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**UNIVERSIDADE ESTADUAL PAULISTA
“JÚLIO DE MESQUITA FILHO”
FACULDADE DE MEDICINA DE BOTUCATU**

Jofer Andree Zamame Ramirez

**Autofagia em células tumorais: um mecanismo de
carcinogênese e resistência aos quimioterápicos**

Botucatu

2017

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Jofer Andree Zamame Ramirez

Dissertação apresentada à Faculdade de Medicina, Universidade Estadual Paulista “Júlio de Mesquita Filho”, Câmpus de Botucatu, como parte dos requisitos necessários para a obtenção do título de Mestre em Patologia.

Orientador: Prof. Dr. Ramon Kaneno

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“Aprender a leer es lo más importante que me ha pasado en la vida”.

Mario Vargas Llosa

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Autophagy in tumor cells: a mechanism of carcinogenesis and resistance to chemotherapeutic agents

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Autophagy in tumor cells: a mechanism of carcinogenesis and resistance to chemotherapeutic agents

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Abstract

Autophagy is a dynamic physiological macromolecular process, whereby intracellular substrates are exposed to lysosomes for degradation and recycling of damaged organelles, alleviating cellular stress conditions. Several studies have shown that autophagy plays a critical role in tumoral cell survival, performing a protective role by correcting carcinogenic damages. However, this physiological process can be subverted in some cells, leading to the promotion of carcinogenesis or allowing cell escape by increasing its resistance to chemotherapeutics. This review covers the basic mechanisms and genes involved in autophagy as well as the controversial findings on their role tumor cells; we also reviewed the processes by which drug resistance may be determined, for a better understanding of how autophagy works and how it can be handled as an antitumor therapeutic intervention.

1. Introduction: Autophagy as a physiological process

The term autophagy was first used by deDuve in 1963(1), and was defined by Klionsky in 2003 as "self-eating" at subcellular level(2). Since then, many scientists have investigated the mechanisms that trigger this process. Current definition of autophagy is a conserved cellular degradation process, in which portions of cytosol and organelles are sequestered into a double-membrane vesicle, called autophagosome and fused with lysosome for breakdown and eventual recycling of resulting macromolecules(3). This process is required for the maintenance of cellular homeostasis and generation of amino acids to sustain viability during periods of nutrient privation, or under other kinds of metabolic stress like intracellular bacterial or viral infections, mutations or exposition to drugs(4-6).

The autophagy process has three different forms: macroautophagy, microautophagy, and chaperone-mediated autophagy(3). Macroautophagy, which is often simply referred as autophagy, is the most characterized form and has been extensively investigated (4, 6, 7), being defined as the sequestration of a bulk of cytoplasm and organelles in a double-membrane vesicle called phagophore. This phagophore is originated from endoplasmic reticulum and formed by expansion of autophagosome, and fuses with lysosome, forming autophagolysosome. Inside autophagolysosome, damaged organelles are degraded by lysosomal hydrolases, and finally, the resultant *ATP* and peptides are used to keep the cell viability (3, 8). In contrast, microautophagy is characterized by direct uptake of cytoplasmic substrates by invagination of lysosomal membrane, while chaperone-mediated autophagy occurs by shuttling soluble proteins into the lysosome, via lysosomal chaperone proteins such as *hsc70*(3, 8).

Survival and homeostatic functions of autophagy have been evolutionarily conserved from yeast to mammalian. When cells have rich nutrient condition, autophagy is produced

at low levels (basal autophagy), providing tissues with a cleaning mechanism, to control intracellular quality through cytoplasmic rotation and removal of damaged or unnecessary organelles (9-12). However, when a cell is under different kinds of stress, such as nutrient deprivation, hypoxia, intracellular infections (13) and exposition to drugs(14), autophagy is rapidly induced in order to maintain the amino acid pool in the cytoplasm, and cell survives possibly through protein neo-synthesis, energy production and gluconeogenesis(12), that maintain cell *ATP* production.

Autophagy involves a concerted action of highly conserved gene products and is controlled by two pathways involved in the regulation of cell growth and metabolism, the *mTOR* (mammalian target of rapamycin) and the *AMPK* (*AMP-activated protein kinase*) /*UVRAG* (*UV irradiation resistance associated gene*) signaling pathways. During normal situations (nutrient availability), *mTOR complex I* (*mTORC1*) inhibits autophagy since it phosphorylates *Atg13*, causing disaggregation of *ULK1 complex* (*ULK1*, *Atg13* and *FIP200*), required to form autophagosome. However, when *mTOR* is inhibited by dephosphorylation (caused by cell starvation), *ULK1* complex works and autophagy is induced (8). Conversely, *AMPK*, is activated during energy deprivation or hypoxia by increased conversion of *ADP* to *ATP*, and can inactivate *mTORC1*, and trigger autophagy (10, 11). In parallel, *AMPK* and *UVRAG* pathways activate the pro-autophagy kinase *Vps34* (*Class III PI3 Kinase*) by phosphorylating and recruiting *beclin-1* (proautophagy and tumor suppressor gene), which produces *PI(3)P* that is essential for to initiate autophagosomes. Autophagy triggering is illustrated in Figure 1.

4. Conclusion

Although the controversy about the action of autophagy as tumor promoter or tumor suppressor, there is evidence that its inhibition helps to kill tumor cells mostly dependent on the kind of tumor and the therapeutic agents to be associated with. Therefore, we need a better understanding of the functional relevance of autophagy in the tumor microenvironment and enhance the ongoing dialogue between the laboratory and clinical researches in order to provide a new focus on therapeutic strategy to prevent resistance and increase the effects of cancer therapies.

References

1. de Duve C. Lysosomes revisited. *European Journal of Biochemistry*. 1983;137(3):391-7. doi: 10.1111/j.1432-1033.1983.tb07841.x.
2. Klionsky DJ, Cregg JM, Dunn Jr WA, Emr SD, Sakai Y, Sandoval IV, et al. A Unified Nomenclature for Yeast Autophagy-Related Genes. *Developmental Cell*. 2003;5(4):539-45. doi: [http://dx.doi.org/10.1016/S1534-5807\(03\)00296-X](http://dx.doi.org/10.1016/S1534-5807(03)00296-X).
3. Mizushima N, Levine B, Cuervo AM, Klionsky DJ. Autophagy fights disease through cellular self-digestion. *Nature*. 2008;451(7182):1069-75. doi: 10.1038/nature06639. PubMed PMID: PMC2670399.
4. Cuervo AM. Autophagy: In sickness and in health. *Trends in Cell Biology*. 2004;14(2):70-7.
5. Ciechanover A. Proteolysis: from the lysosome to ubiquitin and the proteasome. *Nat Rev Mol Cell Biol*. 2005;6(1):79-87.
6. Shoji-Kawata S, Levine B. Autophagy, antiviral immunity, and viral countermeasures. *Biochimica et biophysica acta*. 2009;1793(9):1478-84. doi: 10.1016/j.bbamcr.2009.02.008. PubMed PMID: PMC2739265.

7. Mehrpour M, Esclatine A, Beau I, Codogno P. Overview of macroautophagy regulation in mammalian cells. *Cell Res.* 2010;20(7):748-62.
8. Eskelinen E-L, Saftig P. Autophagy: A lysosomal degradation pathway with a central role in health and disease. *Biochimica et Biophysica Acta (BBA) - Molecular Cell Research.* 2009;1793(4):664-73. doi: <http://dx.doi.org/10.1016/j.bbamcr.2008.07.014>.
9. Maycotte P, Thorburn A. Autophagy and cancer therapy. *Cancer Biology & Therapy.* 2011;11(2):127-37. doi: 10.4161/cbt.11.2.14627. PubMed PMID: PMC3047083.
10. Janku F, McConkey DJ, Hong DS, Kurzrock R. Autophagy as a target for anticancer therapy. *Nat Rev Clin Oncol.* 2011;8(9):528-39. doi: http://www.nature.com/nrclinonc/journal/v8/n9/supinfo/nrclinonc.2011.71_S1.html.
11. Mathew R, Karantza-Wadsworth V, White E. Role of autophagy in cancer. *Nat Rev Cancer.* 2007;7(12):961-7.
12. Alva AS, Gultekin SH, Baehrecke EH. Autophagy in human tumors: cell survival or death? *Cell Death Differ.* 2004;11(9):1046-8.
13. He C, Klionsky DJ. Regulation Mechanisms and Signaling Pathways of Autophagy. *Annual review of genetics.* 2009;43:67-93. doi: 10.1146/annurev-genet-102808-114910. PubMed PMID: PMC2831538.
14. Rubinsztein DC, Codogno P, Levine B. Autophagy modulation as a potential therapeutic target for diverse diseases. *Nature reviews Drug discovery.* 2012;11(9):709-30. doi: 10.1038/nrd3802. PubMed PMID: PMC3518431.
15. Takeshi Noda KS, Yoshinori Ohsumi. Yeast autophagosomes: de novo formation of a membrane structure. *TRENDS in Cell Biology* 2002;12(5):231-5.
16. Reggiori F, Klionsky DJ. Autophagosomes: biogenesis from scratch? *Current Opinion in Cell Biology.* 2005;17(4):415-22. doi: <http://dx.doi.org/10.1016/j.ceb.2005.06.007>.
17. Virgin HW, Levine B. Autophagy genes in immunity. *Nat Immunol.* 2009;10(5):461-70.
18. Filomeni G, De Zio D, Cecconi F. Oxidative stress and autophagy: the clash between damage and metabolic needs. *Cell Death and Differentiation.* 2015;22(3):377-88. doi: 10.1038/cdd.2014.150. PubMed PMID: PMC4326572.
19. Wiseman H, Halliwell B. Damage to DNA by reactive oxygen and nitrogen species: role in inflammatory disease and progression to cancer. *Biochemical Journal.* 1996;313(Pt 1):17-29. PubMed PMID: PMC1216878.
20. Gomes LC, Di Benedetto G, Scorrano L. During autophagy mitochondria elongate, are spared from degradation and sustain cell viability. *Nature cell biology.* 2011;13(5):589-98. doi: 10.1038/ncb2220. PubMed PMID: PMC3088644.
21. Ding W-X, Yin X-M. Mitophagy: mechanisms, pathophysiological roles, and analysis. *Biological chemistry.* 2012;393(7):547-64. doi: 10.1515/hsz-2012-0119. PubMed PMID: PMC3630798.
22. Li T, Kon N, Jiang L, Tan M, Ludwig T, Zhao Y, et al. Tumor suppression in the absence of p53-mediated cell cycle arrest, apoptosis, and senescence. *Cell.* 2012;149(6):1269-83. doi: 10.1016/j.cell.2012.04.026. PubMed PMID: PMC3688046.
23. Berkers Celia R, Maddocks Oliver D, Cheung Eric C, Mor I, Vousden Karen H. Metabolic Regulation by p53 Family Members. *Cell Metabolism.* 2013;18(5):617-33. doi: 10.1016/j.cmet.2013.06.019. PubMed PMID: PMC3824073.
24. Tasdemir E, Maiuri MC, Galluzzi L, Vitale I, Djavaheri-Mergny M, D'Amelio M, et al. Regulation of autophagy by cytoplasmic p53. *Nat Cell Biol.* 2008;10(6):676-87. doi: http://www.nature.com/ncb/journal/v10/n6/supinfo/ncb1730_S1.html.
25. Tom Strachan AR. Cancer Genetics, in *Human Molecular Genetics* 1999.
26. Astle MV, Hannan KM, Ng PY, Lee RS, George AJ, Hsu AK, et al. AKT induces senescence in human cells via mTORC1 and p53 in the absence of DNA damage: implications for targeting mTOR during malignancy. *Oncogene.* 2012;31(15):1949-62. doi: <http://www.nature.com/ncb/journal/v31/n15/supinfo/ncb2011394s1.html>.

27. Tasdemir E, Maiuri MC, Morselli E, Criollo A, D'Amelio M, Djavaheri-Mergny M, et al. A dual role of p53 in the control of autophagy. *Autophagy*. 2008;4(6):810-4. doi: 10.4161/auto.6486.
28. Huo Y, Cai H, Teplova I, Bowman-Colin C, Chen G, Price S, et al. Autophagy opposes p53-mediated tumor barrier to facilitate tumorigenesis in a model of PALB2-associated hereditary breast cancer. *Cancer discovery*. 2013;3(8):894-907. doi: 10.1158/2159-8290.CD-13-0011. PubMed PMID: PMC3740014.
29. Tripathi R, Ash D, Shaha C. Beclin-1–p53 interaction is crucial for cell fate determination in embryonal carcinoma cells. *Journal of Cellular and Molecular Medicine*. 2014;18(11):2275-86. doi: 10.1111/jcmm.12386. PubMed PMID: PMC4224560.
30. Bjørkøy G, Lamark T, Pankiv S, Øvervatn A, Brech A, Johansen T. Chapter 12 Monitoring Autophagic Degradation of p62/SQSTM1. *Methods in Enzymology*. Volume 452: Academic Press; 2009. p. 181-97.
31. Inami Y, Waguri S, Sakamoto A, Kouno T, Nakada K, Hino O, et al. Persistent activation of Nrf2 through p62 in hepatocellular carcinoma cells. *The Journal of Cell Biology*. 2011;193(2):275-84. doi: 10.1083/jcb.201102031. PubMed PMID: PMC3080263.
32. Mathew R, Karp C, Beaudoin B, Vuong N, Chen G, Chen H-Y, et al. Autophagy Suppresses Tumorigenesis Through Elimination of p62. *Cell*. 2009;137(6):1062-75. doi: 10.1016/j.cell.2009.03.048. PubMed PMID: PMC2802318.
33. Liang XH, Jackson S, Seaman M, Brown K, Kempkes B, Hibshoosh H, et al. Induction of autophagy and inhibition of tumorigenesis by beclin 1. *Nature*. 1999;402(6762):672-6.
34. Liu J, Xia H, Kim M, Xu L, Li Y, Zhang L, et al. Beclin1 Controls the Levels of p53 by Regulating the Deubiquitination Activity of USP10 and USP13. *Cell*. 2011;147(1):223-34. doi: 10.1016/j.cell.2011.08.037. PubMed PMID: PMC3441147.
35. Tang MKS, Kwong A, Tam K-F, Cheung ANY, Ngan HYS, Xia W, et al. BRCA1 deficiency induces protective autophagy to mitigate stress and provides a mechanism for BRCA1 haploinsufficiency in tumorigenesis. *Cancer Letters*. 346(1):139-47. doi: 10.1016/j.canlet.2013.12.026.
36. Qu X, Yu J, Bhagat G, Furuya N, Hibshoosh H, Troxel A, et al. Promotion of tumorigenesis by heterozygous disruption of the beclin 1 autophagy gene. *The Journal of Clinical Investigation*. 2003;112(12):1809-20. doi: 10.1172/JCI20039.
37. Yue Z, Jin S, Yang C, Levine AJ, Heintz N. Beclin 1, an autophagy gene essential for early embryonic development, is a haploinsufficient tumor suppressor. *Proceedings of the National Academy of Sciences of the United States of America*. 2003;100(25):15077-82. doi: 10.1073/pnas.2436255100. PubMed PMID: PMC299911.
38. Fu L-I, Cheng Y, Liu B. Beclin-1: autophagic regulator and therapeutic target in cancer. *Int J Biochem Cell Biol*. 2013;45(5):921-4. doi: 10.1016/j.biocel.2013.02.007. PubMed PMID: 23420005.
39. White E. The role for autophagy in cancer. *The Journal of Clinical Investigation*. 2015;125(1):42-6. doi: 10.1172/JCI73941.
40. He S, Zhao Z, Yang Y, O'Connell D, Zhang X, Oh S, et al. Truncating mutation in the autophagy gene UVRAG confers oncogenic properties and chemosensitivity in colorectal cancers. *Nature Communications*. 2015;6:7839. doi: 10.1038/ncomms8839.
41. Pylayeva-Gupta Y, Grabocka E, Bar-Sagi D. RAS oncogenes: weaving a tumorigenic web. *Nature reviews Cancer*. 2011;11(11):761-74. doi: 10.1038/nrc3106. PubMed PMID: PMC3632399.
42. Downward J. Targeting RAS signalling pathways in cancer therapy. *Nat Rev Cancer*. 2003;3(1):11-22.
43. Ge J, Chen Z, Huang J, Chen J, Yuan W, Deng Z, et al. Upregulation of Autophagy-Related Gene-5 (ATG-5) Is Associated with Chemoresistance in Human Gastric Cancer. *PLOS ONE*. 2014;9(10):e110293. doi: 10.1371/journal.pone.0110293.

44. Wei H, Wei S, Gan B, Peng X, Zou W, Guan J-L. Suppression of autophagy by FIP200 deletion inhibits mammary tumorigenesis. *Genes & Development*. 2011;25(14):1510-27. doi: 10.1101/gad.2051011. PubMed PMID: PMC3143941.
45. Othman EQG, Kaur G, Mutee AF, Tengku Muhammad TS, Tan ML. Immunohistochemical expression of MAP1LC3A and MAP1LC3B protein in breast carcinoma tissues. *Journal of Clinical Laboratory Analysis*. 2009;23(4):249-58. doi: 10.1002/jcla.20309.
46. Yun M, Bai H-Y, Zhang J-X, Rong J, Weng H-W, Zheng Z-S, et al. ULK1: A Promising Biomarker in Predicting Poor Prognosis and Therapeutic Response in Human Nasopharyngeal Carcinoma. *PLOS ONE*. 2015;10(2):e0117375. doi: 10.1371/journal.pone.0117375.
47. Cancer multidrug resistance. *Nat Biotech*. 2000;18:18-20.
48. Abdullah LN, Chow EK-H. Mechanisms of chemoresistance in cancer stem cells. *Clinical and Translational Medicine*. 2013;2:3-. doi: 10.1186/2001-1326-2-3. PubMed PMID: PMC3565873.
49. Guan J-L, Simon AK, Prescott M, Menendez JA, Liu F, Wang F, et al. Autophagy in stem cells. *Autophagy*. 2013;9(6):830-49. doi: 10.4161/auto.24132. PubMed PMID: PMC3672294.
50. Kantara C, O'Connell M, Sarkar S, Moya S, Ullrich R, Singh P. Curcumin Promotes Autophagic Survival of a Sub-Set of Colon Cancer Stem Cells, which are Ablated by DCLK1-siRNA. *Cancer research*. 2014;74(9):2487-98. doi: 10.1158/0008-5472.CAN-13-3536. PubMed PMID: PMC4013529.
51. Sanchez CG, Penfornis P, Oskowitz AZ, Boonjindasup AG, Cai DZ, Dhule SS, et al. Activation of autophagy in mesenchymal stem cells provides tumor stromal support. *Carcinogenesis*. 2011;32(7):964-72. doi: 10.1093/carcin/bgr029. PubMed PMID: PMC3128555.
52. Gong C, Bauvy C, Tonelli G, Yue W, Deloménie C, Nicolas V, et al. Beclin 1 and autophagy are required for the tumorigenicity of breast cancer stem-like/progenitor cells. *Oncogene*. 2013;32(18):2261-72. doi: 10.1038/onc.2012.252. PubMed PMID: PMC3679409.
53. Fujii S, Mitsunaga S, Yamazaki M, Hasebe T, Ishii G, Kojima M, et al. Autophagy is activated in pancreatic cancer cells and correlates with poor patient outcome. *Cancer Science*. 2008;99(9):1813-9. doi: 10.1111/j.1349-7006.2008.00893.x.
54. Yang M-C, Wang H-C, Hou Y-C, Tung H-L, Chiu T-J, Shan Y-S. Blockade of autophagy reduces pancreatic cancer stem cell activity and potentiates the tumoricidal effect of gemcitabine. *Molecular Cancer*. 2015;14:179. doi: 10.1186/s12943-015-0449-3. PubMed PMID: PMC4603764.
55. Rangwala R, Chang YC, Hu J, Algazy KM, Evans TL, Fecher LA, et al. Combined MTOR and autophagy inhibition: Phase I trial of hydroxychloroquine and temsirolimus in patients with advanced solid tumors and melanoma. *Autophagy*. 2014;10(8):1391-402. doi: 10.4161/auto.29119. PubMed PMID: PMC4203516.
56. Institute NC. Akt Inhibitor MK2206 and Hydroxychloroquine in Treating Patients With Advanced Solid Tumors, Melanoma, Prostate or Kidney Cancer. *ClinicalTrials.gov* [Internet]: Bethesda (MD): National Library of Medicine (US). ; 2011 [cited 2017 Mar 31]. <https://clinicaltrials.gov/ct2/show/NCT01480154>.
57. Hospital. MG. Hydroxychloroquine With or Without Erlotinib in Advanced Non-small Cell Lung Cancer. *ClinicalTrials.gov* [Internet]: Bethesda (MD): National Library of Medicine (US). 2009 [cited 2017 Mar 31]. <https://clinicaltrials.gov/ct2/show/NCT01026844>.
58. Hospital SKWH-SM. A Phase II Trial of Combined Hydroxychloroquine and Sirolimus in Soft Tissue Sarcoma *ClinicalTrials.gov* [Internet]. Bethesda (MD): National Library of Medicine (US). 2012 [cited 2017 Mar 31]. <https://clinicaltrials.gov/ct2/show/NCT01842594>.
59. Health N. Hydroxychloroquine in Untreated B-CLL Patients *ClinicalTrials.gov* [Internet]. Bethesda (MD): National Library of Medicine (US). 2008 [cited 2017 Mar 31]. <https://clinicaltrials.gov/ct2/show/NCT00771056>.
60. Pennsylvania ACCotUo. FOLFOX/Bevacizumab/Hydroxychloroquine (HCQ) in Colorectal Cancer *ClinicalTrials.gov* [Internet]. : Bethesda (MD): National Library of Medicine (US). ; 2010 [cited 2017 Mar 31]. <https://clinicaltrials.gov/ct2/show/NCT01206530>.