

**UNIVERSIDADE ESTADUAL PAULISTA – UNESP
CÂMPUS JABOTICABAL**

**EFEITO DA UTILIZAÇÃO DE DIFERENTES MATRIZES
GENÔMICAS E PARENTESCO NA AVALIAÇÃO GENÉTICA
DE BOVINOS DE CORTE**

**Michel Marques Farah
Zootecnista**

2014

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Michel Marques Farah

Orientador: Prof. Dr. Ricardo da Fonseca

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TÍTULO: EFEITO DA UTILIZAÇÃO DE DIFERENTES MATRIZES GENÔMICAS E PARENTESCO NA AVALIAÇÃO GENÉTICA DE BOVINOS DE CORTE

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DADOS CURRICULARES DO AUTOR

Michel Marques Farah, filho de Nicolau Wladimir Farah e Elide Marques Farah, nasceu em São Paulo – SP, em 24 de setembro de 1982. Em 2002, iniciou curso de graduação em Zootecnia, pela Universidade Federal dos Vales do Jequitinhonha e Mucuri - MG, graduando-se em julho de 2007. Em março de 2008 iniciou curso de Mestrado em Zootecnia, na área de Melhoramento Animal, pela Universidade Federal dos Vales do Jequitinhonha – MG. Em 15 de julho de 2010 tornou-se Mestre em Zootecnia. Em agosto de 2010 iniciou curso de Doutorado em Genética e Melhoramento Animal, pela Universidade Estadual Paulista “Júlio de Mesquita Filho” – SP. Realizou o programa de sanduiche na University of Queensland - Austrália durante o ano de 2013.

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À minha família
À Camila e a família dela

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Efeito da utilização de diferentes matrizes genômicas de parentesco na avaliação genética de bovinos de corte

RESUMO - No melhoramento genético animal a forma tradicional de realizar seleção é com base no fenótipo dos indivíduos e na informação do parentesco entre estes, porém é um processo lento, sendo assim, programas de melhoramento estão procurando identificar os genes responsáveis pela característica de interesse e assim realizar a seleção dos animais que carregam a informação desejada. Com as informações dos indivíduos genotipados, tornou-se possível a utilização da informação de genes idênticos em estado tornando viável a utilização de uma matriz de parentesco (G) permitindo aumentar a precisão das avaliações genéticas, porém, devido à dificuldade de se obter o genótipo de todos os animais de uma população, foi proposto um método que realiza a integração da matriz G com a matriz de parentesco (A) em uma matriz de parentesco-genômica (H). Embora tenham trabalhos que indiquem uma similaridade no progresso genético utilizando estas diferentes matrizes é importante a avaliação da contribuição da avaliação genômica nos processos de avaliação genética em populações com estruturas de parentesco diferentes, bem como avaliar a metodologia de seleção genômica em populações multirraciais, a fim de atender o sistema de criação de animais cruzados. Assim, o objetivo geral deste trabalho foi estudar os efeitos da informação genômica na avaliação genética animal por meio de diferentes matrizes genômicas, utilizando dados de bovinos de corte com diferentes estruturas populacionais e composições raciais. Primeiramente avaliou-se 3 diferentes metodologias de se obter a matriz H, com a frequência alélica observada (HGOF), menor frequência alélica (HGMF) e uma frequência de 0,5 para todos os SNPs (HG50). Foram feitas comparações entre estas matrizes genômicas e a matriz de parentesco tradicional (A) utilizando uma população de 1695 animais da raça Brahman (BB). De acordo com os resultados obtidos, a HGOF foi a matriz que apresentou melhor similaridade com a matriz A. Porém, as maiores diferenças foram encontradas na classificação dos animais, quando avaliou-se a classificação dos animais utilizando as diferentes matrizes, todas as matrizes genômicas apresentaram diferente classificação da matriz A. Outro trabalho foi feito para investigar a possibilidade de aumentar a acurácia da seleção genômica em animais da raça Tropical Composite (TC), obtida pelo cruzamento principalmente da raça Brahman com animais *Bos taurus*, usando dados de BB. Assim foram criadas duas matrizes genômicas, uma utilizando apenas as informações genômicas da população de TC (GRM_{SB}) e outra utilizando a informação da contribuição da raça BB em cada animal TC (GRM_{XB}). Ambas as matrizes estimaram parâmetros genéticos similares mas maiores que quando utilizando a matriz A. Porém, a GRM_{SB} apresentou maiores acurácias na predição dos valores genéticos, principalmente quando aumenta o uso da informação de BB na população de TC. De maneira geral, o uso da informação genômica para criar matrizes de parentesco contribui para melhorar a predição de relacionamento entre os indivíduos e é uma importante ferramenta para uso em populações de gado composto.

Palavras-chave: *bos indicus*, coeficiente de parentesco, gado de corte, parâmetros genéticos, seleção genômica, validação cruzada

Effect of different genomic relationship matrices on genetic evaluation of beef cattle

ABSTRACT - In animal breeding methodologies, the traditional method of performing selection is based on the phenotype of individuals and information of relationship between them, but it is a slow process, so breeding programs are trying to identify the genes responsible for the trait of interest and thus achieve selection of animals that carry the interesting genes. With the information of genotyped individuals, it became possible to use the information of genes identical in state making it feasible to use a relationship matrix (G) which increase the accuracy of genetic evaluations, however, due to difficulty of obtaining the genotype of all animals in a population, we propose a method that performs the integration of the G matrix with the relationship matrix (A) in a pedigree-genomic relationship matrix (H). Although studies indicating a similarity in genetic progress using these matrices is important to evaluate the contribution of genomic evaluation in the process of genetic evaluation in populations with different structures of kinship, as well as evaluating the methodology of genomic selection in multiracial populations in order to cater to the creation of crossbred system. Thus the objective of this work was to study the effects of genomic information in genetic evaluation through different genomic arrays using data from beef cattle with different population structures and racial compositions. First we evaluated three different methods of obtaining the H matrix with the observed allele frequency (HGOF), lower allele frequency (HGMF) and a frequency of 0.5 for all SNPs (HG50). Comparisons between these genomic arrays and traditional kinship (A) using a population of 1695 animals breed Brahman (BB) matrix were made. According to the results, the HGOF was a matrix that showed the greatest similarity to the matrix A but the greatest differences were found in the classification of animals, when we evaluated the classification of animals using different matrices, all matrices showed different genomic rank of the matrix A. Another study was done to investigate the possibility of increasing the accuracy of genomic selection in animals breed Tropical Composite (TC), which is a breed obtained by crossing Brahman mainly with *Bos taurus*, using data from BB. So two genomic matrices, one using only the genomic information of the population of TC (GRM_{SB}) and another one using the information of the contribution of the BB breed in each animal TC (GRM_{XB}) were created. Both similar but larger matrices estimated genetic parameters when using the matrix A. However, GRM_{SB} showed higher accuracies in the prediction of breeding values, especially when increasing the use of information in the BB population of TC. In general, the use of genomic information to create relationship matrices contributes to an increase of the prediction of relationship between individuals and is an important tool for use in multibreed cattle populations.

Key words: *bos indicus*, relationship coefficient, beef cattle, genetic parameters, genomic selection, cross-validation

CAPÍTULO 1 - CONSIDERAÇÕES GERAIS

INTRODUÇÃO

Tradicionalmente a seleção de características de interesse econômico são realizadas com base no valor fenotípico dos indivíduos e na informação do parentesco entre os animais. Esta seleção é eficiente, porém o processo demanda tempo, principalmente para características que são medidas em apenas um sexo, como produção de leite, ou características medidas após o abate dos animais, como a qualidade da carne, ou ainda medidas mensuradas no final da vida do indivíduo, por exemplo, longevidade. Assim, para realizar programas de melhoramento para estas características, pesquisadores buscam identificar os genes que afetam tais características e a seleção de animais que carregam os alelos desejáveis (MEUWISSEN; GODDARD, 1996).

Os projetos de sequenciamento e geração de informações genômicas de alta qualidade estão cada vez mais sendo utilizados no melhoramento genético animal. A quantidade de nucleotídeos de polimorfismos únicos (SNP) identificados cresce rapidamente em bovinos. E com isso vem crescendo também a quantidade de pesquisadores interessados em utilizar as informações genômicas nos programas de melhoramento genético animal (MEUWISSEN; GODDARD, 1996; CHRISTENSEN; LUND, 2010; GIANOLA *et al.*, 2010; HAYES *et al.*, 2010).

Com o avanço destas novas tecnologias, os pesquisadores também estão procurando novas técnicas de incorporação desta informação na estimação do parentesco dos animais, formando uma matriz de relacionamento genômico (G). A utilização de G nas avaliações genéticas, permitiu aumentar a precisão da avaliação genética dos animais, criando o conceito de Seleção Genômica (SG). Segundo Meuwissen *et al.* (2001), a SG aumenta a taxa de ganho genético e reduz o custo do teste de progênie, permitindo aos criadores pré-selecionar animais que tenham herdado segmentos cromossômicos de maior mérito. Estes valores genéticos podem ser obtidos usando o modelo de equações de modelos mistos (EMM) com a matriz de parentesco A, substituída pela matriz G.

De modo geral, G inclui informações genômicas de poucos animais, devido a impossibilidade de genotipar toda a população ou de se obter o genótipo de alguns

ancestrais. Christensen; Lund (2010) propuseram um método para a predição do genoma de animais não genotipados, tornando possível a integração de todas as informações genômicas no pedigree e levando ao aumento na precisão das estimativas dos componentes de variância.

Entretanto, o método proposto por Christensen; Lund (2010) é complexo, exigindo alta demanda por equipamentos com grande capacidade de processamento e memória. Por fim, outros trabalhos, como em Forni et al. (2011) e Legarra et al. (2009), procuraram uma maneira de integrar esta informação genômica com a informação de parentesco, com o objetivo de aumentar a quantidade de informações no pedigree e assim buscar uma melhor estimativa dos componentes genéticos dos indivíduos e da população.

Além de proporcionar parentesco mais acurado entre os indivíduos, a utilização da informação genômica pode auxiliar na avaliação genética de animais compostos por duas ou mais raças, levando à estimação de relacionamento entre os indivíduos mais acurada devido a informação de parentesco e a real proporção de cada raça no animal que compõe a população em análise.

Já no Brasil, a avaliação genética considerando uma população multirracial pode ser de interesse para os programas de melhoramento genético animal pois aproximadamente 80% da população de bovinos que é destinado ao corte, praticamente 80% da população é composta por raças zebu ou cruzamento de zebu (JOSAHKIAN, 2000). Em consequência, existe um grande número de subpopulações de vários tamanhos, com composição racial *Bos indicus* x *Bos indicus* e *Bos indicus* x *Bos taurus*, as quais se enquadram na descrição de população multirracial (ELZO & BORJAS, 2004).

Assim, o objetivo geral deste trabalho foi estudar os efeitos da informação genômica na avaliação genética animal por meio de diferentes matrizes genômicas utilizando dados de bovinos de corte com diferentes estruturas populacionais.

Para isto foram feitos dois trabalhos, no primeiro objetivou-se avaliar a integração entre as matrizes genômicas, obtidas por variações nas frequências alélicas, e as informações de pedigree formando diferentes matrizes de parentesco.

No segundo trabalho o objetivo principal foi desenvolver métodos de predição genômica para populações cruzadas, utilizando informações da proporção de semelhança genética entre animais compostos e a principal raça formadora.

REVISÃO DE LITERATURA

Predição dos Valores Genéticos

O valor genético de um indivíduo consiste no mérito genético que pode ser transmitido às progênes deste indivíduo. De acordo com Henderson (1975) há diversas maneiras de se predizer este valor genético, sendo o Melhor Preditor Linear Não-Viesado (BLUP) o método mais utilizado pelos melhoristas para a predição dos valores genéticos dos animais.

Este método de predição envolve todos os indivíduos identificados na estrutura genealógica da população para estabelecer os relacionamentos genéticos. Indivíduos relacionados tem uma proporção maior de genes em comum relacionada ao grau de parentesco, que é informado por meio de uma inversa da matriz de parentesco (PEREIRA, 2012), possibilitando assim, a metodologia Equações de Modelos Mistos (MME) para a obtenção do BLUP dos valores genéticos dos animais, proposta por Henderson (1975).

Diversos modelos podem ser especificados para as MME, dependendo da aplicação das características avaliadas e estrutura de dados desenvolvidos, como o Modelo Animal, Modelo Animal Reduzido e Modelo Touro entre outros (PEREIRA, 2012).

A equação básica que descreve estes modelos é:

$$y = X\beta + Za + e$$

em que:

y é um vetor de observações;

β é um vetor de efeitos fixos desconhecidos;

X é uma matriz de incidência dos efeitos fixos;

a é um vetor de efeitos aleatórios genéticos desconhecidos para todos os indivíduos envolvidos na análise;

Z é uma matriz de incidência dos efeitos aleatórios;

e é um vetor de efeitos aleatórios residuais desconhecidos.

Para o modelo Touro, cada reprodutor tem uma equação e o desempenho de todas as progênes ligadas de um determinado reprodutor estão ligados a este por

meio da matriz Z . Já no modelo Animal todos os indivíduos apresentam uma equação e a matriz Z é uma matriz de incidência, associando cada observação ao indivíduo que a produziu. Outra diferença básica entre estes dois modelos é que o primeiro estima a Diferença Esperada na Progenie (DEP) enquanto o segundo estima o valor genético do indivíduo, que corresponde ao dobro da DEP.

O Modelo Animal mudou a forma de pensar na interpretação da covariância entre parentes para a estrutura de modelo linear, onde se determinam variâncias diretamente pelo ajustamento correspondente aos efeitos aleatórios do modelo de análise. As covariâncias entre os efeitos aleatórios para parentes são levadas em conta através da especificação da matriz de variâncias dos efeitos aleatórios. A variância genética aditiva é estimada como a variância do mérito genético aditivo dos animais. Da mesma forma, os componentes genéticos não-aditivos podem ser estimados pelo ajustamento de um efeito aleatório correspondente, como a dominância ou efeito genético materno, para cada animal (VAYEGO, 2007).

A partir do modelo de predição do valor genético dos indivíduos desenvolveu-se novas metodologias de seleção, com base nas informações fenotípicas e correlações entre os indivíduos, porém, com o avanço das tecnologias e possibilidade de conhecer o genótipo dos animais uma nova ferramenta está atualmente disponível e amplamente utilizada pelos pesquisadores, conhecida como Seleção Genômica.

Seleção Genômica

Seleção genômica (SG) é um método que usa a informação genômica para prever os valores genéticos e os indivíduos candidatos à seleção nos programas de melhoramento genético (CLARK et al., 2012). A SG foi proposta inicialmente por Meuwissen et al. (2001) que tem como principal objetivo a utilização direta das informações de marcadores moleculares e informações do DNA na seleção.

Este método apresenta uma grande vantagem em relação à seleção tradicional, pois permite uma alta eficiência seletiva, principalmente em características de difícil mensuração, como características de carcaça, fertilidade, longevidade e eficiência alimentar, pois são características com alto custo para medir, medidas apenas em um sexo ou necessita de informações de seus parentes para obter estimativa do animal (BOLORMAA et al., 2013a). A SG também pode ser definida como seleção simultânea para centenas ou milhares de marcadores, os quais

cobrem o genoma de uma maneira densa fazendo com que os genes de uma característica quantitativa estejam em desequilíbrio de ligação com pelo menos uma parte dos marcadores utilizados (VANRADEN, 2008).

Esta metodologia pode ser aplicada em todas as famílias com informações de fenótipo e genótipo, bem como combinando dados de diferentes raças (BOLORMAA et al., 2013b). Esta avaliação apresenta alta acurácia seletiva para seleção baseada exclusivamente em marcadores e não exige prévio conhecimento das posições dos “quantitative trait loci” (QTL) (RESENDE et al. 2008), além de reduzir o número de medidas fenotípicas em cada geração (MUIR, 2007) e possibilitar uma predição mais acurada entre diferentes raças, desde que tenha uma densidade suficiente de marcadores (GODDARD, 2009).

A implementação da SG segue, basicamente, dois passos: 1) estimativa dos efeitos dos SNPs em uma população de referência e 2) predição dos valores genéticos genômicos (“Genomic Estimated Breeding Values” - GEBV) para animais que não estão na população de referência (candidatos à seleção).

A questão chave da predição genômica está na estimativa do efeito individual de um SNP em uma característica de interesse. Para isso é necessário a utilização de uma população de referência, também conhecida como população de treinamento (MEUWISSEN, 2007).

Esta população de treinamento contém indivíduos com informação fenotípica confiável, bem como informação do genótipo de cada indivíduo desta população (CALUS, 2010). Esta população é usada para obter informações sobre os fenótipos e genótipos importantes para que os GEBVs tenham uma alta acurácia nos indivíduos candidatos à seleção (CLARK et al., 2012).

Para predizer os valores genéticos a partir de informações genômicas, diversos métodos são utilizados como: Mínimos Quadrados, gBLUP, BayesA, BayesB (MEUWISSEN, 2001), LASSO (TIBSHIRANI, 1996), entre outros. Esta vasta gama de métodos de estimativa de valores genéticos assume desde um pequeno número de *loci* tenham efeito, como no caso do BayesB, até modelo que assume igual variância em todos os *loci*, como no caso do gBLUP e todas elas seguem abordagens multi passos (“multi-steps”) e um único passo (“single-step”) (DUCROCQ et al., 2009, VANRADEN et al. 2009, HARRIS & JOHNSON et al. 2010 e SU et al., 2012).

Atualmente, o método “single-step” tem sido mais utilizado por obter maior acurácia do GEBV do que a abordagem “multi-steps” (SU et al., 2012). A base da

abordagem “single-step” consiste na integração de uma matriz de relacionamento genômico (“Genomic Relationship Matrix” - GRM) com a matriz de pedigree (“Numerator Relationship Matrix” - NRM) utilizando, simultaneamente, informações de indivíduos genotipados e não genotipados (LEGARRA et al., 2009; CHRISTENSEN; LUND, 2010).

Matrizes de Relacionamento

A maioria dos métodos de seleção utilizados necessita de parentesco, ou relacionamento, entre os indivíduos de uma população obtendo uma melhor acurácia de predição (Henderson, 1975). Para a estimação destes parentescos foi desenvolvido métodos propostos por Wright (1917) e Malécot (1948), os quais definiram conceitos e métodos para calcular genes idênticos por descendência (IBD), que são usados para indicar a probabilidade de que dois alelos homólogos tenham sido herdados a partir de um ancestral comum (POWELL et al., 2010).

Tradicionalmente, a probabilidade de que dois alelos sejam IBD pode ser estimada utilizando informações de pedigree da população. Assim, os programas de melhoramento genético utilizam-se esta informação de pedigree para calcular a probabilidade de que dois indivíduos compartilhem o mesmo alelo proveniente de um ancestral em comum, montando a matriz de parentesco conhecida como “Numerator Relationship Matrix” (NRM).

A partir da definição desta matriz NRM, tornou-se possível a obtenção de componentes de variância para uma população-base e a predição de valores genéticos de indivíduos de qualquer geração, por meio do Método da Máxima Verossimilhança Restrita (REML) proposto por Patterson e Thompson (1971).

Este processo é eficiente, porém lento, principalmente para características de difícil mensuração ou mensuradas em apenas um sexo, como produção de leite e características de carcaça (MEUWISSEN; GODDARD, 1996). A principal limitação desta metodologia está no cálculo do parentesco entre os indivíduos, o qual é calculado como uma probabilidade destes animais apresentarem genes em comum, porém muitos alelos podem ser idênticos por estado (IBS) podendo tornar os indivíduos mais aparentados que a média da população (POWELL et al., 2010).

Conforme definido anteriormente, as covariâncias genéticas (parentesco genético) entre os indivíduos são derivadas das probabilidades de que pares de

genes compartilhados entre os indivíduos são idênticos por descendência (LYNCH & WALSH, 1998), assim, espera-se, por exemplo, que dois irmãos germanos apresentem 50% de seus alelos IBD. No entanto, esta metodologia baseada nas informações do pedigree ignora os efeitos aleatórios devido à meiose no processo de gametogênese, esta variação é definida como Amostragem Mendeliana (AVENDAÑO et al., 2005).

Assim, com o avanço das técnicas utilizadas no melhoramento genético animal e a possibilidade de genotipar indivíduos, tornou-se possível a utilização de informações mais precisas sobre os genes IBD e IBS que podem ser compartilhados através de ancestrais comuns, ausentes no pedigree tornando possível a utilização de uma matriz de parentesco genômica denominada “Genomic Relationship Matrix” (GRM) (FORNI et al. 2011). Diversas metodologias são usadas para calcular uma matriz GRM, como observado em VanRaden et al. (2008), Harris and Johnson (2010) e Yang et al. (2010). Porém, o principal objetivo destes métodos é tornar os coeficientes da matriz de parentesco genômico o mais próximo da matriz de parentesco tradicional.

A GRM pode substituir a matriz NRM na tradicional metodologia BLUP e de acordo com Clark et al. (2012) é esperado que a GRM forneça estimativas mais acurada da covariância entre os indivíduos, entretanto, é importante entender o quanto de ganho de acurácia será atribuída ao conhecimento mais preciso do parentesco e quanto se ganha com a adição de informações sobre parentes distantes, anteriormente ignorados pela matriz de parentesco.

Outra possível vantagem da utilização de uma GRM pode ser em se obter coeficientes de parentesco dos indivíduos mais acurados, por exemplo, em uma população multirracial. Algumas pesquisas em gado de leite estão utilizando métodos de cálculo destes parentescos genômicos através de uma estimativa da proporção de raças que compõe os indivíduos sob avaliação genética (ERBE et al., 2012; HARRIS & JOHNSON, 2010; OLSON et al., 2012).

Determinação da proporção racial (Proporção *Bos indicus*)

Os bovinos podem ser divididos em dois diferentes grupos, ambos descendentes do agora extinto *Bos primigenius*. Estas duas subespécies foram separadas há centenas de milhares de anos com independentes domesticações,

resultando nas subespécies *Bos taurus* e *Bos indicus* (MCTAVISH et al., 2013). Hoje estes dois grupos apresentam características distintas tais como adaptabilidade a específicos ambientes, fertilidade e qualidades de produção (TEASDALE et al., 2012).

Estas duas subespécies geralmente são cruzadas formando um animal comumente conhecido como mestiço ou composto que pode ser utilizado para a formação de raças compostas, aproveitando as características de produção dos *Bos taurus* e adaptação aos ambientes tropicais do *Bos indicus* (KUEHN et al., 2011).

A Austrália está entre os maiores produtores de carne do mundo, de acordo com o site da Meat & Livestock Austrália (MLA), as previsões para o rebanho bovino em junho de 2014 serão em torno de 27,5 milhões de cabeça, desta população total pode-se dividir a população, basicamente, em animais da raça Brahman, aproximadamente 39% e raça Tropical Composite, representando aproximadamente 30% da população total.

Como pode ser observado, a raça Brahman é predominante na Austrália e vem crescendo significativamente no Brasil. Esta raça foi criada no Estados Unidos, derivada de quatro raças *Bos indicus* (Guzerá, Nelore, Gir e Krishna Valley). Na Austrália, sua importação teve início no começo do século passado, porém, de acordo com o “Departamento of Primary Industries of New South Wales”, a raça só teve importância econômica a partir do ano 1933 quando uma grande quantidade de animais foi importado pelo Sindicato de criadores de gado de “Queensland” que realizou mais duas importantes importações de animais dos Estados Unidos entre os anos de 1950 e 1954.

Esta raça é caracterizada por sua docilidade, vivacidade e curiosidade. Apresenta porte médio com resistência a doenças e parasitas e boa adaptação a variações de ambiente (MARQUES, 2003) e, de acordo com a Associação de Criadores de Brahman da Austrália, apesar de apresentar maturidade mais tardia, a raça é adequada para cruzamentos, dando excelente vigor híbrido nas progênes.

A raça Tropical “Composite” é um dos principais compostos, obtido pelo cruzamento de Brahman com outras raças (*Bos taurus*) não adaptadas aos trópicos, como “Hereford”, “Shorthorn”, “Red Angus”, “Red Pull” e Charolês (PORTO-NETO et al., 2013). Este composto foi criado no norte da Austrália na tentativa de aumentar o vigor híbrido de várias características reprodutivas e adaptativas utilizando as raças estabelecidas no país, assim, resultaram na formação de raças compostas a partir de raças tropicais adaptadas e raças britânicas ou européias (BOLORMAA et al., 2013a).

A determinação da proporção de genes de uma raça específica em um indivíduo composto pode ser uma ferramenta auxiliar na seleção dos animais com habilidades específicas, principalmente em sistemas de manejo onde se adota uma estrutura de reprodutor múltiplo, assim, a composição de raças em um indivíduo é desconhecida. Outra aplicação das estimativas genômicas da composição de raças é para certificar a proporção de raça em programas que certificam a qualidade da carne e a raça produzida, por exemplo, o esquema de certificação da “Australian Angus beef”, o qual as progênies necessitam ser provenientes de reprodutores exclusivamente da raça Angus e rastreados através de amostras de DNA obtidas nas análises da carcaça (Australian Angus Society, 2013).

O mercado australiano também beneficia os produtores pela qualidade da carcaça e de acordo com o “Meet & Livestock Australia”, que é um programa de pesquisas e “marketing” do governo australiano, a proporção de *Bos indicus* no animal tem impacto negativo sobre uma série de cortes comuns neste país. Assim, o grau de *Bos indicus* em uma carcaça poderia ser mais exato com o auxílio de ferramentas genômicas (THOMPSON, 2002).

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CAPÍTULO 2 - ACCURACY OF GENOMIC SELECTION PREDICTIONS FOR STATURE IN CATTLE USING HD CHIP GENOTYPES: COMPARING RELATIONSHIP MATRICES ESTIMATED FROM PEDIGREE WITH GENOMIC DERIVED MATRICES

Accuracy of genomic selection predictions for hip height in Brahman cattle using HD chip genotypes: comparing relationship matrices estimated from pedigree with genomic derived matrices

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RUNNING HEAD: Genomic selection with different relationship matrices

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Summary (80 words)

We compared 3 variations of genomic relationship matrices (G) with each other and with the pedigree matrix (NRM). The use of G resulted in accuracies higher than 70%. The top 20% animals (higher breeding values) were similar across methods. The use of the observed allele frequency was the option for estimating G that gave variance and heritability results most similar to the pedigree matrix and resulted in the higher accuracy of prediction.

Abstract (250 words)

Cattle selection is based on the phenotype of individuals and information of kinship, which is traditionally derived from pedigree records. It is possible to predict kinship from genomic information. Potential advantages of using a genomic relationship matrix (G) are reduced generation interval and increased genetic evaluation accuracy. The objective of this study was to evaluate the effects of genomic information in genetic evaluation, using different matrices built from genomic and pedigree data in Brahman cattle. Hip height measurements from 1,695 animals were used. Cattle were genotyped with high-density BeadChip or imputed (569,620 markers after quality control). The pedigree matrix NRM was compared to the H matrix, which incorporated NRM and G matrices. Genotypes were used to estimate 3 versions of G : observed allele frequency of each SNP (H_{GOF}), average minor allele frequency (H_{GMF}), and 0.5 for all markers (H_{G50}). For matrices comparisons, animal data were either used in full or divided in calibration (80% older animals) and validation (20% younger animals) datasets. All matrices had similar accuracies close to 0.80. Minor variances, diagonal and off-diagonal elements, and estimated breeding values for NRM and H_{GOF} were very similar. The use of genomic information resulted in very similar relationship estimates when compared to pedigree-based relationships. The top 20% animals were very similar for all matrices, but ranking within these varied depending on the method used. The use of H_{GOF} resulted in the higher accuracy of prediction for hip height estimated breeding values.

Key words: genomics, *Bos indicus*, beef cattle, hip height, rare alleles

Introduction

Traditionally, animal selection studies target traits of interest and use the phenotype of individuals and information of kinship derived from pedigree records. Recorded pedigree information is the basis for building the relationship matrix NRM . This animal breeding and selection method is efficient, but the process can be slow, especially for traits that are measured only in one sex such as milk production, traits measured after the slaughter of animals, such as meat quality, or traits measured late in life, for example, longevity. To enhance or accelerate selection programs focussed on such traits, researchers seek to identify genes or genetic markers associated to the traits, enabling the selection for animals carrying desirable alleles (Meuwissen and Goddard 1996).

A growing number of researchers are interested in the use of genomic information in animal breeding programs (Meuwissen and Goddard, 1996; Christensen and Lund, 2010; Gianola *et al.*, 2010; Hayes *et al.*, 2010; Erbe *et al.*, 2012; Bolormaa *et al.*, 2013). Advancement of technology and the opportunity of genotyping a high number of individuals made possible to use information more precise on alleles identical by state that can be shared through common ancestors in the pedigree (including ancestors that may be missing from pedigree or not genotyped). This technology made the use a genomic relationship matrix G feasible (Meuwissen *et al.*, 2001; Forni *et al.*, 2011), allowing to increase accuracy of predicted breeding values in genetic evaluations. According to Meuwissen *et al.* (2001), genomic selection (GS) using G increases the rate of genetic improvement and reduces the cost of testing progeny. This model of “pre-selection” contributed greatly to the rapid implementation of GS in dairy cattle, despite claims it may create bias (Petry and Ducrocq, 2011).

Breeding values are obtained, traditionally, using mixed model equations (MME) that use the NRM relationship matrix (pedigree information). In one form of GS, NRM or G represent the additive genetic matrix. However, in most circumstances, G includes genomic information of fewer animals. So, Legarra *et al.* (2009) and Misztal *et al.* (2009) proposed a method that performs a integration of the NRM and G matrices in a single H matrix, enabling genetic evaluation based on

Best Linear Unbiased Prediction (BLUP), which was successfully applied to dairy cattle (Aguilar *et al.*, 2010). Forni *et al.* (2011) used different ways to create the genomic relationship G matrix and subsequent integration with the NRM matrix by varying the population allele frequencies used. Forni *et al.* (2011) concluded that varying population allele frequencies to build G did not affect estimated breeding values and variance components in a population of pigs. Despite the result in pigs, however different outcomes may be obtained in other populations or species that present with a different relationship structure. The pig industry is quite unique in its breeding practices and it is different from beef cattle breeding. Thus, it is important to evaluate the contribution of genomic information in genetic evaluation processes in different species and different population structures.

The objective of this study was to evaluate the effects of genomic information in genetic evaluation of beef cattle, using different matrices built from genomic and pedigree data. The population under investigation in this study is a population of Brahman cattle, with predominantly (90%) *Bos indicus* genes (Bolormaa *et al.*, 2011).

Methods

Animal Care and Use Committee approval was not required for this study because the data were obtained from existing phenotypic databases and DNA storage banks as described in the following section.

Phenotype and genotype data:

Height measurements taken from 1,695 Brahman animals between 15 and 18 months of age were used in the current study. These cattle represent a subset of the extensively phenotyped population bred by the Cooperative Research Centre for Beef Genetic Technologies (Beef CRC, Australia) that has been described in detail previously (Barwick *et al.*, 2009; Johnston *et al.*, 2009; Corbet *et al.*, 2011; Fortes *et al.*, 2011; Hawken *et al.*, 2012). All individuals in this population have genotype information for 777,000 SNP, and these high-density SNP data were genotyped or imputed. Animals

were genotyped using three different SNP chips: the BovineSNP50 bead chip (Matukumalli *et al.*, 2009) version 1 was used to genotype females, version 2 was used to genotype males (that combined are the 1,695 phenotyped animals), and the high-density SNP chip was used to genotype 917 samples. These 917 samples were from sires and selected representative animals of the Beef CRC populations, which were genotyped with the high-density SNP chip to allow for genotype imputation, using the BEAGLE program (Browning and Browning, 2011) with average of imputation accuracy of 0.90. Further detail on genotyping, imputation and quality control was described previously (Bolormaa *et al.*, 2013). All SNP chips were processed according to the manufacturer's protocols (Illumina Inc., San Diego, CA). Repeated samples were included in the genotyping for quality assurance, and BEAD STUDIO software (Illumina Inc., San Diego, CA) was used to determine genotype calls.

In quality control analysis, SNP was excluded if: the minor allele frequency was smaller than 0.05 or the correlation between SNP genotypes was bigger than 0.95. After quality control procedures, 569,620 SNPs remained and were used to estimate genomic relationship coefficients in the G matrices.

The pedigree information used to build the matrix NRM was composed by 3,030 animals, including the genotyped animals that corresponded to 55.94% of the total population.

Statistical data analysis:

Estimated breeding values for hip height (HH) were calculated following the animal model represented below, in matrix notation:

$$y = X\beta + Za + e$$

where y is the vector of observations; X is a incidence matrix of the fixed effects that included information of sex, cohort (interaction between year of birth and farm), and age at HH measurement was fitted as covariate; β is a vector of the fixed effects; Z is a incidence matrix of the genetics random effects; a is a vector of the animal random effects, representing the additive genetic values

of each animal; and e is a vector of the residual random effects. The vectors y , a and e follow the assumptions below:

$$\begin{bmatrix} y \\ a \\ e \end{bmatrix} \sim N \left\{ \begin{bmatrix} X\beta \\ 0 \\ 0 \end{bmatrix}, \begin{bmatrix} ZAZ' + R & ZA & R \\ & AZ' & A & \Phi \\ & R & \Phi & R \end{bmatrix} \right\},$$

where, Φ is a zero matrix; 0 is a zero vector; R is a residual matrix; A is an additive genetic matrix that composes the observations.

To obtain the estimated breeding values, the matrix NRM used a traditional method, wherein the relationships between individuals were calculated with pedigree information. The combined pedigree-genomic relationship matrix H , was calculated using both pedigree and genomic information (Aguilar *et al.*, 2010):

$$H = \begin{bmatrix} NRM_{11} & NRM_{21} \\ NRM_{12} & NRM_{\Delta} \end{bmatrix}$$

where, NRM_{11} , NRM_{12} , NRM_{21} represent the relationships between animals with no genotypes, and $NRM_{\Delta} = NRM_{22} - G$, is the difference between pedigree-based (NRM_{22}) and genomic-based (G) relationships for the genotyped individuals, thus the H matrix had dimension equal NRM matrix (n=3030), including genotyped and no genotyped animals. G was obtained using the method of VanRaden (2008):

$$G = \frac{(M - P)(M - P)'}{2 \sum_{j=1}^m p_j(1 - p_j)'}$$

where, M is a matrix that specifies which marker alleles each individual inherited with m columns (m is the total number of markers) and n rows (n is the total number of genotyped individuals); and P is a matrix with the frequency of the second allele (p_j), expressed as $2p_j$. M_{ij} was 0 if the genotype of individual i for SNP j was homozygous AA, was 1 if heterozygous, or 2 if genotype was homozygous BB. The frequencies used to obtain P were according Forni *et al.* (2011): observed allele frequency of each SNP (GOF), the average minor allele frequency (GMF), and 0.5 for all markers (G50).

To avoid problems with inversion in MME, we also used the method proposed by VanRaden (2008) that includes a weighting between G and NRM_{22} matrices:

$$Gw = wG + (1 - w)NRM_{22},$$

where, Gw is a genomic matrix used to obtain the inverse of H matrix; G is an initial genomic matrix, before weighting; w is a weighting factor equal to 0.95, Aguilar *et al.* (2010) reported negligible differences in GEBV using w between 0.95 and 0.98; and NRM_{22} is the subset of the pedigree relationship matrix with the genotyped animals.

After obtaining the weighted Gw matrix, we used the method developed by Aguilar *et al.* (2010) and Christensen & Lund (2010) to calculate the inverse of H :

$$H^{-1} = NRM^{-1} + \begin{bmatrix} 0 & 0 \\ 0 & Gw^{-1} - NRM_{22}^{-1} \end{bmatrix},$$

where, H^{-1} is the inverse of the pedigree-genomic relationship matrix; NRM^{-1} is the inverse of the pedigree relationship matrix; Gw^{-1} is the inverse of the genomic matrix; and NRM_{22}^{-1} is the inverse of the pedigree relationship matrix of the genotyped individuals. Related to the variations in allele frequencies used to build the G matrices, we built 3 versions of the H matrix: H_{GOF} , H_{GMF} , and H_{G50} .

Thus, obtained the variations of H matrix, the additive genetic matrix, NRM or G , on MME can be replaced by H and obtain the genomic breeding values (GEBV).

To obtain the inversions of these matrices, the estimates of the variance components and genetic parameters, we used restricted maximum likelihood (REML) methods in Wombat (Meyer 2007).

To compare the accuracies of GEBVs obtained with each H matrix, the mean accuracy was estimated using the prediction error variance (PEV):

$$r_i = \sqrt{1 - \frac{PEV_j}{\sigma_{a_i}^2}}$$

where, r_i is the accuracy of mean additive value for each matrix i ; $\sigma_{a_i}^2$ is the additive variance estimated for each matrix i ; PEV_j is the prediction error variance for each animal j estimated by the matrix i . These PEV was obtained by Wombat, which provides approximate sampling errors.

Mean accuracies of GEBV based on 1,695 GEBVs were calculated using phenotypes of all the genotyped animals for the prediction (GEN) and using 80% of the phenotype information (OLD, subset of data corresponding to the oldest animals in the dataset).

To compare the accuracy of prediction was used the OLD subset to predict the GEBVs of the 20% youngest animals (YOUNG) was also estimated by omitting the phenotypes of these younger animals from the prediction. Thus, as an alternative “accuracy” metric, correlations between the adjusted phenotype $Phen_{adj}$ and genomic estimated breeding values (GEBVs) were calculated following:

$$r = \frac{cov(Phen_{adj}, GEBV_j)}{\sqrt{h_i^2}}$$

where, the h_i^2 is the heritability estimated for HH by using each matrix i (H_{GOF} , H_{GMF} , and H_{G50}). The correlation between GEBVs estimated with and without including the phenotypes of YOUNG animals in the prediction was also calculated.

Another comparison between the 3 versions of the H matrix considered the ranking of the animals based on estimated GEBVs. To compare the rankings, animals that had the higher GEBVs for HH (top 20% of the population, TOP20%, $n = 339$) were investigated. We used a spearman rank coefficient (ρ) to compare these TOP20% that is defined as the Pearson correlation coefficient between ranked variables (Yitzhaki 2013), using the alternative formula proposed in Conover (1999).

$$\rho = 1 - \frac{6 \sum d_i^2}{n(n^2 - 1)}$$

Where, d_i^2 is the difference between the ranks of each observation on the two variables and n is the number of observations. The standard Pearson correlation between rankings of animals in different matrices was also estimated.

Results

Relationship coefficients

Descriptive statistics of the relationship coefficients estimated for genotyped animals are provided in Table 1. Minor variances and both diagonal elements and off-diagonal elements were obtained for H_{GOF} , H_{GMF} , and H_{G50} and the NRM matrix. For the diagonal elements, the NRM matrix had smaller variance, probably because the inbreeding value of this population is very small, how indicated on mean of diagonal to NRM , indicating that there is low relationship between studied families. In addition, it can be explained because the NRM is incomplete. In this population the no genotyped animals represent 55.94% of all animals. Also, the NRM matrix calculates the probability of kinship, decreasing the variances of the elements. However, when genomic information was used these families did shared common alleles and the estimated relationship coefficients were different (Table 1). For off-diagonal elements, the matrices A and H_{GOF} were very similar. The greatest variance and relationship coefficients were found in H_{GMF} , followed by H_{G50} , both of these matrices have used the same allele frequency for all markers: 0.50 or 0.27 (the average minor allele frequency was 0.27). Observed allele frequencies were distant from 0.5 for many markers (Fig. 1), which may be an effect of SNP chip development, based mostly on *Bos taurus* data not *Bos indicus* (Gibbs *et al.* 2009).

(Insert Table 1 about here)

(Insert Fig 1 about here)

Variance components

The estimates of variance components are presented in Table 2. The data used to compare variance components were either the full phenotype dataset of genotyped animals (GEN, $n = 1,695$) or a subset that included 80% of the oldest animals data (OLD, $n = 1,356$). In both GEN and OLD datasets the variance components were similar when matrices estimated with the same methodology were compared (i.e. the A matrix of GEN was similar to the A matrix of OLD). However, when

matrices estimated with difference methodologies were compared the variance components were different. For example, H_{G50} resulted in higher additive variances while A resulted in smaller. These differences between matrices are in contrast to the data presented by Forni *et al.* (2011), who detected that the additive variance was higher when the difference between the average diagonal and the off-diagonal elements of the matrix was smaller. In our study, the differences in of the diagonal and off-digonal elemens estimated with A , H_{GOF} and H_{GMF} were not important (0.99, 1.03 and 0.93 respectively), but the additive variances were different. Only for H_{G50} this relation found in Forni *et al.* (2011) was true. For H_{G50} , the difference between the coefficients was 0.68.

(Insert Table 2 about here)

Breeding values and accuracies

Average GEBVs of genotyped animals were similar for the matrices A and H_{GOF} . Average GEBVs were also similar for the matrices H_{GMF} and H_{G50} (Table 3). When phenotypes of the 20% youngest animals (YOUNG) were omitted, GEBVs remained similar (Table 3).

(Insert Table 3 about here)

Correlations between GEBVs of all genotyped animals estimated using different matrices are presented in Fig. 2. On average, the choice of relationship matrix did not influence GEBVs, as correlations were high. However, when validation phenotypes were omitted (20% YOUNG omitted), the GEBVs estimated for the youngest animals in the population varied and correlations between GEBV from H matrices and A were lower (Fig. 3).

(Insert Fig 2 about here)

(Insert Fig 3 about here)

The average accuracies, using GEN phenotype information ($n = 1,695$), 80% of the phenotype information represented by the oldest animals (OLD, $n = 1356$) and just for the 20% of youngest

animals that the phenotype was omitted for validation (YOUNG, n = 339) are shown in Table 4. This Table represents the accuracies of prediction in YOUNG population and correlations for GEN and OLD based in GEBVs estimated with the adjusted phenotype. To YOUNG subset, the accuracies of prediction were based on 339 GEBVs and the correlations were made with the GEBVs estimated with and without the phenotypic information. The GEBVs predicted for GEN and OLD in all matrices did not have significant difference. However, the accuracy of GEBVs when YOUNG phenotypes were omitted decreased, as expected, but the accuracy was less to *NRM* matrix when compared with the inclusion of genomic information (Table 4). In the present study, the average accuracy reflects more variance components estimates than predictive ability, thus, H_{GOF} provided a better rate $PEV_i / \sigma_{a_i}^2$ than others matrix. Because this, the average accuracy for H_{GOF} was highest in all population scenarios.

(Insert Table 4 about here)

All the matrices estimated a high correlation (predictive ability) in GEN and OLD scenarios (Table 4). These correlations were calculated using the GEBVs estimated and the adjusted phenotype. The correlations showed in YOUNG scenario were calculated between the GEBVs estimated with and without the phenotype information and for all genomic matrices this correlation was bigger than *NRM* matrix.

Other difference between matrices is in the ranking of individual animals (Supplementary Table S1). Table 5 shows the number of common animals when the 20% genotyped animals with higher GEBVs were selected (TOP20%, n = 339). From this TOP20%, 87% of the animals were the same when comparing *NRM* with any of the *H* matrices. Between different *H* matrices 99% of the TOP20% animals were the same (Fig. 4, Fig. 5). However, the ranking of these TOP20% animals was different between matrices, and these differences in ranking impact on the correlations between matrices (Fig. 3). In the comparisons between *H* matrices almost all TOP20% animals were the same

and the Spearman coefficient between ranking positions were higher. In the comparisons between *NRM* and the *H* matrices, the correlations between ranking of animals were also similar, around 0.83.

(Insert Table 5 about here)

(Insert Fig 4 about here)

(Insert Fig 5 about here)

Discussion

Relationships using the observed allele frequencies can provide more accurate GEBV predictions, when compared to pedigree derived relationships. It is possible that the increased accuracy observed results from more precise estimates of genetic covariance between relatives (Clark *et al.* 2012). Estimates of genetic covariance in *G* matrices are influenced by allele frequencies in the population. Ideally, *G* matrices should be estimated using the allele frequencies from the unselected base population, which is not available. In real situation is practically impossible to obtain this information and the three methods tested alternative solutions: using the observed allele frequencies (H_{GOF}), the minor allele frequencies (H_{GMF}) and a fixed frequency (H_{G50}). In our study, using H_{GOF} seemed advantageous as this matrix presented a greater similarity to *NRM* in terms of the variance components and resulted in higher accuracies for predicted GEBVs, an artefact of inflated additive variance. It is possible that H_{GOF} was the best option in our study for two reasons: the presence of extreme allele frequencies observed for many markers and the fact that the validation population was not independent from the calibration dataset. As the YOUNG animals used for validation are related to the OLD animals (calibration), it is expected that observed allele frequencies are similar in both subgroups of this Brahman population.

The variance components obtained using H_{GOF} and *NRM* were quite similar in this study. This similarity is consistent with the findings of Riley *et al.* (2007). Variance components in H_{GMF} and H_{G50} were less similar to *NRM* than those in H_{GOF} and may have been inflated with the use of fixed

allele frequencies. Several researcher related problems with inflated estimates of variance components (Aguilar *et al.* 2010; Forni *et al.* 2011; Chen *et al.* 2011) due to false kinship coefficients, in this case in H_{GMF} and H_{G50} matrices, that showed a higher values than NRM or H_{GOF} .

When observed allele frequencies are distant from 0.5, “rare” alleles have greater influence in the relationship estimated and this may be the underlying reason approximating H_{G50} to H_{GMF} and distancing these from NRM and H_{GOF} . This difference between NRM and H_{G50} or H_{GMF} was not observed in a previous study that tested the same variations of H in a population of pigs (Forni *et al.* 2011). Average MAF in our population was similar to that observed in the pig population studied by Forni *et al.* (2011): 0.24 and 0.27, respectively. However, the distribution of allele frequencies was different: while in pig population allele frequencies were all close to 0.5, in the Brahman cattle population many markers had allele frequencies distant from 0.5. Presence of these markers that are “rare” (allele frequency distant from 0.5) may reflect the fact that the families in this population can be distinct, whereas that the high density SNP chip was developed using markers selected from *Bos Taurus* animals and *Bos indicus*. And the animals of current population were genotyped or inputted to high density SNP chip.

In addition, using the same allele frequency for all SNPs increased the correlation between the animals, also the estimates of variance components in the population and PEV for each animal were increased (Table 2). In the case of H_{GMF} these PEVs were bigger than additive variance, thus, the accuracies were not calculated because generated a negative numbers.

The difference between the elements of the diagonal and off-diagonal elements were approximately one for all matrices, disagreeing with the (Forni *et al.* 2011) who concluded that the inflation of genetic values can be related to this difference between how much individuals are more closely related (off-diagonal elements) and the average inbreeding of the population (diagonal elements). These genetic values inflated can be explained by the alleles frequencies, when the same frequency was used the animals unrelated were more related because decrease the importance of rare alleles.

Our results support the idea of observing and evaluating population allele frequencies prior to construction of G matrices for improved accuracies. The pig industry is quite unique in its breeding practices and it is different from beef cattle breeding. Therefore, H matrices that were used with no apparent difference to predictions in pigs (i.e. H_{G50} and H_{GMF}) may not be ideal for the studied Brahman population. Nonetheless, correlations between GEBVs and adjusted phenotypes were similar regardless of the H matrix used.

Other point, is that need be observed is that these correlations, accuracies and prediction ability, following the formulas described above, and are influenced by the additive variance estimated for each matrix and consequently the heritability. So, if the estimated additive variance was inflated may be these results were sub estimated. Bijma (2012) showed that the ordinary accuracies of estimated breeding values (EBVs) obtained from genetic evaluations may deviate very substantially from the correlation between true and EBVs.

The TOP20% animals (339 animals with higher GEBVs) were a similar group irrespective of which H or NRM matrix formulation was used. However, within this TOP20% the individual rankings of animals varied. Variation in ranking of animals may be a problematic issue for practical application of genomic selection, because of commercial implications. In some countries, bull ranking is used as a marketing tool and the bull ranked number one could sell more doses of semen, or achieve a higher price on an auction and finally sire a higher number of offspring in the following generation. Evidently, if the use of different methods (NRM , H_{GOF} , H_{GMF} and H_{G50}) leads to a different bull ranked, there is room for discussion and conflict of interest. In the dairy industry, this issue seems more openly discussed or overcome by a standardization of the genomic method used. In the beef industry, this is not resolved yet. The TOP20% as a group is very similar between methods and in most industries, but specially where artificial insemination (AI) is not so common this is probably enough to avoid any conflict, as all TOP20% are equally likely to sire the next generation. Ideally, for the top bull to be in fact the “best” sire of future generations, a progeny test of the best group of animals (TOP20%) would be performed.

Conclusions

In this study, the use of genomic information resulted in very similar relationship estimates when compared to pedigree based relationships in beef cattle. The use of the observed allele frequency seems to be the best option for estimating G ; this method (H_{GOF}) estimated relationships most similar to those of the NRM matrix and resulted in the higher accuracy of predictions, in the studied population allele frequencies were distant from 0.5 for many markers. Was a clear the differences between the ranking presented in TOP20%, despite all genomic matrices resulted in similar animals being selected, more studies are necessary to choose how matrix (NRM or Genomic matrices) selected the rank more accurate. This variation may have implications for cattle breeding commercial practices. Matrices H_{GMF} and H_{G50} can be a good alternative to selection method but not to evaluate the genetic progress in this beef cattle population.

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Table 1. Statistics of relationship coefficients estimated using pedigree and genomic data*

	Diagonal Elements			
	Mean	Min.	Max.	Var.
<i>NRM</i>	1.0003	1.0000	1.1250	3.7×10^{-5}
<i>H_{GOF}</i>	1.0281	0.8971	1.2588	3.4×10^{-3}
<i>H_{GMF}</i>	2.8420	2.5718	3.0816	3.6×10^{-3}
<i>H_{G50}</i>	1.3576	1.1979	1.5244	1.54×10^{-3}
	Off-diagonal elements			
	Mean	Min.	Max.	Var
<i>NRM</i>	0.0086	0.0000	0.6250	1.4×10^{-3}
<i>H_{GOF}</i>	-0.0006	-0.1062	0.6614	1.9×10^{-3}
<i>H_{GMF}</i>	1.9121	1.5498	2.5818	5.7×10^{-3}
<i>H_{G50}</i>	0.6776	0.4453	1.1599	2.6×10^{-3}

**NRM* (relationship matrix pedigree-based); *H_{GOF}* (genomic relationship matrix with observed frequency); *H_{GMF}* (genomic relationship matrix with averaged minor allele frequency); *H_{G50}* (genomic relationship matrix with frequency 0.5 for all alleles). These elements were calculated using the full dataset.

Table 2. Additive and residual variances and heritability estimates using pedigree and genomic matrices built with data from all genotyped animals (GEN) or 80% (OLD) of these.

Additive Variance		
	GEN (n=1695)	OLD (n=1356)
<i>NRM</i>	7.96(±1.10)	7.91(±1.32)
<i>H_{GOF}</i>	8.52(±0.94)	8.57(±1.12)
<i>H_{GMF}</i>	9.40(±1.04)	9.45(±1.24)
<i>H_{G50}</i>	12.71(±1.40)	12.80(±1.67)
Residual Variance		
	GEN	OLD
<i>NRM</i>	6.47(±0.76)	6.96(±0.95)
<i>H_{GOF}</i>	5.84(±0.58)	6.26(±0.72)
<i>H_{GMF}</i>	5.82(±0.58)	6.24(±0.72)
<i>H_{G50}</i>	5.76(±0.58)	6.17(±0.73)
Heritability		
	GEN	OLD
<i>NRM</i>	0.55(±0.06)	0.53(±0.07)
<i>H_{GOF}</i>	0.59(±0.05)	0.58(±0.06)
<i>H_{GMF}</i>	0.62(±0.05)	0.60(±0.06)
<i>H_{G50}</i>	0.69(±0.04)	0.67(±0.04)
Average PEV		
	GEN	OLD
<i>NRM</i>	3.168	3.338
<i>H_{GOF}</i>	2.890	3.100
<i>H_{GMF}</i>	16.200	16.657
<i>H_{G50}</i>	8.221	8.207

**NRM* (relationship matrix pedigree-based); *H_{GOF}* (genomic relationship matrix with observed frequency); *H_{GMF}* (genomic relationship matrix with averaged minor allele frequency); *H_{G50}* (genomic relationship matrix with frequency 0.5 for all alleles); PEV (approximated prediction error variance for each animal).

Table 3. Averages and variances of estimated breeding values (EBVs) obtained with the pedigree matrix (*NRM*) and 3 variations of the *H* matrix (combined pedigree and genomic relationships).

	Average			
	FULL	GEN	OLD	YOUNG
<i>NRM</i>	-0.01	-0.03	0.00	0.00
<i>H_{GOF}</i>	0.00	0.00	0.00	0.00
<i>H_{GMF}</i>	-1.06	-1.41	-0.86	-0.86
<i>H_{G50}</i>	-0.88	-1.17	-0.70	-0.71
	Variance			
	FULL	GEN	80%	20%
<i>NRM</i>	2.99	3.33	1.00	3.33
<i>H_{GOF}</i>	3.64	4.31	1.98	4.31
<i>H_{GMF}</i>	3.97	4.32	1.98	4.32
<i>H_{G50}</i>	3.91	4.38	1.98	4.38

**NRM* (relationship matrix pedigree-based); *H_{GOF}* (genomic relationship matrix with observed frequency); *H_{GMF}* (genomic relationship matrix with averaged minor allele frequency); *H_{G50}* (genomic relationship matrix with frequency 0.5 for all alleles); FULL (n = 3,030 animals, including not genotyped animals that were in the pedigree); GEN all the genotyped animals (n = 1,695); OLD 80% of the population represented by the oldest animals (n = 1,356); YOUNG 20% of the population represented by the youngest animals that had the phenotypes omitted for validation (n = 339).

Table 4. Average accuracies of estimated breeding values (EBVs) and correlations between EBVs and adjusted phenotypes*

	Accuracies			Correlations		
	GEN	OLD	YOUNG	GEN	OLD	YOUNG
<i>NRM</i>	0.776	0.699	0.457	0.969	0.900	0.479
<i>H_{GOF}</i>	0.813	0.746	0.536	0.938	0.868	0.613
<i>H_{GMF}</i>	-	-	-	0.916	0.853	0.612
<i>H_{G50}</i>	0.594	0.598	0.594	0.870	0.882	1

**NRM* (relationship matrix pedigree-based); *H_{GOF}* (genomic relationship matrix with observed frequency); *H_{GMF}* (genomic relationship matrix with averaged minor allele frequency); *H_{G50}* (genomic relationship matrix with frequency 0.5 for all alleles); GEN all the genotyped animals (n = 1,695); OLD 80% of the population represented by the oldest animals (n = 1,356); YOUNG 20% of the population represented by the youngest animals that had the phenotypes omitted for validation (n = 339). Accuracies of GEBVs and correlations for GEN and OLD are based on 1,695 GEBVs, estimated with phenotypic data from all genotyped animals (GEN) or with 80% of the phenotypic data (OLD). Accuracies (prediction ability) for YOUNG are based on the 339 EBVs estimated for the 20% younger animals when their phenotypic data was omitted. Correlations reported for YOUNG were based on 339 EBVs, calculated with and without the phenotype information of the 20% younger animals.

Table 5. Number of highest GEBV (TOP20%, n = 339) animals in common between the different matrices, and Pearson correlations between EBVs, above diagonal. Below diagonal, Spearman coefficients calculated between the rank position of each animal*

	<i>NRM</i>	<i>H_{GOF}</i>	<i>H_{GMF}</i>	<i>H_{G50}</i>
<i>NRM</i>		296(0.996)	296(0.996)	296(0.996)
<i>H_{GOF}</i>	0.834		339(0.999)	337(0.999)
<i>H_{GMF}</i>	0.836	0.999		337(0.999)
<i>H_{G50}</i>	0.837	0.999	0.999	

**A* (pedigree-based relationship matrix); *H_{GOF}* (genomic relationship matrix with observed allele frequencies); *H_{GMF}* (genomic relationship matrix with averaged minor allele frequency); *H_{G50}* (genomic relationship matrix with allele frequency 0.5 for all markers).

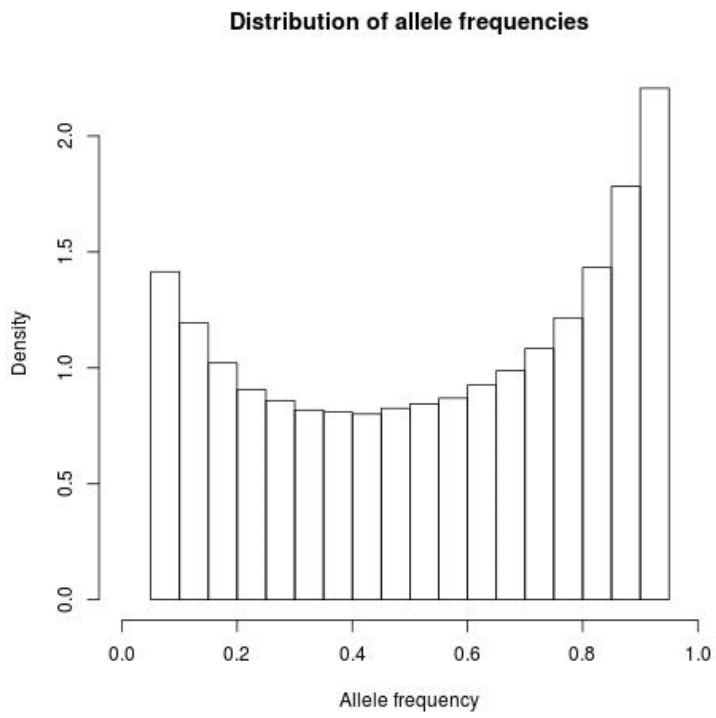


Fig. 1. Distribution of observed frequencies for the second allele

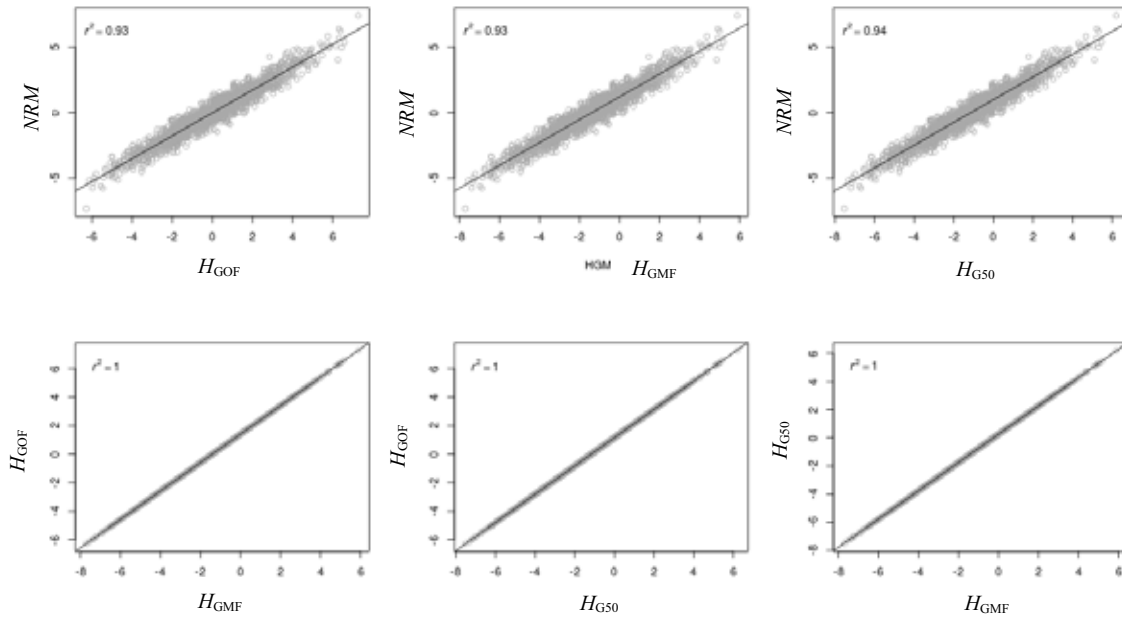


Fig. 2. Correlations between estimated breeding values using pedigree (NRM) and genomic relationship coefficients with observed allele frequency (H_{GOF}), average of minor allele frequency (H_{GMF}) and frequency 0.5 for all alleles (H_{G50}), using phenotypes from all genotyped animals ($n = 1,695$).

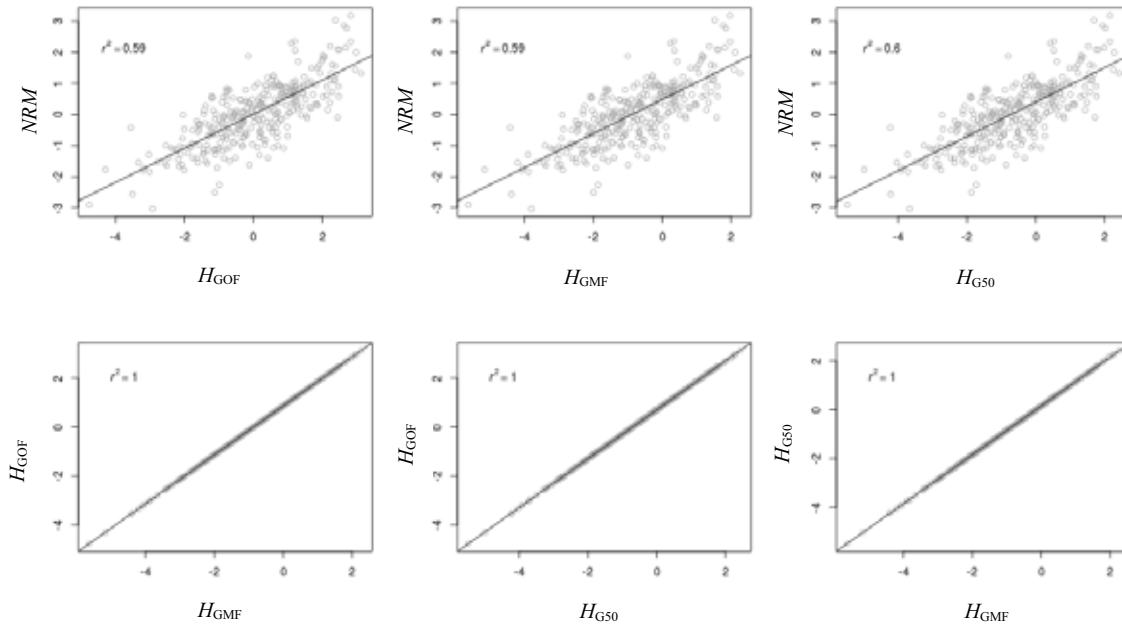


Fig. 3. Correlation between estimated breeding values using pedigree (NRM) and genomic relationship coefficients with observed allele frequency (H_{GOF}), average of minor allele frequency (H_{GMF}) and frequency 0.5 for all alleles (H_{G50}) for all genotyped animals, but omitting 20% of the phenotypic information for validation. These correlations are based on 1,695 animals that were genotyped, with 1,356 phenotypes informed and 339 animals with just genotype information (omitted phenotypes of the 20% youngest animals).

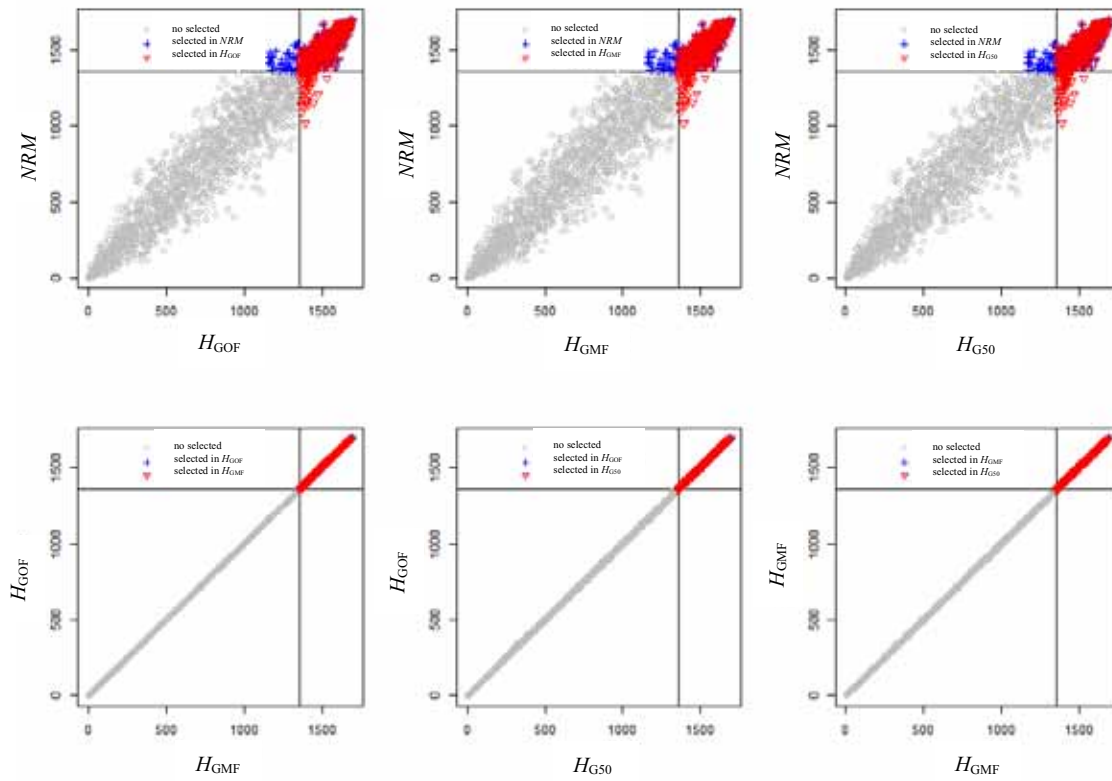


Fig. 4. Correlations between rankings of genotyped animals estimated with different relationship matrices. Rankings were based on EBVs of 1,695 animals (all genotyped population). Abbreviations in figure are: NRM (relationship matrix pedigree-based); H_{GOF} (genomic relationship matrix with observed frequency); H_{GMF} (genomic relationship matrix with averaged minor allele frequency); H_{G50} (genomic relationship matrix with frequency 0.5 for all alleles).

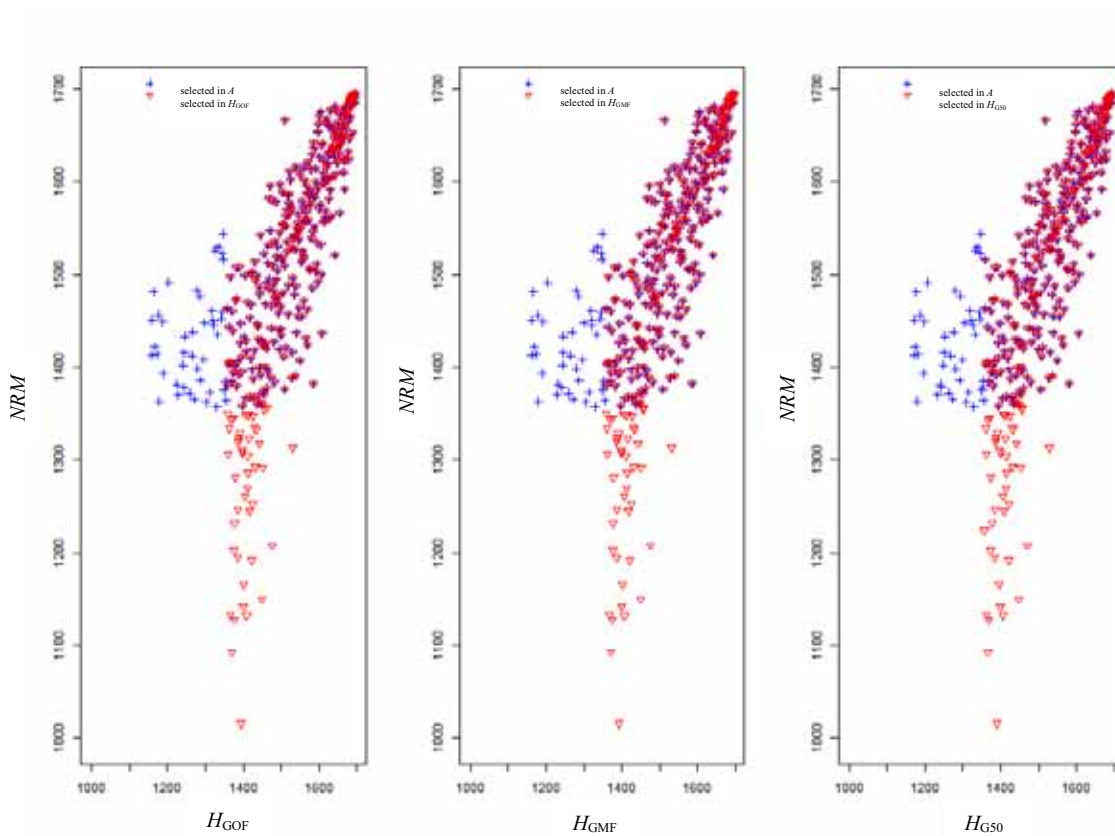


Fig. 5. Correlations between rankings of the top 20% of the genotyped animals with highest estimated breeding values obtained with different relationship matrices: pedigree-based (NRM) and genomic enhanced matrices (H_{GOF} , H_{GMF} , H_{G50}). These correlations are based on the results for 339 animals. Abbreviations in the figure are: NRM (pedigree-based relationship matrix); H_{GOF} (genomic relationship matrix with observed allele frequencies); H_{GMF} (genomic relationship matrix with averaged minor allele frequency); H_{G50} (genomic relationship matrix with allele frequency of 0.5 for all markers).

Supplementary Table

S 1. Ranking of animals that had the highest estimated breeding values (EBVs) for hip height, top 20% of the genotyped animals (TOP20%, $n = 339^*$), based on EBVs calculated with different relationship matrices: pedigree-based relationship matrix (NRM) and genomic enhanced matrices based on observed allele frequencies (H_{GOF}), minor allele frequency (H_{GMF}) and allele frequency equal to 0.50 for all markers (H_{G50}).

Animal ID	Rank			
	NRM	H_{GOF}	H_{GMF}	H_{G50}
16	14	22	22	22
19	91	72	72	71
20	304	308	307	307
21	126	168	166	165
22	166	205	204	202
26	31	20	20	20
27	219	-	-	-
28	7	12	12	12
30	252	257	257	259
32	138	124	125	124
667	174	242	241	234
669	34	35	35	35
673	53	39	40	42
674	2	5	5	5
677	194	272	272	270
679	52	45	45	44
680	225	136	136	140
684	313	214	215	217
687	189	315	315	312
689	231	198	199	201
690	152	-	-	-
691	151	200	200	200
695	186	218	219	220
699	147	154	155	155
731	89	146	146	146
739	298	-	-	-
740	208	177	177	179
741	281	-	-	-
744	261	-	-	-
745	210	253	253	250
749	273	243	243	245
750	13	10	10	10
756	96	79	79	79
769	101	225	224	219
784	306	159	159	159

799	135	83	83	86
807	187	188	187	187
834	163	222	222	221
836	311	-	-	-
838	69	94	94	94
840	213	-	-	-
845	110	176	176	175
848	15	54	53	51
850	109	128	128	129
854	288	-	-	-
860	112	121	121	123
866	178	197	195	193
867	250	260	259	261
870	182	265	265	265
875	8	16	16	16
876	198	262	262	260
880	190	120	120	121
881	67	115	115	113
889	103	123	122	122
890	66	44	44	45
899	155	163	163	163
910	84	145	145	141
912	173	-	-	-
915	87	85	86	87
918	263	-	-	-
934	188	307	306	305
945	240	-	-	-
946	258	-	-	-
952	282	-	-	-
957	321	-	-	-
966	245	-	-	-
969	29	77	76	74
970	235	-	-	-
971	-	322	321	324
989	97	104	104	105
992	319	234	236	236
993	176	139	139	139
997	241	152	152	157
1036	227	211	210	208
1039	-	266	266	266
1061	107	193	193	189
1090	333	-	-	-
1111	339	-	-	-
1112	93	81	81	81
1141	284	-	-	-
1145	259	156	157	158
1151	64	19	19	19
1155	60	63	63	62
1158	54	110	110	109

1164	-	317	318	322
1175	247	-	-	-
1178	324	-	-	-
1183	39	37	37	37
1226	175	55	56	58
1231	114	229	229	228
1240	256	204	205	211
1262	131	151	151	153
1273	18	34	34	34
1279	-	296	297	297
1280	-	292	292	290
1284	95	134	133	126
1287	146	144	144	144
1288	293	335	336	337
1290	38	62	62	63
1292	330	290	290	292
1293	257	324	323	319
1302	317	313	314	315
1313	142	192	192	192
1321	4	11	11	11
1354	294	-	-	-
1355	43	100	100	99
1367	295	318	317	316
1624	133	185	186	185
1626	275	183	185	186
1629	6	2	2	2
1630	160	212	212	209
1631	216	190	190	191
1637	300	232	232	238
1638	75	125	123	120
1639	153	155	154	152
1641	211	113	114	117
1644	11	9	9	9
1655	-	311	311	311
1669	3	4	4	3
1674	22	23	23	23
1679	20	50	50	46
1681	179	137	137	138
1683	206	235	233	233
1685	-	282	283	284
1693	16	13	13	13
1697	312	327	327	326
1699	332	-	-	-
1713	303	-	-	-
1714	30	186	184	178
1717	82	106	105	104
1728	203	263	261	254
1736	76	49	49	47
1749	26	95	95	95

1752	37	65	64	65
1773	315	-	-	-
1776	299	273	273	274
1777	177	208	208	204
1789	65	73	73	73
1799	318	-	-	-
1806	290	312	312	314
1811	-	254	254	255
1814	272	127	127	131
1824	72	18	18	18
1827	41	61	60	56
1852	44	7	7	7
1858	180	-	-	338
1863	-	-	-	339
1864	35	98	97	97
1865	243	279	278	276
1870	80	158	158	151
1871	310	-	-	-
1893	307	224	226	226
1900	291	332	332	332
1901	251	-	-	-
1906	238	203	203	206
1907	124	138	138	137
1912	168	170	170	170
1919	-	321	320	323
1920	9	8	8	8
1932	264	295	294	294
1934	79	132	130	125
1938	335	249	249	252
1947	-	319	319	318
1972	274	-	-	-
1983	215	-	-	-
1986	118	209	207	203
2023	149	226	225	222
2024	286	301	301	301
2029	144	184	183	183
2037	-	330	330	333
2041	40	25	26	26
2045	90	58	57	57
2046	314	111	111	115
2051	127	182	180	177
2052	325	241	242	242
2055	10	15	14	14
2056	218	161	162	161
2059	70	53	54	55
2061	-	291	291	291
2065	172	180	182	184
2066	-	165	165	167
2068	-	309	309	309

2071	115	70	70	70
2076	-	283	282	279
2086	297	255	255	253
2087	104	29	29	29
2089	334	-	-	-
2112	134	64	66	69
2117	-	281	281	283
2121	125	96	96	96
2123	296	239	239	241
2124	220	202	202	205
2133	248	-	-	-
2136	167	-	-	-
2139	309	270	270	269
2143	253	334	333	330
2144	59	59	59	59
2147	17	26	25	25
2148	150	112	112	114
2154	12	3	3	4
2155	-	298	298	299
2157	-	261	263	263
2166	287	277	277	278
2168	88	51	51	53
2171	181	194	194	196
2175	-	294	295	300
2182	73	87	88	88
2184	137	135	135	134
2188	255	196	196	197
2191	102	97	98	98
2192	121	93	93	93
2201	145	108	108	108
2205	192	206	206	207
2206	228	237	234	232
2207	254	223	223	224
2218	212	228	228	229
2222	162	101	101	102
2231	32	47	46	49
2238	116	219	218	218
2243	207	217	217	215
2245	308	169	169	168
2246	242	142	142	142
2254	140	84	84	85
2255	156	162	161	160
2256	130	43	43	43
2257	185	103	103	103
2263	27	14	15	15
2266	221	213	213	210
2267	209	157	156	154
2272	83	117	117	112
2277	120	118	118	116

2279	81	76	77	76
2280	260	88	89	90
2287	271	238	237	235
2288	269	215	214	214
2296	337	246	246	246
2303	-	336	335	334
2304	249	303	302	298
2306	-	240	240	240
2309	62	56	55	54
2312	36	32	32	32
2313	265	131	132	135
2317	327	258	258	258
2326	277	201	201	198
2328	58	42	42	41
2329	159	149	149	148
2336	5	6	6	6
2339	336	259	260	262
2340	98	82	82	82
2359	78	75	75	75
2366	224	316	316	317
2368	85	173	174	172
2370	100	130	131	132
2373	323	-	-	-
2374	105	67	67	66
2378	63	60	61	61
2380	322	244	244	244
2386	270	304	304	303
2389	329	268	268	268
2393	161	181	181	180
2405	237	-	-	-
2409	47	21	21	21
2410	244	-	-	-
2425	246	-	-	-
2429	57	89	87	84
2435	71	33	33	33
2449	1	1	1	1
2452	232	148	148	149
2459	276	236	238	237
2472	267	248	248	248
2479	197	293	293	293
2481	42	46	47	50
2482	193	105	106	107
2495	-	274	275	275
2496	-	302	303	306
2501	-	245	245	243
2504	86	31	31	31
2507	113	107	107	106
2516	196	90	90	89
2517	-	328	328	331

2522	92	68	68	68
2524	117	141	140	136
2526	-	305	305	304
2530	199	331	331	325
2540	200	166	168	169
2541	204	99	99	100
2549	223	314	313	313
2560	154	252	250	247
2563	230	221	220	223
2567	143	191	191	195
2569	222	207	209	213
2571	23	41	41	38
2572	-	247	247	249
2577	-	306	308	308
2580	164	91	91	92
2583	266	256	256	257
2588	-	320	322	328
2601	111	179	179	182
2606	108	114	113	111
2608	184	231	230	230
2609	283	-	-	-
2621	94	171	171	173
2625	123	172	172	174
2626	128	80	80	80
2630	-	269	269	271
2634	236	325	325	321
2648	229	275	276	277
2651	195	276	274	272
2654	21	92	92	91
2659	279	289	287	285
2660	25	27	27	27
2664	51	38	38	39
2669	33	78	78	78
2670	28	28	28	28
2677	302	323	324	320
2678	202	210	211	212
2683	320	187	188	190
2687	141	153	153	156
2688	338	299	300	302
2699	-	297	296	295
2704	-	284	284	281
2707	-	310	310	310
2708	45	36	36	36
2711	268	288	288	286
2713	-	271	271	273
2714	316	338	339	-
2716	217	251	252	251
2721	77	74	74	77
2723	169	160	160	162

2741	-	264	264	264
2743	326	-	-	-
2750	24	24	24	24
2759	233	339	337	336
2765	49	57	58	60
2768	148	150	150	150
2770	-	326	326	329
2780	68	69	69	67
2782	50	17	17	17
2783	48	48	48	48
2786	56	40	39	40
2797	46	52	52	52
2799	106	102	102	101
2805	-	285	285	282
2806	262	195	198	199
2819	239	178	178	181
2820	-	233	235	239
2824	280	167	167	166
2827	214	216	216	216
2834	171	140	141	145
2837	285	189	189	188
2838	-	278	279	287
2839	157	116	116	119
2841	226	286	286	288
2845	132	174	173	171
2849	139	133	134	133
2850	301	300	299	296
2873	129	129	129	130
2876	289	147	147	147
2877	170	-	-	-
2885	19	66	65	64
2894	158	164	164	164
2898	55	86	85	83
2904	99	199	197	194
2906	292	329	329	327
2907	191	119	119	118
2922	61	109	109	110
2933	122	122	124	127
2934	205	-	-	-
2943	119	175	175	176
2949	278	267	267	267
2952	305	250	251	256
2961	165	230	231	231
2962	-	337	338	-
2984	201	227	227	227
2992	183	143	143	143
2993	-	220	221	225
3000	328	280	280	280
3003	234	126	126	128

3005	74	30	30	30
3008	-	287	289	289
3019	136	71	71	72
3020	-	333	334	335
3023	331	-	-	-

*This table show 383 animals because have animals that was selected in just one matrix.

CAPÍTULO 3 - ACCURACY OF GENOMIC SELECTION FOR AGE AT PUBERTY IN A MULTI BREED POPULATION OF TROPICALLY ADAPTED BEEF CATTLE

Short title: Genomic selection in a multi-breed population

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Summary

Genomic selection is becoming a standard tool in livestock breeding programs, particularly for traits that are hard to measure. Accuracy of genomic selection can be improved by increasing quantity and quality of data and potentially by improving analytical methods. Adding genotypes and phenotypes from additional breeds or crosses often improves the accuracy of genomic predictions, but will require specific methodology. A model was developed to incorporate breed composition estimated from genotypes into genomic selection models. This method was applied to age at puberty data (as estimated from age at first observation of a *corpus luteum*) from a mix of Brahman and Tropical Composite beef cattle. In this data set the new model incorporating breed composition did not increase the accuracy of genomic selection. However the breeding values exhibited slightly less bias (as assessed by deviation of regression of phenotype and genomic breeding values from the expected value of 1). Adding additional Brahman animals to the Tropical Composite analysis increased the accuracy of genomic predictions and did not affect the accuracy of the Brahman predictions.

Keywords: *Bos taurus*, Brahman, cross validation, Tropical Composite

Introduction

Improved genomic selection for fertility and other economically important traits associated to beef production will be reliant on the availability of genotyped reference populations with accurate phenotypes, and the development of better analytical methods. There is a need to test alternative methods of genomic prediction and estimation of individual marker effects, given the multi-breed scenario that is typical of the beef industry in Northern Australia. Most of the methods used to date have been based on those implemented in the dairy industry, and have therefore been developed and tested within a single breed (Holstein). The Australian beef industry in contrast consists of a mix of breeds, especially in tropical regions where adaption traits are important and animals with varying degrees of *Bos indicus* genetics are widely used (Bolormaa *et al.* 2013b; Burrow 2012). Tropical Composite is a term used to define a breed that is a stable cross of Zebu

(*Bos indicus*) and Taurine (*Bos taurus*) breeds, which is prominent in Northern Australia (Burns *et al.* 2013; Prayaga *et al.* 2009). Recent studies have analysed Tropical Composite cattle and have considered them to be a single population (Corbet *et al.* 2013). Alternative prediction methods have been proposed for use in multi-breed dairy cattle populations (Erbe *et al.* 2012; Harris & Johnson 2010; Olson *et al.* 2012). These methods were shown to increase accuracy of genomic selection for Jerseys where additional Holstein data were added to the analysis. Both studies also suggest that the methods could be further modified to account for crossbred animals. Accordingly methods have been proposed by Harris & Johnson (2010) that will accommodate both multiple breeds and crosses.

Because of the complexity of multi-breed populations, there is increased potential for biases in genomic breeding values if models do not account for breed of origin (Misztal *et al.* 2013). Better understanding of the factors that degrade predictive power in multi-breed populations is necessary in order to increase the accuracy of estimated genomic breeding values. Therefore, the aim of this paper was to develop genomic prediction methods to model the diverse nature of the population.

Material and Methods

Phenotype and genotype data

The trait used for this study was age at the first *corpus luteum* (AGECL, days) recorded on 2054 genotyped females that consisted of Brahman (BB, n=980) and Tropical Composite breeds (TC, n=1074). AGECL is used as an indicator of the age at puberty in beef cattle. Actual mean AGECL in days (\pm s.d.) of females on each breed was 750.6 ± 141.8 for BB and 652.2 ± 119.4 for TC. These cattle represent a subset of the population established by the Cooperative Research Centre for Beef Genetic Technologies (Beef CRC). This population and its phenotypes have been described in detail previously (Barwick *et al.* 2009; Burns *et al.* 2013; Hawken *et al.* 2012; Johnston *et al.* 2009). A key feature of the population structure relevant to our study is that the Tropical Composite animals used were formed by crossing *Bos indicus* (Brahman) and *Bos taurus* breeds. The relative

contribution from genes of each group (*Bos indicus* and *Bos taurus*) was established for the Tropical Composite animals in our study, and used as a central component of the analyses.

All individuals have high density SNP genotypes available, either directly genotyped or imputed from lower density genotypes. Animals were genotyped using the Illumina Bovine SNP50 bead chip (Matukumalli *et al.* 2009) version 1 (containing approximately 50,000 SNP). Imputation was performed using a reference set of 917 animals genotyped with the high density BovineHD. The imputation was performed using BEAGLE and the methods, number of animals used and accuracy is described in detail in (Bolormaa *et al.* 2013a). All SNP chips were processed according to the manufacturer's protocols. Repeated samples were included in the genotyping for quality assurance, and Bead Studio software (Illumina, Inc.) was used to determine genotype calls. Quality control analysis methods and results have been reported previously (Hawken *et al.* 2012).

Genomic analysis methods

Genomic breeding values were estimated using GBLUP, based on the following general mixed model:

$$y = X\beta + Zu + e$$

where y is the vector of AGECL phenotypes; \mathbf{X} is an incidence matrix for fixed effects; β is a vector of fixed effects; \mathbf{Z} is an incidence matrix for genomic breeding values; u is a vector of random genomic breeding values for each animal ($Var(u) = G\sigma_u^2$ where \mathbf{G} is a genomic relationship matrix as described below and σ_u^2 is the variance of genomic breeding values), and e is a vector of residual random effects ($Var(e) = I\sigma_e^2$ where \mathbf{I} is an identity matrix and σ_e^2 is the residual variance).

The model was fitted with one of two genomic relationship matrices (GRM), genomic relationships using allele frequencies calculated as a single breed group \mathbf{GRM}_{SB} and \mathbf{GRM}_{XB} with allele frequency adjusted for breed, for the 2054 recorded and genotyped females. The GRMs were calculated following an adaptation of the methods described by Harris & Johnson (2010); VanRaden *et al.* (2011); Yang *et al.* (2010):

$$Gr = \frac{(WW')}{n}$$

where $W = M - 2P$, in which \mathbf{M} is the $n \times m$ matrix of genotypes for $n=2054$ animals and m SNP, with values of 0 for the homozygous genotype of the first allele, 1 for the heterozygous genotype, and 2 for the homozygous genotype of the second allele. \mathbf{P} is the $n \times m$ matrix containing the frequencies of the second allele of each SNP (p_i) expressed as the frequency multiplied by 2.

For \mathbf{GRM}_{SB} , allele frequencies for each SNP in \mathbf{P} were calculated from the group of 2054 analysis females, irrespective of breed. Therefore, rows of \mathbf{P} are the same for all animals.

For \mathbf{GRM}_{XB} , \mathbf{P} was calculated as QC , where \mathbf{Q} is a $n \times 2$ matrix describing the fraction of genes of Brahman and *Bos taurus* origin (columns) for each of the 2054 analysis animals (rows). Each row of \mathbf{Q} sums to 1. \mathbf{C} is a $2 \times m$ matrix containing the allele frequencies of each SNP (columns) for BB and *Bos taurus* populations (rows). Both \mathbf{Q} and \mathbf{C} were derived from analyses using the software package Admixture (Alexander & Lange, 2011; Alexander *et al.* 2009), as described below. Apart from the multi-breed formulation of QC a key difference between \mathbf{GRM}_{XB} and \mathbf{GRM}_{SB} is that allele frequencies in \mathbf{GRM}_{XB} were estimated in the Admixture analysis from animals of known breed not including the analyzed animals, whereas allele frequencies in \mathbf{GRM}_{SB} were estimated directly from the analyzed animals. Harris & Johnson (2010) described a similar method for deriving a multi-breed GRM, although in their study the breed fractions (\mathbf{Q}) were derived from pedigree rather than genomic information.

Such genomic relationships matrices are positive semi-definite, and often singular (Forni *et al.* 2011). So, to enable inversion, genomic relationship matrices were weighted following (VanRaden 2008):

$$G = wGr + (1 - w)A_{22},$$

where, \mathbf{G} is the final genomic relationship matrix to be used in the analysis; \mathbf{Gr} is the initial genomic relationship matrix as described above and based only on genotypic information, w is a weighting factor equal to 0.95 (Aguilar *et al.* 2010); and \mathbf{A}_{22} is the subset of the pedigree based numerator relationship matrix (\mathbf{NRM}) for the genotyped females in the analysis.

Estimation of Brahman content

The Brahman and *Bos taurus* content (\mathbf{Q}) for each animal was estimated using a supervised Admixture analysis as described previously in (Alexander & Lange 2011; Alexander *et al.* 2009). The dataset used to estimate Brahman content (BB%) consisted of training animals from five *Bos taurus* breeds (Angus, Murray Grey, Charolais, Hereford, and Shorthorn) with 2,000, 200, 400, 500 and 500 cattle respectively, totaling 3,600 animals in training group. The *Bos indicus* training set included 2000 Brahman cattle. Both groups are part of the same Beef CRC experimental population, but excluded the 2054 analyzed females used in this study. To obtain the estimates of breed content required for \mathbf{Q} the analyzed females were added to the Admixture analysis with their breed masked. The analysis was performed considering the six *Bos taurus* breeds as a single breed, and compared with the Brahman animals. Thus the number of breeds (the 'k' parameter) in Admixture was set to 2, and all other parameters set to their default values (Alexander & Lange 2011; Alexander *et al.* 2009).

Estimation of genomic breeding values

Variance components for σ_u^2 and σ_e^2 used in GBLUP analyses were estimated by restricted maximum likelihood (REML) using the Wombat software package (Meyer 2007). The variance estimates used in GBLUP were calculated based on all animals with phenotype and genotype data using an animal model fitted with the inverse of the pedigree based numerator relationship matrix. Fixed effects fitted included cohort (year of birth and farm, n=14), origin (O, n=8), month of birth (BM, n=9), sire breed (Sg, n=7), dam breed (Dg, n=9) and the interactions between BM*O (n=34), cohort*O (n=30), Sg*Dg (n=34), BM*Sg (n=35) and has been tested the inclusion or not of BB% in the model as a covariate. Variance estimates from these models are presented in Table 4 and were used in the estimation of breeding values for the GBLUP cross validation analysis. The GBLUP analyses were also fitted in Wombat using the same fixed effects and the two GRM previously described (\mathbf{GRM}_{SB} and \mathbf{GRM}_{XB}).

Scenarios tested

Cross validation was used to evaluate the impact of data and model factors on accuracy and bias of genomic evaluation. To study the impact of data on Tropical Composite predictions, increasing amounts of records on Brahman females were added to the analyses. The model factors studied were: fitting **GRM_{SB}** compared to **GRM_{XB}**, fitting Brahman content (BB%) as a covariate, and pre-adjustment (rescale) of data by breed to the same phenotypic variance dividing the phenotype values by the variance.

A series of cross validation analysis were performed to estimate the effect of each of the three factors on accuracy and bias of genomic predictions. Cross validation groups were formed within each breed group (Brahmans and Tropical Composites) by randomly selecting sire families into one of four groups, stratified by number of sibs with genotypes to ensure reasonably similar sized groups.

The cross validation strategies are described in Table 1. Standard cross validation where one of the four groups was omitted from the analysis to use as a validation group was performed within Brahman and Tropical Composites (denoted 3BB and 3TC, respectively). A series of cross validation analysis was then run where additional groups were added to the Tropical Composite cross validation. In each case all, possible combinations of BB groups were run in cross validation. At the end of the analysis for each of the cross validation runs the correlation between adjusted phenotype and genomic estimated breeding value (GEBV) was estimated for animals that were not included in training the model for each combination. The mean correlation and regression was then estimated from the group estimates.

Results

Figure 1 represents the absolute value of the difference in allele frequency between Brahman (BB) and *Bos taurus* (BT). The smaller difference between the frequencies show similarity between

the frequencies in both population. This Figure shows that a high proportion of SNP have similar frequencies in both Brahmans and *Bos taurus*.

The proportion of BB% and BT% in all animals was estimated using the Admixture software package on a reference population of 2000 Brahman and 3600 *Bos taurus* cattle. For the animals included in training the estimated breed proportions were fixed at 1 for their respective breeds (Table 2). The estimated BB% of Brahman and *Bos taurus* animals not included in training was slightly lower with averages of 0.974 and 0.002 respectively. The average BB% of Tropical Composite animals was 0.41, but the estimated proportions for individual animals covered a wide range (Figure 2).

Comparison of different GRM methods

Statistics of relationship coefficients are represented in Table 3. For the diagonal elements both genomic matrices (**GRM_{SB}**, **GRM_{XB}**) were similar and were smaller than the pedigree relationship matrix (**NRM**). The variances of these elements were very small (close to zero) for all matrices. The off-diagonals were impacted by the different GRM methods. The average, minimum and maximum off-diagonal was smaller when allele frequencies were adjusted for breed composition (**GRM_{XB}**) in both the Tropical Composites and the Brahmans. The off diagonals linking BB and TC animals were increased slightly by adjusting for breed composition.

Table 4 presents variance component estimates from each breed group and for the combined dataset using each of the relationship matrices. The variance components from the full model were used in the estimation of genomic breeding values (GEBV).

Accuracy and precision of genomic selection

Table 5 presents the correlations between phenotype and GEBV predicted using a range of models and including different numbers of cross validation groups. The accuracy of predicting Tropical Composites from Brahmans alone was similar to that when predicting Tropical Composites from Tropical Composites alone. Adding Brahmans groups increased the accuracy (from 0.14 to

0.22). There was no difference in the correlations observed between the two GRMs (<0.003), adding the covariate BB% (<0.03), or rescaling the phenotypes (<0.03).

The accuracy of predicting Brahman animals from Tropical Composites was low. Adding as little as one BB group into the analysis increased the accuracy substantially (from 0.086 to 0.242). Additional groups increased the accuracy to around 0.33. The accuracy using three groups from both breeds was similar to the results from the Brahman only analysis, although adding Tropical Composite data to Brahman analysis did not reduce accuracy of prediction within Brahmans. There was no difference in the accuracy between the two relationships matrices. In contrast to the Tropical Composite results, adding BB% had a small impact in some scenarios, but when three groups of Brahmans were included in the analysis there was no difference (scenarios 3BB and 3TC + 3BB). However, if less than three BB groups were included in the analysis the inclusion of BB% increased the correlation. The correlation was increased by 0.04-0.05 for the TC only analysis and by a smaller amount for the other training scenarios (0.01-0.03). Rescaling the phenotypes had no impact on the correlation.

Table 6 presents the slope of the regression coefficients between GEBV and adjusted phenotypes. In general the regression coefficients were closer to 1 for the Tropical Composite animals and well above 1 for the BB animals. Within the Tropical Composite animals adding Brahmans increased the regression coefficient when BB% was not included in the model. When BB% was included in the model the regression coefficient was either stable when phenotypes were rescaled, or decreasing when not rescaled. Lastly, the regression coefficient was slightly more stable when considering **GRM_{XB}** compared to **GRM_{SB}**.

Within the BB animals the regression coefficients were lowest (and closest to 1) when no Brahmans were included in training. Adding Brahman animals increased the regression coefficients. Adding BB% as a covariate reduced the range in regression coefficients across all other scenarios, particularly when no BB animals were included in the analysis. There was little difference in the regression coefficients between the two GRMs.

The principal difference when used a bivariate analysis were represented on Table 9 and Table 10, that represent the regression coefficient between the GEBV and adjusted phenotype to AGECL-BB and AGECL-TC. In these scenarios, the regression coefficients increased when compared with Table 6. The principal difference was AGECL-TC trait that showed highest results when compared with a univariate analysis. Just when added 3 BB groups that these values decreased, that can be occur because a highest correlation between these family groups. And observing the AGECL-BB in Brahman these values showed very high because increase the Brahman phenotype information leaving a better estimation of GEBV than compared with others scenarios.

Discussion

Genetic evaluation in mixed or admixed breed populations is complicated by the estimation of the effect of the ancestral breeds on each trait. The breed proportion in traditional analysis is calculated by tracing the parental breed through the pedigree. Using this approach each animal is given the average proportion of its parents, however through recombination the actual proportion inherited may vary from this due to Mendelian segregation. It has been proposed that breed component should be estimated from genomic information to use in genetic evaluation (Porto-Neto *et al.* 2013; Thomasen *et al.* 2013). Accuracy of breed composition estimated from high density genotype SNP panels are high (Frkonja *et al.* 2012; Kuehn *et al.* 2011) thus it would be expected that using these values in place of pedigree based estimates of breed proportions may increase accuracy. Accordingly, Thomasen *et al.* (2013) added breed proportion as a covariate in analysis of genomic data using random regression. In this case the accuracy of genomic selection was not improved, however in this study the divergence between the breeds was rather small as the two breeds (Danish and US Jersey populations) had only been separated for 100 years (Thomasen *et al.* 2013). This is in contrast to Brahman and the *Bos taurus* component of Tropical Composites which are estimated to have diverged hundreds of thousands of years ago. Accordingly, Porto-Neto *et al.*

(2013) suggested that the Zebu content could be added to genetic evaluation programs that include Tropical Composites.

Genomic predictions across breeds have low accuracy, particularly for breeds not represented within the training population (Erbe *et al.* 2012; Garrick 2011). However, when a minor breed is represented in both the training and validation populations the accuracy is often similar to or slightly better than training on the smaller population. For example (Erbe *et al.* 2012; Pryce *et al.* 2012) found that adding Holstein animals to a Jersey reference increased accuracy with either no reduction or a small reduction in Holstein accuracies depending on the trait. Similarly, Zhang *et al.* (2014) found that adding Brahman animals to TC increases accuracy for Tropical Composites, and this also was observed in our analysis: adding additional groups of Brahmans to the training population lead to consistent increases in realised accuracy.

This study confirmed that adding BB information can lead to increases in accuracy of TC using genomic evaluations. Adding breed specific GRMs did not improve the accuracy of genomic evaluation however it did improve the regression coefficient for TC animals, considering no covariate scenario (Q=No). This impact will be particularly important when there are animals that do not have links to animals in the current genetic evaluation. Such animals need to be placed into appropriate genetic groups. The effect of incorrect genetic grouping can have substantial impact on breeding value estimates (Misztal *et al.* 2013).

As noted it was observed that the Brahman regression coefficient was inflated when the value for the Tropical Composite regression coefficient was around 1 in all scenarios studied. So, an additional analysis was performed where the variances were adjusted so the Brahman regression coefficient was closer to 1, however under these parameters the Tropical Composite regressions were well below 1 (data not shown). Thus it does not seem possible to obtain correct regressions for both traits under a univariate model

Porto-Neto *et al.* (2013) estimated the Zebu content of this population using a different set of reference animals and a larger validation population: in their study 81 Angus and 29 Nelore were

used as reference animals. The Brahman animals used in our study would contain a proportion of *Bos taurus* genes as a consequence of the grading up process, where a small number of imported Brahman sires were crossed to Australian *Bos taurus* animals to produce the current industry Brahman herds. This is reflected in the Zebu average content of 95% in the analysis of (Porto-Neto *et al.* 2013). The contrasts with the estimate using Brahman animals as reference population (BB%=98) the estimate of BB% in the Tropical Composites was also slightly lower (43%) than the estimate of Porto-Neto *et al.* (2013).

All models does influence the precision of genomic evaluations, maybe the model used had a problem of multicollinearity, principally when include the sire and dam breed and Brahman proportion, and therefore highlights the importance of correctly accounting for breed in genetic evaluation. It is suggested that future work would examine the effect of BB% on multibreed GEBVs in more detail and examine the effect in additional data sets. However, the model used did not have an impact on the accuracy of prediction, but showed that adding Brahman information increase the predictive capacity in training population.

Conclusions

There was a clear benefit in adding Brahman animals to Tropical Composite genomic evaluations. The Brahman information with an accurate and high correlated between these two breeds is appropriated to evaluate the genomic breeding values in Tropical Composite breed. Considering the two breeds as separate traits for AGECL can be a strategy for obtain more precise information in prediction of genomic estimate breeding values.

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Table 1. Example of cross validation strategy used for each scenario examined. All possible combinations of groups were run within BB when < 3 groups were included in training (T) and validation (V)

Training Strategy Name	TC groups				Number of TC CV groups	BB groups				Number of BB CV groups	Number of analysis
	1	2	3	4		1	2	3	4		
0TC+3BB	V	V	V	V	0	T	T	T	V	3	4
3TC	T	T	T	V	3	V	V	V	V	0	4
3TC+1BB	T	T	T	V	3	T	V	V	V	1	4*4=16
3TC+2BB	T	T	T	V	3	T	T	V	V	2	4*6=24
3TC+3BB	T	T	T	V	3	T	T	T	V	3	4*4=16
3TC+4BB	T	T	T	V	3	T	T	T	T	4	4

Table 2. Average and standard deviation of estimated Brahman content results from Admixture, training animals were used in development of predictions and testing animals were excluded from training analysis.

Training			
Population	Mean	SD	N
BB	1.000	0.000	2000
BT	0.000	0.000	3650
Testing			
Population	Mean	SD	N
BB	0.974	0.048	3045
BT	0.002	0.011	1435
TC	0.412	0.086	1788

BB is a Brahman population; BT is a *Bos taurus* population; TC is a Tropical Composite population; SD is a Standard Deviation; and N is the total of animals used in Admixture

Table 3. Statistics of relationship coefficients for Brahman (BB), Tropical Composite (TC), between Brahman and Tropical Composite (BBTC) and all the population (FULL) using pedigree and genomic information

	Diagonal											
	FULL			BB			TC			BBTC		
	NRM	GRM _{SB}	GRM _{XB}	NRM	GRM _{SB}	GRM _{XB}	NRM	GRM _{SB}	GRM _{XB}	NRM	GRM _{SB}	GRM _{XB}
Average	1.002	0.766	0.760	1.002	0.796	0.789	1.001	0.738	0.734	-	-	-
Min.	1.000	0.689	0.692	1.000	0.742	0.741	1.000	0.689	0.692	-	-	-
Max.	1.266	0.899	0.888	1.266	0.899	0.888	1.158	0.864	0.861	-	-	-
Var.	0.000	0.001	0.001	0.000	0.000	0.000	0.000	0.000	0.000	-	-	-
Off-Diagonal												
	FULL			BB			TC			BBTC		
	NRM	GRM _{SB}	GRM _{XB}	NRM	GRM _{SB}	GRM _{XB}	NRM	GRM _{SB}	GRM _{XB}	NRM	GRM _{SB}	GRM _{XB}
Average	0.004	0.338	0.339	0.008	0.473	0.465	0.008	0.319	0.316	0.000	0.286	0.294
Min.	0.004	0.338	0.339	0.008	0.473	0.465	0.008	0.319	0.316	0.000	0.286	0.294
Max.	0.511	0.654	0.643	0.511	0.654	0.643	0.454	0.553	0.550	0.000	0.433	0.430
Var.	0.001	0.006	0.005	0.001	0.001	0.001	0.002	0.001	0.001	0.000	0.001	0.001

NRM – Pedigree based relationship matrix; GRM_{SB} elements adjusted by average allele frequency of the singlebreed dataset; GRM_{XB} elements of the GRM adjusted by individual animals breed proportion thus including breed allele frequencies.

Table 4. Averages of heritability and genetic parameters to 4 validation family groups for Brahman (BB), Tropical Composite (TC) and both (FULL) breeds using the numerator relationship matrix (NRM) and two genomic relationship matrices, with single breed (GRM_{SB}) and multi-breed (GRM_{XB})

NRM			
	h^2	σ_e^2	σ_u^2
FULL	0.546	5543.300	6671.800
BB	0.661	4636.425	9058.200
TC	0.464	5388.600	5706.450
GRM_{SB}			
	h^2	σ_e^2	σ_u^2
FULL	0.747	5847.000	17300.000
BB	0.841	4871.275	26248.000
TC	0.679	5866.550	12658.950
GRM_{XB}			
	h^2	σ_e^2	σ_u^2
FULL	0.747	5859.300	17263.000
BB	0.840	4890.500	26170.750
TC	0.678	5869.275	12650.650

Table 5. Realized correlations between genomic breeding values (GEBV) and adjusted phenotypes considering increasing numbers of Brahman animals in training (Row Q No/ Yes indicates BB% included as covariate in analysis; Rescale Yes/No indicates phenotypes Brahman and Tropical Composite animals rescaled to the same phenotypic variance; BD SB and XB indicate Single breed allele frequency and adjusted for breed specific allele frequency respectively)

Q	No				Yes			
	No		Yes		No		Yes	
	SB	XB	SB	XB	SB	XB	SB	XB
Rescale								
GRM								
Tropical Composites								
3BB	0.142	0.144	0.142	0.144	0.131	0.137	0.131	0.137
TC	0.151	0.151	0.151	0.151	0.178	0.177	0.178	0.177
TC+1BB	0.174	0.174	0.173	0.173	0.191	0.191	0.191	0.191
TC+2BB	0.196	0.195	0.194	0.193	0.205	0.206	0.205	0.206
TC+3BB	0.213	0.212	0.211	0.210	0.217	0.219	0.217	0.218
TC+4BB	0.227	0.226	0.225	0.223	0.226	0.230	0.226	0.229
Brahman								
3BB	0.335	0.334	0.335	0.334	0.336	0.335	0.336	0.335
TC	0.086	0.091	0.086	0.091	0.135	0.133	0.135	0.133
TC+1BB	0.242	0.243	0.237	0.238	0.266	0.265	0.263	0.262
TC+2BB	0.316	0.316	0.312	0.312	0.330	0.329	0.328	0.327
TC+3BB	0.334	0.333	0.332	0.332	0.344	0.343	0.344	0.342

*TC is cross validation with 3 groups included in training; Number preceding BB represents the number of BB cross validation groups included in training

Table 6. Regression coefficient between genomic breeding values (GEBV) and adjusted phenotypes considering increasing numbers of Brahman animals in training (Row Q No/ Yes indicates BB% included as covariate in analysis; Rescale Yes/No indicates phenotypes Brahman and Tropical Composite animals rescaled to the same phenotypic variance; BD SB and XB indicate Single breed allele frequency and adjusted for breed specific allele frequency respectively)

Q	No				Yes			
	Rescale No		Rescale Yes		Rescale No		Rescale Yes	
	BD	XB	SB	XB	BD	XB	SB	XB
Tropical Composites								
TC	0.783	0.789	0.746	0.752	1.018	1.015	0.971	0.968
TC+1BB	0.885	0.881	0.844	0.841	0.992	1.007	0.963	0.974
TC+2BB	0.944	0.936	0.908	0.900	0.972	0.995	0.955	0.975
TC+3BB	0.971	0.964	0.943	0.936	0.948	0.978	0.941	0.968
3BB	0.970	1.006	1.027	1.065	0.830	0.895	0.879	0.948
TC+4BB	0.976	0.972	0.957	0.952	0.919	0.956	0.921	0.954
Brahman								
TC	1.036	1.101	0.987	1.049	1.704	1.680	1.624	1.601
TC+1BB	1.895	1.906	1.879	1.892	2.054	2.051	2.063	2.059
TC+2BB	1.921	1.920	1.965	1.966	1.945	1.950	2.004	2.008
TC+3BB	1.690	1.687	1.757	1.754	1.693	1.696	1.767	1.769
3BB	1.749	1.750	1.852	1.853	1.729	1.734	1.831	1.835

*TC is cross validation with 3 groups included in training; Number preceding BB represents the number of BB cross validation groups included in training

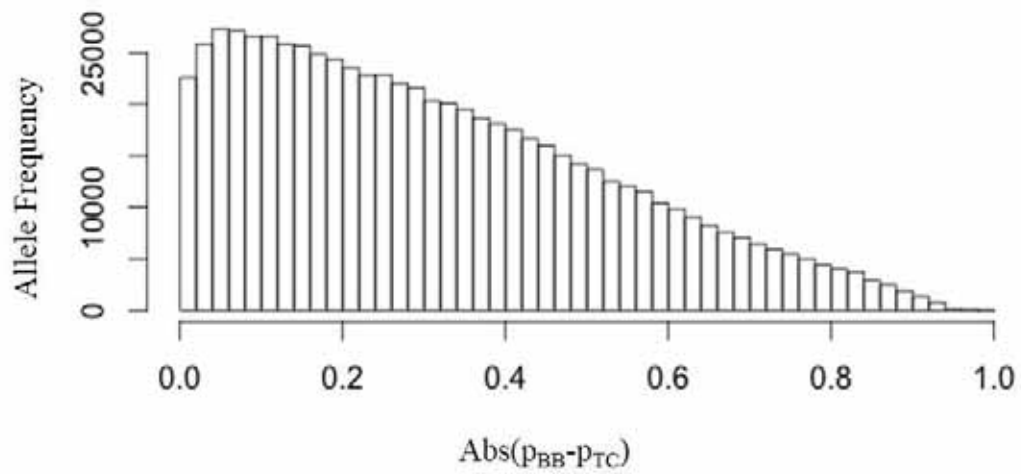


Figure 1 Histogram of the absolute value of the difference in allele frequency between Brahman (p_{BB}) and Bos Taurus (p_{TC}) for individual SNP (calculated across 6 BT breeds)

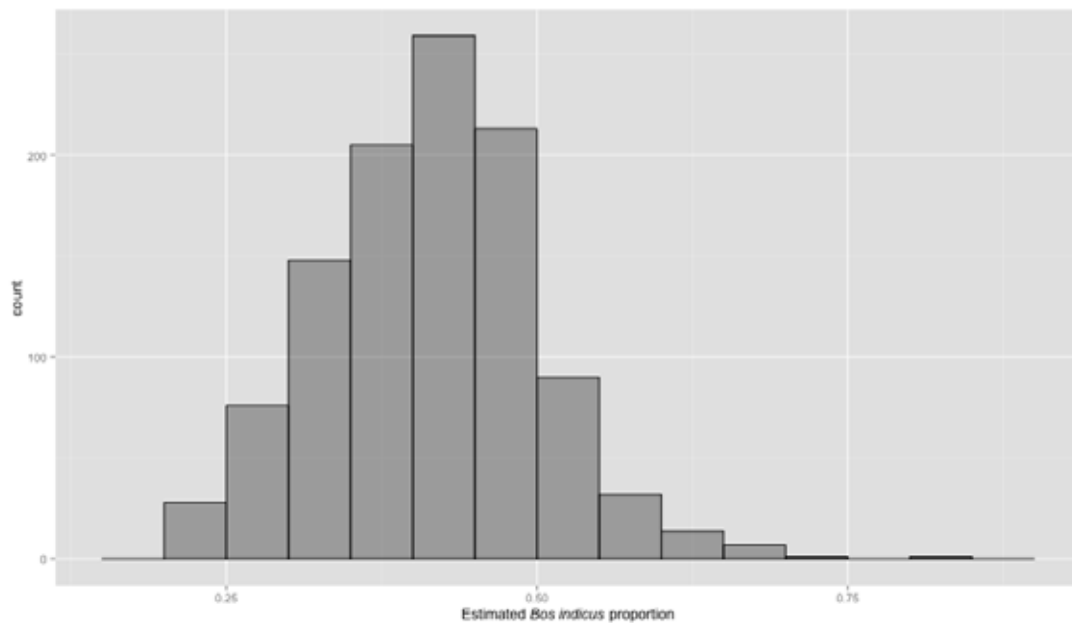


Figure 2 Histogram demonstrating the diversity of *Bos indicus* proportion estimates within Tropical Composite beef cattle.

CAPÍTULO 4 - CONSIDERAÇÕES FINAIS

A partir dos resultados encontrados neste estudo observa-se que as matrizes de parentesco utilizando as informações de dados genômicos podem ser uma importante informação para auxiliar na avaliação e seleção de gado de corte.

Mesmo não detectando diferença significativa na estimação dos parâmetros genéticos populacionais utilizando as diferentes matrizes, é possível notar a diferença da classificação dos animais em cada metodologia. Isto pode não ser de extrema importância para o melhoramento genético animal, principalmente se for selecionar grupos de animais, porém pode ter uma grande influência econômica já que existe diferença na posição de classificação dos animais melhores classificados. Por exemplo, o sêmen de um touro tem maior valor quanto melhor sua classificação na população e o número de doses vendidas também poderá alterar-se. Este trabalho indicou que existem diferenças neste ranqueamento dos indivíduos usando diferentes matrizes de relacionamento, porém seria interessante uma melhor investigação de qual matriz de relacionamento apresenta uma classificação mais acurada dos animais.

Para características de moderada a alta herdabilidade, a seleção genômica pode não ser viável quando comparada ao método tradicional devido ao alto custo de implementação e o baixo ganho na acurácia mesmo adicionando informações de relacionamento genético entre indivíduos não correlacionados pelo pedigree. Porém deve-se levar em conta que os bancos de dados com estas novas informações vêm crescendo e acredita-se que, no futuro, com o domínio da tecnologia e a redução do custo de genotipagem dos animais, esta nova metodologia poderá trazer grandes vantagens para características que podem ser medidas com precisão e que não tenham alta herdabilidade.

Quanto a avaliação multirracial no Brasil, apesar da população bovina de gado de corte ser predominante zebuína, cada vez mais vem sendo utilizado cruzamento entre raças devido à crescente exigência do mercado por cortes de melhor qualidade e para maior adaptação dos animais.

Os coeficientes de parentesco genômico podem levar a uma melhor avaliação genética dos animais e as novas metodologias propostas neste trabalho podem ser uma ferramenta importante para esta avaliação, pois como observado em alguns

trabalhos, animais *Bos taurus* apresentam melhor classificação de carcaça, aliado com as características já conhecidas do zebuino brasileiro pode-se atender as exigências do mercado sem perder a qualidade genética obtida nestes longos anos de melhoramento genético animal brasileiro.

Finalmente, deve-se considerar que todas as análises realizadas nestes trabalhos foram univariadas, ou seja, para uma característica. Portanto, seria interessante comparar estes resultados com outras análises utilizando várias características e também deve-se considerar a utilização de outras fontes de informação, como índices de seleção, podendo, assim, detectar algumas diferenças significativas nas metodologias descritas, tanto na determinação de qual matriz de parentesco é a mais adequada para a população em análise quanto na adição de informações de proporção de *Bos indicus* em populações multirraciais.