

# RESSALVA

Atendendo solicitação do(a) autor(a), o texto completo desta dissertação será disponibilizado somente a partir de 20/09/2026.

**SÃO PAULO STATE UNIVERSITY (UNESP)**

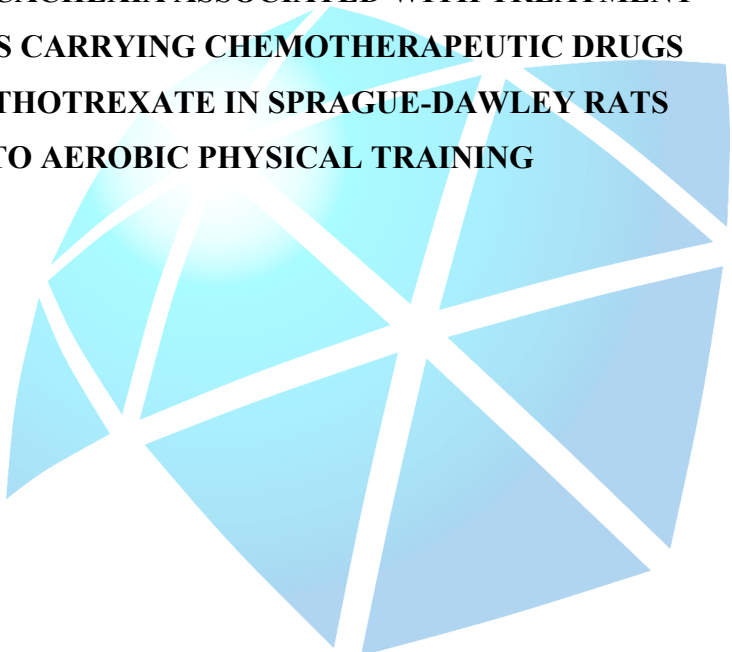
**“JÚLIO DE MESQUITA FILHO”**

**SCHOOL OF DENTISTRY, ARAÇATUBA**

**MASTER’S THESIS**

**RAFAEL RIBEIRO CORREIA**

**ANALYSIS OF CANCER CACHEXIA ASSOCIATED WITH TREATMENT  
WITH NANOPARTICLES CARRYING CHEMOTHERAPEUTIC DRUGS  
DOCETAXEL AND METHOTREXATE IN SPRAGUE-DAWLEY RATS  
SUBMITTED TO AEROBIC PHYSICAL TRAINING**



**ARAÇATUBA – SP**

**2024**

**SÃO PAULO STATE UNIVERSITY (UNESP)**

**“JÚLIO DE MESQUITA FILHO”**

**SCHOOL OF DENTISTRY, ARAÇATUBA**

**MASTER’S THESIS**

**RAFAEL RIBEIRO CORREIA**

**ANALYSIS OF CANCER CACHEXIA ASSOCIATED WITH TREATMENT  
WITH NANOPARTICLES CARRYING CHEMOTHERAPEUTIC DRUGS  
DOCETAXEL AND METHOTREXATE IN SPRAGUE-DAWLEY RATS  
SUBMITTED TO AEROBIC PHYSICAL TRAINING**

Thesis presented to the Multicenter Graduate Program in Physiological Sciences, of the School of Dentistry, Araçatuba, São Paulo State University “Júlio de Mesquita Filho”, to obtain the title of Master in Physiological Sciences.

Supervisor: Giovana Rampazzo Teixeira, Ph.D.

**ARAÇATUBA – SP**

**2024**

Catálogo na Publicação (CIP)

Diretoria Técnica de Biblioteca e Documentação – FOA / UNESP

Correia, Rafael Ribeiro.

C824a      Analysis of cancer cachexia associated with treatment with nanoparticles carrying chemotherapeutic drugs Docetaxel and Methotrexate in Sprague-Dawley rats submitted to aerobic physical training / Rafael Ribeiro Correia. - Araçatuba, 2024  
164 f. : il. ; tab.

Dissertação (Mestrado) – Universidade Estadual Paulista, Faculdade de Odontologia, Araçatuba

Orientadora: Profa. Giovana Rampazzo Teixeira

1. Cachexia 2. Physical Exercise 3. Docetaxel 4. Methotrexate  
I. T.

CDD 612

Claudio Hideo Matsumoto CRB-8/5550

## **CURRICULAR DATA**

**Born:** 12/22/1999, Monte Aprazível/SP, Brazil.

**Affiliation:** Clodoaldo Correia Primo

Jusceli Aparecida Ribeiro Correia

**2018-2022:** Bachelor's Degree in Physical Education, São Paulo State University (Unesp), School of Technology and Sciences, Presidente Prudente, São Paulo, Brazil.

**2022-2024:** Master's degree in Multicenter Graduate Program in Physiological Sciences (PPGMCF), São Paulo State University (Unesp), School of Dentistry (FOA), Araçatuba, São Paulo, Brazil.

**Names in bibliographic citations:**

CORREIA, R. R.; CORREIA, RAFAEL RIBEIRO; CORREIA, RAFAEL R.

## **DEDICATION**

To all those for whom this work, a mere piece of a gigantic scientific puzzle, may be useful.

To all those who came before us, whether in recent times or in remote times, facing difficulties and persecutions, and who nevertheless dedicated their lives to science.

To all those who will succeed us, whether shortly or even in the distant future, perhaps even in a better world than mine, where creativity does not encounter bureaucratic, temporal, and ideological barriers, and who are not judged by the number of articles published, but by their quality.



## **ACKNOWLEDGEMENTS**

First, on yet another occasion, I would like to take this opportunity to express my sincere gratitude to the staff at the FCT/Unesp student housing. I remember it as if it were yesterday: on February 12, 2018, I was arriving to visit the city that would be my home for four long years, and I was welcomed by wonderful people who took the time to show me everything. So, I would like to thank Laís, Eric, Jeferson, Thaís, Gabriel (Renato), Natanael (Mathias), Dielme, Taís, Cláudio, Moisés, Rafael (Di Maria), Lucas, Marcelo, Fagner, Jediael, Nilo and all the staff at B2, A3 and other housing units. You were my family, and without you, I am sure I would not have finished my undergraduate degree, much less my master's degree.

I would like to thank my colleagues who arrived with me on February 19, 2018, Thiago and Eduardo, and who share their moments with me to this day, each on their path. Thank you for all the learning and growth we had. It was difficult, but we managed.

I would also like to thank all the student permanence policies at FCT/Unesp, which made it possible for me to stay at the University. I would also like to take this opportunity to express my wish that all the policies become increasingly accessible and democratic so that no one is prevented from studying for financial reasons.

I would like to thank all my friends from home (all of whom are graduate students) who live in the same house and share the same difficulties: José, Ivete, Eduardo, Victor, and Tatiana. We mustn't forget our kittens Da Vince and Maria, who are our respite from our tiring academic life. I extend my thanks to Jeferson, my academic pupil.

I would like to thank my friends from my undergraduate studies. During the 4 years of our lives (unfortunately, 2 of them were spent remotely), we shared unforgettable moments that will probably become just stories told to future generations when we are asked about our youth. I am very grateful to my friends inside and outside UNESP who, directly or indirectly, shared experiences with me and helped me become who I am today. Class LVI of Physical Education, I will never forget you. You are eternal!

I would like to thank my advisor, Prof. Dr. Giovana Rampazzo Teixeira, for all the patience she had with me and for all the time she spent guiding me since 2018/2019. Her teachings, provocations, and ear-pulling were and continue to be of great value to my scientific and personal development. Thank you for your trust and patience during all this time.

I would also like to thank all the members of the Experimental Laboratory of Exercise Biology (LEBioEx), of the São Paulo State University (UNESP), School of Technology and Sciences, Presidente Prudente, Department of Physical Education,

Alice, Duda, Victor, Rebeca, Mateus, Wolfgang and João, for the opportunity to develop the project on their premises and for all the help and commitment they had in helping me carry out this work. During all this time, especially during the pandemic period, we became much more than work colleagues. We had disagreements, shared joys, and very rich discussions, situations that qualified us as a family.

I would like to express my deep gratitude to PhD student Caique de Figueiredo, a member of the Laboratory of Cellular Physiology of Exercise (LaFICE). In addition to all his help, Caique was always ready to help us in any way possible, sometimes leaving home on holidays and weekends just to open the freezer for us. He is a fantastic person and deserves all the affection possible. I would also like to take this opportunity to thank the current professor, Dr. Rafael Jesus Gonçalves Rubira, a professor at UNESP in Rio Claro, who, like Caique, was a former resident of the student housing at UNESP in Presidente Prudente. You two are my examples of life within the academy.

I would like to thank Prof. Dr. Antonio Hernandes Chaves Neto (and all his students), coordinator of the Saliva Biochemistry Laboratory (LABIQS), São Paulo State University (UNESP), School of Dentistry, Araçatuba, for the kindness and eagerness they showed in receiving us at their facilities and helping us conduct and analyze the oxidative data.

I would like to thank all the technical and administrative staff at FCT/Unesp, especially the cleaning, maintenance, security, and logistics professionals. Without you, our research would not be possible.

I would like to thank Dr. Teresa Audrey Zimmers, who warmly welcomed me to her country, giving me all the academic support possible and, above all, experiences that I never thought I deserved. I would also like to thank her students, who taught me a lot and delighted me during the time I was there. I would like to take this opportunity to extend my thanks to Oregon Health and Science University (OHSU).

I would like to thank Nick Toda, a North American who welcomed me into his home and who, outraged by the possibility that a Brazilian had never seen the sea, took me to accomplish this feat, even in an ocean in the northern hemisphere, too cold to stay in for at least 30 seconds.

I would like to thank Prof. Dr. Robson Francisco Carvalho and his valuable team at the Cancer Cachexia Omics Lab, Department of Structural and Functional Biology, Institute of Biosciences (IBB) of the São Paulo State University (UNESP), Botucatu,

whom I had the honor of meeting in Scotland, and who later helped me enormously, and I have no doubt that they will continue to do so.

I would like to thank my family for all the support they gave me throughout this process. You taught me to be strong and resilient, and here I am. Every choice and step I took when I left home was with the idea of one day giving back everything, you gave me. I am not yet the son I want to be, but I hope to get there one day!

I would like to especially thank Jade, my old classmate, the most charming Physical Education Teacher and Professional the world has ever seen, who shared difficult, joyful, and challenging moments with me. After all, she agreed to share her life with me, becoming my wife. More than that, she agreed to embark on a new adventure with me many miles away from here. I hope I live up to everything you represent to me. I love you and thank you very much!

I would like to thank the São Paulo Research Foundation (FAPESP) for the financial support (process number: 2021/14514-2 and 2022/13577-3).

Finally, I am immensely grateful to this country where I was born and raised. A country that has suffered, with a beautiful origin, but that still bleeds from a painful past, marked by European invasions and the terror of slavery. Even so, it is still known for the joy of its people, who do not let themselves be shaken by the difficulties, which are many. Today I say goodbye to it, with a heavy heart, without any guarantee of my return, but with unconditional love. Wherever I go, I will always be, proudly, a Brazilian boy who loves to play at being a scientist.

**EPIGRAPH**

“Peace for our time”

Neville Chamberlain

***RESUMO***

A caquexia associada ao câncer é uma síndrome multifatorial caracterizada pela perda progressiva de massa muscular esquelética, resistente à reversão por suporte nutricional convencional. Este estudo investigou os efeitos dos quimioterápicos docetaxel e metotrexato, emulsificados em nanopartículas LDE, em associação ao exercício aeróbico, em um modelo de ratos com caquexia do câncer. No Capítulo 1, a análise dos efeitos de um protocolo de exercício moderado revelou que a atividade aeróbica agravou a caquexia, aumentando a massa tumoral, promovendo significativa atrofia muscular e alterando vias metabólicas como o ciclo do TCA, fosforilação oxidativa e gliconeogênese. Observou-se também ativação de vias de neurodegeneração no músculo esquelético e modificação na expressão gênica tumoral relacionada à organização da matriz extracelular, sugerindo que o exercício após a instalação do tumor pode resultar em sobrecarga e piora nos desfechos. O Capítulo 2 avaliou a combinação de LDE-docetaxel com exercício, que não foi eficaz na preservação do músculo tibial anterior. A redução da tolerância ao exercício e a regulação negativa de genes críticos para a regeneração muscular, como *Cdk2*, *Nr4a1* e *Atf3*, indicaram comprometimento na recuperação muscular, evidenciando a complexa interação entre massa tumoral, o tratamento e a integridade muscular. No Capítulo 3, a investigação dos efeitos de LDE-metotrexato com exercício demonstrou que, embora houvesse redução no peso corporal total, a atrofia muscular foi acentuada, com desregulação de genes envolvidos na organização do citoesqueleto e na função mitótica, comprometendo a capacidade de regeneração muscular. Esses resultados indicam que, apesar da redução tumoral, as intervenções terapêuticas combinadas ao exercício impactaram negativamente a saúde muscular, ressaltando os desafios na gestão da caquexia em contextos oncológicos.

**Palavras-chave:** caquexia; exercício aeróbico; LDE; docetaxel; metotrexato; transcriptômica.



## **ABSTRACT**

Cancer cachexia is a multifactorial syndrome characterized by progressive skeletal muscle loss that remains unresponsive to conventional nutritional support. This study investigated the effects of the chemotherapeutic agents docetaxel and methotrexate, emulsified in LDE nanoparticles, in combination with aerobic exercise in a rat model of cancer cachexia. In Chapter 1, the analysis of a moderate exercise protocol revealed that aerobic activity exacerbated cachexia, increasing tumor mass, inducing significant muscle atrophy, and disrupting metabolic pathways such as the TCA cycle, oxidative phosphorylation, and gluconeogenesis. Neurodegenerative pathways were also activated in skeletal muscle, and tumor gene expression related to extracellular matrix organization was altered, suggesting that exercise following tumor onset may lead to overtraining and worsening outcomes. Chapter 2 evaluated the combination of LDE-docetaxel with exercise, which failed to preserve the tibialis anterior muscle. The observed reduction in exercise tolerance and downregulation of key genes involved in muscle regeneration, such as *Cdk2*, *Nr4a1*, and *Atf3*, indicated impaired muscle recovery, highlighting the complex interplay between tumor mass, treatment, and muscle integrity. Chapter 3 explored the effects of LDE-methotrexate combined with exercise, showing that while overall body weight decreased, muscle atrophy was pronounced, with significant gene dysregulation affecting cytoskeleton organization and mitotic function, thereby compromising muscle regenerative capacity. These findings suggest that although tumor reduction was achieved, the combined therapeutic and exercise interventions negatively impacted muscle health, underscoring the challenges in managing cachexia in oncological contexts.

**Keywords:** cachexia; aerobic exercise; LDE; docetaxel; methotrexate; transcriptomics.

## **LIST OF FIGURES**

<b>Figure 1. Summary of cancer cachexia.</b> Source: Own authorship. <b>a)</b> The cachexia word origin and actual definition of cancer cachexia were based on previous studies [8,60]. <b>b)</b> The incidence graph of cachexia in different types of cancer was used with the previous reference [7]. <b>c)</b> The arrow representing the three stages of cancer cachexia was based on the drawing presented previously [8,72]. <b>d)</b> The cachexia sex differences in the table were based on previous references [71–74]. <b>e)</b> The mechanisms governing cachexia were based on previous reviews [8,10,14,68,69].	37
<b>Figure 2. Cachexia canonical pathways.</b> (adapted from Argilés, et al, 2024 [68]).	39
<b>Figure 3. Ubiquitin-proteasome system.</b> (Original figure from Deshaies and Joazeiro, 2009 [92]).	40
<b>Figure 4. Activation of proteolysis by ROS.</b> Cytokines from tumors can activate intracellular pathways using ROS, such as H <sub>2</sub> O <sub>2</sub> , and contribute to protein breakdown through the ubiquitin-proteasome system. (Original figure from Assi and Rébillard, 2016 [100]).	41
<b>Figure 5. LDE nanoemulsion.</b> When injected into the bloodstream, LDE incorporates ApoE, and now is allowed to bind the LDLR in the cancer cell carrying the drug. (Original figure from Perez, et al, 2024 [121])	43
<b>Figure 6. Mechanisms of docetaxel in cancer cells.</b> (Original figure from Mikuła-Pietrasik, et al, [137]).	45
<b>Figure 7. Mechanisms of methotrexate in cancer cells.</b> (Original figure from Zarou, et al, 2021 [27]).	47
<b>Figure 8. Extramuscular and intramuscular energy resources in skeletal muscle during physical exercise.</b> (Original figure from Hargreaves and Spriet, 2020 [176]).	51
<b>Figure 9. Effects of physical exercise on cancer cachexia.</b> (Original figure from Halle, et al, 2020 [183]).	52
<b>Figure 10. Intramuscular mechanisms of overtraining.</b> (Original figure from Cheng, et al, 2020 [207]).	54
<b>Figure 11. Aerobic exercise exacerbated body weight loss, muscle loss, adipose tissue loss, and increased tumor weight.</b> <b>a)</b> Experimental design. <b>b)</b> Body weight over time, initial body weight, and tumor-free body weight at euthanasia. <b>c)</b> tumor weight as a fraction of total body weight. <b>d)</b> skeletal muscle weight normalized to IBW. <b>e)</b> adipose tissue weight. <b>f)</b> tibialis anterior histology. # = Significant difference between the Cancer+Ex and Cancer groups using paired t-test. Non-Cancer (n=10); Cancer (n=13) until the week 13. After that, the cancer group was divided into two groups: Cancer (n=7); and Cancer+Ex (n=6).	73
<b>Figure 12. The tibialis transcriptome revealed activation of inflammation, inhibition of oxidative metabolism, and induction of known cachexia markers.</b> <b>a)</b> Muscle transcriptome principal component analysis. Volcano plots of tibialis gene expression in <b>b)</b> Cancer vs Non-Cancer, and Cancer+Ex vs Non-Cancer. <b>c)</b> Heatmap of differentially expressed genes. <b>d)</b> Functional Enrichment Analysis of the first cluster found in the heatmap showing altered pathways ranked by significance. <b>e)</b> Functional enrichment analysis of the second cluster in the heatmap ranked by significance. <b>f)</b> Meta-analysis of the transcriptome between Cancer and Cancer+Ex and contrast of Gene Ontology pathway differences. Non-Cancer (n=4); Cancer (n=4); Cancer+Ex (n=4).	75
<b>Figure 13. Cancer-induced cachexia pathways in non-wasted muscle generally enhanced by exercise.</b> <b>a)</b> Cancer cachexia-related genes according to the Skeletal Muscle Wasting in Cancer Cachexia dataset provided by the Elsevier_Pathway_Collection library. <b>b)</b> Autophagy-related genes according to the Reactome_2022 library. <b>c)</b> Oxidative Stress-related genes according to the BioPlanet_2019 library. All the plotted genes are significantly different from the Control for both Cancer and Cancer+Ex. Non-Cancer (n=4); Cancer (n=4); Cancer+Ex (n=4).	76

**Figure 14. The reduction of exercise tolerance time was associated with the disruption of energy resource pathways and evidence of overtraining.** **a)** Decreasing exercise tolerance in the Cancer+Ex group over time. **b)** Heatmap of Oxidative Phosphorylation (KEGG\_2019\_Mouse library) and **c)** Glycolysis/Gluconeogenesis (KEGG\_2019\_Mouse library) genes in tibialis. Non-Cancer (n=4); Cancer (n=4); Cancer+Ex (n=4). ..... 79

**Figure 15. Tumor identity analysis suggests a rhabdomyosarcoma with transcriptome only modestly affected by exercise.** **a)** Histology of tumors. Bar=20µm, 400x magnification. **b)** Tumor top 1000 genes enrichment for cell type. **c)** Volcano plot and **d)** heatmap for all genes  $FC \geq |2|$  and  $FDR \leq 0.05$ . **e)** Functional enrichment analysis of the first gene cluster of the heatmap. **f)** Functional enrichment analysis of the second gene cluster of the heatmap. Cancer (n=4); Cancer+Ex (n=4). ..... 80

**Figure 16. Alterations in serum proteins versus tumor and tibialis transcriptomes.** **a)** Cytokine array of serum chemokines with more than 2-fold change. **b)** Respective gene expression in tumor and tibialis. The values were log-transformed for plotting. Non-Cancer (n=4); Cancer (n=4); Cancer+Ex (n=4). ..... 81

**Figure 17. Interactome analysis reveals possible crosstalk within and among tumor and tibialis.** **a)** Possible involved pathways in autocrine communication in the Cancer+Ex vs Cancer comparison for tumor and tibialis anterior tissues. **b)** Functional enrichment analysis of the genes presented in **a**. **c)** Possible pathways involved in paracrine communication in the Cancer+Ex vs Cancer comparison from tumor to tibialis. The node thickness represents the communication score. **d)** Functional enrichment analysis of the genes presented in **c** as an Alluvial Plot. The node thickness represents the communication score. **e)** Possible involved pathways in paracrine communication in the Cancer+Ex vs Cancer comparison from tibialis to tumor. The node thickness represents the communication score. **f)** Functional enrichment analysis of the genes presented in **f** as an Alluvial Plot. Cancer (n=4); Cancer+Ex (n=4). ..... 83

**Figure 18. Graphical abstract of the tumor-muscle crosstalk.** This figure illustrates the interactions between chemokines, cytokines, and growth factors between tumor and skeletal muscle. This interaction was efficient in inducing a cachexia phenotype by upregulation of cachexia canonical genes, such as *Fbxo32*, and *Trim63*. Aerobic exercise exacerbated the cachexia phenotype, increasing the tumor weight and accelerating muscle wasting. The autocrine communication in tumor and tibialis was mainly mediated by chemokines. In tumors, the *Cxcl6* and *Cxcr2* receptor cascade were candidates to mediate the effects. In tibialis, *Ccl19* and *Cxcl11* ligands were candidates for binding the *Cxcr3* receptor, as well as the *Ccl5* ligand to binding the *Ccr3* receptor. .... 88

**Figure 19. Morphometric analysis.** **a)** experimental design. BIA = bioelectrical impedance. **b)** heatmap for anatomical data of tumor, initial body weight (IBW), final tumor-free body weight (FBW), adipose tissue (AT), Liver, Tibialis Anterior (TA), gastrocnemius muscle (GAS), kidney, pancreas, Extensor digitorum longus muscle (EDL), soleus muscle (SOL). **c)** IBW in week 13. **d)** FBW in week 21. **e)** TA weight. **f)** relative tumor weight (%). All organs and muscle weights were presented as a relative weight by FBW. .... 106

**Figure 20. Tibialis RNA-Sequencing.** Using the filter  $FC > 1.5$  and  $FDR < 0.05$ , we identified the DEG for CA+LDE vs CA (**a**), CA+LDE-DTX vs CA+LDE (**b**), CA+LDE-DTX+Ex vs CA+LDE-DTX (**c**), CA+LDE-DTX+Ex vs CA+LDE-Ex (**d**), and CA+LDE-DTX+Ex vs CA (**e**). **f)** PCA plot representing all the 18,664 transcripts. **g)** Venn Diagram of the DEG. **h)** heatmap for all DEG. **i)** functional enrichment analysis of all genes into the first heatmap cluster. **j)** functional enrichment analysis of all genes into the second heatmap cluster. All pathways were ranked using the Log10 of the Combined Score, and the libraries used were GO\_Biological\_Process\_2023, and Reactome\_2022. .... 108

**Figure 21. Protein-protein interaction.** **a)** Interaction of CA+LDE-DTX+Ex vs CA+LDE-DTX differently expressed genes. **b)** Communication of CA+LDE-DTX+Ex vs CA differently

expressed genes. The communication was done using the web tool STRING (STRING: functional protein association networks (string-db.org)). The minimum required interaction score selected was 0.700 (high confidence). The disconnected nodes in the network were hidden for better visualization. The line color indicates the type of interaction evidence. Light blue = curated databases, pink = experimentally determined, green = gene neighborhood, blue = gene co-occurrence, light green = text mining, black = co-expression, matte blue = protein homology. The functional enrichment analysis was made using the web tool EnrichR (Enrichr (maayanlab.cloud)), and the biological process involved in each communication was obtained according to the GO\_Biological\_Process\_2023 library..... 110

**Figure 22. Fatigue resistance.** **a)** Exercise maximal capacity test in the CA+LDE+Ex and the CA+LDE-DTX+Ex group. \* = significant difference in the 2<sup>nd</sup> assessment compared to the 1<sup>st</sup> assessment. # = significant difference in the 3<sup>rd</sup> assessment compared to the 2<sup>nd</sup> assessment. \$ = significant difference in the 3<sup>rd</sup> assessment compared to the 1<sup>st</sup> assessment. We used the One-Way ANOVA test followed by the Tukey test, and the p-value adopted was p<0.05. **b)** heatmap showing the oxidative phosphorylation genes using the genes available in the GO\_Biological\_Process\_2023 library and applied to our normalized count's gene list. .... 111

**Figure 23. CSA analysis.** **a)** distribution of fiber enzymatic activity and the CSA of the CA group. **b)** distribution of fiber enzymatic activity and the CSA of the CA+LDE group. **c)** distribution of fiber enzymatic activity and the CSA of the CA+LDE+Ex group. **d)** distribution of fiber enzymatic activity and the CSA of the CA+LDE-DTX group. **e)** distribution of fiber enzymatic activity and the CSA of the CA+LDE-DTX+Ex group. CSA across the groups and fiber enzymatic activity, being high enzymatic activity (**f**), intermediate enzymatic activity (**g**), and low enzymatic activity (**h**). The One-Way ANOVA test was followed by the Tukey test, and the p-value adopted was p<0.05. .... 113

**Figure 24. Bioelectrical Impedance.** The paired BIA analysis showing the comparisons from 1<sup>st</sup> to 2<sup>nd</sup> assessment and 1<sup>st</sup> to 3<sup>rd</sup> assessment in the group CA (**a**), CA+LDE (**b**), CA+LDE+Ex (**c**), CA+LDE-DTX (**d**), and CA+LDE-DTX+Ex (**e**). The y-axes mean the reactance values and the x-axes mean the resistance values. We used the PhA values to correlate with TA weight, tumor weight, and FBW in the groups CA (**f**, **g**, **h**), CA+LDE (**j**, **k**, **l**), CA+LDE+Ex (**m**, **n**, **o**), CA+LDE-DTX (**p**, **q**, **r**), and CA+LDE-DTX+Ex (**s**, **t**, **u**). All of the correlations were made using the Pearson linear correlation. **i)** PhA value of all groups. **v)** heatmap for genes involved in Membrane Raft Organization (GO:0031579), and Protein Localization To Membrane Raft (GO:1903044) and applied to our gene normalized counts list..... 115

**Figure 25. Effects of MTX on the transcriptome of the Tibialis Anterior muscle.** **a)** Unsupervised PCA of a total of 19,134 transcripts. **b)** Venn diagram of DEGs. **c)** A total number of DEGs in each comparison. **d)** functional enrichment analysis of downregulated genes in CA+LDE-MTX+Ex vs CA according to the GO\_Biological\_Process\_2023 library. **e)** functional enrichment analysis of downregulated genes in CA+LDE-MTX+Ex vs CA+Ex according to the GO\_Biological\_Process\_2023 library. **f)** heatmap of 20 gene markers using a comparison to check the effect of aerobic exercise (CA+Ex and CA+LDE-MTX+Ex) vs the groups without exercise (CA and CA+LDE-MTX). .... 135

**Supplementary Figure 1. Food intake, correlations, and oxidative stress.** **a)** Food intake over time. **b)** Correlation of muscle wasting and tumor mass in the Cancer group, and Cancer+Ex group. **c)** heatmap of IBW, final body weight (FBW), and normalized TA, Liver, Kidney, Pancreas, GAS, EDL, SOL, epididymal adipose tissue (AT), and tumor weights (gray bars). **d)** The principal component analysis plot for morphometric data is presented in the heatmap. **e)** Biochemical assay of oxidative stress related enzymes. # = Significant difference between the Cancer+Ex group and the Cancer group using paired *t*-test. Non-Cancer (n=10);

Cancer (n=13) until the week 13. After that, the cancer group was divided into two groups: Cancer (n=7); and Cancer+Ex (n=6).....	90
<b>Supplementary Figure 2. Tumor identity, protein, and gene expression. a)</b> Dot plot of rhabdomyosarcoma marker genes. <b>b)</b> Localization of proteins in the tumor tissue protein array. <b>c)</b> Bar plot of tumor proteins increased > 2  FC with exercise. <b>d)</b> Dot plot of tumor Dlk1 gene expression, the only differentially expressed gene with a corresponding protein in <b>d)</b> . <b>e)</b> MA plot of tumor genes differentially expressed between Cancer and Cancer+Ex. Cancer (n=4); Cancer+Ex (n=4).....	91
<b>Supplementary Figure 3. Serum proteins versus tumor and tibialis transcriptomes. a)</b> Cytokine array of serum proteins with more than 2-fold change. <b>b)</b> Respective gene expression in tumor and tibialis. The values were log-transformed for plotting. Non-Cancer (n=4); Cancer (n=4); Cancer+Ex (n=4).....	92
<b>Supplementary Figure 4. Morphometric PCA and cachexia genes. a)</b> PCA representing the anatomical data. <b>b)</b> genes involved in Skeletal Muscle Wasting in Cancer Cachexia pathway according to the Elsevier_Pathway_Collection library.....	120
<b>Supplementary Figure 5. DTX-related genes and LDE uptake. a)</b> heatmap for docetaxel-involved genes according to the DSigDB library, also added the Ldlr and Vldlr genes. <b>b)</b> dot plot for the Ldlr gene in the CA+LDE and CA+LDE+Ex groups. <b>c)</b> table showing the LDE uptake by glycolytic muscles and tumor tissue. The CA+LDE and CA+LDE+Ex groups were used to represent the LDE uptake with or without exercise. cpm = counts per minute. ....	121
<b>Supplementary Figure 6. Exclusive and shared pathways. a)</b> Analysis of all altered pathways in all comparisons. Red intensities represent the Log10 of the Combined Score. The larger the circles, the bigger the -Log10 of the adjusted p-value. ....	122
<b>Supplementary Figure 7. Protein-protein interaction of CA+LDE-DTX+Ex vs CA+LDE+Ex comparison. a)</b> Interaction of CA+LDE-DTX+Ex vs CA+LDE-LDE+Ex differently expressed genes. The communication was done using the web tool STRING (STRING: functional protein association networks (string-db.org)). The minimum required interaction score selected was 0.700 (high confidence). The disconnected nodes in the network were hidden for better visualization. The line color indicates the type of interaction evidence. Light blue = curated databases, pink = experimentally determined, green = gene neighborhood, blue = gene co-occurrence, light green = text mining, black = co-expression, matte blue = protein homology. ....	123
<b>Supplementary Figure 8. Oxidative stress. a)</b> heatmap for genes involved in Cellular Response To Oxidative Stress (GO0034599) according to the GO_Biological_Process_2023 library. Biochemical assay for FRAP ( <b>b)</b> ), CAT ( <b>c)</b> ), SOD ( <b>d)</b> ), GPx ( <b>e)</b> ), GSH ( <b>f)</b> ), TBARS ( <b>g)</b> ), and Carbonylated Proteins ( <b>h)</b> ). ....	124
<b>Supplementary Figure 9. Histological analysis. a)</b> Tibialis Anterior histology. <b>b)</b> mast cells graph. <b>c)</b> collagen quantification graph. <b>d)</b> heatmap for genes involved in Muscle-mast Cell according to the Tabula_Sapiens library. <b>e)</b> heatmap for genes involved in Mast Cell Degranulation (GO:0043303) and Activation Involved In Immune Response (GO:0002279) according to the GO_Biological_Process_2023 library. <b>f)</b> heatmap for genes involved in Collagen Formation R-HAS-1474290 according to the Reactome_2022 library. <b>g)</b> heatmap for genes involved in Collagen-Containing Extracellular Matrix (GO:0062023) according to the GO_Cellular_Component_2023 library.....	125
<b>Supplementary Figure 10. Muscle fiber genes involved. a)</b> MHC genes. The statistical analysis was made using the One-Way ANOVA test followed by the Tukey test, and the p-value adopted was p<0.05.....	126

## **LIST OF TABLES**



<b>Table 1.</b> Complete list of DEGs.....	107
<b>Table 2.</b> Exclusively expressed genes among the comparisons.....	107
<b>Table 3.</b> Morphometric data .....	133
<b>Table 4.</b> Exclusively expressed genes among the comparisons.....	134
<b>Table 5.</b> Complete list of DEGs.....	134
<b>Supplementary Table 1.</b> Body weight raw data.....	92
<b>Supplementary Table 2.</b> Complete list of DEGs .....	92

## **ABBREVIATIONS**

**ADP** - Adenosine Diphosphate  
**AICAR** - Aminoimidazole-4-Carboxamide Ribonucleotide  
**AMPK** - AMP-Activated Protein Kinase  
**AP-1** - Activator Protein 1  
**ATP** - Adenosine Triphosphate  
**AT** - Adipose Tissue  
**BAX** - BCL2 Associated X Apoptosis Regulator  
**BAX** - Bcl-2-associated X protein  
**BCE** - Before the Common Era  
**BCL-2** - B-Cell Lymphoma 2  
**BCL-xL** - B-cell lymphoma-extra large  
**Ca<sup>+</sup>** - Calcium Ion  
**CAT** - Catalase  
**CC** - Cancer Cachexia  
**CD8+ T cells** - Cluster of Differentiation 8 Positive T Cells  
**CEMIB/UNICAMP** - Animal Center of the University of Campinas  
**CIFs** - Cancer-Inducing Factors  
**CK** - Creatine Kinase  
**Ckm** - Creatine Kinase, M-type  
**COX-2** - Cyclooxygenase-2  
**Des** - Desmin  
**DNA** - Deoxyribonucleic Acid  
**DMBA** - 7,2-Dimethylbenz(a)anthracene  
**DTX** - Docetaxel  
**E1** - Ubiquitin-Activating Enzyme  
**E2** - Ubiquitin-Carrier Enzyme  
**E3** - Ubiquitin-Protein Ligase  
**EACH** - Exercise Anti-Cachectic Hypothetical  
**EDL** - Extensor Digitorum Longus  
**ERK** - Extracellular Signal-Regulated Kinase  
**FADD** - Fas-Associated Death Domain  
**Fas receptor** - Fas Cell Surface Death Receptor  
**FBW** - Final Body Weight  
**FBXO32** - F-Box Protein 32 (also known as Atrogin-1)  
**FOXO3a** - Forkhead Box O3a  
**GAS** - Gastrocnemius  
**GLUT4** - Glucose Transporter Type 4  
**GPx** - Glutathione Peroxidase  
**HDL-c** - High-Density Lipoprotein Cholesterol  
**HR** - Heart Rate  
**Hras** - Harvey Rat Sarcoma Viral Oncogene  
**IBW** - Initial Body Weight  
**IFN- $\gamma$**  - Interferon-Gamma  
**IGF-1** - Insulin-Like Growth Factor-1  
**IL-1 $\beta$**  - Interleukin-1 Beta  
**IL-1ra** - Interleukin-1 Receptor Antagonist  
**IL-4** - Interleukin-4  
**IL-6** - Interleukin-6  
**IL-10** - Interleukin-10  
**LDE** - Lipid Nanoemulsion like Low-Density Lipoprotein (LDL)  
**LDL** - Low-Density Lipoprotein  
**LDL-c** - Low-Density Lipoprotein Cholesterol  
**LDLR** - LDL Receptor  
**LLC** - Lewis Lung Carcinoma  
**Mb** - Myoglobin  
**MCT4** - Monocarboxylate Transporter 4  
**NF- $\kappa$ B** - Nuclear Factor Kappa B  
**NK cells** - Natural Killer Cells  
**NP** - Nanoparticle  
**NR4A1** - Nuclear Receptor Subfamily 4 Group A  
**NRF-1** - Nuclear Respiratory Factor 1  
**NRF-2** - Nuclear Respiratory Factor 2  
**O<sub>2</sub>** - Oxygen  
**PAHs** - Polycyclic Aromatic Hydrocarbons  
**PDH** - Pyruvate Dehydrogenase  
**PGC-1 $\alpha$**  - Peroxisome Proliferator-Activated Receptor Gamma Coactivator 1-alpha  
**Pi** - Inorganic Phosphate  
**RNA-seq** - RNA Sequencing  
**ROS** - Reactive Oxygen Species  
**SC** - Satellite Cell  
**SD** - Sprague-Dawley  
**SOD** - Superoxide Dismutase  
**SOL** - Soleus  
**STAT3** - Signal Transducer and

Activator of Transcription 3

**TA** - Tibialis Anterior

**TCA cycle** - Tricarboxylic Acid Cycle

**Tfam** - Mitochondrial Transcription  
Factor A

**TGF- $\beta$**  - Transforming Growth Factor-  
Beta

**TNF- $\alpha$**  - Tumor Necrosis Factor-Alpha

**TRIM63** - Tripartite Motif Containing  
63

**TWEAK** - TNF-Related Weak Inducer

of Apoptosis

**Ub** - Ubiquitin

**UCP-3** - Uncoupling Protein 3

**VEGF** - Vascular Endothelial Growth  
Factor

**VO<sub>2</sub> Max** - Maximum Oxygen  
Consumption

**ZMP** - 5-Aminoimidazole-4-  
Carboxamide-1- $\beta$ -D-Ribofuranosyl-5'-  
Monophosphate

## **SUMMARY**

## CONTENTS

<b>RESUMO</b> .....	<b>14</b>
<b>ABSTRACT</b> .....	<b>16</b>
<b>LIST OF FIGURES</b> .....	<b>18</b>
<b>LIST OF TABLES</b> .....	<b>23</b>
<b>ABBREVIATIONS</b> .....	<b>25</b>
<b>SUMMARY</b> .....	<b>28</b>
<b>INTRODUCTION</b> .....	<b>30</b>
<b>THEORETICAL FRAMEWORK</b> .....	<b>34</b>
HISTORICAL FACTS ABOUT SKELETAL MUSCLE AND CANCER	
CACHEXIA .....	35
CLASSICAL MOLECULAR PATHWAYS IN CACHEXIA.....	37
CHOLESTEROL-RICH LIPID NANOEMULSION (LDE), CARRIER OF	
CHEMOTHERAPY DRUGS DOCETAXEL (DTX) AND METHOTREXATE	
(MTX).....	41
AEROBIC PHYSICAL EXERCISE .....	48
ANIMAL MODELING FOR CACHEXIA AND CARCINOGEN.....	55
<b>JUSTIFICATION</b> .....	<b>58</b>
<b>AIMS</b> .....	<b>60</b>
<b>CHAPTER 1</b> .....	<b>62</b>
INTRODUCTION .....	66
METHODS .....	67
RESULTS .....	72
DISCUSSION.....	84
DATA AVAILABILITY STATEMENT .....	88
AUTHOR CONTRIBUTIONS.....	88
CONFLICT OF INTEREST .....	89
ACKNOWLEDGMENTS .....	89
SUPPLEMENTARY FIGURES.....	89
SUPPLEMENTARY TABLES .....	92
<b>CHAPTER 2</b> .....	<b>93</b>
INTRODUCTION .....	97
METHODS .....	98
RESULTS .....	105
DISCUSSION.....	116
CONCLUSIONS .....	118
AUTHORS CONTRIBUTIONS .....	119
ACKNOWLEDGMENT .....	119
SUPPLEMENTARY FIGURES.....	119
<b>CHAPTER 3</b> .....	<b>127</b>
INTRODUCTION .....	130
METHODS .....	130
RESULTS .....	132
DISCUSSION.....	135
ACKNOWLEDGEMENTS.....	137
CONFLICT OF INTEREST .....	137
<b>FINAL CONSIDERATIONS</b> .....	<b>138</b>
<b>REFERENCES</b> .....	<b>141</b>
<b>ANNEXES</b> .....	<b>167</b>

## **INTRODUCTION**

Cancer is one of the leading causes of death today [1]. It is estimated that in the period 2026-2030, 690,394 new cases will be diagnosed among women, while men will account for 757,806 new cases [2]. Worldwide, lung cancer is the most diagnosed type of cancer, accounting for around 12.4% of all cases globally (2.5 million new cases in 2022 alone), followed by breast cancer (11.6%), colorectal cancer (9.6%), prostate cancer (7.3%) and stomach cancer (4.9%) [3]. Risk factors for the development of cancer can be divided into 3 categories, namely 1) behavioral risks (smoking and alcoholism), 2) environmental and occupational risks (environmental pollution and occupational exposure to asbestos), and 3) metabolic risks (obesity and high glycemic index) [4].

Approximately 80% of patients with advanced cancer have cancer cachexia (CC) [5,6]. CC can manifest itself through weight loss, muscle weakness, anorexia, fatigue, and other multifactorial symptoms [7,8]. CC is defined as a multifactorial syndrome characterized by continuous loss of skeletal muscle mass (with or without loss of adipose tissue) that is not fully reversed by conventional nutritional support [9]. CC promotes a decrease in functional capacity, reduced quality of life and survival, constituting one of the main prognostic factors in the advanced stage of cancer [10]. It is estimated that 22-30% of causes of death from cancer are related to cancer cachexia [10]. However, due to the severity of each type of cancer, different incidences of cancer cachexia can be found, with 45% of lung cancer patients developing cachexia [11] and 25% with breast cancer [12], 28-80% of people with kidney disorders [13] and 85% for pancreatic cancer [14].

Nowadays, cachexia is an underestimated and under-researched muscle disease when compared to its causes. In general, it is estimated that 50% of cancer patients develop cachexia [6]. The etiology of cancer-induced cachexia is linked to metabolic disorders originating from the tumor, causing increased muscle catabolism and decreased protein synthesis [15]. The syndrome presents with 3 stages of severity, namely: 1) pre-cachexia, where there is a loss of body mass of up to 5% resulting from factors such as anorexia and insulin resistance; 2) cachexia characterized by a loss of body mass greater than 5% in the last 6 months; 3) refractory cachexia, a result of very advanced cancer (preterminal) associated with active catabolism, low-performance status and a life expectancy of fewer than 3 months [9]. However, there is still no consensus on the exact distinction between non-cachectic and pre-cachectic individuals [16,17].

The use of nanoemulsions has become an alternative in the biomedical field for the transport of drugs [18]. The encapsulation of drugs in their lipid interior protects them from degradation and reduces toxicity, as well as favoring their internalization into target



cells [19]. In this sense, Maranhão et al [20,21] developed a lipid nanoemulsion like LDL called LDE with impregnation in solid tumors after its administration into the bloodstream [20,21]. It was also successfully demonstrated by the group of Professor Maranhão, RC., that the use of the chemotherapy drug docetaxel (DTX) associated with LDE promoted significant reductions in aortic atheroma in rabbits [22], without demonstrating cytotoxic effects [22]. In this regard, the LDE nanoemulsion associated with DTX has also been shown to be effective in reducing prostate cancer [23]. Due to the versatility of LDE in drug delivery, the use of methotrexate associated with LDE has already been shown to be effective [24,25]. Methotrexate is indicated for the treatment of autoimmune diseases and chemotherapy for the treatment of cancer [26]. However, when used alone, it presents high cytotoxicity [27]. On the other hand, Maranhão et al. showed that the administration of LDE-MTX presents lower hematologic cytotoxicity [28]. It was found that the use of LDE associated with MTX reduces approximately 67% of macrophage infiltration in atherosclerotic plaques in rabbits [25]. However, we still do not know the effects of the use of LDE associated with chemotherapeutics (DTX and MTX) in cancer cachexia.

Physiological responses induced by physical exercise can mitigate the effects caused by senility and comorbidities, as well as minimize the impact of progressive loss of muscle mass associated with loss of strength and reduced physical performance, characterized as a syndrome called sarcopenia [29]. Furthermore, it is known in the current literature that physical exercise is useful for patients who develop cancer cachexia [30,31]. Skeletal muscle, an organ affected by cancer cachexia, improves with physical exercise, as it increases protein synthesis and mitochondrial biogenesis [32]. Moderate physical exercise (50 to 70% max HR) promotes physiological and muscular adaptations that are well described in the literature [33,34], inducing the release of anti-inflammatory cytokines derived from muscle contraction [35,36]. These cytokines are involved in inhibiting inflammation and regulating muscle energy metabolism [37,38]. These are the foundations that support the Exercise Anti-Cachectic Hypothetical – EACH [39], since, due to the endocrine characteristic of skeletal muscle during exercise [40,41] and the release of anti-inflammatory myokines, such as IL-1ra, IL-4, and IL-10, exercise becomes a tool against the progression of cachexia [42].

Given the advanced development of cancer and traditional treatments (chemotherapies, surgeries, and radiotherapy), attenuating cachexia in cancer patients has become an essential challenge. In this sense, studies that focus on the discovery of new

therapies are of utmost importance. In addition, studies on the efficacy and safety of physical exercise as a strategy associated with pharmacological action in the prevention of cancer cachexia are still inconclusive. To identify new paths for the treatment of cachexia, the present study characterized the genetic, molecular, and biochemical effects of LDE nanoemulsion carrying the chemotherapeutic agent docetaxel and methotrexate, associated with aerobic physical training. These results shed light on cancer cachexia and establish possible mechanisms of tumor action.

## **FINAL CONSIDERATIONS**

This thesis investigated the complex interactions between cancer cachexia, aerobic exercise, and chemotherapy drugs (docetaxel and methotrexate) emulsified in lipid-based nanoparticles (LDE) on skeletal muscle health in a rat model of cancer. The findings across the three chapters provide valuable insights into how these factors interplay at the molecular, transcriptomic, and morphological levels.

In **Chapter 1**, we demonstrated that while moderate aerobic exercise is often considered beneficial for skeletal muscle adaptations, in the context of cancer cachexia, it can have detrimental effects. The exercise protocol implemented in this study resulted in overtraining, evidenced by reduced exercise tolerance and exacerbated cachexia, as seen by decreased tibialis anterior muscle weight and increased tumor weight. Additionally, there was notable chemokine-mediated crosstalk between the tumor and muscle, particularly involving *Cxcl6/Cxcr2* genes, which suggests that exercise under these conditions can potentially worsen outcomes by promoting a cachexia-like transcriptome and disrupting muscle energy metabolism.

**Chapter 2** expanded on this by investigating the effects of docetaxel (DTX) delivered via LDE nanoparticles in combination with exercise. The results showed that the LDE-DTX combination was not sufficient to prevent muscle wasting. This was accompanied by downregulation of genes critical for muscle regeneration, such as *Cdk2*, *Nr4a1*, and *Atf3*, further highlighting compromised muscle recovery. The use of electrical bioimpedance as a non-invasive monitoring tool showed promise in tracking the progression of cachexia and muscle loss over time, offering a potential clinical application for managing cachexia in cancer patients.

**Chapter 3** focused on the effects of methotrexate (MTX) carried by LDE nanoparticles in conjunction with exercise. The findings indicated that aerobic exercise exacerbated body weight loss, while all treatment groups involving LDE-MTX experienced muscle wasting compared to controls. The downregulation of key pathways involved in spindle microtubule attachment and organization in the CA+LDE-MTX+Ex group further suggests a mechanistic basis for the observed muscle atrophy. These results point to the complex and often counterintuitive effects of combining chemotherapy and exercise, particularly in the presence of cachexia.

**Overall Conclusion:** Collectively, these findings underscore the complexity of managing cancer cachexia, particularly about physical exercise and chemotherapeutic interventions. Both docetaxel and methotrexate, when delivered via LDE nanoparticles, and their combination with moderate exercise appears to compromise muscle health,

especially in the context of advanced cachexia. This thesis provides critical insights into the biological processes affected by these interventions, revealing potential biomarkers and therapeutic targets for better managing cachexia in cancer patients. Future research should focus on optimizing exercise protocols and exploring adjunctive treatments that can mitigate muscle loss while enhancing chemotherapy efficacy.

## **REFERENCES**

- [1] Silva GAE, Jardim BC, Ferreira V de M, Junger WL, Girianelli VR. Cancer mortality in the Capitals and in the interior of Brazil: a four-decade analysis. *J Public Health (Bangkok)* 2020;54:126. <https://doi.org/10.11606/s1518-8787.2020054002255>.
- [2] Barbosa IR, De Souza DLB, Bernal MM, Do C.c. Costa Í. Cancer mortality in Brazil: Temporal trends and predictions for the year 2030. *Medicine (United States)* 2015;94:1–6. <https://doi.org/10.1097/MD.0000000000000746>.
- [3] Bray F, Laversanne M, Sung H, Ferlay J, Siegel RL, Soerjomataram I, et al. Global cancer statistics 2022: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin* 2024;74:229–63. <https://doi.org/10.3322/caac.21834>.
- [4] Tran KB, Lang JJ, Compton K, Xu R, Acheson AR, Henrikson HJ, et al. The global burden of cancer attributable to risk factors, 2010–19: a systematic analysis for the Global Burden of Disease Study 2019. *The Lancet* 2022;400:563–91. [https://doi.org/10.1016/S0140-6736\(22\)01438-6](https://doi.org/10.1016/S0140-6736(22)01438-6).
- [5] Tisdale MJ. Reversing Cachexia. *Cell* 2010;142:511–2. <https://doi.org/10.1016/j.cell.2010.08.004>.
- [6] von Haehling S, Anker SD. Cachexia as a major underestimated and unmet medical need: Facts and numbers. *J Cachexia Sarcopenia Muscle* 2010;1:1–5. <https://doi.org/10.1007/S13539-010-0002-6>.
- [7] Esper DH, Harb WA. The cancer cachexia syndrome: A review of metabolic and clinical manifestations. *Nutrition in Clinical Practice* 2005;20:369–76. <https://doi.org/10.1177/0115426505020004369>.
- [8] Baracos VE, Martin L, Korc M, Guttridge DC, Fearon KCH. Cancer-associated cachexia. *Nat Rev Dis Primers* 2018;4:1–18. <https://doi.org/10.1038/nrdp.2017.105>.
- [9] Fearon K, Strasser F, Anker SD, Bosaeus I, Bruera E, Fainsinger RL, et al. Definition and classification of cancer cachexia: An international consensus. *Lancet Oncol* 2011;12:489–95. [https://doi.org/10.1016/S1470-2045\(10\)70218-7](https://doi.org/10.1016/S1470-2045(10)70218-7).
- [10] Lim S, Brown JL, Washington TA, Greene NP. Development and progression of cancer cachexia: Perspectives from bench to bedside. *Sports Medicine and Health Science* 2020;2:177–85. <https://doi.org/10.1016/j.smhs.2020.10.003>.
- [11] Zhu R, Liu Z, Jiao R, Zhang C, Yu Q, Han S, et al. Updates on the pathogenesis of advanced lung cancer-induced cachexia. *Thorac Cancer* 2019;10:8–16. <https://doi.org/10.1111/1759-7714.12910>.
- [12] Simon Gallot Y, Costelli P, Blaauw B, Rébillard A, Pagano AF, Mallard J, et al. Skeletal Muscle Deconditioning in Breast Cancer Patients Undergoing Chemotherapy: Current Knowledge and Insights From Other Cancers 2021. <https://doi.org/10.3389/fcell.2021.719643>.
- [13] Koppe L, Fouque D, Kalantar-Zadeh K. Kidney cachexia or protein-energy wasting in chronic kidney disease: facts and numbers. *J Cachexia Sarcopenia Muscle* 2019;10:479–84. <https://doi.org/10.1002/jcsm.12421>.

- [14] Henderson SE, Makhijani N, Mace TA. Pancreatic Cancer-Induced Cachexia and Relevant Mouse Models. *Pancreas* 2018;47:937–45. <https://doi.org/10.1097/MPA.0000000000001124>.
- [15] Baracos VE, Mazurak VC, Bhullar AS. Cancer cachexia is defined by an ongoing loss of skeletal muscle mass. *Ann Palliat Med* 2019;8:3–12. <https://doi.org/10.21037/APM.2018.12.01>.
- [16] Blum D, Stene GB, Solheim TS, Fayers P, Hjermstad MJ, Baracos VE, et al. Validation of the Consensus-Definition for Cancer Cachexia and evaluation of a classification model—a study based on data from an international multicentre project (EPCRC-CSA). *Annals of Oncology* 2014;25:1635–42. <https://doi.org/10.1093/annonc/mdu086>.
- [17] Blauwhoff-Buskermolen S, de van der Schueren MAE, Verheul HMW, Langius JAE. “Pre-cachexia”: a non-existing phenomenon in cancer? *Annals of Oncology* 2014;25:1668–9. <https://doi.org/10.1093/annonc/mdu178>.
- [18] Elzayat A, Adam-Cervera I, Álvarez-Bermúdez O, Muñoz-Espí R. Nanoemulsions for synthesis of biomedical nanocarriers. *Colloids Surf B Biointerfaces* 2021;203. <https://doi.org/10.1016/j.colsurfb.2021.111764>.
- [19] Jaiswal M, Dudhe R, Sharma PK. Nanoemulsion: an advanced mode of drug delivery system. *3 Biotech* 2015;5:123–7. <https://doi.org/10.1007/s13205-014-0214-0>.
- [20] Maranhão RC, Graziani SR, Yamaguchi N, Melo RF, Latrilha MC, Rodrigues DG, et al. Association of carmustine with a lipid emulsion: In vitro, in vivo and preliminary studies in cancer patients. *Cancer Chemother Pharmacol* 2002;49:487–98. <https://doi.org/10.1007/s00280-002-0437-3>.
- [21] Maranhão RC, Garicochea B, Silva EL, Dorlhiac-Llacer P, Cadena SMS, Coelho IJC, et al. Plasma Kinetics and Biodistribution of a Lipid Emulsion Resembling Low-Density Lipoprotein in Patients with Acute Leukemia. *Cancer Res* 1994;54:4660–6.
- [22] Meneghini BC, Tavares ER, Guido MC, Tavoni TM, Stefani HA, Kalil-Filho R, et al. Lipid core nanoparticles as vehicle for docetaxel reduces atherosclerotic lesion, inflammation, cell death and proliferation in an atherosclerosis rabbit model. *Vascul Pharmacol* 2019;115:46–54. <https://doi.org/10.1016/j.vph.2019.02.003>.
- [23] Veras ASC, Batista VRG, Correia RR, de Almeida Tavares ME, Rubira RJG, Tavares ER, et al. Integrated aerobic exercise with LDE-docetaxel treatment: a novel approach to combat prostate cancer progression. *Sci Rep* 2024;14:9626. <https://doi.org/10.1038/S41598-024-60138-Y>.
- [24] Maranhão RC, Guido MC, de Lima AD, Tavares ER, Marques AF, de Melo MDT, et al. Methotrexate carried in lipid core nanoparticles reduces myocardial infarction size and improves cardiac function in rats. *Int J Nanomedicine* 2017;12:3767–84. <https://doi.org/10.2147/IJN.S129324>.
- [25] Bulgarelli A, Leite ACA, Dias AAM, Maranhão RC. Anti-atherogenic effects of methotrexate carried by a lipid nanoemulsion that binds to ldl receptors in cholesterol-fed rabbits. *Cardiovasc Drugs Ther* 2013;27:531–9. <https://doi.org/10.1007/s10557-013-6488-3>.



- [26] Koźmiński P, Halik PK, Chesori R, Gniazdowska E. Overview of dual-acting drug methotrexate in different neurological diseases, autoimmune pathologies and cancers. *Int J Mol Sci* 2020;21. <https://doi.org/10.3390/ijms21103483>.
- [27] Zarou MM, Vazquez A, Vignir Helgason G. Folate metabolism: a re-emerging therapeutic target in haematological cancers. *Leukemia* 2021;35:1539–51. <https://doi.org/10.1038/s41375-021-01189-2>.
- [28] Moura JA, Valduga CJ, Tavares ER, Kretzer IF, Maria DA, Maranhão RC. Novel formulation of a methotrexate derivative with a lipid nanoemulsion. *Int J Nanomedicine* 2011;2285–95. <https://doi.org/10.2147/IJN.S18039>.
- [29] Simioni C, Zauli G, Martelli AM, Vitale M, Sacchetti G, Gonelli A, et al. Oxidative stress: Role of physical exercise and antioxidant nutraceuticals in adulthood and aging. *Oncotarget* 2018;9:17181–98. <https://doi.org/10.18632/oncotarget.24729>.
- [30] Sadeghi M, Keshavarz-Fathi M, Baracos V, Arends J, Mahmoudi M, Rezaei N. Cancer cachexia: Diagnosis, assessment, and treatment. *Crit Rev Oncol Hematol* 2018;127:91–104. <https://doi.org/10.1016/j.critrevonc.2018.05.006>.
- [31] Lira FS, Antunes BDMM, Seelaender M, Neto JCR. The therapeutic potential of exercise to treat cachexia. *Curr Opin Support Palliat Care* 2015;9:317–24. <https://doi.org/10.1097/SPC.000000000000170>.
- [32] Alves CRR, Da Cunha TF, Da Paixão NA, Brum PC. Aerobic exercise training as therapy for cardiac and cancer cachexia. *Life Sci* 2015;125:9–14. <https://doi.org/10.1016/j.lfs.2014.11.029>.
- [33] Tomiya S, Kikuchi N, Nakazato K. Moderate Intensity Cycling Exercise after Upper Extremity Resistance Training Interferes Response to Muscle Hypertrophy but Not Strength Gains. *J Sports Sci Med* 2017;16:391–5.
- [34] Kirby TJ, McCarthy JJ. MicroRNAs in skeletal muscle biology and exercise adaptation 2013:95–105. <https://doi.org/10.1016/j.freeradbiomed.2013.07.004>.
- [35] Pedersen BK. Anti-inflammatory effects of exercise: role in diabetes and cardiovascular disease. *Eur J Clin Invest* 2017;47:600–11. <https://doi.org/10.1111/eci.12781>.
- [36] Fiuza-Luces C, Garatachea N, Berger NA, Lucia A. Exercise is the real polypill. *Physiology* 2013;28:330–58. <https://doi.org/10.1152/physiol.00019.2013>.
- [37] Saraiva M, Saraiva M, Vieira P, Vieira P, Vieira P, O’Garra A, et al. Biology and therapeutic potential of interleukin-10. *Journal of Experimental Medicine* 2020;217:1–19. [https://doi.org/10.1084/jem\\_20190418](https://doi.org/10.1084/jem_20190418).
- [38] Muñoz-Cánoves P, Scheele C, Pedersen BK, Serrano AL. Interleukin-6 myokine signaling in skeletal muscle: A double-edged sword? *FEBS Journal* 2013;280:4131–48. <https://doi.org/10.1111/febs.12338>.
- [39] Battaglini CL, Hackney AC, Goodwin ML. Cancer Cachexia: Muscle Physiology and Exercise Training. *Cancers (Basel)* 2012;4:1247–51. <https://doi.org/10.3390/cancers4041247>.

- [40] Pedersen BK, Febbraio MA. Muscle as an Endocrine Organ: Focus on Muscle-Derived Interleukin-6. *Physiol Rev* 2008;88:1379–406. <https://doi.org/10.1152/physrev.90100.2007>.
- [41] Pedersen BK, Febbraio MA. Muscles, exercise and obesity: Skeletal muscle as a secretory organ. *Nat Rev Endocrinol* 2012;8:457–65. <https://doi.org/10.1038/nrendo.2012.49>.
- [42] Daou HN. Exercise as an anti-inflammatory therapy for cancer cachexia: a focus on interleukin-6 regulation. *Am J Physiol Regul Integr Comp Physiol* 2019;318. <https://doi.org/10.1152/ajpregu.00147>.
- [43] Musil V, Suchomel Z, Malinova P, Stingl J, Vlcek M, Vacha M. The history of Latin terminology of human skeletal muscles (from Vesalius to the present). *Surgical and Radiologic Anatomy* 2015;37:33–41. <https://doi.org/10.1007/s00276-014-1305-7>.
- [44] Blanc Y, Dimanico U. History of the Study of Skeletal Muscle Function with Emphasis on Kinesiological Electromyography. *Open Rehabil J* 2010;3:84–93. <https://doi.org/10.2174/1874943701003010084>.
- [45] Huxley AF, Niedergerke R. Structural changes in muscle during contraction - interference microscopy of living muscle fibres. *Nature* 1954;173:971–3. <https://doi.org/10.1038/173971a0>.
- [46] Huxley Hugh, Hanson Jean. Changes in the cross-striations of muscle during contraction and stretch and their structural interpretation. *Nature* 1954;173:973–6. <https://doi.org/10.1038/173973a0>.
- [47] Mauro A. Satellite cell of skeletal muscle fibers. *J Biophys Biochem Cytol* 1961;9:493–5. <https://doi.org/10.1083/jcb.9.2.493>.
- [48] Schultz E, Gibson MC, Champion T. Satellite cells are mitotically quiescent in mature mouse muscle: An EM and radioautographic study. *Journal of Experimental Zoology* 1978;206:451–6. <https://doi.org/10.1002/jez.1402060314>.
- [49] Schultz E. A quantitative study of the satellite cell population in postnatal mouse lumbrical muscle. *Anat Rec* 1974;180:589–95. <https://doi.org/10.1002/ar.1091800405>.
- [50] Buckingham M, Bajard L, Chang T, Daubas P, Hadchouel J, Meilhac S, et al. The formation of skeletal muscle: from somite to limb. *J Anat* 2003;202:59–68. <https://doi.org/10.1046/j.1469-7580.2003.00139.x>.
- [51] Gros J, Manceau M, Thomé V, Marcelle C. A common somitic origin for embryonic muscle progenitors and satellite cells. *Nature* 2005;435:954–8. <https://doi.org/10.1038/nature03572>.
- [52] Mckinnell IW, Rudnicki MA. Developmental biology: one source for muscle. *Nature* 2005;435:898–9. <https://doi.org/10.1038/435898a>.
- [53] White RB, Biérinx AS, Gnocchi VF, Zammit PS. Dynamics of muscle fibre growth during postnatal mouse development. *BMC Dev Biol* 2010;10. <https://doi.org/10.1186/1471-213X-10-21>.

- [54] Cheung TH, Rando TA. Molecular regulation of stem cell quiescence. *Nat Rev Mol Cell Biol* 2013;14:329–40. <https://doi.org/10.1038/nrm3591>.
- [55] Forbes SJ, Rosenthal N. Preparing the ground for tissue regeneration: From mechanism to therapy. *Nat Med* 2014;20:857–69. <https://doi.org/10.1038/nm.3653>.
- [56] Wosczyzna MN, Rando TA. A Muscle Stem Cell Support Group: Coordinated Cellular Responses in Muscle Regeneration. *Dev Cell* 2018;46:135–43. <https://doi.org/10.1016/j.devcel.2018.06.018>.
- [57] Roman W, Pinheiro H, Pimentel MR, Segalés J, Oliveira LM, García-Domínguez E, et al. Muscle repair after physiological damage relies on nuclear migration for cellular reconstruction. *Science (1979)* 2021;374:355–9. <https://doi.org/10.1126/science.abe5620>.
- [58] Visser J, Vianney De Jong JMB, De Visser M. The history of progressive muscular atrophy: syndrome or disease? *Neurology* 2008;70:723–7. <https://doi.org/10.1212/01.wnl.0000302187.20239.93>.
- [59] Hancock J. Remarks on the Common Cachexia, or Leucophlegmasia, Called Mal d’Estomac in the Colonies, and Its Kindred Affections, as Dropsy, &c. *Edinb Med Surg J* 1831;35:67–76.
- [60] Ni J, Zhang L. Cancer cachexia: Definition, staging, and emerging treatments. *Cancer Manag Res* 2020;12:5597–605. <https://doi.org/10.2147/CMAR.S261585>.
- [61] Katz AM, Katz PB. DISEASES OF THE HEART IN THE WORKS OF HIPPOCRATES. *Br Heart J* 1962;257–64. <https://doi.org/10.1136/hrt.24.3.257>.
- [62] Petruzzelli M, Wagner EF. Mechanisms of metabolic dysfunction in cancer-associated cachexia. *Genes Dev* 2016;30:489–501. <https://doi.org/10.1101/gad.276733.115>.
- [63] Baracos V, Rodemann HP, Dinarello CA, Goldberg AL. Stimulation of muscle protein degradation and prostaglandin E2 release by leukocytic pyrogen (interleukin-1). A mechanism for the increased degradation of muscle proteins during fever. *N Engl J Med* 1983;308:553–8. <https://doi.org/10.1056/NEJM198303103081002>.
- [64] Beutler B, Mahoney J, Le Trang N, Pekala P, Cerami A. PURIFICATION OF CACHECTIN, A LIPOPROTEIN LIPASE-SUPPRESSING HORMONE SECRETED BY ENDOTOXIN-INDUCED RAW 264.7 CELLS. *J Exp Med* 1985;161:984–95. <https://doi.org/10.1084/jem.161.5.984>.
- [65] Pennica D, Hayflick JS, Bringman TS, Palladinot MA, Goeddel D V. Cloning and expression in *Escherichia coli* of the cDNA for murine tumor necrosis factor. *Proc Natl Acad Sci USA* 1985;82:6060–4. <https://doi.org/10.1073/pnas.82.18.6060>.
- [66] Zimmers TA, Davies M V, Koniaris LG, Haynes P, Esquela AF, Tomkinson KN, et al. Induction of Cachexia in Mice by Systemically Administered Myostatin. *Science (1979)* 2002;296:1486–8. <https://doi.org/10.1126/science.1069525>.
- [67] von Haehling S, Coats AJS, Anker SD. Ethical guidelines for publishing in the Journal of Cachexia, Sarcopenia and Muscle: update 2021. *J Cachexia Sarcopenia Muscle* 2021;12:2259–61. <https://doi.org/10.1002/jcsm.12899>.

- [68] Argilés JM, Busquets S, Stemmler B, López-Soriano FJ. Cancer cachexia: Understanding the molecular basis. *Nat Rev Cancer* 2014;14:754–62. <https://doi.org/10.1038/nrc3829>.
- [69] Mendes MCS, Pimentel GD, Costa FO, Carvalheira JBC. Molecular and neuroendocrine mechanisms of cancer cachexia. *Journal of Endocrinology* 2015;226:R29–43. <https://doi.org/10.1530/JOE-15-0170>.
- [70] Yeom E, Yu K. Understanding the molecular basis of anorexia and tissue wasting in cancer cachexia. *Exp Mol Med* 2022;54:426–32. <https://doi.org/10.1038/s12276-022-00752-w>.
- [71] Haizlip KM, Harrison BC, Leinwand LA. Sex-based differences in skeletal muscle kinetics and fiber-type composition. *Physiology* 2015;30:30–9. <https://doi.org/10.1152/physiol.00024.2014>.
- [72] Jeon Y, Choi J, Kim HJ, Lee H, Lim JY, Choi SJ. Sex- and fiber-type-related contractile properties in human single muscle fiber. *J Exerc Rehabil* 2019;15:537–45. <https://doi.org/10.12965/jer.1938336.168>.
- [73] Zhong X, Zimmers TA. Sex Differences in Cancer Cachexia. *Curr Osteoporos Rep* 2020;18:646–54. <https://doi.org/10.1007/s11914-020-00628-w>.
- [74] Hetzler KL, Hardee JP, Puppa MJ, Narsale AA, Sato S, Davis JM, et al. Sex differences in the relationship of IL-6 signaling to cancer cachexia progression. *Biochim Biophys Acta Mol Basis Dis* 2015;1852:816–25. <https://doi.org/10.1016/j.bbadis.2014.12.015>.
- [75] Nicolini A, Ferrari P, Masoni MC, Fini M, Pagani S, Giampietro O, et al. Malnutrition, anorexia and cachexia in cancer patients: A mini-review on pathogenesis and treatment. *Biomedicine and Pharmacotherapy* 2013;67:807–17. <https://doi.org/10.1016/j.biopha.2013.08.005>.
- [76] Gaafer OU, Zimmers TA. Nutrition challenges of cancer cachexia. *Journal of Parenteral and Enteral Nutrition* 2021;45:16–25. <https://doi.org/10.1002/jpen.2287>.
- [77] Agustsson T, D'souza MA, Nowak G, Isaksson B. Mechanisms for skeletal muscle insulin resistance in patients with pancreatic ductal adenocarcinoma. *Nutrition* 2011;27:796–801. <https://doi.org/10.1016/j.nut.2010.08.022>.
- [78] Rodriguez J, Vernus B, Chelh I, Cassar-Malek I, Gabillard JC, Hadj Sassi A, et al. Myostatin and the skeletal muscle atrophy and hypertrophy signaling pathways. *Cellular and Molecular Life Sciences* 2014;71:4361–71. <https://doi.org/10.1007/s00018-014-1689-x>.
- [79] Abrigo J, Elorza AA, Riedel CA, Vilos C, Simon F, Cabrera D, et al. Role of Oxidative Stress as Key Regulator of Muscle Wasting during Cachexia. *Oxid Med Cell Longev* 2018;2018. <https://doi.org/10.1155/2018/2063179>.
- [80] del Fabbro E, Hui D, Nooruddin ZI, Dalal S, Dev R, Freer G, et al. Associations Among Hypogonadism, C-Reactive Protein, Symptom Burden, and Survival in Male Cancer Patients with Cachexia: A Preliminary Report. *J Pain Symptom Manage* 2010;39:1016–24. <https://doi.org/10.1016/j.jpainsymman.2009.09.021>.

- [81] das Neves RX, Yamashita AS, Riccardi DMR, Köhn-Gaone J, Camargo RG, Neto NI, et al. Cachexia causes time-dependent activation of the inflammasome in the liver. *J Cachexia Sarcopenia Muscle* 2023. <https://doi.org/10.1002/jcsm.13236>.
- [82] Zimmers TA, Fishel ML, Bonetto A. STAT3 in the systemic inflammation of cancer cachexia. *Semin Cell Dev Biol* 2016;54:28–41. <https://doi.org/10.1016/j.semcdb.2016.02.009>.
- [83] Macciò A, Madeddu C, Lai E, Scartozzi M. Cancer cachexia and chronic inflammation: an unbreakable bond. *Br J Cancer* 2023;128:1609–10. <https://doi.org/10.1038/s41416-023-02200-6>.
- [84] Ezeoke CC, Morley JE. Pathophysiology of anorexia in the cancer cachexia syndrome. *J Cachexia Sarcopenia Muscle* 2015;6:287–302. <https://doi.org/10.1002/jcsm.12059>.
- [85] Laviano A, Meguid MM, Inui A, Muscaritoli M, Rossi-Fanelli F. Therapy insight: Cancer anorexia-cachexia syndrome - When all you can eat is yourself. *Nat Clin Pract Oncol* 2005;2:158–65. <https://doi.org/10.1038/ncponc0112>.
- [86] Xie T, Lv T, Zhang T, Feng D, Zhu F, Xu Y, et al. Interleukin-6 promotes skeletal muscle catabolism by activating tryptophan–indoleamine 2,3-dioxygenase 1–kynurenine pathway during intra-abdominal sepsis. *J Cachexia Sarcopenia Muscle* 2023;14:1046–59. <https://doi.org/10.1002/jcsm.13193>.
- [87] Mehl KA, Davis JM, Clements JM, Berger FG, Pena MM, Carson JA, et al. Decreased intestinal polyp multiplicity is related to exercise mode and gender in *Apc Min/* mice. *J Appl Physiol* 2005;98:2219–25. <https://doi.org/10.1152/jappphysiol.00975.2004>.
- [88] Loumays A, De Barse M, Nachit M, Lause P, Frateur L, Van Maanen A, et al. Role of activin A and myostatin in human cancer cachexia. *Journal of Clinical Endocrinology and Metabolism* 2015;100:2030–8. <https://doi.org/10.1210/jc.2014-4318>.
- [89] Zimmers TA, Davies M V, Koniaris LG, Haynes P, Esquela AF, Tomkinson KN, et al. Induction of Cachexia in Mice by Systemically Administered Myostatin. *Science* (1979) 2002;287:1486–8. <https://doi.org/10.1126/science.1069525>.
- [90] Vanderveen BN, Fix DK, Carson JA. Disrupted Skeletal Muscle Mitochondrial Dynamics , Mitophagy , and Biogenesis during Cancer Cachexia : A Role for Inflammation. *Oxid Med Cell Longev* 2017;2017:24–7. <https://doi.org/https://doi.org/10.1155/2017/3292087> Review.
- [91] Lecker SH, Solomon V, Mitch WE, Goldberg AL. Muscle protein breakdown and the critical role of the ubiquitin-proteasome pathway in normal and disease states. *J Nutr* 1999;129:227S–237S. <https://doi.org/10.1093/jn/129.1.227S>.
- [92] Deshaies RJ, Joazeiro CAP. RING domain E3 ubiquitin ligases. *Annu Rev Biochem* 2009;78:399–434. <https://doi.org/10.1146/annurev.biochem.78.101807.093809>.
- [93] Egerman MA, Glass DJ. Signaling pathways controlling skeletal muscle mass. *Crit Rev Biochem Mol Biol* 2014;49:59–68. <https://doi.org/10.3109/10409238.2013.857291>.
- [94] Rosa-Caldwell ME, Benson CA, Lee DE, Brown JL, Washington TA, Greene NP, et al. Mitochondrial Function and Protein Turnover in the Diaphragm are Altered in LLC Tumor Model of Cancer Cachexia n.d. <https://doi.org/10.3390/ijms21217841>.

- [95] Collins P, Bing C, Mcculloch P, Williams G. Muscle UCP-3 mRNA levels are elevated in weight loss associated with gastrointestinal adenocarcinoma in humans. *Br J Cancer* 2002;86:372–5. <https://doi.org/10.1038/sj/bjc/6600074>.
- [96] Smith HJ, Mukerji P, Tisdale MJ. Attenuation of Proteasome-Induced Proteolysis in Skeletal Muscle by  $\beta$ -Hydroxy- $\beta$ -Methylbutyrate in Cancer-Induced Muscle Loss by  $\beta$ -Hydroxy- $\beta$ -Methylbutyrate in Cancer-Induced Muscle Loss. *American Association for Cancer Research Journal* 2005;65:277–83.
- [97] Tisdale MJ. *Leading Edge Previews Reversing Cachexia* 2010. <https://doi.org/10.1016/j.cell.2010.08.004>.
- [98] Li Y-P, Reid MB. NF- $\kappa$ B mediates the protein loss induced by TNF- $\alpha$  in differentiated skeletal muscle myotubes. *Am J Physiol Regul Integr Comp Physiol* 2000;279:1165–70. <https://doi.org/10.1152/ajpregu.2000.279.4.R1165>.
- [99] Li Y, Schwartz RJ, Waddell ID, Holloway BR, Reid MB. Skeletal muscle myocytes undergo protein loss and reactive oxygen-mediated NF- $\kappa$ B activation in response to tumor necrosis factor $\alpha$ . *The FASEB Journal* 1998;12:871–80. <https://doi.org/10.1096/fasebj.12.10.971>.
- [100] Assi M, Rébillard A. The Janus-Faced Role of Antioxidants in Cancer Cachexia: New Insights on the Established Concepts. *Oxid Med Cell Longev* 2016;2016. <https://doi.org/10.1155/2016/9579868>.
- [101] Gueritat J, Lefevre-Orfila L, Vincent S, Cretual A, Ravanat JL, Gratas-Delamarche A, et al. Exercise training combined with antioxidant supplementation prevents the antiproliferative activity of their single treatment in prostate cancer through inhibition of redox adaptation. *Free Radic Biol Med* 2014;77:95–105. <https://doi.org/10.1016/j.freeradbiomed.2014.09.009>.
- [102] Assi M, Derbré F, Lefevre-Orfila L, Rébillard A. Antioxidant supplementation accelerates cachexia development by promoting tumor growth in C26 tumor-bearing mice. *Free Radic Biol Med* 2016;91:204–14. <https://doi.org/10.1016/j.freeradbiomed.2015.12.019>.
- [103] Feynman RP. There's Plenty of Room at the Bottom. *Eng Sci* 1960:22–36.
- [104] Khan I, Saeed K, Khan I. Nanoparticles: Properties, applications and toxicities. *Arabian Journal of Chemistry* 2019;12:908–31. <https://doi.org/10.1016/j.arabjc.2017.05.011>.
- [105] Karthik P, Ezhilarasi PN, Anandharamakrishnan C. Challenges associated in stability of food grade nanoemulsions. *Crit Rev Food Sci Nutr* 2017;57:1435–50. <https://doi.org/10.1080/10408398.2015.1006767>.
- [106] McClements DJ. Nanoemulsions versus microemulsions: Terminology, differences, and similarities. *Soft Matter* 2012;8:1719–29. <https://doi.org/10.1039/C2SM06903B>.
- [107] Lee JE, Lee N, Kim T, Kim J, Hyeon T. Multifunctional mesoporous silica nanocomposite nanoparticles for theranostic applications. *Acc Chem Res* 2011;44:893–902. <https://doi.org/10.1021/ar2000259>.
- [108] Marchettini P, Stuart OA, Mohamed F, Yoo D, Sugarbaker PH. Docetaxel: pharmacokinetics and tissue levels after intraperitoneal and intravenous administration

- in a rat model. *Cancer Chemother Pharmacol* 2002;49:499–503.  
<https://doi.org/10.1007/S00280-002-0439-1>.
- [109] Sparreboom A, Van Tellingen O, Nooijen WJ, Beijnen JH. Preclinical pharmacokinetics of paclitaxel and docetaxel. *Anticancer Drugs* 1998;9:1–17.  
<https://doi.org/10.1097/00001813-199801000-00001>.
- [110] Baker J, Ajani J, Scotté F, Winther D, Martin M, Aapro MS, et al. Docetaxel-related side effects and their management. *European Journal of Oncology Nursing* 2009;13:49–59.  
<https://doi.org/10.1016/j.ejon.2008.10.003>.
- [111] Verweij J, Catimel G, Sulkes A, Sternberg C, Wolff I, Aamdal S, et al. Phase II studies of docetaxel in the treatment of various solid tumours. *Eur J Cancer* 1995;31:21–4.  
[https://doi.org/10.1016/0959-8049\(95\)00362-M](https://doi.org/10.1016/0959-8049(95)00362-M).
- [112] Zhang L, Zhang N. How nanotechnology can enhance docetaxel therapy. *Int J Nanomedicine* 2013;8:2927–41. <https://doi.org/10.2147/IJN.S46921>.
- [113] Maranhão RC, Garicochea B, Silva EL, Llacer PD, Pileggi FJ, Chamone DA. Increased plasma removal of microemulsions resembling the lipid phase of low-density lipoproteins (LDL) in patients with acute myeloid leukemia: a possible new strategy for the treatment of the disease. *Braz J Med Biol Res* 1992;25:1003–7.
- [114] Maranhão RC, Garicochea B, Silva EL, Maranhao RC, Dorlhiac-Ilacer P, Cadena SMS, et al. Plasma Kinetics and Biodistribution of a Lipid Emulsion Resembling Low-Density Lipoprotein in Patients with Acute Leukemia Plasma Kinetics and Biodistribution of a Lipid Emulsion Resembling Low-Density Lipoprotein in Patients with Acute Leukemia 1994:4660–6.
- [115] Hevonoja T, Pentikainen MO, Hyvonen MT, Kovanen PT, Ala-Korpela M. Structure of low density lipoprotein (LDL) particles: Basis for understanding molecular changes in modified LDL. *Biochim Biophys Acta* 2000;1488:189–210.  
[https://doi.org/10.1016/s1388-1981\(00\)00123-2](https://doi.org/10.1016/s1388-1981(00)00123-2).
- [116] Maranhão RC, Cesar TB, Pedroso-Mariani SR, Hirata MH, Mesquita CH. Metabolic behavior in rats of a nonprotein microemulsion resembling low-density lipoprotein. *Lipids* 1993;28:691–6.
- [117] Loeffler B, Heeren J, Blaeser M, Radner H, Kayser D, Aydin B, et al. Lipoprotein lipase-facilitated uptake of LDL is mediated by the LDL receptor. *J Lipid Res* 2006;48:288–98.  
<https://doi.org/10.1194/jlr.M600292-JLR200>.
- [118] Goldstein JL, Brown MS. The LDL receptor. *Arterioscler Thromb Vasc Biol* 2009;29:431–8. <https://doi.org/10.1161/ATVBAHA.108.179564>.
- [119] Ho YK, Smith RG, Brown MS, Goldstein JL. Low-Density Lipoprotein (LDL) Receptor Activity in Human Acute Myelogenous Leukemia Cells. *Blood* 1978;52:1099–112.  
<https://doi.org/10.1182/blood.V52.6.1099.1099>.
- [120] Kuzu OF, MA N, GP R. The role of cholesterol in cancer 2016;70:773–9.  
<https://doi.org/10.1097/OGX.0000000000000256>. Prenatal.

- [121] Perez AS, Morikawa AT, Maranhão RC, Figueiredo Neto AM. Structural characterization of cholesterol-rich nanoemulsion (LDE). *Chem Phys Lipids* 2024;263. <https://doi.org/10.1016/j.chemphyslip.2024.105418>.
- [122] Ades A, Carvalho JP, Graziani SR, Amancio RF, Souen JS, Pinotti JA, et al. Uptake of a cholesterol-rich emulsion by neoplastic ovarian tissues. *Gynecol Oncol* 2001;82:84–7. <https://doi.org/10.1006/gyno.2001.6203>.
- [123] Pires LA, Hegg R, Valduga CJ, Graziani SR, Rodrigues DG, Maranhão RC. Use of cholesterol-rich nanoparticles that bind to lipoprotein receptors as a vehicle to paclitaxel in the treatment of breast cancer: Pharmacokinetics, tumor uptake and a pilot clinical study. *Cancer Chemother Pharmacol* 2009;63:281–7. <https://doi.org/10.1007/s00280-008-0738-2>.
- [124] Pires LA, Hegg R, Freitas FR, Tavares ER, Almeida CP, Baracat EC, et al. Effect of neoadjuvant chemotherapy on low-density lipoprotein (LDL) receptor and LDL receptor-related protein 1 (LRP-1) receptor in locally advanced breast cancer. *Brazilian Journal of Medical and Biological Research* 2012;45:557–64. <https://doi.org/10.1590/S0100-879X2012007500068>.
- [125] Vital CG, Maranhão RC, Freitas FR, Van Eyll BM, Graziani SR. Use of paclitaxel carried in lipid core nanoparticles in patients with late-stage solid cancers with bone metastases: Lack of toxicity and therapeutic benefits. *J Bone Oncol* 2022;34. <https://doi.org/10.1016/j.jbo.2022.100431>.
- [126] Pozzi FS, Maranhão RC, Guedes LK, Borba EF, Laurindo IMM, Bonfa E, et al. Plasma kinetics of an LDL-like non-protein nanoemulsion and transfer of lipids to high-density lipoprotein (HDL) in patients with rheumatoid arthritis. *J Clin Lipidol* 2015;9:72–80. <https://doi.org/10.1016/j.jacl.2014.10.004>.
- [127] Graziani SR, Vital CG, Morikawa AT, Van Eyll BM, Fernandes Junior HJ, Kalil Filho R, et al. Phase II study of paclitaxel associated with lipid core nanoparticles (LDE) as third-line treatment of patients with epithelial ovarian carcinoma. *Medical Oncology* 2017;34. <https://doi.org/10.1007/s12032-017-1009-z>.
- [128] Kretzer IF, Maria DA, Maranhão RC. Drug-targeting in combined cancer chemotherapy: Tumor growth inhibition in mice by association of paclitaxel and etoposide with a cholesterol-rich nanoemulsion. *Cellular Oncology* 2012;35:451–60. <https://doi.org/10.1007/s13402-012-0104-6>.
- [129] Gordon B, Chen S, Durstine JL. The Effects of Exercise Training on the Traditional Lipid Profile and Beyond 2014:253–9.
- [130] Ficker ES, Maranhão RC, Chacra APM, Neves VC, Negrão CE, Martins VCN, et al. Exercise training accelerates the removal from plasma of LDL-like nanoemulsion in moderately hypercholesterolemic subjects. *Atherosclerosis* 2010;212:230–6. <https://doi.org/10.1016/j.atherosclerosis.2010.04.030>.
- [131] Vinagre CGC, Ficker ES, Finazzo C, Alves MJN, de Angelis K, Irigoyen MC, et al. Enhanced removal from the plasma of LDL-like nanoemulsion cholesteryl ester in trained men compared with sedentary healthy men. *J Appl Physiol* 2007;103:1166–71. <https://doi.org/10.1152/jappphysiol.01176.2006>.



- [132] da Silva JL, Vinagre CGCM, Morikawa AT, Alves MJNN, Mesquita CH, Maranhão RC. Resistance training changes LDL metabolism in normolipidemic subjects: A study with a nanoemulsion mimetic of LDL. *Atherosclerosis* 2011;219:532–7. <https://doi.org/10.1016/j.atherosclerosis.2011.08.014>.
- [133] De Weger VA, Beijnenb JH, Schellensa JHM. Cellular and clinical pharmacology of the taxanes docetaxel and paclitaxel -A review. *Anticancer Drugs* 2014;25:488–94. <https://doi.org/10.1097/CAD.000000000000093>.
- [134] Pienta KJ. Preclinical mechanisms of action of docetaxel and docetaxel combinations in prostate cancer. *Semin Oncol* 2001;28:3–7. <https://doi.org/10.1053/sonc.2001.26892>.
- [135] Herbst RS, Khuri FR. Mode of action of docetaxel - A basis for combination with novel anticancer agents. *Cancer Treat Rev* 2003;29:407–15. [https://doi.org/10.1016/S0305-7372\(03\)00097-5](https://doi.org/10.1016/S0305-7372(03)00097-5).
- [136] Cushen SJ, Power DG, Murphy KP, McDermott R, Griffin BT, Lim M, et al. Impact of body composition parameters on clinical outcomes in patients with metastatic castrate-resistant prostate cancer treated with docetaxel. *Clin Nutr ESPEN* 2016;13:e39–45. <https://doi.org/10.1016/j.clnesp.2016.04.001>.
- [137] Mikuła-Pietrasik J, Witucka A, Pakuła M, Uruski P, Begier-Krasińska B, Niklas A, et al. Comprehensive review on how platinum- and taxane-based chemotherapy of ovarian cancer affects biology of normal cells. *Cellular and Molecular Life Sciences* 2019;76:681–97. <https://doi.org/10.1007/s00018-018-2954-1>.
- [138] Harvey V, Mouridsen H, Semiglazov V, Jakobsen E, Voznyi E, Robinson BA, et al. Phase III trial comparing three doses of docetaxel for second-line treatment of advanced breast cancer. *Journal of Clinical Oncology* 2006;24:4963–70. <https://doi.org/10.1200/JCO.2005.05.0294>.
- [139] Imran M, Saleem S, Chaudhuri A, Ali J, Baboota S. Docetaxel: An update on its molecular mechanisms, therapeutic trajectory and nanotechnology in the treatment of breast, lung and prostate cancer. *J Drug Deliv Sci Technol* 2020;60. <https://doi.org/10.1016/j.jddst.2020.101959>.
- [140] Van Zuylen L, Verweij J, Sparreboom A. Role of formulation vehicles in taxane pharmacology. *Invest New Drugs* 2001;19:125–41. <https://doi.org/10.1023/A:1010618632738>.
- [141] Deeken JF, Road R, Weiss GJ. A phase I study of liposomal-encapsulated docetaxel (LE-DT) in patients with advanced solid tumor malignancies 2013;71:627–33. <https://doi.org/10.1007/s00280-012-2048-y.A>.
- [142] Mahalingam D, Nemunaitis JJ, Malik L, Sarantopoulos J, Weitman S, Sankhala K, et al. Phase i study of intravenously administered ATI-1123, a liposomal docetaxel formulation in patients with advanced solid tumors. *Cancer Chemother Pharmacol* 2014;74:1241–50. <https://doi.org/10.1007/s00280-014-2602-x>.
- [143] Chaillou T, McPeck A, Lanner JT. Docetaxel does not impair skeletal muscle force production in a murine model of cancer chemotherapy. *Physiol Rep* 2017;5:1–9. <https://doi.org/10.14814/phy2.13261>.

- [144] Braun J, Rau R. An update on methotrexate. *Curr Opin Rheumatol* 2009;21:216–23. <https://doi.org/10.1097/BOR.0b013e328329c79d>.
- [145] Chan ESL, Cronstein BN. Methotrexate how does it really work? *Nat Rev Rheumatol* 2010;6:175–8. <https://doi.org/10.1038/nrrheum.2010.5>.
- [146] Headrick JP, Ashton KJ, Rose Meyer RB, Peart JN. Cardiovascular adenosine receptors: Expression, actions and interactions. *Pharmacol Ther* 2013;140:92–111. <https://doi.org/10.1016/j.pharmthera.2013.06.002>.
- [147] Haskó G, Cronstein B, Filep JG, Ludwig A, Taub DD. Regulation of inflammation by adenosine 2013. <https://doi.org/10.3389/fimmu.2013.00085>.
- [148] Russo GT, Minutoli L, Bitto A, Altavilla D, Alessi E, Giandalia A, et al. Methotrexate increases skeletal muscle GLUT4 expression and improves metabolic control in experimental diabetes. *J Nutr Metab* 2012;2012:1–7. <https://doi.org/10.1155/2012/132056>.
- [149] Pirkmajer S, Kulkarni SS, Tom RZ, Ross FA, Hawley SA, Hardie DG, et al. Methotrexate promotes glucose uptake and lipid oxidation in skeletal muscle via AMPK activation. *Diabetes* 2015;64:360–9. <https://doi.org/10.2337/db14-0508>.
- [150] Stocks B, Zierath JR. Post-translational modifications: The signals at the intersection of exercise, glucose uptake and insulin sensitivity. *Endocr Rev* 2021. <https://doi.org/10.1210/edrev/bnab038/6420158>.
- [151] Chabner BA, Allegra CJ, Curt GA, Clendeninn NJ, Baram J, Koizumi S, et al. Perspectives Polyglutamation of Methotrexate Is Methotrexate a Prodrug? *J Clin Invest* 1985;76:907–12. <https://doi.org/10.1172/JCI112088>.
- [152] Day P, Sharff A, Parra L, Cleasby A, Williams M, Hörer S, et al. Structure of a CBS-domain pair from the regulatory c1 subunit of human AMPK in complex with AMP and ZMP. *Research Papers Acta Cryst* 2007;63:587–96. <https://doi.org/10.1107/S0907444907009110>.
- [153] Corton JM, Gillespie JG, Hawley SA, Grahame D, Hardie DG. 5-Aminoimidazole-4-carboxamide ribonucleoside A specific method for activating AMP-activated protein kinase in intact cells? *Eur J Biochem* 1995;229. <https://doi.org/10.1111/j.1432-1033.1995.tb20498.x>.
- [154] Birk JB, Wojtaszewski JFP. Predominant  $\alpha 2/\beta 2/\gamma 3$  AMPK activation during exercise in human skeletal muscle. *J Physiol* 2006;577:1021–32. <https://doi.org/10.1113/jphysiol.2006.120972>.
- [155] Popoff MR, Vandewalle A. AMPK: A Key Sensor of Fuel and Energy Status in Skeletal Muscle. *Physiology* 2007:48–60.
- [156] Alves CRR, Eichelberger EJ, das Neves W, Ribeiro MAC, Bechara LRG, Voltarelli VA, et al. Cancer-induced muscle atrophy is determined by intrinsic muscle oxidative capacity. *FASEB Journal* 2021;35. <https://doi.org/10.1096/fj.202100263R>.
- [157] Tipton CM. Historical Perspectives The history of “Exercise Is Medicine” in ancient civilizations. *Adv Physiol Educ* 2014;38:109–17. <https://doi.org/10.1152/advan.00136.2013>.

- [158] Caspersen CJ, Powell KE, Christenson GM. Physical Activity, Exercise, and Physical Fitness: Definitions and Distinctions for Health-Related Research Synopsis. *Public Health Rep* 1985;100:126–31.
- [159] Dasso NA. How is exercise different from physical activity? A concept analysis. *Nurs Forum (Auckl)* 2019;54:45–52. <https://doi.org/10.1111/nuf.12296>.
- [160] Patel H, Alkhawam H, Madanieh R, Shah N, Kosmas CE, Vittorio TJ. Aerobic vs anaerobic exercise training effects on the cardiovascular system. *World J Cardiol* 2017;9:134–8. <https://doi.org/10.4330/wjc.v9.i2.134>.
- [161] Riddell MC, Scott SN, Fournier PA, Colberg SR, Gallen IW, Moser O, et al. The competitive athlete with type 1 diabetes. *Diabetologia* 2020;63:1475–90. <https://doi.org/10.1007/s00125-020-05183-8>.
- [162] Gualdi-Russo E, Zaccagni L. Physical activity for health and wellness. *Int J Environ Res Public Health* 2021;18. <https://doi.org/10.3390/ijerph18157823>.
- [163] Kapoor G, Chauhan P, Singh G, Malhotra N, Chahal A. Physical Activity for Health and Fitness: Past, Present and Future. *J Lifestyle Med* 2022;12:9–14. <https://doi.org/10.15280/jlm.2022.12.1.9>.
- [164] Wang Y, Ashokan K. Physical Exercise: An Overview of Benefits From Psychological Level to Genetics and Beyond. *Front Physiol* 2021;12. <https://doi.org/10.3389/fphys.2021.731858>.
- [165] Correia RR, Veras ASC, Tebar WR, Rufino JC, Batista VRG, Teixeira GR. Strength training for arterial hypertension treatment: a systematic review and meta-analysis of randomized clinical trials. *Sci Rep* 2023;13. <https://doi.org/10.1038/s41598-022-26583-3>.
- [166] Correia RR, Batista VRG, Veras ASC, Tavares ME de A, Souza FLA, Pacagnelli FL, et al. High-intensity interval training attenuates the effects caused by arterial hypertension in the ventral prostate. *Prostate* 2022;82:373–87. <https://doi.org/10.1002/pros.24285>.
- [167] Tavares MEA, Pinto AP, da Rocha AL, Sampaio LV, Correia RR, Batista VRG, et al. Combined physical exercise re-synchronizes expression of Bmal1 and REV-ERB $\alpha$  and up-regulates apoptosis and metabolism in the prostate during aging. *Life Sci* 2024;351. <https://doi.org/10.1016/J.LFS.2024.122800>.
- [168] Batista VRG, Correia RR, Fernandes VS, Veras ASC, Tavares MEA, Chaves-Neto AH, et al. High-Intensity Interval Training Minimizes the Deleterious Effects of Arterial Hypertension on the Urinary Bladder of Spontaneously Hypertensive Rats. *Oxid Med Cell Longev* 2023. <https://doi.org/10.1155/2023/9979397>.
- [169] Veras ASC, Correia RR, Batista VRG, Tavares ME de A, Rubira RJG, Fiais GA, et al. Aerobic physical exercise modifies the prostate tumoral environment. *Life Sci* 2023;332. <https://doi.org/10.1016/J.LFS.2023.122097>.
- [170] dos Santos GF, Veras ASC, de Freitas MC, McCabe J, Seraphim PM, Teixeira GR. Strength training reduces lipid accumulation in liver of obese Wistar rats. *Life Sci* 2019;235:116834. <https://doi.org/10.1016/J.LFS.2019.116834>.

- [171] Poole DC, Wilkerson DP, Jones AM. Validity of criteria for establishing maximal O<sub>2</sub> uptake during ramp exercise tests. *Eur J Appl Physiol* 2008;102:403–10. <https://doi.org/10.1007/s00421-007-0596-3>.
- [172] Levine BD. VO<sub>2</sub>: What do we know, and what do we still need to know? *Journal of Physiology* 2008;586:25–34. <https://doi.org/10.1113/jphysiol.2007.147629>.
- [173] Rey-López JP, Lee DH, Ferrari G, Giovannucci E, Rezende LFM. Physical Activity Intensity Measurement and Health: State of the Art and Future Directions for Scientific Research. *Int J Environ Res Public Health* 2023;20. <https://doi.org/10.3390/ijerph20116027>.
- [174] Ministério da Saúde. Guia de Atividade Física para a População Brasileira 2021.
- [175] Muscella A, Stefàno E, Lunetti P, Capobianco L, Marsigliante S. The regulation of fat metabolism during aerobic exercise. *Biomolecules* 2020;10:1–29. <https://doi.org/10.3390/biom10121699>.
- [176] Hargreaves M, Spriet LL. Skeletal muscle energy metabolism during exercise. *Nat Metab* 2020;2:817–28. <https://doi.org/10.1038/s42255-020-0251-4>.
- [177] Hood DA. Mechanisms of exercise-induced mitochondrial biogenesis in skeletal muscle. *Applied Physiology, Nutrition and Metabolism* 2009;34:465–72. <https://doi.org/10.1139/H09-045>.
- [178] Ballard-Barbash R, Hunsberger S, Alciati MH, Blair SN, Goodwin PJ, McTiernan A, et al. Physical activity, weight control, and breast cancer risk and survival: clinical trial rationale and design considerations. *J Natl Cancer Inst* 2009;101:630–43. <https://doi.org/10.1093/jnci/djp068>.
- [179] Friedenreich CM, Shaw E, Neilson HK, Brenner DR. Epidemiology and biology of physical activity and cancer recurrence 2017. <https://doi.org/10.1007/s00109-017-1558-9>.
- [180] Ruiz-Casado A, Martín-Ruiz A, Pérez LM, Provencio M, Fiuza-Luces C, Lucia A. Exercise and the Hallmarks of Cancer. *Trends Cancer* 2017;3:423–41. <https://doi.org/10.1016/j.trecan.2017.04.007>.
- [181] Katzmarzyk PT, Leon AS, Rankinen T, Gagnon J, Skinner JS, Wilmore JH, et al. Changes in blood lipids consequent to aerobic exercise training related to changes in body fatness and aerobic fitness. *Metabolism* 2001;50:841–8. <https://doi.org/10.1053/meta.2001.24190>.
- [182] Gallagher EJ, Zelenko Z, Neel BM, Antoniou IM, Rajan L, Kase N, et al. Elevated tumor LDLR expression accelerates LDL cholesterol-mediated breast cancer growth in mouse models of hyperlipidemia HHS Public Access. *Oncogene* 2017;36:6462–71. <https://doi.org/10.1038/onc.2017.247>.
- [183] Halle JL, Counts BR, Carson JA. Exercise as a therapy for cancer-induced muscle wasting. *Sports Medicine and Health Science* 2020;2:186–94. <https://doi.org/10.1016/j.smhs.2020.11.004>.
- [184] Halle JL, Counts BR, Zhang Q, Carson JA. Short duration treadmill exercise improves physical function and skeletal muscle mitochondria protein expression after recovery from FOLFOX chemotherapy in male mice. *FASEB Journal* 2022;36. <https://doi.org/10.1096/FJ.202200460R>.

- [185] Cannioto RA, Hutson A, Dighe S, McCann W, McCann SE, Zirpoli GR, et al. Physical activity before, during, and after chemotherapy for high-risk breast cancer: Relationships with survival. *J Natl Cancer Inst* 2021;113:54–63. <https://doi.org/10.1093/JNCI/DJAA046>.
- [186] Pigna E, Berardi E, Aulino P, Rizzuto E, Zampieri S, Carraro U, et al. Aerobic Exercise and Pharmacological Treatments Counteract Cachexia by Modulating Autophagy in Colon Cancer. *Sci Rep* 2016;6. <https://doi.org/10.1038/srep26991>.
- [187] Hardee JP, Counts BR, Carson JA. Understanding the Role of Exercise in Cancer Cachexia Therapy. *Am J Lifestyle Med* 2019;13:46–60. <https://doi.org/10.1177/1559827617725283>.
- [188] Leal LG, Lopes MA, Peres SB, Batista ML. Exercise Training as Therapeutic Approach in Cancer Cachexia: A Review of Potential Anti-inflammatory Effect on Muscle Wasting. *Front Physiol* 2021;11. <https://doi.org/10.3389/fphys.2020.570170>.
- [189] Tanaka M, Sugimoto K, Fujimoto T, Xie K, Takahashi T, Akasaka H, et al. Differential effects of pre-exercise on cancer cachexia-induced muscle atrophy in fast- and slow-twitch muscles. *FASEB Journal* 2020;34:14389–406. <https://doi.org/10.1096/fj.202001330R>.
- [190] Pedersen BK. Muscles and their myokines. *Journal of Experimental Biology* 2011;214:337–46. <https://doi.org/10.1242/jeb.048074>.
- [191] Frydelund-Larsen L, Penkowa M, Akerstrom T, Zankari A, Nielsen S, Pedersen BK. Exercise induces interleukin-8 receptor (CXCR2) expression in human skeletal muscle. *Exp Physiol* 2007;92:233–40. <https://doi.org/10.1113/expphysiol.2006.034769>.
- [192] Papadopetraki A, Maridaki M, Zagouri F, Dimopoulos, Koutsilieris M, Philippou A. Physical Exercise Restrains Cancer Progression through Muscle-Derived Factors. *Cancers (Basel)* 2022;14. <https://doi.org/10.3390/cancers14081892>.
- [193] Pin F, Bonewald LF, Bonetto A. Role of myokines and osteokines in cancer cachexia. *Exp Biol Med* 2021;246:2118–27. <https://doi.org/10.1177/15353702211009213>.
- [194] Kim JS, Galvão DA, Newton RU, Gray E, Taaffe DR. Exercise-induced myokines and their effect on prostate cancer. *Nat Rev Urol* 2021;18:519–42. <https://doi.org/10.1038/s41585-021-00476-y>.
- [195] Rupert JE, Narasimhan A, Jengelly DHA, Jiang Y, Liu J, Au E, et al. Tumor-derived IL-6 and trans-signaling among tumor, fat, and muscle mediate pancreatic cancer cachexia. *J Exp Med* 2021;218. <https://doi.org/10.1084/JEM.20190450>.
- [196] Trovato E, Di Felice V, Barone R. Extracellular vesicles: Delivery vehicles of myokines. *Front Physiol* 2019;10. <https://doi.org/10.3389/fphys.2019.00522>.
- [197] Cocucci E, Meldolesi J. Ectosomes and exosomes: Shedding the confusion between extracellular vesicles. *Trends Cell Biol* 2015;25:364–72. <https://doi.org/10.1016/j.tcb.2015.01.004>.
- [198] Shelke GV, Lässer C, Gho YS, Lötvall J. Importance of exosome depletion protocols to eliminate functional and RNA-containing extracellular vesicles from fetal bovine serum. *J Extracell Vesicles* 2014;3. <https://doi.org/10.3402/jev.v3.24783>.

- [199] Lovett JAC, Durcan PJ, Myburgh KH. Investigation of circulating extracellular vesicle microRNA following two consecutive bouts of muscle-damaging exercise. *Front Physiol* 2018;9. <https://doi.org/10.3389/fphys.2018.01149>.
- [200] Rundqvist H, Veliça P, Barbieri L, Gameiro PA, Bargiela D, Gojkovic M, et al. Cytotoxic t-cells mediate exercise-induced reductions in tumor growth. *Elife* 2020;9:1–25. <https://doi.org/10.7554/eLife.59996>.
- [201] Wang B, Xu H, Hu X, Ma W, Zhang J, Li Y, et al. Synergetic inhibition of daidzein and regular exercise on breast cancer in bearing-4T1 mice by regulating NK cells and apoptosis pathway. *Life Sci* 2020;245. <https://doi.org/10.1016/j.lfs.2020.117387>.
- [202] Idorn M, thor Straten P. Exercise and cancer: from “healthy” to “therapeutic”? *Cancer Immunology, Immunotherapy* 2017;66:667–71. <https://doi.org/10.1007/s00262-017-1985-z>.
- [203] Xiao W, Chen P, Dong J. Effects of overtraining on skeletal muscle growth and gene expression. *Int J Sports Med* 2012;33:846–53. <https://doi.org/10.1055/s-0032-1311585>.
- [204] Pereira BC, Pauli JR, Antunes LMG, De Freitas EC, De Almeida MR, De Paula Venâncio V, et al. Overtraining is associated with DNA damage in blood and skeletal muscle cells of Swiss mice. *BMC Physiol* 2013;13. <https://doi.org/10.1186/1472-6793-13-11>.
- [205] Kreher JB, Schwartz JB. Overtraining Syndrome: A Practical Guide. *Sports Health* 2012;4:128–38. <https://doi.org/10.1177/1941738111434406>.
- [206] Dos G, Cunha S, Ribeiro JL, Reischak De Oliveira A. Overtraining: theories, diagnosis and markers. *Rev Bras Med Esporte* 2006;12. <https://doi.org/10.1590/S1517-86922006000500014>.
- [207] Cheng AJ, Jude B, Lanner JT. Intramuscular mechanisms of overtraining. *Redox Biol* 2020;35. <https://doi.org/10.1016/j.redox.2020.101480>.
- [208] Kerdelhué B, Forest C, Coumoul X. Dimethyl-Benz(a)anthracene: A mammary carcinogen and a neuroendocrine disruptor. *Biochim Open* 2016;3:49–55. <https://doi.org/10.1016/j.biopen.2016.09.003>.
- [209] Russo J, Tait L, Russo IH. Susceptibility of the Mammary Gland to Carcinogenesis III. The Cell of Origin of Rat Mammary Carcinoma. *Am J Pathol* 1983;113:50–66.
- [210] Marion SL, Watson J, Sen N, Brewer MA, Barton JK, Hoyer PB. 7,12-Dimethylbenz[a]anthracene-Induced Malignancies in a Mouse Model of Menopause. *Comp Med* 2013;63:6–12.
- [211] Jimenez RE, Z'graggen K, Hartwig W, Graeme-Cook F, Warshaw AL, Castillo CF del. Immunohistochemical characterization of pancreatic tumors induced by dimethylbenzanthracene in rats. *American Journal of Pathology* 1999;154:1223–9. [https://doi.org/10.1016/S0002-9440\(10\)65374-6](https://doi.org/10.1016/S0002-9440(10)65374-6).
- [212] Niazi SK, Hussain M. Disposition kinetics of 7,12-dimethylbenz(a)anthracene in rats. *Biopharm Drug Dispos* 1992;13:591–6. <https://doi.org/10.1002/bdd.2510130805>.

- [213] Brookes P, Heidelberger C. Isolation and Degradation of DNA from Cells Treated with Tritium-labeled 7,12-Dimethylbenz(a)anthracene: Studies on the Nature of the Binding of This Carcinogen to DNA. *Cancer Res* 1969;29:157–65.
- [214] Abdalla DR, Rocha Aleixo AA, Murta EFC, Michelin MA. Innate immune response adaptation in mice subjected to administration of DMBA and physical activity. *Oncol Lett* 2014;7:886–90. <https://doi.org/10.3892/ol.2013.1774>.
- [215] Kellen JA. Topographical Distribution of Dimethylbenz(a)anthracene-Induced Rat Mammary Tumors. *Ann Clin Lab Sci* 1983;13:289–90.
- [216] Woodhead AD, Bornbusch A. The pattern of distribution of dimethylbenz(a)anthracene in the Amazon molly, *Poecilia formosa* (Girard), after intraperitoneal injection\*. *J Fish Biol* 1983;22:241–8. <https://doi.org/10.1111/j.1095-8649.1983.tb04744.x>.
- [217] Huggins C, Fukunishi R. Mammary and Peritoneal Tumors Induced by Intraperitoneal Administration of 7,12-Dimethylbenz[a]anthracene in Newborn and Adult Rats\*. *Cancer Res* 1963;23:785–9.
- [218] Ward JM, Rehm S, Devor D, Hennings H, Wenkt ML. Differential Carcinogenic Effects of Intraperitoneal Initiation with 7,12-Dimethylbenz(a)anthracene or Urethane and Topical Promotion with 12-O-Tetradecanoylphorbol-13-acetate in Skin and Internal Tissues of Female SENCAR and BALB/c Mice. *Environ Health Perspect* 1986;68:61–8. <https://doi.org/10.1289/ehp.866861>.
- [219] Daniel FB, Joyce NJ. DNA Adduct Formation by 7, 12-Dimethylbenz[a]anthracene and Its Noncarcinogenic 2-Fluoro Analogue in Female Sprague-Dawley Rats. *J Natl Cancer Inst* 1983;70:111–8. <https://doi.org/10.1093/jnci/70.1.111>.
- [220] Wang Z, Zhang X. Chemopreventive Activity of Honokiol against 7, 12 - Dimethylbenz[a]anthracene-Induced Mammary Cancer in Female Sprague Dawley Rats. *Front Pharmacol* 2017;8. <https://doi.org/10.3389/fphar.2017.00320>.
- [221] Rajakumar T, Pugalendhi P, Thilagavathi S, Ananthkrishnan D, Gunasekaran K. Allyl isothiocyanate, a potent chemopreventive agent targets AhR/Nrf2 signaling pathway in chemically induced mammary carcinogenesis. *Mol Cell Biochem* 2018;437:1–12. <https://doi.org/10.1007/s11010-017-3091-0>.
- [222] Machida Y, Imai T. Different properties of mammary carcinogenesis induced by two chemical carcinogens, DMBA and PhIP, in heterozygous BALB/c Trp53 knockout mice. *Oncol Lett* 2021;22. <https://doi.org/10.3892/ol.2021.12999>.
- [223] Guo W, Wu X, Li Y, Gao J, Wang F, Jin Y, et al. Evaluation of biophysical as well as biochemical potential of curcumin and resveratrol during prostate cancer. *J Drug Target* 2020;28:41–5. <https://doi.org/10.1080/1061186X.2019.1601199>.
- [224] Christou M, Moore CJ, Gould MN, Jefcoate CR. Induction of mammary cytochromes P-450: An essential first step in the metabolism of 7,12-dimethylbenz[a]anthracene by rat mammary epithelial cells. *Carcinogenesis* 1987;8:73–80. <https://doi.org/10.1093/carcin/8.1.73>.
- [225] Manoharan S, Panjamurthy K, Pugalendi P, Balakrishnan S, Rajalingam K, Vellaichamy L, et al. Protective role of withaferin-A on red blood cell integrity during 7,12-

- dimethylbenz[A]anthracene induced oral carcinogenesis. *African Journal of Traditional, Complementary and Alternative Medicines* 2009;6:94–102.  
<https://doi.org/10.4314/ajtcam.v6i1.57079>.
- [226] Katayama S, Fiala E, Reddy BS, Rivenson A, Silverman J, Williams GM, et al. Prostate adenocarcinoma in rats: Induction by 3,2'-dimethyl-4-aminobiphenyl. *J Natl Cancer Inst* 1982;68:867–73. <https://doi.org/10.1093/jnci/68.5.867>.
- [227] Ito N, Hirose M, Fukushima S, Tsuda H, Shirai T, Tatematsu M. Studies on antioxidants: Their carcinogenic and modifying effects on chemical carcinogenesis. *Food and Chemical Toxicology* 1986;24:1071–82. [https://doi.org/10.1016/0278-6915\(86\)90291-7](https://doi.org/10.1016/0278-6915(86)90291-7).
- [228] Janvier Labs. SPRAGUE DAWLEY® Rat. Janvier Labs 2019. [https://janvier-labs.com/en/fiche\\_produit/sprague\\_dawley\\_rat/](https://janvier-labs.com/en/fiche_produit/sprague_dawley_rat/) (accessed March 29, 2022).
- [229] Foster JR, Frost D. The History of the Rat. *Boorman's Pathology of the Rat*, Elsevier; 2018, p. 7–12. <https://doi.org/10.1016/b978-0-12-391448-4.00003-4>.
- [230] Alemán CL, Más RM, Rodeiro I, Noa M, Hernández C, Menéndez R, et al. Reference database of the main physiological parameters in Sprague-Dawley rats from 6 to 32 months 1998;32:457–66. <https://doi.org/10.1258/002367798780599802>.
- [231] Gileta AF, Fitzpatrick CJ, Chitre AS, St Pierre CL, Joyce E V, Maguire RJ, et al. Genetic characterization of outbred Sprague Dawley rats and utility for 4 genome-wide association studies. *BioRxiv* 2021. <https://doi.org/10.1101/412924>.
- [232] Crossley LM, Allison JB, Muenzen JB. II. Treatment to Flexner-Jobling carcinoma in Sprague-Dawley rats with 2,4,6-triethylenimino-s-triazine (TEM). *Proceedings of the Society for Experimental Biology and Medicine* 1952;80:452–5. <https://doi.org/10.3181/00379727-80-19654>.
- [233] Petterino C, Argentino-Storino A. Clinical chemistry and haematology historical data in control Sprague-Dawley rats from pre-clinical toxicity studies. *Experimental and Toxicologic Pathology* 2006;57:213–9. <https://doi.org/10.1016/j.etp.2005.10.002>.
- [234] Baumans V. Use of animals in experimental research: An ethical dilemma? *Gene Ther* 2004;11:64–6. <https://doi.org/10.1038/sj.gt.3302371>.
- [235] Szpirer C. Cancer research in rat models. *Methods Mol Biol* 2010;597:445–58. [https://doi.org/10.1007/978-1-60327-389-3\\_30](https://doi.org/10.1007/978-1-60327-389-3_30).
- [236] Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, et al. Global Cancer Statistics 2020: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. *CA Cancer J Clin* 2021;71:209–49. <https://doi.org/10.3322/caac.21660>.
- [237] Alves CRR, Neves W das, de Almeida NR, Eichelberger EJ, Jannig PR, Voltarelli VA, et al. Exercise training reverses cancer-induced oxidative stress and decrease in muscle COPS2/TRIP15/ALIEN. *Mol Metab* 2020;39:1–17. <https://doi.org/10.1016/J.MOLMET.2020.101012>.



- [238] Honors MA, Kinzig KP. Characterization of the Yoshida sarcoma: A model of cancer cachexia. *Supportive Care in Cancer* 2013;21:2687–94. <https://doi.org/10.1007/s00520-013-1839-y>.
- [239] Betancourt A, Busquets S, Ponce M, Toledo M, Guàrdia-Olmos J, Peró-Cebollero M, et al. The animal cachexia score (ACASCO). *Animal Model Exp Med* 2019;2:201–9. <https://doi.org/10.1002/ame2.12082>.
- [240] Brower M, Grace M, Kotz CM, Koya V. Comparative analysis of growth characteristics of Sprague Dawley rats obtained from different sources. *Lab Anim Res* 2015;31:166. <https://doi.org/10.5625/lar.2015.31.4.166>.
- [241] Robinson NB, Krieger K, Khan F, Huffman W, Chang M, Naik A, et al. The current state of animal models in research: A review. *International Journal of Surgery* 2019;72:9–13. <https://doi.org/10.1016/j.ijssu.2019.10.015>.
- [242] Cole CL, Bachman JF, Ye J, Murphy J, Gerber SA, Beck CA, et al. Increased myocellular lipid and IGF1BP-3 expression in a pre-clinical model of pancreatic cancer-related skeletal muscle wasting. *J Cachexia Sarcopenia Muscle* 2021;12:731–45. <https://doi.org/10.1002/jcsm.12699>.
- [243] Neyroud D, Laitano O, Dasgupta A, Lopez C, Schmitt R, Schneider J, et al. Blocking muscle wasting via deletion of the muscle-specific E3 ubiquitin ligase MuRF1 impedes pancreatic tumor growth. *Res Sq* 2023. <https://doi.org/10.21203/rs.3.rs-2524562/v1>.
- [244] Eng CM, Smallwood LH, Rainiero MP, Lahey M, Ward SR, Lieber RL. Scaling of muscle architecture and fiber types in the rat hindlimb. *Journal of Experimental Biology* 2008;211:2336–45. <https://doi.org/10.1242/jeb.017640>.
- [245] Karauda P, Podgórski M, Paulsen F, Polguy M, Olewnik Ł. Anatomical variations of the tibialis anterior tendon. *Clinical Anatomy* 2021;34:397–404. <https://doi.org/10.1002/ca.23663>.
- [246] Mintz EL, Passipieri JA, Lovell DY, Christ GJ. Applications of in vivo functional testing of the rat tibialis anterior for evaluating tissue engineered skeletal muscle repair. *Journal of Visualized Experiments* 2016;2016. <https://doi.org/10.3791/54487>.
- [247] Vanderveen BN, Fix DK, Counts BR, Carson JA. The Effect of Wheel Exercise on Functional Indices of Cachexia in Tumor-bearing Mice. *Med Sci Sports Exerc* 2020;52:2320–30. <https://doi.org/10.1249/MSS.0000000000002393>.
- [248] Alves CRR, das Neves W, Tobias GC, de Almeida NR, Barreto RF, Melo CM, et al. High-intensity interval training slows down tumor progression in mice bearing Lewis lung carcinoma. *JCSM Rapid Commun* 2018;1:1–10. <https://doi.org/10.1002/j.2617-1619.2018.tb00008.x>.
- [249] Chung Y, Hsiao YT, Huang WC. Physiological and psychological effects of treadmill overtraining implementation. *Biology (Basel)* 2021;10. <https://doi.org/10.3390/biology10060515>.
- [250] Ferreira JCB, Rolim NPL, Bartholomeu JB, Gobatto CA, Kokubun E, Brum PC. Maximal lactate steady state in running mice: Effect of exercise training. *Clin Exp Pharmacol Physiol* 2007;34:760–5. <https://doi.org/10.1111/j.1440-1681.2007.04635.x>.

- [251] Ramilowski JA, Goldberg T, Harshbarger J, Kloppman E, Lizio M, Satagopam VP, et al. A draft network of ligand-receptor-mediated multicellular signalling in human. *Nat Commun* 2015;6. <https://doi.org/10.1038/ncomms8866>.
- [252] Armingol E, Officer A, Harismendy O, Lewis NE. Deciphering cell–cell interactions and communication from gene expression. *Nat Rev Genet* 2021;22:71–88. <https://doi.org/10.1038/s41576-020-00292-x>.
- [253] Parham DM. Pathologic Classification of Rhabdomyosarcomas and Correlations with Molecular Studies. *Mod Pathol* 2001;14:506–14. <https://doi.org/10.1038/modpathol.3880339>.
- [254] Gallagher IJ, Stephens NA, MacDonald AJ, Skipworth RJE, Husi H, Greig CA, et al. Suppression of skeletal muscle turnover in cancer cachexia: Evidence from the transcriptome in sequential human muscle biopsies. *Clinical Cancer Research* 2012;18:2817–27. <https://doi.org/10.1158/1078-0432.CCR-11-2133>.
- [255] Prill K, Carlisle C, Stannard M, Windsor Reid PJ, Pilgrim DB. Myomesin is part of an integrity pathway that responds to sarcomere damage and disease. *PLoS One* 2019;14. <https://doi.org/10.1371/journal.pone.0224206>.
- [256] Rouillon J, Poupiot J, Zocevic A, Amor F, Léger T, Garcia C, et al. Serum proteomic profiling reveals fragments of MYOM3 as potential biomarkers for monitoring the outcome of therapeutic interventions in muscular dystrophies. *Hum Mol Genet* 2015;24:4916–32. <https://doi.org/10.1093/hmg/ddv214>.
- [257] Lee M, Shin J, Kato T, Kanda K, Oikawa S, Sakuma J, et al. An acute eccentric exercise increases circulating myomesin 3 fragments. *Journal of Physiological Sciences* 2021;71. <https://doi.org/10.1186/s12576-021-00789-y>.
- [258] Fernández-Verdejo R, Vanwynsberghe AM, Essaghir A, Demoulin JB, Hai T, Deldicque L, et al. Activating transcription factor 3 attenuates chemokine and cytokine expression in mouse skeletal muscle after exercise and facilitates molecular adaptation to endurance training. *FASEB Journal* 2017;31:840–51. <https://doi.org/10.1096/fj.201600987R>.
- [259] Dunkelberger JR, Song WC. Complement and its role in innate and adaptive immune responses. *Cell Res* 2010;20:34–50. <https://doi.org/10.1038/cr.2009.139>.
- [260] Zhang C, Wang C, Li Y, Miwa T, Liu C, Cui W, et al. Complement C3a signaling facilitates skeletal muscle regeneration by regulating monocyte function and trafficking. *Nat Commun* 2017;8. <https://doi.org/10.1038/s41467-017-01526-z>.
- [261] Pillon NJ, Bilan PJ, Fink LN, Klip A. Cross-talk between skeletal muscle and immune cells: muscle-derived mediators and metabolic implications. *Am J Physiol Endocrinol Metab* 2013;304:453–65. <https://doi.org/10.1152/ajpendo.00553.2012>.
- [262] Ballarò R, Beltrà M, De Lucia S, Pin F, Ranjbar K, Hulmi JJ, et al. Moderate exercise in mice improves cancer plus chemotherapy-induced muscle wasting and mitochondrial alterations. *FASEB Journal* 2019;33:5482–94. <https://doi.org/10.1096/fj.201801862R>.
- [263] Pedersen L, Idorn M, Olofsson GH, Pedersen BK, Straten T, Hojman P. Voluntary Running Suppresses Tumor Growth through Epinephrine-and IL-6-Dependent NK Cell

- Mobilization and Redistribution. *Cell Metab* 2016;23:554–62.  
<https://doi.org/10.1016/j.cmet.2016.01.011>.
- [264] Deminice R, Rezende LFM, Rosa DD, Cangussu R, Garcia LMT, Riera R, et al. Physical activity recommendations for cancer prevention and control: a Brazilian consortium. *Brazilian Journal of Oncology* 2022;18. <https://doi.org/10.5935/2526-8732.20220311>.
- [265] Rodríguez-Cañamero S, Cobo-Cuenca AI, Carmona-Torres JM, Pozuelo-Carrascosa DP, Santacruz-Salas E, Rabanales-Sotos JA, et al. Impact of physical exercise in advanced-stage cancer patients: Systematic review and meta-analysis. *Cancer Med* 2022;11:3714–27. <https://doi.org/10.1002/cam4.4746>.
- [266] Rosenkranz AA, Slastnikova TA. Epidermal Growth Factor Receptor: Key to Selective Intracellular Delivery. *Biochemistry (Moscow)* 2020;85:967–93.  
<https://doi.org/10.1134/S0006297920090011>.
- [267] Hsu JL, Hung MC. The role of HER2, EGFR, and other receptor tyrosine kinases in breast cancer. *Cancer and Metastasis Reviews* 2016;35:575–88.  
<https://doi.org/10.1007/s10555-016-9649-6>.
- [268] Hynes NE, MacDonald G. ErbB receptors and signaling pathways in cancer. *Curr Opin Cell Biol* 2009;21:177–84. <https://doi.org/10.1016/j.ceb.2008.12.010>.
- [269] Scaltriti M, Baselga J. The epidermal growth factor receptor pathway: A model for targeted therapy. *Clinical Cancer Research* 2006;12:5268–72.  
<https://doi.org/10.1158/1078-0432.CCR-05-1554>.
- [270] Accattato F, Greco M, Pullano SA, Caré I, Fiorillo AS, Pujia A, et al. Effects of acute physical exercise on oxidative stress and inflammatory status in young, sedentary obese subjects. *PLoS One* 2017;12. <https://doi.org/10.1371/journal.pone.0178900>.
- [271] Coffey Jr RJ, Meise KS, Matsui Y, Hogan BL, Dempsey PJ, Halter SA. Acceleration of mammary neoplasia in transforming growth factor alpha transgenic mice by 7,12-dimethylbenzanthracene. *Cancer Res* 1994;1:1678–83.
- [272] Awwad RA, Sergina N, Yang H, Ziober B, Willson JK V, Zborowska E, et al. The role of transforming growth factor alpha in determining growth factor independence. *Cancer Res* 2003;63:4731–8.
- [273] Ma X, Zheng J, He K, Wang L, Wang Z, Wang K, et al. TGFA expression is associated with poor prognosis and promotes the development of cervical cancer. *J Cell Mol Med* 2024;28. <https://doi.org/10.1111/jcmm.18086>.
- [274] Liu G, An L, Zhang H, Du P, Sheng Y. Activation of CXCL6/CXCR1/2 axis promotes the growth and metastasis of osteosarcoma cells in vitro and in vivo. *Front Pharmacol* 2019;10. <https://doi.org/10.3389/fphar.2019.00307>.
- [275] Callaway CS, Delitto AE, Patel R, Nosacka RL, D’Lugos AC, Delitto D, et al. IL-8 released from human pancreatic cancer and tumor-associated stromal cells signals through a CXCR2-ERK1/2 axis to induce muscle atrophy. *Cancers (Basel)* 2019;11.  
<https://doi.org/10.3390/cancers11121863>.

- [276] McGee SL, Hargreaves M. Exercise adaptations: molecular mechanisms and potential targets for therapeutic benefit. *Nat Rev Endocrinol* 2020;16:495–505. <https://doi.org/10.1038/s41574-020-0377-1>.
- [277] Rausch V, Sala V, Penna F, Porporato PE, Ghigo A. Understanding the common mechanisms of heart and skeletal muscle wasting in cancer cachexia. *Oncogenesis* 2021;10. <https://doi.org/10.1038/s41389-020-00288-6>.
- [278] Janssen I, Heymsfield SB, Wang Z, Ross R. Skeletal muscle mass and distribution in 468 men and women aged 18–88 yr. *J Appl Physiol* 2000;89:81–8. <https://doi.org/10.1152/jappl.2000.89.1.81>.
- [279] Siegel RL, Giaquinto AN, Jemal A. Cancer statistics, 2024. *CA Cancer J Clin* 2024;74:12–49. <https://doi.org/10.3322/CAAC.21820>.
- [280] Roeland EJ, Bohlke K, Baracos VE, Bruera E, del Fabbro E, Dixon S, et al. Management of Cancer Cachexia: ASCO Guideline. *Journal of Clinical Oncology* 2020;38:2438–53. <https://doi.org/10.1200/JCO.20.00611>.
- [281] Rogers ES, Ormiston W, Heron R, Pontré B, Macleod R, Doyle A. Body composition skeletal muscle analysis in cancer cachexia studies: Is there a place for 3T MRI analysis? *J Cachexia Sarcopenia Muscle* 2018;3:1–12. <https://doi.org/10.17987/jcsm-cr.v3i2.59>.
- [282] Buffa R, Floris G, Marini E. Migration of the bioelectrical impedance vector in healthy elderly subjects. *Nutrition* 2003;19:917–21. [https://doi.org/10.1016/s0899-9007\(03\)00180-1](https://doi.org/10.1016/s0899-9007(03)00180-1).
- [283] Tomeleri CM, Ribeiro AS, Cavaglieri CR, Deminice R, Schoenfeld BJ, Schiavoni D, et al. Correlations between resistance training-induced changes on phase angle and biochemical markers in older women. *Scand J Med Sci Sports* 2018;28:2173–82. <https://doi.org/10.1111/sms.13232>.
- [284] Trutschnigg B, Kilgour RD, Reinglas J, Rosenthal L, Hornby L, Morais JA, et al. Precision and reliability of strength (Jamar vs. Biodex handgrip) and body composition (dual-energy X-ray absorptiometry vs. bioimpedance analysis) measurements in advanced cancer patients. *Applied Physiology, Nutrition and Metabolism* 2008;33:1232–9. <https://doi.org/10.1139/H08-122>.
- [285] Maranhão RC, Cesara TB, Pedrosa-madani SR, Hirata MH, Mesquita CH. Metabolic Behavior in Rats of a Nonprotein Microemulsion Resembling Low-Density Lipoprotein. *Lipids* 1993;28:691–6.
- [286] You T, Disanzo BL, Arsenis NC. Aerobic exercise training attenuates obesity-related hypogonadism in male rats. *Med Sci Sports Exerc* 2013;45:1244–51. <https://doi.org/10.1249/MSS.0b013e318285816c>.
- [287] Mantzorou M, Tolia M, Poultsidi A, Pavlidou E, Papadopoulou SK, Papandreou D, et al. Can bioelectrical impedance analysis and BMI be a prognostic tool in head and neck cancer patients? A review of the evidence. *Cancers (Basel)* 2020;12:1–16. <https://doi.org/10.3390/cancers12030557>.

- [288] Oliveira Filho JM de, Bernardes PS, Serpa GHC, Siqueira GD de J, Noll M, Venâncio PEM, et al. Bioelectrical Vector Analysis in Obese Adolescents. *Rev Paul Pediatr* 2020;38:e2019017. <https://doi.org/10.1590/1984-0462/2020/38/2019017>.
- [289] Koury JC, de Oliveira-Junior AV, Portugal MRC, de Oliveira K de JF, Donangelo CM. Bioimpedance parameters in adolescent athletes in relation to bone maturity and biochemical zinc indices. *Journal of Trace Elements in Medicine and Biology* 2018;46:26–31. <https://doi.org/10.1016/j.jtemb.2017.11.003>.
- [290] Watanabe H, Nakano T, Saito R, Akasaka D, Saito K, Ogasawara H, et al. Serotonin improves high fat diet induced obesity in mice. *PLoS One* 2016;11:1–14. <https://doi.org/10.1371/journal.pone.0147143>.
- [291] Brooke MH, Kaiser KK. Muscle Fiber Types: How Many and What Kind? *Arch Neurol* 1970;23:369–79. <https://doi.org/10.1001/archneur.1970.00480280083010>.
- [292] Folch J, Lees M, Sloane Stanley GH. A simple method for the isolation and purification of total lipides from animal tissues. *J Biol Chem* 1956;226:497–509. [https://doi.org/10.1016/s0021-9258\(18\)64849-5](https://doi.org/10.1016/s0021-9258(18)64849-5).
- [293] Mesquita CS, Oliveira R, Bento F, Geraldo D, Rodrigues J V., Marcos JC. Simplified 2,4-dinitrophenylhydrazine spectrophotometric assay for quantification of carbonyls in oxidized proteins. *Anal Biochem* 2014;458:69–71. <https://doi.org/10.1016/j.ab.2014.04.034>.
- [294] Buege JA, Aust SD. Microsomal Lipid Peroxidation. *Methods Enzymol* 1978;52:302–10. [https://doi.org/https://doi.org/10.1016/s0076-6879\(78\)52032-6](https://doi.org/https://doi.org/10.1016/s0076-6879(78)52032-6).
- [295] Benzie IFF, Strain JJ. The Ferric Reducing Ability of Plasma (FRAP) as a Measure of “Antioxidant Power”: The FRAP Assay. *Anal Biochem* 1996;8:70–6. <https://doi.org/https://doi.org/10.1006/abio.1996.0292>.
- [296] Beutler E, Duron O, Kelly BM. Improved method for the determination of blood glutathione. *J Lab Clin Med* 1963.
- [297] MARKLUND S, MARKLUND G. Involvement of the Superoxide Anion Radical in the Autoxidation of Pyrogallol and a Convenient Assay for Superoxide Dismutase. *Eur J Biochem* 1974;47:469–74. <https://doi.org/10.1111/j.1432-1033.1974.tb03714.x>.
- [298] Aebi H. Catalase in vitro. *Methods Enzymol* 1984;105:121–6. [https://doi.org/10.1016/S0076-6879\(84\)05016-3](https://doi.org/10.1016/S0076-6879(84)05016-3).
- [299] Schindelin J, Arganda-Carreras I, Frise E, Kaynig V, Longair M, Pietzsch T, et al. Fiji: An open-source platform for biological-image analysis. *Nat Methods* 2012;9:676–82. <https://doi.org/10.1038/nmeth.2019>.
- [300] Starruß J, De Back W, Bruschi L, Deutsch A. Morpheus: A user-friendly modeling environment for multiscale and multicellular systems biology. *Bioinformatics* 2014;30:1331–2. <https://doi.org/10.1093/bioinformatics/btt772>.
- [301] Dobin A, Davis CA, Schlesinger F, Drenkow J, Zaleski C, Jha S, et al. STAR: ultrafast universal RNA-seq aligner. *Bioinformatics* 2013;29:15–21. <https://doi.org/10.1093/BIOINFORMATICS/BTS635>.

- [302] Breese MR, Liu Y. NGSUtils: a software suite for analyzing and manipulating next-generation sequencing datasets. *Bioinformatics* 2013;29:494–6. <https://doi.org/10.1093/BIOINFORMATICS/BTS731>.
- [303] Liao Y, Smyth GK, Shi W. featureCounts: an efficient general purpose program for assigning sequence reads to genomic features. *Bioinformatics* 2014;30:923–30. <https://doi.org/10.1093/BIOINFORMATICS/BTT656>.
- [304] Goedhart J, Luijsterburg MS. VolcanoR is a web app for creating, exploring, labeling and sharing volcano plots. *Sci Rep* 2020;10. <https://doi.org/10.1038/s41598-020-76603-3>.
- [305] Kuleshov M V., Jones MR, Rouillard AD, Fernandez NF, Duan Q, Wang Z, et al. Enrichr: a comprehensive gene set enrichment analysis web server 2016 update. *Nucleic Acids Res* 2016;44:W90–7. <https://doi.org/10.1093/nar/gkw377>.
- [306] Chen EY, Tan CM, Kou Y, Duan Q, Wang Z, Meirelles GV, et al. Enrichr: interactive and collaborative HTML5 gene list enrichment analysis tool. *BMC Bioinformatics* 2013;14. <https://doi.org/10.1186/1471-2105-14-128>.
- [307] Xie Z, Bailey A, Kuleshov M V., Clarke DJB, Evangelista JE, Jenkins SL, et al. Gene Set Knowledge Discovery with Enrichr. *Curr Protoc* 2021;1:1–3. <https://doi.org/10.1002/cpz1.90>.
- [308] Khan A, Mathelier A. Intervene: A tool for intersection and visualization of multiple gene or genomic region sets. *BMC Bioinformatics* 2017;18. <https://doi.org/10.1186/s12859-017-1708-7>.
- [309] Barde P, Barde M. What to use to express the variability of data: Standard deviation or standard error of mean? *Perspect Clin Res* 2012;3:113. <https://doi.org/10.4103/2229-3485.100662>.
- [310] Altman DG, Martin Bland J. Standard deviations and standard errors. *BMJ* 2005;331:903–903. <https://doi.org/10.1136/bmj.331.7521.903>.
- [311] Beaudry AG, Law ML. Variable body and tissue weight reporting in preclinical cachexia literature may alter study outcomes and interpretation. *Dis Model Mech* 2023. <https://doi.org/10.1242/dmm.050148>.
- [312] Zahedi P, De Souza R, Piquette-Miller M, Allen C. Docetaxel Distribution Following Intraperitoneal Administration in Mice. *Journal of Pharmacy & Pharmaceutical Sciences* 2011;14:90–9. <https://doi.org/10.18433/j3qw26>.
- [313] Summermatter S, Bouzan A, Pierrel E, Melly S, Stauffer D, Gutzwiller S, et al. Blockade of Metallothioneins 1 and 2 Increases Skeletal Muscle Mass and Strength. *Mol Cell Biol* 2017;37. <https://doi.org/10.1128/mcb.00305-16>.
- [314] Kadota Y, Yamanokuchi R, Ohnishi N, Matsuoka M, Kawakami T, Sato M, et al. Metallothionein Gene Deficiency Facilitates the Differentiation of C2C12 Myoblasts into Slow-Twitch Myotubes. *Biol Pharm Bull* 2023;46:1240–8. <https://doi.org/10.1248/bpb.b23-00165>.

- [315] Zhang S, Yang F, Huang Y, He L, Li Y, Ching Y, et al. ATF3 induction prevents precocious activation of skeletal muscle stem cell by regulating H2B expression. *Nat Commun* 2023;14. <https://doi.org/10.1038/s41467-023-40465-w>.
- [316] Liu Y, Liu N, Yu Y, Wang D. Nr4a1 promotes cell adhesion and fusion by regulating Zeb1 transcript levels in myoblasts. *Biochem Biophys Res Commun* 2021;556:127–33. <https://doi.org/10.1016/j.bbrc.2021.03.153>.
- [317] Mohan A, Asakura A. CDK inhibitors for muscle stem cell differentiation and self-renewal. *J Phys Fit Sports Med* 2017;6:65–74. <https://doi.org/10.7600/jpfsm.6.65>.
- [318] Ballak SB, Degens H, Busé-Pot T, de Haan A, Jaspers RT. Plantaris muscle weakness in old mice: relative contributions of changes in specific force, muscle mass, myofiber cross-sectional area, and number. *Age (Omaha)* 2014;36. <https://doi.org/10.1007/s11357-014-9726-0>.
- [319] Sánchez-Iglesias A, Fernández-Lucas M, Teruel JL. The electrical basis of bioimpedance. *Nefrologia* 2012;32:133–5. <https://doi.org/10.3265/Nefrologia.pre2012.Jan.11310>.
- [320] Trobec K, Kos MK, Von Haehling S, Springer J, Anker SD, Lainscak M. Pharmacokinetics of drugs in cachectic patients: A systematic review. *PLoS One* 2013;8. <https://doi.org/10.1371/journal.pone.0079603>.
- [321] Bardou P, Mariette J, Escudié F, Djemiel C, Klopp C. jvenn: an interactive Venn diagram viewer. *BMC Bioinformatics* 2014;15. <https://doi.org/10.1186/1471-2105-15-293>.