunction in mouse ischemic kidneys. **The American Journal of Pathology**, v. 166, n. 6, p. 1895–1905, jun. 2005.

MUNN, D. H. et al. GCN2 Kinase in T Cells Mediates Proliferative Arrest and Anergy Induction in Response to Indoleamine 2,3-Dioxygenase. **Immunity**, v. 22, n. 5, p. 633–642, 1 maio 2005.

MUNN, D. H.; MELLOR, A. L. Indoleamine 2,3 dioxygenase and metabolic control of immune responses. **Trends in Immunology**, v. 34, n. 3, p. 137–143, 1 mar. 2013.

NAJAR, M. et al. Immune-Related Antigens, Surface Molecules and Regulatory Factors in Human-Derived Mesenchymal Stromal Cells: The Expression and Impact of Inflammatory Priming. **Stem Cell Reviews and Reports**, v. 8, n. 4, p. 1188–1198, 1 dez. 2012.

NAJAR, M. et al. Mesenchymal stromal cells and immunomodulation: A gathering of regulatory immune cells. **Cytotherapy**, v. 18, n. 2, p. 160–171, fev. 2016.

NAKAMURA, T.; MIZUNO, S. The discovery of Hepatocyte Growth Factor (HGF) and its significance for cell biology, life sciences and clinical medicine. **Proceedings of the Japan Academy. Series B, Physical and Biological Sciences**, v. 86, n. 6, p. 588–610, 11 jun. 2010.

NAKANISHI, M.; ROSENBERG, D. W. Multifaceted roles of PGE2 in inflammation and cancer. **Seminars in Immunopathology**, v. 35, n. 2, p. 123–137, 1 mar. 2013.

NGUYEN, T. M. et al. EphB and Ephrin-B interactions mediate human mesenchymal stem cell suppression of activated T-cells. **Stem Cells and Development**, v. 22, n. 20, p. 2751–2764, 15 out. 2013.

NICOTRA, T. et al. Mesenchymal stem/stromal cell quality control: validation of mixed lymphocyte reaction assay using flow cytometry according to ICH Q2(R1). **Stem Cell Research & Therapy**, v. 11, n. 1, p. 426, 1 out. 2020.

ODINAK, M. M. et al. [Transplantation of mesenchymal stem cells in multiple sclerosis]. **Zhurnal nevrologii i psikiatrii imeni S.S Korsakova**, v. 111, n. 2 Pt 2, p. 72–76, 1 jan. 2011.

OHMICH, H.; MATSUMOTO, K.; NAKAMURA, T. In vivo mitogenic action of HGF on lung epithelial cells: pulmotrophic role in lung regeneration. **The American Journal of Physiology**, v. 270, n. 6 Pt 1, p. L1031-1039, jun. 1996.

PACHLER, K. et al. An In Vitro Potency Assay for Monitoring the Immunomodulatory Potential of Stromal Cell-Derived Extracellular Vesicles. **International Journal of Molecular Sciences**, v. 18, n. 7, p. 1413, 1 jul. 2017.

PARK, E.-S.; UCHIDA, K.; NAKAYAMA, H. Th1-, Th2-, and Th17-related cytokine and chemokine receptor mRNA and protein expression in the brain tissues, T cells, and macrophages of dogs with necrotizing and granulomatous meningoencephalitis. **Veterinary Pathology**, v. 50, n. 6, p. 1127–1134, nov. 2013.

PARR, A. M.; TATOR, C. H.; KEATING, A. Bone marrow-derived mesenchymal stromal cells for the repair of central nervous system injury. **Bone Marrow Transplantation**, v. 40, n. 7, p. 609–619, out. 2007.

PAUL, G.; ANISIMOV, S. V. The secretome of mesenchymal stem cells: potential implications for neuroregeneration. **Biochimie**, v. 95, n. 12, p. 2246–2256, dez. 2013.

PÉREZ-MERINO, E. M. et al. Safety and efficacy of allogeneic adipose tissue-derived mesenchymal stem cells for treatment of dogs with inflammatory bowel disease: Endoscopic and histological outcomes. **Veterinary Journal (London, England: 1997)**, v. 206, n. 3, p. 391–397, dez. 2015.

PETCHDEE, S.; SOMPEEWONG, S. Intravenous administration of puppy deciduous teeth stem cells in degenerative valve disease. **Veterinary World**, v. 9, n. 12, p. 1429–1434, dez. 2016.

PICCIRILLO, C. A.; THORNTON, A. M. Cornerstone of peripheral tolerance: Naturally occurring CD4 +CD25+ regulatory T cells. **Trends in Immunology**, v. 25, n. 7, p. 374–380, 2004.

PLUMAS, J. et al. Mesenchymal stem cells induce apoptosis of activated T cells. **Leukemia**, v. 19, n. 9, p. 1597–1604, set. 2005.

RAMASAMY, R. et al. The immunosuppressive effects of human bone marrow-derived mesenchymal stem cells target T cell proliferation but not its effector function.

Cellular Immunology, v. 251, n. 2, p. 131–136, fev. 2008.

RONCAROLO, M. G. et al. Tr1 cells and the counter-regulation of immunity: Natural mechanisms and therapeutic applications.

Current Topics in Microbiology and Immunology, v. 380, p. 39–68, 2014.

SATO, K. et al. Nitric oxide plays a critical role in suppression of T-cell proliferation by mesenchymal stem cells.

Blood, v. 109, n. 1, p. 228–234, 19 set. 2006.

SCHMASSMANN, A. et al. Roles of hepatocyte growth factor and its receptor Met during gastric ulcer healing in rats.

Gastroenterology, v. 113, n. 6, p. 1858–1872, dez. 1997.

SCHORL, C.; SEDIVY, J. M. Analysis of cell cycle phases and progression in cultured mammalian cells.

Methods, Methods in Cell Cycle Research. v. 41, n. 2, p. 143–150, 1 fev. 2007.

SCHWARTZ, R. H. Historical Overview of Immunological Tolerance.

Cold Spring Harbor Perspectives in Biology, v. 4, n. 4, p. a006908, 4 jan. 2012.

SGRIGNOLI, M. R. et al. Reduction in the inflammatory markers CD4, IL-1, IL-6 and TNF α in dogs with keratoconjunctivitis sicca treated topically with mesenchymal stem cells.

Stem cell research, v. 39, p. 101525, 1 ago. 2019.

SOLCHAGA, L. A.; ZALE, E. A. Prostaglandin E2: a putative potency indicator of the immunosuppressive activity of human mesenchymal stem cells.

American Journal of Stem Cells, v. 1, n. 2, p. 138–145, 30 jun. 2012.

TAJIMA, H. et al. Tissue distribution of hepatocyte growth factor receptor and its exclusive down-regulation in a regenerating organ after injury.

Journal of Biochemistry, v. 111, n. 3, p. 401–406, mar. 1992.

TANAKA, T.; KISHIMOTO, T. Targeting interleukin-6: All the way to treat autoimmune and inflammatory diseases. **International Journal of Biological Sciences**, v. 8, n. 9, p. 1227–1236, 2012.

UCCELLI, A.; MORETTA, L.; PISTOIA, V. Mesenchymal stem cells in health and disease. **Nature Reviews. Immunology**, v. 8, n. 9, p. 726–736, set. 2008.

VAN VELTHOVEN, C. T. J. et al. Mesenchymal Stem Cell Transplantation Attenuates Brain Injury After Neonatal Stroke. **Stroke**, v. 44, n. 5, p. 1426–1432, maio 2013.

VIEIRA-POTTER, V. J. Inflammation and macrophage modulation in adipose tissues. **Cellular Microbiology**, v. 16, n. 10, p. 1484–1492, out. 2014.

VIGNALI, D. A. A.; COLLISON, L. W.; WORKMAN, C. J. How regulatory T cells work. **Nature Reviews. Immunology**, v. 8, n. 7, p. 523–532, jul. 2008.

VILLATORO, A. J. et al. Allogeneic adipose-derived mesenchymal stem cell therapy in dogs with refractory atopic dermatitis: clinical efficacy and safety. **The Veterinary Record**, v. 183, n. 21, p. 654, 1 dez. 2018.

VOLK, S. W.; THEORET, C. Translating stem cell therapies: the role of companion animals in regenerative medicine. **Wound repair and regeneration: official publication of the Wound Healing Society [and] the European Tissue Repair Society**, v. 21, n. 3, p. 382–394, maio 2013.

WANG, L.; ZHAO, Y.; SHI, S. Interplay between Mesenchymal Stem Cells and Lymphocytes: Implications for Immunotherapy and Tissue Regeneration. **Journal of Dental Research**, v. 91, n. 11, p. 1003–1010, 1 nov. 2012.

WANG, Y. et al. Plasticity of mesenchymal stem cells in immunomodulation: pathological and therapeutic implications. **Nature Immunology**, v. 15, n. 11, p. 1009–1016, nov. 2014.

WANG, Z. et al. Safety of neural stem cell transplantation in patients with severe traumatic brain injury. **Experimental and Therapeutic Medicine**, v. 13, n. 6, p. 3613–3618, 1 jun. 2017.

WATANABE, N. et al. BTLA is a lymphocyte inhibitory receptor with similarities to CTLA-4 and PD-1. **Nature Immunology**, v. 4, n. 7, p. 670–679, 2003.

YAMOUT, B. et al. Bone marrow mesenchymal stem cell transplantation in patients with multiple sclerosis: a pilot study. **Journal of Neuroimmunology**, v. 227, n. 1–2, p. 185–189, 8 out. 2010.

YANG, L. et al. Lack of TIM-3 immunoregulation in multiple sclerosis. **Journal of Immunology**, v. 180, n. 7, p. 4409–4414, 2008.

YEUNG, R. S. M. et al. Enhancing translational research in paediatric rheumatology through standardization. **Nature Reviews. Rheumatology**, v. 12, n. 11, p. 684–690, nov. 2016.

YOSHIDA, S. et al. Neutralization of hepatocyte growth factor leads to retarded cutaneous wound healing associated with decreased neovascularization and granulation tissue formation. **The Journal of Investigative Dermatology**, v. 120, n. 2, p. 335–343, fev. 2003.

ZHANG, J. et al. Human bone marrow stromal cell treatment improves neurological functional recovery in EAE mice. **Experimental Neurology**, v. 195, n. 1, p. 16–26, 1 set. 2005.

ZHANG, Q.; VIGNALI, D. A. A. Co-stimulatory and Co-inhibitory Pathways in Autoimmunity. **Immunity**, v. 44, n. 5, p. 1034–1051, 17 maio 2016.

ZHAO, Q.; CHEN, G. Role of IL-33 and its receptor in T cell-mediated autoimmune diseases. **BioMed Research International**, v. 2014, 2014.

ZEIRA, O. et al. Adult autologous mesenchymal stem cells for the treatment of suspected non-infectious inflammatory diseases of the canine central nervous system: safety, feasibility and preliminary clinical findings. **Journal of Neuroinflammation**, v. 12, n. 1, p. 181, 29 set. 2015.

ABSTRACT

Mesenchymal stem cells (MSCs) therapy has been shown to be a promising biotechnology in regenerative medicine, mainly due to its immunomodulatory potential. Its action occurs through several mechanisms, such as secretion of soluble factors and direct interaction between cells. Among the various interactions, the reduction of T lymphocyte proliferation is fundamental for the control of autoimmune diseases in general. Among the ways to evaluate the immunomodulatory potential of MSCs in vitro, the mixed leukocyte reaction test (MLR) is one of the most important, which evaluates the inhibition of leukocyte proliferation, particularly lymphocytes. The aim of this study was to evaluate in vitro the cell cycle and proliferation inhibition potential of lymphocytes stimulated with concanavalin A, when in co-culture (in direct and indirect contact) with canine adipose tissue-derived MSCs (AT-MSCs) and peripheral blood mononuclear cells (PBMC). For this purpose, MLR tests were performed using PBMCs from clinically healthy donor dogs obtained through separation gradient with Histopaque 1077, in co-culture with canine AT-MSCs (n=4). Cultures were performed in duplicates of PBMCs (10^6 cells per well), PBMCs stimulated with concanavalin A (10^6 cells + 5ug/ml ConA), PBMCs stimulated with concanavalin A in direct co-culture with AT-MSCs (10^6 PBMCs + 5ug/ mL of ConA + 2×10^5 AT-MSCs), and PBMCs stimulated with concanavalin A in indirect co-culture (transwell membranes) with AT-MSCs (10^6 PBMCs + 5ug/mL of ConA + 2×10^5 AT-MSCs). The plates were kept in culture for 96h, with BrdU being added in the last 24h. Then, culture samples were incubated with anti-CD3 canine, anti-BrdU and 7-AAD antibodies for evaluation in flow cytometry. Canine AT-MSCs in co-culture with PBMCs induced a reduction in the proliferative phase of CD3+ lymphocytes, possibly shifting them to the G0/G1 phase of the cell cycle. The reduction in the proliferative phase of CD3+ lymphocytes was evidenced both in co-cultures of AT-MSCs and PBMCs with direct and indirect contact, indicating that the mechanisms of action also depend on chemical mediators secreted in the medium. Although we did not observe any statistical difference, there was a tendency to inhibit the proliferation of CD3+ lymphocytes stimulated with concanavalin A when in direct and indirect contact with AT-MSCs. Our results indicate that canine AT-MSCs have immunomodulatory

potential. New studies must be carried out to elucidate the immunomodulatory mechanisms of action of canine AT-MSCs on lymphocytes.

Keywords: PBMC, mixed lymphocyte reaction, autoimmune, regeneration

CONCLUSION

The Mixed Leukocyte Reaction test proved to be an important tool to investigate in vitro the immunomodulatory potential of MSCs, through the evaluation of the ability to inhibit lymphocyte proliferation.

Canine AT-MSCs induced a reduction in the proliferative phase (S phase of the cell cycle) of CD3+ lymphocytes, in co-cultures with and without direct contact.

Although we did not observe any statistical difference, there was a tendency towards a reduction in the proliferation of CD3+ lymphocytes stimulated with concanavalin A when in direct and indirect co-culture with AT-MSCs.

Our results indicate that canine AT-MSCs have immunomodulatory potential. New studies must be carried out to elucidate the immunomodulatory mechanisms of action of canine AT-MSCs on lymphocytes.

Data Availability Statement

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Author Disclosure Statement

The authors declare no potential conflicts of interest.

Funding Information

This study was financed in part by the Coordenação de Aperfeiçoamento de Pessoal de Nível Superior – Brazil (CAPES) – Finance Code 001.

REFERENCES

1. Amorim, R.M., Clark, K.C., Walker, N.J., Kumar, P., Herout, K., Borjesson, D.L., Wang, A., 2020. Placenta-derived multipotent mesenchymal stromal cells: a promising potential cell-based therapy for canine inflammatory brain disease. *Stem Cell Res Ther* 11, 304. <https://doi.org/10.1186/s13287-020-01799-0>
2. Bernardo, M.E., Fibbe, W.E., 2013. Mesenchymal stromal cells: sensors and switchers of inflammation. *Cell Stem Cell* 13, 392–402. <https://doi.org/10.1016/j.stem.2013.09.006>
3. Bulati, M., Miceli, V., Gallo, A., Amico, G., Carcione, C., Pampalone, M., Conaldi, P.G., 2020. The Immunomodulatory Properties of the Human Amnion-Derived Mesenchymal Stromal/Stem Cells Are Induced by INF- γ Produced by Activated Lymphomonocytes and Are Mediated by Cell-To-Cell Contact and Soluble Factors. *Frontiers in Immunology* 11.
4. Carrade, D.D., Borjesson, D.L., 2013. Immunomodulation by Mesenchymal Stem Cells in Veterinary Species. *Comp Med* 63, 207–217.
5. Clark, K.C., Amador, A., Wang, A., 2023. Convergence of human and veterinary medicine: leveraging canine naturally occurring neurological disorders to develop regenerative treatments. *Neural Regeneration Research* 18, 541–542. <https://doi.org/10.4103/1673-5374.350195>
6. Dominici, M., Le Blanc, K., Mueller, I., Slaper-Cortenbach, I., Marini, F., Krause, D., Deans, R., Keating, A., Prockop, D., Horwitz, E., 2006. Minimal criteria for defining multipotent mesenchymal stromal cells. The International Society for Cellular Therapy position statement. *Cytotherapy* 8, 315–317. <https://doi.org/10.1080/14653240600855905>
7. Jiang, W., Xu, J., 2020. Immune modulation by mesenchymal stem cells. *Cell Proliferation* 53, e12712. <https://doi.org/10.1111/cpr.12712>
8. Khan, U., Ghazanfar, H., 2018. Chapter Three - T Lymphocytes and Autoimmunity, in: Galluzzi, L., Rudqvist, N.-P. (Eds.), *International Review of Cell and Molecular Biology, Biology of T Cells - Part A*. Academic Press, pp. 125–168. <https://doi.org/10.1016/bs.ircmb.2018.05.008>
9. Kol, A., Arzi, B., Athanasiou, K.A., Farmer, D.L., Nolte, J.A., Rebhun, R.B., Chen, X., Griffiths, L.G., Verstraete, F.J.M., Murphy, C.J., Borjesson, D.L., 2015. Companion animals: Translational scientist's new best friends. *Science Translational Medicine* 7, 308ps21-308ps21. <https://doi.org/10.1126/scitranslmed.aaa9116>

10. Kyurkchiev, D., Bochev, I., Ivanova-Todorova, E., Mourdjeva, M., Oreshkova, T., Belemmezova, K., Kyurkchiev, S., 2014. Secretion of immunoregulatory cytokines by mesenchymal stem cells. *World J Stem Cells* 6, 552–570. <https://doi.org/10.4252/wjsc.v6.i5.552>
11. Najar, M., Raicevic, G., Fayyad-Kazan, H., Bron, D., Tounouz, M., Lagneaux, L., 2016. Mesenchymal stromal cells and immunomodulation: A gathering of regulatory immune cells. *Cytotherapy* 18, 160–171. <https://doi.org/10.1016/j.jcyt.2015.10.011>
12. Sato, K., Ozaki, K., Oh, I., Meguro, A., Hatanaka, K., Nagai, T., Muroi, K., Ozawa, K., 2006. Nitric oxide plays a critical role in suppression of T-cell proliferation by mesenchymal stem cells. *Blood* 109, 228–234. <https://doi.org/10.1182/blood-2006-02-002246>
13. Taechangam, N., Iyer, S.S., Walker, N.J., Arzi, B., Borjesson, D.L., 2019. Mechanisms utilized by feline adipose-derived mesenchymal stem cells to inhibit T lymphocyte proliferation. *Stem Cell Research & Therapy* 10, 188. <https://doi.org/10.1186/s13287-019-1300-3>
14. Uccelli, A., Moretta, L., Pistoia, V., 2008. Mesenchymal stem cells in health and disease. *Nat Rev Immunol* 8, 726–736. <https://doi.org/10.1038/nri2395>
15. Wang, Y., Chen, X., Cao, W., Shi, Y., 2014. Plasticity of mesenchymal stem cells in immunomodulation: pathological and therapeutic implications. *Nat Immunol* 15, 1009–1016. <https://doi.org/10.1038/ni.3002>