

# RESSALVA

Atendendo solicitação do(a) autor(a), o texto completo desta tese será disponibilizado somente a partir de 08/02/2025.

SÃO PAULO STATE UNIVERSITY  
INSTITUTE OF BIOSCIENCES  
POSTGRADUATE PROGRAM IN PHARMACOLOGY AND BIO-  
TECHNOLOGY

**Study of genes stimulated by interferon- $\tau$  in immune  
cells as candidates for pregnancy markers in bovine fe-  
males**

**ISABELLA RIO FELTRIN**

Botucatu – SP  
2024

SÃO PAULO STATE UNIVERSITY  
INSTITUTE OF BIOSCIENCES  
POSTGRADUATE PROGRAM IN PHARMACOLOGY AND BIO-  
TECHNOLOGY

**Study of genes stimulated by interferon- $\tau$  in immune  
cells as candidates for pregnancy markers in bovine fe-  
males**

**ISABELLA RIO FELTRIN**

Thesis submitted to the Postgraduate Program in  
Pharmacology and Biotechnology of the Institute  
of Biosciences at the São Paulo State University to  
obtain the PhD degree in Pharmacology and Bio-  
technology.

**Advisor:** Claudia Maria Bertan Membrive, PhD.

**Co-Advisor:** Guilherme Pugliesi, PhD.

Botucatu – SP

2024

FICHA CATALOGRÁFICA ELABORADA PELA SEÇÃO TÉC. AQUIS. TRATAMENTO DA INFORM.  
DIVISÃO TÉCNICA DE BIBLIOTECA E DOCUMENTAÇÃO - CÂMPUS DE BOTUCATU - UNESP  
BIBLIOTECÁRIA RESPONSÁVEL: MARIA CAROLINA A. CRUZ E SANTOS-CRB 8/10188

Feltrin, Isabella Rio.

Study of genes stimulated by interferon-T in immune cells as candidates for pregnancy markers in bovine females / Isabella Rio Feltrin. - Botucatu, 2024

Tese (doutorado) - Universidade Estadual Paulista "Júlio de Mesquita Filho", Instituto de Biociências de Botucatu

Orientador: Claudia Maria Bertan Membrive

Coorientador: Guilherme Pugliesi

Capes: 50504002

1. Interferon. 2. Immunity, Cellular. 3. Genes Expression.  
4. Animal Pregnancy. 5. Cattle Reproduction.

Palavras-chave: ISG; Immune cells; Interferon-T; Pregnancy diagnosis; Uterine flush.

**Author:** Isabella Rio Feltrin

**Title:** Study of genes stimulated by interferon- $\tau$  in immune cells as candidates for pregnancy markers in bovine females

**Committee Members**

*Prof. Dr. Guilherme Pugliesi*

President and Co-Advisor.

University of São Paulo – Pirassununga/SP.

*Prof. Dr. Fernando Silveira Mesquita*

Titular member.

Federal University of Pampa – Uruguaiana/RS.

*Prof. Dr. Alfredo Quites Antoniazzi*

Titular member.

Federal University of Santa Maria – Santa Maria/RS.

*Prof. Dr. Mateus José Sudano*

Titular member.

Federal University of São Carlos – São Carlos/SP.

*Prof. Dr. Luciano Andrade Silva*

Titular member.

University of São Paulo – Pirassununga/SP.

**Date:** 08/02/2024



## CERTIFICADO

Certificamos que a proposta intitulada "Inovações no diagnóstico precoce da gestação em bovinos", protocolada sob o CEUA nº 8192280317, sob a responsabilidade de **Guilherme Pugliesi** - que envolve a produção, manutenção e/ou utilização de animais pertencentes ao filo Chordata, subfilo Vertebrata (exceto o homem), para fins de pesquisa científica ou ensino - está de acordo com os preceitos da Lei 11.794 de 8 de outubro de 2008, com o Decreto 6.899 de 15 de julho de 2009, bem como com as normas editadas pelo Conselho Nacional de Controle da Experimentação Animal (CONCEA), e foi **aprovada** pela Comissão de Ética no Uso de Animais da Faculdade de Medicina Veterinária e Zootecnia da Universidade de São Paulo (CEUA/FMVZ) na reunião de 10/05/2017.

We certify that the proposal "Innovations in the early pregnancy diagnosis in cattle", utilizing 90 Bovines (90 females), protocol number CEUA 8192280317, under the responsibility of **Guilherme Pugliesi** - which involves the production, maintenance and/or use of animals belonging to the phylum Chordata, subphylum Vertebrata (except human beings), for scientific research purposes or teaching - is in accordance with Law 11.794 of October 8, 2008, Decree 6899 of July 15, 2009, as well as with the rules issued by the National Council for Control of Animal Experimentation (CONCEA), and was **approved** by the Ethic Committee on Animal Use of the School of Veterinary Medicine and Animal Science (University of São Paulo) (CEUA/FMVZ) in the meeting of 05/10/2017.

Finalidade da Proposta: **Pesquisa**

Vigência da Proposta: de **05/2017 a 04/2018** Área: **Reprodução Animal**

Origem:	Prefeitura do Campus da USP de Pirassununga		
Espécie:	Bovinos	sexo:	Fêmeas
Linhagem:	Nelore	idade:	3 a 9 anos
		Peso:	400 a 700 kg
		N:	30
Origem:	Prefeitura do Campus da USP de Pirassununga		
Espécie:	Bovinos	sexo:	Fêmeas
Linhagem:	Nelore	idade:	12 a 24 anos
		Peso:	300 a 400 kg
		N:	30
Origem:	Prefeitura do Campus da USP de Pirassununga		
Espécie:	Bovinos	sexo:	Fêmeas
Linhagem:	Holandesa	idade:	2 a 7 anos
		Peso:	450 a 700 kg
		N:	30

Resumo: A identificação do estado gestacional mais precocemente aos métodos atuais realizados por volta do 30º dia pós-inseminação permite uma redução no intervalo de inseminações, o que pode resultar em menor período para concepção e maior rentabilidade ao sistema de produção pecuário. A tese central do projeto é que biomarcadores no sangue estimulados pelo conceito são detectáveis no início da gestação. Isto pode permitir o desenvolvimento de um método precoce e inovador de diagnóstico da gestação em bovinos. Assim, tem-se como objetivo principal desenvolver um método de diagnóstico através da quantificação da abundância de genes estimulados por IFN-tau (ISGs) já conhecidos ou por novos marcadores aliado a uma identificação precoce dos animais não-gestantes pela avaliação da função luteal. Para isso, no Estudo 1 novos marcadores do estado gestacional poderão ser identificados através do estudo transcriptômico das células imunes do sangue no dia 14 e 18 pós-inseminação. Pretende-se caracterizar a abundância de ISGs já conhecidos e de novos transcritos estimulados pela presença do conceito, no sangue total, nas células imunes polimorfonucleares (PMNs) e em células imunes do leite entre os dias 12 e 20 de gestação em vacas de corte e leite. A luteólise será detectada por ultrassonografia Doppler através das mudanças no tamanho e vascularização do corpo lúteo (CL) e pelas concentrações circulantes de progesterona (P4) para detecção de vacas com CL não-funcional. Para mensuração das concentrações de P4 objetiva-se desenvolver e validar um ensaio imunoenzimático com alta sensibilidade e especificidade para evitar o uso de radioisótopos. No Estudo 2 será determinado a acurácia de métodos de diagnóstico da gestação realizados antes do dia 20 pós-IA, e baseados no uso isolado ou combinado da expressão de ISGs e da funcionalidade luteal (pela concentração de P4 ou ultrassonografia Doppler). Os novos conhecimentos e métodos inovadores aqui propostos embasarão o desenvolvimento de tecnologias para a pecuária de leite e corte. A exequibilidade será garantida pelo trabalho integrado de especialistas em cada etapa analítica e pela infraestrutura das instituições envolvidas.

Local do experimento: **CBRA/VRA/FMVZ**

São Paulo, 12 de maio de 2017



Profa. Dra. Denise Tabacchi Fantoni  
Presidente da Comissão de Ética no Uso de Animais  
Faculdade de Medicina Veterinária e Zootecnia da Universidade  
de São Paulo

Roseli da Costa Gomes  
Secretaria Executiva da Comissão de Ética no Uso de Animais  
Faculdade de Medicina Veterinária e Zootecnia da Universidade  
de São Paulo

*I dedicate this thesis to my parents, Ana and Feltrin, who  
are the basis of my life and my greatest supporters. Everything I  
am today is thanks to you!  
All my love for you!*

## ACKNOWLEDGMENTS

Firstly, my sincere thanks to my parents, *Ana and Feltrin*, for all their unconditional support dedicated to my dreams and me. You are the foundation of my life and my greatest example of person's and character. Without you, none of this would be possible. All my love to you!

To my life partner, *Vinicius*, for all the love, affection, and patience dedicated to my dreams and me. Thank you for being by my side every moment of my life. All my love for you!

To the *Postgraduate program and the professors* of the Pharmacology and Biotechnology program at the Institute of Biosciences of the São Paulo State University, I extend my gratitude for the opportunity provided to pursue my PhD and for the courses taught that significantly contributed to my professional and academic qualification.

To the *São Paulo State Research Support Foundation (FAPESP)*, for the financial support granted for the project (grant number 2015/10606-9).

To the *Coordination for the Improvement of Higher Education Personnel (CAPES)* for the PhD scholarship that facilitated my scientific qualification.

To the *Center for Biotechnology in Animal Reproduction (CBRA)* of the Fernando Costa Campus of the Faculty of Veterinary Medicine and Zootecnics of the University of São Paulo (FMVZ, USP – Pirassununga), for providing the animals used in this study, as well as for the infrastructure and logistics that enabled the successful completion of this project.

To the *members of the evaluation committee*, Prof. Fernando Silveira Mesquita, Prof. Alfredo Quites Antoniazzi, Prof. Mateus José Sudano, and Prof. Luciano Andrade Silva, for their valuable scientific contributions to this thesis.

My sincere thanks to my advisor, ***Prof. Claudia M. Bertan Membrive***, for granting me the opportunity to pursue my master's and PhD degrees. Your support, teachings, and knowledge shared over the years have significantly contributed to my personal and intellectual growth. My eternal gratitude and admiration for you and for what you represent for science!

My sincere thanks to my co-advisor, ***Prof. Guilherme Pugliesi***, for being part of my academic journey. For all the knowledge, teachings, and scientific contributions that have made me the professional I am today; and for the opportunity to be part of the LFEM family, which has certainly changed my life forever. My eternal gratitude. You are an inspiration to us all!

To my dear friends from the Postgraduate - ***Amanda Guimarães, Ana Clara Degan, Karine Morelli, Thais Sayuri, and Cecília Rocha***; I am thankful for your unconditional support, companionship, and affection throughout these years. Your presence was indispensable in my journey. Thank you for everything, and I love you all!

Special thanks to ***my friends and members of LFEM*** - Amanda, Ana Clara, Karine, Thiago, Danilo, Igor, Diego, Ana Laura, Lucas, Samuel, and the scientific initiation students and trainees; for their assistance and time dedicated to the execution and logistics of this project.

Finally, I extend my gratitude to everyone who directly or indirectly contributed to the planning and execution of this study, leading to the development of my PhD. Thank you very much.

## RESUMO

FELTRIN, I.R. **Estudo de genes estimulados por interferon- $\tau$  em células imunes como candidatos à marcadores de prenhez em fêmeas bovinas.** [Study of genes stimulated by interferon- $\tau$  in immune cells as candidates for pregnancy markers in bovine females]. 2024. 99 f. Tese (Doutorado em Farmacologia e Biotecnologia) – Instituto de Biociências, Universidade Estadual Paulista, Botucatu, 2024.

Nas fêmeas bovinas, as perdas embrionárias ocorridas entre o 14º e o 19º dia após o estro afetam significativamente a eficiência reprodutiva dos rebanhos. Portanto, o desenvolvimento de métodos diagnósticos capazes de detectar precocemente a gestação ou não gestação é um ponto importante para melhorar o desempenho reprodutivo dos bovinos. O interferon- $\tau$  (IFN- $\tau$ ) produzido pelo concepto desempenha um papel importante durante o reconhecimento materno da prenhez (MPR) em ruminantes, que envolve mais do que a inibição dos pulsos luteolíticos da prostaglandina F2 alfa (PGF<sub>2 $\alpha$</sub> ) para manter a funcionalidade do corpo lúteo (CL). Para uma prenhez bem-sucedida, o concepto semi-alogênico deve evitar a sua rejeição pelo sistema imunológico materno, destacando a importância de elucidar os mecanismos envolvidos durante o início da prenhez. Além disso, neste momento, o IFN- $\tau$  é liberado na circulação sanguínea e estimula a expressão de genes (ISGs). O estudo da expressão gênica dos ISGs clássicos (*ISG15*, *OAS1*, *MX1* e *MX2*) em células imunes do sangue periférico, como células mono (PBMC) e polimorfonucleares (PMN) já vem sendo utilizado como ferramenta de diagnóstico precoce da prenhez, porém apresentou baixa acurácia. Neste contexto, nossos objetivos neste estudo foram: 1) caracterizar a magnitude da resposta dos ISGs clássicos (*ISG15*) e não clássicos (*RSAD2* e *IFI44*); bem como o perfil das citocinas pró- (*IL1 $\beta$* ) e anti-inflamatórias (*IL10*) em PBMC e PMN estimuladas com IFN- $\tau$  (**Experimento 1**) ou lavados uterinos (UF) de vacas gestantes (**Experimento 2**); e 2) avaliar a acurácia desses biomarcadores recentemente identificados (*RSAD2* e *IFI44*) como preditores da prenhez precoce em fêmeas bovinas (**Experimento 3**). Nos Experimentos 1 e 2, PBMC e PMN foram isoladas do sangue de vacas Nelore não gestantes (N=9) entre 10-12 dias pós-ovulação (D0 = dia da ovulação), e estimuladas com 100 ng/mL de roIFNT ou UF de vacas no dia 18 de gestação. A expressão gênica foi determinada por qPCR. Para o Experimento 3, fêmeas Nelore (núlparas, N=103; primíparas, N=53; pluríparas, N=91) foram submetidas à coleta de

sangue para isolamento das PMN, e foi realizada ultrassonografia Doppler para avaliar a funcionalidade do CL no D20 após inseminação artificial em tempo fixo (D0 = dia da IATF). A expressão gênica também foi determinada por qPCR. Além disso, foram realizadas curvas ROC para determinar a acurácia dos preditores da prenhez no D20 (ISGs e Doppler). Nos Experimentos 1 e 2, a expressão de todos os ISGs foi maior ( $P < 0,05$ ) em ambas as células tratadas com roIFNT e UF-Conceptus do que nos grupos Controle. O fold change indicou que o *ISG15* e *RSAD2* foram os genes mais estimulados ( $P < 0,05$ ) em PBMC e PMN. A expressão de *IL1 $\beta$*  foi menor ( $P < 0,05$ ) em PBMC e PMN tratadas com UF-Conceptus; no entanto, nenhuma diferença ( $P > 0,1$ ) foi observada entre os outros grupos de tratamento. Para o Experimento 3, todas as combinações genéticas foram testadas, e a melhor associação para aumento da acurácia (92,7%) e redução de resultados falsos negativos (0,9%, 2/233) foi obtida quando os animais foram considerados prenhes se um dos os quatro ISGs (*ISG15*, *OAS1*, *RSAD2* e *IFI44*) foram estimulados em fêmeas com CL ativo ( $> 25\%$  de perfusão sanguínea) no D20. Concluindo, o *ISG15* e *RSAD2* foram os ISGs mais estimulados em PBMC e PMN, indicando que uma associação entre ISGs clássicos e não clássicos pode ser utilizada como uma ferramenta precoce para melhorar a predição da prenhez no D20 em fêmeas bovinas com CL ativo determinado através da US-Doppler.

**Palavras-chave:** ISG, células imunes, interferon- $\tau$ , lavado uterino, diagnóstico de prenhez.

## ABSTRACT

FELTRIN, I.R. **Study of genes stimulated by interferon- $\tau$  in immune cells as candidates for pregnancy markers in bovine females.** [Estudo de genes estimulados por interferon- $\tau$  em células imunes como candidatos à marcadores de prenhez em fêmeas bovinas]. 2024. 99 f. Tese (Doutorado em Farmacologia e Biotecnologia) – Instituto de Biociências, Universidade Estadual Paulista, Botucatu, 2024.

In bovine females, embryonic losses occurring between the 14th and 19th day after estrus significantly affect the reproductive efficiency of herds. Therefore, the development of diagnostic methods capable of early detection of pregnancy or non-pregnancy is an important point for improving the reproductive performance of cattle. Interferon- $\tau$  (IFN- $\tau$ ) produced by the conceptus plays an important role during maternal recognition of pregnancy (MPR) in ruminants, which involves more than the inhibition of luteolytic pulses of prostaglandin F2 alpha (PGF<sub>2 $\alpha$</sub> ) to maintain the function of corpus luteum (CL). For a successful pregnancy, the semi-allogeneic conceptus must avoid rejection by the maternal immune system, highlighting the importance of elucidating the mechanisms involved during early pregnancy. Furthermore, at this moment, IFN- $\tau$  is released into the blood circulation and stimulates the expression of genes (ISGs). The study of gene expression of the classic ISGs (*ISG15*, *OAS1*, *MX1*, and *MX2*) in peripheral blood immune cells, such as mono (PBMC) and polymorphonuclear (PMN) cells has already been used as a form of early diagnosis of pregnancy, but showed low accuracy. In this context, our objective were: 1) characterize the response magnitude of classic (*ISG15*) and non-classic (*RSAD2* and *IFI44*) ISGs; as well as pro- (*IL1 $\beta$* ) and anti-inflammatory (*IL10*) cytokines in PBMC and PMN stimulated by IFN- $\tau$  (**Experiment 1**) or uterine flush (UF) from pregnant cows (**Experiment 2**); and 2) evaluate the accuracy of these recently identified biomarkers (*RSAD2* and *IFI44*) as early pregnancy predictors in bovine females (**Experiment 3**). In the *Experiment 1 and 2*, PBMC and PMN were isolated from the blood of non-pregnant Nelore cows (N=9) between 10-12 days post-ovulation (D0 = day of ovulation) and stimulated with 100 ng/mL of roIFNT or UF from day 18 of pregnancy cows. Gene expression was determined by qPCR. For the *Experiment 3*, Nelore females (nulliparous, N=103;

primiparous, N=53; pluriparous, N=91) were submitted to blood collection for PMN isolation, and Doppler ultrasonography was performed to assess the functionality of the corpus luteum (CL) on D20 after timed-artificial insemination (D0 = TAI day). Gene expression was also determined by qPCR. In addition, ROC curves were performed to determine the accuracy of pregnancy predictors on D20 (ISGs and Doppler). In *Experiments 1 and 2*, expression of all ISGs was greater ( $P < 0.05$ ) in both cells treated with roIFNT and UF-Conceptus than its controls. The fold change indicated that *ISG15* and *RSAD2* were the most stimulated genes ( $P < 0.05$ ) in PBMC and PMN. Expression of *IL1 $\beta$*  was lesser ( $P < 0.05$ ) in PBMC and PMN treated with UF-Conceptus; however, no difference ( $P > 0.1$ ) was observed between the other treatment groups. For the *Experiment 3*, all gene combinations were tested, and the best association for an increase in accuracy (92.7%) and reduction of false negative results (0.9%, 2/233) was obtained when pregnant animals were considered if one of the four ISGs (*ISG15*, *OAS1*, *RSAD2*, and *IFI44*) were stimulated in females with an active CL (> 25% blood perfusion) on D20. In conclusion, *ISG15* and *RSAD2* were the most stimulated ISGs in PBMC and PMN, indicating that an association between classic and non-classic ISGs can be used as an early tool to improve pregnancy prediction at D20 in bovine females with active CL determined by US-Doppler.

**Keywords:** ISG, immune cells, interferon- $\tau$ , uterine flush, pregnancy diagnosis.

## SUMMARY

<b>1. GENERAL INTRODUCTION</b> .....	17
<b>2. CHAPTER 1: LITERATURE REVIEW</b> .....	21
2.1 PRODUCTION OF IFN- $\tau$ BY EMBRYO AND BOVINE CONCEPTUS. ....	21
2.2 AUTOCRINE AND PARACRINE EFFECTS OF IFN- $\tau$ DURING MATERNAL RECOGNITION OF PREGNANCY .....	22
2.3 ENDOCRINE EFFECTS OF IFN- $\tau$ ON THE CORPUS LUTEUM AND IMMUNE CELLS DURING MATERNAL RECOGNITION OF PREGNANCY.....	24
2.4 MODULATION OF THE IMMUNE SYSTEM AT THE ONSET OF PREGNANCY.....	29
2.5 USE OF ISGS IN EARLY DIAGNOSIS OF PREGNANCY IN BOVINE FEMALES.....	32
2.6 REFERENCES .....	35
<b>3. CHAPTER 2: BOVINE CONCEPTUS INDUCES ACCURATE BIOMARKERS     IN IMMUNE CELLS FOR EARLY PREGNANCY DIAGNOSIS.....</b>	<b>51</b>
3.1 INTRODUCTION .....	51
3.2 MATERIAL AND METHODS .....	54
<b>3.2.1 Ethics statement</b> .....	54
<b>3.2.3 Experimental design of Experiments 1 and 2</b> .....	55
3.2.3.1 <i>Isolation of immune cells from peripheral blood</i> .....	56
3.2.3.2 <i>Collection of UF on day 18 of the estrous cycle</i> .....	56

3.2.3.3 <i>Experiment 1: Stimulation of immune cells with roIFNT</i> .....	57
3.2.3.4 <i>Experiment 2: Culture of immune cells in UF</i> .....	57
<b>3.2.4 Experimental design of Experiment 3: accuracy of pregnancy markers in PMN</b> .....	58
<b>3.2.5 RNA extraction, cDNA synthesis, and quantitative polymerase chain reaction (qPCR)</b> .....	58
<b>3.2.6 Statistical analyses</b> .....	60
3.3 RESULTS.....	61
<b>3.3.1 Experiment 1 and 2</b> .....	61
3.3.1.1 <i>Effects of roIFNT on the expression of ISG and cytokines in immune cells</i> .....	61
3.3.1.2 <i>Effects of UF from Day 18 on the expression of ISG and cytokines in immune cells</i> .....	62
3.3.1.3 <i>Correlations between ISG and cytokines in immune cells</i> .....	62
<b>3.3.2 Experiment 3</b> .....	63
3.3.2.1 <i>Interferon stimulated genes – RSAD2 and IFI44</i> .....	63
3.3.2.2 <i>Correlations between ISG in PMN</i> .....	64
3.3.2.3 <i>Accuracy of pregnancy predictors</i> .....	64
3.4 DISCUSSION.....	66
3.5 REFERENCES.....	74
3.6 FINAL CONSIDERATIONS.....	83
3.7 FIGURES AND LEGENDS.....	85

3.8 TABLES .....	92
3.9 SUPPLEMENTARY TABLES.....	97

## 1           **1. GENERAL INTRODUCTION**

2           Brazil has the second-largest cattle herd in the world, totaling around 202 million  
3 animals, representing 12.18% of the global herd (ABIEC, 2023). Although the Brazilian  
4 beef industry is on the rise, the reproductive efficiency of herds is still low and deserves  
5 attention. For example, the calving interval in Brazilian beef cattle operations is around  
6 16.3 months, far from the ideal calving period of 12-13 months (BARUSELLI et al.,  
7 2016). Therefore, developing and improving technologies that help increase the produc-  
8 tivity of properties and optimize breeding systems and herd profitability are essential.

9           To achieve enhanced genetic and production gains, reproductive strategies must  
10 prioritize improving service rates and reducing the interval between inseminations,  
11 without compromising the viability of the already-established pregnancy (SÁ FILHO et  
12 al., 2014). Based on this concept, protocols for resynchronizing ovulation in females who  
13 did not become pregnant were developed. Thus, the success in reducing prolonged cal-  
14 ving intervals is associated with an early pregnancy diagnosis combined with an efficient  
15 resynchronization protocol. In this context, early detection of embryonic loss is essential  
16 - which occurs between days 8 and 17 of pregnancy in 40% of cases (THATCHER et al.,  
17 2001) - or early detection of conception failure.

18           Embryonic losses significantly affect reproductive efficiency and result from fai-  
19 lure in maternal pregnancy recognition (MPR). MPR occurs between days 15 and 19 post-  
20 ovulation in bovine females, through the release of the interferon- $\tau$  (IFN- $\tau$ ), which is pro-  
21 duced and secreted by the trophoblastic cells of the conceptus in the uterine environment,  
22 which blocks the mechanisms involved in pulsatile secretion of prostaglandin F2 alpha  
23 (PGF<sub>2 $\alpha$</sub> ) by the endometrium (MARQUES et al., 2007). IFN- $\tau$  increases on day 15, peaks  
24 on day 20, and decreases to basal concentrations near day 25 (BAZER et al., 2009). In  
25 addition, IFN- $\tau$  acts on endometrial cells downregulating the estradiol (ER) and oxytocin

26 (OXTR) receptors (SPENCER et al., 2007). As a consequence, the pulsatile release of  
27  $\text{PGF}_{2\alpha}$ , induced by the binding of oxytocin (OT) to its endometrial receptor, does not  
28 occur, and the corpus luteum (CL) remains active producing high concentrations of pro-  
29 gesterone (P4) to maintain pregnancy.

30         Among the pregnancy diagnosis methods applied to cattle, the most used techni-  
31 que is B-mode ultrasonography between days 28 and 32 after timed artificial insemination  
32 (TAI) (MARQUES et al, 2012; STEVENSON et al., 2003). However, this method makes  
33 it impossible to identify non-pregnant cows immediately after MRP and before returning  
34 to estrus (FORDE et al., 2011). Actually, other techniques have become available for the  
35 diagnosis of pregnancy, one of which is the detection of structural luteolysis in non-preg-  
36 nant females using Doppler ultrasonography (Doppler-US). The detection of structural  
37 luteolysis has been used in beef cows between 20 and 22 days after TAI (PUGLIESI et  
38 al., 2014) with accuracy and sensitivity exceeding 90%. However, this technique has a  
39 limiting factor: the high number of false positives.

40         In the last years, advances have been achieved, generating the possibility of per-  
41 forming an indirect pregnancy diagnosis through the expression of genes stimulated by  
42  $\text{IFN-}\tau$  (ISGs) on days 18 and 20 of pregnancy in peripheral blood immune cells. The most  
43 traditional ISGs used to detect pregnancy in immune cells are Ubiquitin-like Modifier 15  
44 (*ISG15*), 2'-5'-Oligoadenylate Synthetase 1 (*OAS1*), MX dynamin as GTPase 1 (*MX1*),  
45 and MX dynamin as GTPase 2 (*MX2*) (KIZAKI et al. al., 2013; PUGLIESI et al., 2014).  
46 The increase of ISGs abundance in peripheral blood mononuclear cells (PBMC) has been  
47 reported in ewes (ANTONIAZZI et al., 2013), dairy cows (SHIRASUNA et al., 2012,  
48 FERRAZ et al., 2021), and beef cattle (PUGLIESI et al., 2014; DALMASO DE MELO  
49 et al., 2020). The expression of ISGs follows the  $\text{IFN-}\tau$  release pattern, which may indi-

50 cate pregnancy earlier than other conventional diagnostic methods. Furthermore, expres-  
51 sion of classic ISGs has also been reported in peripheral blood polymorphonuclear cells  
52 (PMN) in cattle (KIZAKI et al., 2013; TOJI et al., 2017; MELO et al., 2020), and there  
53 is evidence that PMNs are more sensitive to IFN- $\tau$  stimulation than PBMCs (KIZAKI et  
54 al., 2013).

55 The expression of classic ISGs in PBMC and PMN has been used in a few studies  
56 for prospective diagnosis of pregnancy on the 20th day post-TAI in beef cows; however,  
57 the maximum accuracy achieved was 87% (PUGLIESI et al., 2014; DALMASO de  
58 MELO et al., 2020). Therefore, in a previous study conducted by Rocha et al. (2020) using  
59 RNA sequencing, novel early-pregnancy-induced genes were identified in PBMC (*IFI6*,  
60 *RSAD2*, *IFI44*, *IFITM2*, *CLEC3B*, *OAS2*, *TNFSF13B*, *DMKN* and *LGALS3BP*) and  
61 PMN (*IFI44*, *RSAD2*, *OAS2*, *LGALS3BP*, *IFI6* and *CIR*) on the 18th day after TAI.  
62 These genes have the potential to be used as pregnancy biomarkers, offering an alterna-  
63 tive for greater accuracy in the early diagnosis of pregnancy compared to the classic ISGs  
64 already in used. Thus, a better understanding of the transcriptional stimulus of each bi-  
65 omarker in immune system cells is a necessary aspect to be explored to deepen the selec-  
66 tion criteria. Moreover, in the aforementioned study (Rocha et al., 2020), a limited sample  
67 size was used (N=6 animals/group), and as pregnancy assessment is a binary test, a larger  
68 number of animals must be explored to correctly select a pregnancy biomarker.

69 The herein study aims to determine novel pregnancy markers in peripheral blood  
70 immune cells that can serve as a basis for developing and improvement of molecular  
71 methods for pregnancy diagnosis in cattle. Our central hypothesis is that biomarkers in  
72 peripheral blood, stimulated by the presence of the conceptus, may be detectable at the  
73 early stages of pregnancy in circulating immune cells. This condition would enable the  
74 development of a method for early pregnancy diagnosis in bovine females 20 days after

75 mating. To test our hypothesis, the specific objectives of the current study were: 1) char-  
76 acterize the response magnitude of classic (*ISG15*) and non-classic (*RSAD2* and *IFI44*)  
77 ISGs; as well as pro- (*IL1 $\beta$* ) and anti-inflammatory (*IL10*) cytokines in PBMC and PMN  
78 stimulated by IFN- $\tau$  (*Experiment 1*) or uterine flush (UF) from pregnant cows (*Experi-*  
79 *ment 2*); and 2) evaluate the accuracy of these recently identified biomarkers (*RSAD2* and  
80 *IFI44*) as early pregnancy predictors in bovine females (*Experiment 3*).

81 This thesis is divided into two chapters. Chapter 1 reviews relevant topics related  
82 to the experiments conducted in the subsequent chapter. Chapter 2 describes the magni-  
83 tude of the transcriptional response of each ISG and cytokine due to challenge by direct  
84 stimulation of IFN- $\tau$  or UF in PBMC and PMN, using an *in vitro* study model consisting  
85 of two experiments. Based on the results obtained in these studies, we conducted the third  
86 experiment. In the third experiment, we tested the accuracy of ISGs expression in PMN  
87 as early pregnancy predictors in a large number of animals. Chapter 2 was written in  
88 accordance with the standards of the journal *Biology of Reproduction*, to which the man-  
89 uscript will be submitted. The final considerations at the end of this thesis summarize the  
90 most important findings of each experiment and discuss future perspectives for further  
91 studies.

92 We believe that the present study introduces an innovative aspect as it corresponds  
93 to the development of new techniques to diagnose pregnancy earlier than current meth-  
94 ods, in addition to generating the possibility of identifying new molecular markers in  
95 peripheral blood immune cells, in addition to classic ISGs. Such advances can directly  
96 affect the efficiency of reproductive programs in cattle, especially those that use TAI.

97  
98  
99

1378           **3.5 REFERENCES**

1379

1380   [1] Forde N, Bazer FW, Spencer TE, Lonergan P. 'Conceptualizing' the Endometrium:  
1381   Identification of Conceptus-Derived Proteins During Early Pregnancy in Cattle. *Biology*  
1382   *of Reproduction* 2015; **92**(6):156.

1383

1384   [2] Imakawa K, Anthony RV, Kazemi M, Marotti KR, Polites HG, Roberts RM. Inter-  
1385   feron-like sequence of ovine trophoblast protein secreted by embryonic trophectoderm.  
1386   *Nature* 1987; **330**:377-379.

1387

1388   [3] Bazer FW, Spencer TE, Ott TL. Interferon tau: a novel pregnancy recognition signal.  
1389   *Am. J. Reprod. Immunol.* 1997; **37**:412-420.

1390

1391 [4] Reese ST, Franco GA, Poole RK, Hood R, Fernandez Montero L, Oliveira Filho RV,  
1392 Cooke RF, Pohler KG. Pregnancy loss in beef cattle: A meta-analysis. *Anim Reprod Sci.*  
1393 2020; **212**:106251.  
1394  
1395 [5] Morelli SS, Mandal M, Goldsmith LT, Kashani BN, Ponzio NM. The maternal im-  
1396 mune system during pregnancy and its influence on fetal development. *Res. Rep. Biol.*  
1397 2015; **6**:171–189.  
1398  
1399 [6] Bonney EA. Immune regulation in pregnancy: a matter of perspective? *Obstet. Gyne-*  
1400 *col. Clin.* 2016; **43**: 679–698.  
1401  
1402 [7] Talukder AK, Rashid MB, Yousef MS, Kusama K, Shimizu T, Shimada M, Suarez  
1403 SS, Imakawa K, Miyamoto A. Oviduct epithelium induces interferon-tau in bovine day-  
1404 4 embryos, which generates an anti-inflammatory response in immune cells. *Sci. Rep.*  
1405 2018; **8**:7850.  
1406  
1407 [8] Talukder AK, Yousef MS, Rashid MB, Awai K, Acosta TJ, Shimizu T, Okuda K,  
1408 Shimada M, Imakawa K, Miyamoto A. Bovine embryo induces an anti-inflammatory re-  
1409 sponse in uterine epithelial cells and immune cells *in vitro*: possible involvement of in-  
1410 terferon tau as an intermediary. *J. Reprod. Dev.* 2017; **63**:425-434.  
1411  
1412 [9] Meyerholz MM, Mense K, Knaack H, Sandra O, Schmicke M. Pregnancy-Induced  
1413 ISG-15 and MX-1 Gene expression is detected in the liver of holstein–friesian heifers  
1414 during late peri-implantation period. *Reprod Domest Anim.* 2015; **51**:175-7.  
1415

1416 [10] Sponchiado M, Gomes NS, Fontes PK, Martins T, del Collado M, Pastore AdA,  
1417 Pugliesi G, Nogueira MFG, Binelli M. Pre-hatching embryo-dependent and – independ-  
1418 ent programming of endometrial function in cattle. *PLoS ONE* 2017; **12**(4):e0175954.  
1419

1420 [11] Bridi A, Bertolin K, Rissi VB, Mujica LKS, Glanzner WG, Macedo MP, Comim  
1421 FV, Gonçalves PBD, Antoniazzi AQ. Parthenogenetic bovine embryos secrete type I in-  
1422 terferon capable of stimulating ISG15 in luteal cell culture. *Anim Reprod.* 2018;  
1423 **15**(4):1268-77.  
1424

1425 [12] Pugliesi G, Miagawa BT, Paiva YN, França MR, Silva LA, Binelli M. Conceptus-  
1426 induced changes in the gene expression of blood immune cells and the ultrasound-ac-  
1427 cessed luteal function in beef cattle: How early can we detect pregnancy? *Biol. Reprod.*  
1428 2014; **91**(4): 1-12.  
1429

1430 [13] Soumya, N. P. Differential expression of ISG 15 mRNA in peripheral blood mono-  
1431 nuclear cells of nulliparous and multiparous pregnant versus non-pregnant *Bos indicus*  
1432 cattle. *Reprod. Domest. Anim.* 2017; **52**: 97–106.  
1433

1434 [14] Forde N, Carter F, Spencer TE, Bazer FW, Sandra O, Mansouri-Attia N, Okumu LA,  
1435 McGettigan PA, Mehta JP, McBride R, O'Gaora P, Roche JF, Lonergan P. Conceptus-  
1436 induced changes in the endometrial transcriptome: how soon does the cow know she is  
1437 pregnant? *Biol Reprod.* 2011; **85**(1):144-56.  
1438

- 1439 [15] Motta IG, Rocha CC, Bisinotto DZ, Melo GD, Ataíde Júnior GA, Silva AG, Gonzaga  
1440 VHG, Santos JA, Freitas BG, Lemes KM, Madureira EH, Pugliesi G. Increased preg-  
1441 nancy rate in beef heifers resynchronized with estradiol at 14 days after TAI. *Theriogenol-*  
1442 *ogy* 2020; **15**(147):62-70.
- 1443
- 1444 [16] Siqueira LG, Areas VS, Ghetti AM, Fonseca JF, Palhao MP, Fernandes CA, Viana  
1445 JH. Color Doppler flow imaging for the early detection of nonpregnant cattle at 20 days  
1446 after timed artificial insemination. *J Dairy Sci.* 2013; **96**(10):6461-72.
- 1447
- 1448 [17] Pugliesi G, de Melo GD, Ataíde GA, Pellegrino CAG, Silva JB, Rocha CC, Motta  
1449 IG, Vasconcelos JLM, Binelli M. Use of Doppler ultrasonography in embryo transfer  
1450 programs: feasibility and field results. *Animal Reprod.* 2018; **15**: 239-246.
- 1451
- 1452 [18] Dalmaso de Melo G, Mello BP, Ferreira CA, Souto Godoy Filho CA, Rocha CC,  
1453 Silva AG, Reese ST, Madureira EH, Pohler KG, Pugliesi G. Applied use of interferon-  
1454 tau stimulated genes expression in polymorphonuclear cells to detect pregnancy com-  
1455 pared to other early predictors in beef cattle. *Theriogenology* 2020; **152**: 94-105.
- 1456
- 1457 [19] Rocha CC, da Silva Andrade SC, de Melo GD, Motta IG, Coutinho LL, Gonella-  
1458 Diaza AM, Binelli M, Pugliesi G. Early pregnancy-induced transcripts in peripheral  
1459 blood immune cells in *Bos indicus* heifers. *Sci Rep.* 2020; **10**:13733.
- 1460
- 1461 [20] Houghton PL, Lemenager RP, Hendrix KS, Moss GE, Stewart TS. Effects of body  
1462 composition, pre- and postpartum energy intake and stage of production of energy utili-  
1463 zation by beef cows. *J. Anim Sci.* 1990; **68**(5)1447-56.

1464 [21] Jiemtaweeboon S, Shirasuna K, Nitta A, Kobayashi A, Schuberth HJ, Shimizu T,  
1465 Miyamoto A. Evidence that polymorphonuclear neutrophils infiltrate into the developing  
1466 corpus luteum and promote angiogenesis with interleukin-8 in the cow. *Reprod Biol En-*  
1467 *docrinol.* 2011; **8**(9):79.

1468

1469 [22] Antoniazzi AQ, Webb BT, Romero JJ, Ashley RL, Smirnova N P, Henkes LE, Bott  
1470 RC, Oliveira JF, Niswender GD, Bazer FW, Hansen TR. Endocrine delivery of interferon  
1471 tau protects the corpus luteum from prostaglandin F2 alpha-induced luteolysis in ewes.  
1472 *Biol Reprod.* 2013; **88**(6):144.

1473

1474 [23] Shirasuna K, Matsumoto H, Kobayashi E, Nitta A, Haneda S, Matsui M, Kawashima  
1475 C, Kida K, Shimizu T, Miyamoto A. Upregulation of interferon-stimulated genes and  
1476 interleukin-10 in peripheral blood immune cells during early pregnancy in dairy cows. *J*  
1477 *Reprod Dev.* 2012; **58**(1):84-90.

1478

1479 [24] Oliveira JF, Henkes LE, Ashley RL, Purcell SH, Smirnova NP, Veeramachaneni  
1480 DNR, Anthony RV, Hansen TR. Expression of interferon (IFN)-stimulated genes in ex-  
1481 trauterine tissues during early pregnancy in sheep is the consequence of endocrine IFN-t  
1482 release from the uterine vein. *Endocrinology* 2008; **149**:1252–1259.

1483

1484 [25] Araújo ER, Sponchiado M, Pugliesi G, Van Hoeck V, Mesquita FS, Membrive  
1485 CMB, Binelli M. Spatio-specific regulation of endocrine-responsive gene transcription  
1486 by periovulatory endocrine profiles in the bovine reproductive tract. *Reprod. Fertil.* 2016;  
1487 **28**: 1533–1544.

1488

1489 [26] Pfaffl M W. A new mathematical model for relative quantification in real-time RT-  
1490 PCR. *Nucleic Acids Res.* 2001; **29**:e45.  
1491

1492 [27] Melo GD, Pinto LMF, Rocha CC, Motta IG, Silva LA, da Silveira JC, Gonella-Diaza  
1493 AM, Binelli M, Pugliesi G. Type I interferon receptors and interferon- $\tau$ -stimulated genes  
1494 in peripheral blood mononuclear cells and polymorphonuclear leucocytes during early  
1495 pregnancy in beef heifers. *Reprod Fertil Dev.* 2020; **32**(11):953-966.  
1496

1497 [28] Gifford CA, Racicot K, Clark DS, Austin KJ, Hansen TR, Lucy MC, Davies CJ, Ott  
1498 TL. Regulation of interferon-stimulated genes in peripheral blood leukocytes in pregnant  
1499 and bred, nonpregnant dairy cows. *J Dairy Sci.* 2007; **90**(1):274-80.  
1500

1501 [29] Yankey SJ, Hicks BA, Carnahan KG, Assiri AM, Sinor SJ, Kodali K, Stellflug JN,  
1502 Stellflug JN, Ott TL. Expression of the antiviral protein Mx in peripheral blood mononu-  
1503 clear cells of pregnant and bred, non-pregnant ewes. *J Endocrinol.* 2001; **170**(2):R7-11.  
1504

1505 [30] Rashid MB, Talukder AK, Kusama K, Haneda S, Takedomi T, Yoshino H, Moriyasu  
1506 S, Matsui M, Shimada M, Imakawa K, Miyamoto A. Evidence that interferon-tau secreted  
1507 from Day-7 embryo *in vivo* generates anti-inflammatory immune response in the bovine  
1508 uterus. *Biochem Biophys Res Commun.* 2018; **500**(4):879-884.  
1509

1510 [31] Wegmann TG, Lin H, Guilbert L, Mosmann TR. Bidirectional cytokine interactions  
1511 in the maternal-fetal relationship: is successful pregnancy a TH2 phenomenon? *Immunol*  
1512 *Today.* 1993; **14**(7):353-6.  
1513

1514 [32] Sykes L, MacIntyre DA, Yap XJ, Ponnampalam S, Teoh TG, Bennett PR. Changes  
1515 in the Th1:Th2 cytokine bias in pregnancy and the effects of the anti-inflammatory cy-  
1516 clopenteone prostaglandin 15-deoxy- $\Delta$ (12,14)-prostaglandin J2. *Mediators Inflamm.*  
1517 2012; 2012:416739.

1518

1519 [33] Fiorenza MF, Marey MA, Rashid MB, Zinnah MA, Ma D, Morillo VA, Kusama K,  
1520 Shimada M, Imakawa K, Antoniazzi AQ, Miyamoto A. Neutrophils recognize and am-  
1521 plify IFNT signals derived from day 7 bovine embryo for stimulation of ISGs expression  
1522 *in vitro*: A possible implication for the early maternal recognition of pregnancy. *Biochem*  
1523 *Biophys Res Commun.* 2021; **553**:37-43.

1524

1525 [34] Sponchiado M, Gonella-Diaza AM, Rocha CC, Lo Turco EG, Pugliesi G, Leroy  
1526 JLMR, Binelli M. The pre-hatching bovine embryo transforms the uterine luminal me-  
1527 tabolite composition *in vivo*. *Sci Rep.* 2019; **9**:8354.

1528

1529 [35] Ledgard AM, Lee RS, Peterson AJ. Bovine endometrial legumain and TIMP-2 reg-  
1530 ulation in response to presence of a conceptus. *Mol Reprod Dev.* 2009; **76**(1):65-74.

1531

1532 [36] Berendt FJ, Frohlich T, Schmidt SEM, Reichenbach H-D, Wolf E,  
1533 Arnold GJ. Holistic differential analysis of embryo-induced alterations in the proteome  
1534 of bovine endometrium in the preattachment period. *Proteomics.* 2005; **5**:2551–2560.

1535

1536 [37] Watson AL, Skepper JN, Jauniaux E, Burton GJ. Susceptibility of human placental  
1537 syncytiotrophoblastic mitochondria to oxygen-mediated damage in relation to gestational  
1538 age. *J Clin Endo Metab.* 1998; **83**:1697–1705.

1539 [38] Thatcher WW, Meyer MD, Danet-Desnoyers G. Maternal recognition of pregnancy.  
1540 *J. Reprod. Fertil. Suppl.* 1995; **49**:15–28.  
1541

1542 [39] Roberts RM, Chen Y, Ezashi T, Walker AM. Interferons and the maternal–conceptus  
1543 dialog in mammals. *Semin. Cell Dev. Biol.* 2008; **19**:170–177.  
1544

1545 [40] Toji N, Shigeno S, Kizaki K, Koshi K, Matsuda H, Hashiyada Y, Imai K, Takahashi  
1546 T, Ishiguro-Oonuma T, Hashizume K. Evaluation of interferon-stimulated genes in pe-  
1547 ripheral blood granulocytes as sensitive responders to bovine early conceptus signals. *Vet*  
1548 *J.* 2017; **229**:37-44.  
1549

1550 [41] Han H, Austin K J, Rempel LA, Hansen TR. Low blood ISG15 mRNA and progesterone  
1551 levels are predictive of non-pregnant dairy cows. *J. Endocrinol.* 2006; **191**: 505–  
1552 512.  
1553

1554 [42] Green JC, Okamura CS, Poock SE, Lucy MC. Measurement of interferon-tau (IFN-  
1555 tau) stimulated gene expression in blood leukocytes for pregnancy diagnosis within 18-  
1556 20d after insemination in dairy cattle. *Anim Reprod Sci.* 2010; **121**(1-2):24-33.  
1557

1558 [43] Berg DK, Van Leeuwen J, Beaumont S, Berg M, Pfeffer P.L. Embryo loss in cattle  
1559 between days 7 and 16 of pregnancy. *Theriogenology* 2010; **73**:250–260.  
1560

1561 [44] National Academies of Sciences, Engineering, and Medicine. Nutrient Requirements  
1562 of Beef Cattle: Eighth Revised Edition. Washington, DC: The National Academies Press.  
1563 2016.

1564 [45] Gray CA, Abbey CA, Beremand PD, Choi Y, Farmer JL, Adelson DL, Thomas TL,  
1565 Bazer FW, Spencer TE. Identification of endometrial genes regulated by early pregnancy,  
1566 progesterone, and interferon tau in the ovine uterus. *Biol Reprod.* 2006; **74**(2):383-94.  
1567

1568 [46] Rocha CC, Martins T, Silva FACC, Sponchiado M, Pohler KG, Binelli M. Viperin  
1569 (RSAD2) gene expression in peripheral blood mononuclear cells of pregnant crossbred  
1570 beef cows is altered by *Bos indicus* genetics. *Theriogenology.* 2023; **209**:226-233.  
1571

1572 [47] Yoshino H, Toji N, Sasaki K, Koshi K, Yamagishi N, Takahashi T, Ishiguro-  
1573 Oonuma T, Matsuda H, Yamanouchi T, Hashiyada Y, Imai K, Izaike Y, Kizaki K,  
1574 Hashizume K. A predictive threshold value for the diagnosis of early pregnancy in cows  
1575 using interferon-stimulated genes in granulocytes. *Theriogenology.* 2018; **107**:188-193.  
1576

1577 [48] Pugliesi G, Guimarães da Silva A, Viana JHM, Siqueira LGB. Review: Current sta-  
1578 tus of corpus luteum assessment by Doppler ultrasonography to diagnose non-pregnancy  
1579 and select embryo recipients in cattle. *Animal.* 2023; 1:100752.  
1580

1581 [49] Spencer TE, Forde N, Lonergan P. Insights into conceptus elongation and establish-  
1582 ment of pregnancy in ruminants. *Reprod Fertil Dev.* 2016; **29**(1):84-100.  
1583

1584 [50] Ferraz PA, Filho CASG, Rocha CC, Neto AL, de Andrade Bruni G, Oshiro TSI,  
1585 Baruselli PS, Lima FS, Pugliesi G. Feasibility and accuracy of using different methods to  
1586 detect pregnancy by conceptus-stimulated genes in dairy cattle. *JDS Commun.* 2021;  
1587 **2**(3):153-158.  
1588

1589           **3.6 FINAL CONSIDERATIONS**

1590           The development of methods for early detection of pregnancy or embryonic/fetal  
1591 mortality can contribute to improving the efficiency of reproductive programs in cattle  
1592 worldwide. In this context, modulating the maternal immune system during early preg-  
1593 nancy has been reported as one of the main factors for gestational success and a signifi-  
1594 cant cause of embryonic loss.

1595           In the first studies (*Experiments 1 and 2*) described in Chapter 2, we initially  
1596 sought to understand the response of immune cells from *Bos taurus indicus* heifers, stim-  
1597 ulated by treatments with exogenous IFN- $\tau$ , or conditioned medium by the conceptus,  
1598 through an *in vitro* cell culture system. In this model, we identified that the presence of a  
1599 viable bovine conceptus modulates maternal immunological patterns in the uterus in favor  
1600 of a TH2 anti-inflammatory response, through the downregulation of the cytokine *IL1 $\beta$* ,  
1601 favoring the establishment of pregnancy. However, further studies are needed to elucidate  
1602 which molecules secreted or induced by the conceptus, in addition to IFN- $\tau$ , may be in-  
1603 ducing these maternal immune responses. Furthermore, we also observed that both treat-  
1604 ments induced the expression of classic and novel ISGs, with *ISG15* and *RSAD2* being  
1605 the most stimulated genes in both cell types, indicating that these markers have the po-  
1606 tential to be evaluated for their accuracy as early pregnancy predictors on day 20 post-  
1607 TAI. These findings may influence the improvement of molecular-based tests using ISGs  
1608 to detect pregnancy in cattle, in addition to generating improvements in production sys-  
1609 tems, especially those that use fixed-time programs.

1610           Although several reports describe a more pronounced response of PMNs to stim-  
1611 ulation with IFN- $\tau$ , in the present study, we found no significant difference in the expres-  
1612 sion of ISGs between cell groups. Therefore, the response of PMNs was similar to that  
1613 described in PBMCs, leading us to believe that both groups of cells can be used with

1614 similar efficacy for the diagnosis of pregnancy. Therefore, based on the results obtained  
1615 in the *in vitro* studies, we conducted an accuracy experiment (*Experiment 3*) using only  
1616 PMNs.

1617         To better select circulating transcripts for use as potential markers, larger numbers  
1618 of animals under field conditions are necessary. For this reason, we conducted an *in vivo*  
1619 study, where we analyzed the accuracy of the ISGs selected in the *Experiments 1 and 2*,  
1620 as early pregnancy predictors in bovine females of different parity categories. Although  
1621 all ISGs evaluated were classified as accurate predictors of pregnancy on D20 post-TAI,  
1622 the use of these genes for pregnancy diagnosis still needs to overcome some barriers for  
1623 practical application in the area. Firstly, although the expression of ISGs when associated  
1624 with Doppler-US has generated satisfactory accuracy, the considerable number of false  
1625 positive results, and mainly false negatives, significantly reduces the precision of the  
1626 technique. Secondly, the technique for isolating immune cells and subsequent analysis of  
1627 transcripts by qPCR is expensive and time-consuming, which would not justify the choice  
1628 of this technique over others, using current methodologies. Furthermore, it is known that  
1629 there is a difference in the expression patterns of ISGs between younger animals (heifers)  
1630 compared to older (cows), making the use of this method even more limited.

1631         Despite the obstacles raised, we believe that research using ISGs as pregnancy  
1632 markers should be encouraged, as these genes respond early to a direct stimulus from the  
1633 conceptus during early pregnancy. Furthermore, technology is constantly evolving, and  
1634 today more sensitive and faster techniques are available, such as gene expression analysis  
1635 by digital PCR, and more efficient forms of cell isolation. Moreover, the abundance of  
1636 ISGs in the whole blood fraction could be explored, as we conclude that the cellular frac-  
1637 tion does not affect the expression of these genes. This opens the way for the development

1638 of rapid commercial kits, as well as metabolite analysis techniques through metabolom-  
1639 ics, which can facilitate the application of this technique in the field.  
1640